ORIGINAL PAPER



UV/Vis absorption spectroelectrochemistry of folic acid

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Received: 1 April 2021 / Revised: 9 July 2021 / Accepted: 9 August 2021 / Published online: 20 August 2021 © The Author(s) 2021

Abstract

UV/Vis absorption spectroelectrochemistry is a very promising analytical technique due to the complementary information that is simultaneously obtained from electrochemistry and spectroscopy. In this work, this technique is used in a parallel configuration to study the oxidation of folic acid in alkaline medium. Herein, UV/Vis absorption spectroelectrochemistry has been used to detect both the oxidation products and the folic acid consumed at the electrode/solution interface, allowing us to develop an analytical protocol to quantify vitamin B9 in pharmaceutical tablets. Linear ranges of three orders of magnitude have been achieved in basic medium (pH = 12.9), obtaining high repeatability and low detection limits. The spectroelectrochemical determination of folic acid in pharmaceutical tablets at alkaline pH values is particularly interesting because of the changes that occur in the optical signal during the electrochemical oxidation of FA, providing results with very good figures of merit and demonstrating the utility and versatility of this hyphenated technique, UV/Vis absorption spectroelectrochemistry.

Keywords Vitamin B9 · Pharmaceutical drug · Electrochemistry · UV/Vis absorption spectroscopy

Introduction

Instrumental analytical techniques commonly used to identify and quantify a specific molecule are typically based on the use of a single signal characteristic of the compounds under study. However, it is well-known that the use of two or more techniques can be much more helpful in the determination of a molecule. Techniques such as spectroelectrochemistry (SEC) are extremely useful analytical tools in the quantification of molecules and have proven to be very useful when dealing with complex samples or when there is a large matrix effect [1–5]. SEC encompasses a group of analytical techniques that are characterized by the simultaneous recording of electrochemical and spectroscopic signals

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¹ Department of Chemistry, Universidad de Burgos, Pza. Misael Bañuelos s/n, 09001 Burgos, Spain [6–11]. Among all the SEC techniques, one of the most widely used is UV/Vis absorption SEC (UV/Vis-SEC), in which during the electrochemical oxidation or reduction of a molecule, changes in the molecular absorption spectrum of the studied analytes are observed due to the electrogeneration of the reaction products and/or the consumption of the native molecule [7–9, 12, 13].

SEC in general, and UV/Vis-SEC in particular, is a group of instrumental techniques commonly used in the characterization of new materials and/or molecules, in the study of electron transfer processes, or in the study of reaction mechanisms [6, 14–22]. However, the great potential of these multi-response techniques for quantitative analysis has not been enough explored [1, 2, 4, 5, 12, 23-26]. Among all SEC techniques, the coupling of electrochemistry and UV/Vis absorption spectroscopy is the most successful one in the field of electroanalysis. Significant improvements in instrumentation and SEC devices, such as SEC cells, have significantly contributed to the fruitful use of these operando and multi-response techniques for analytical purposes [1, 9, 10, 15, 27-29]. From a quantitative point of view, it should be remarked that UV/Vis-SEC can provide a dual determination of the analyte studied, using both the electrochemical and the spectroscopic signals, providing much more reliable and fully validated results. In some cases, the electrochemical signal is not good enough for quantitative purposes, but a good optical signal is obtained during the electrochemical process. Furthermore, UV/Vis-SEC results are characterized by a high reproducibility and very low relative standard deviation values (RSD), parameters required by all analytical techniques used for quantitative purposes. Lastly, it should be pointed out that the information gathered in this type of experiments, concerning the reaction mechanism involved in the electron transfer process studied, helps significantly not only to detect outliers but also to obtain an explanation for why that point is an outlier [28].

Vitamin B9 belongs to the group of water-soluble vitamins, and its structure is derived from folic acid (FA). FA is a cofactor in methylation reactions, in biochemical reactions related to the synthesis of some nitrogen bases, such as adenine, guanine, or thymine, and in amino acid metabolism. Therefore, deficiency of this vitamin is directly related to improper DNA replication and, consequently, to improper cell division. Increased metabolic demand for folate is common in pregnancy due to megaloblastic anemia. A deficiency of this vitamin in pregnancy can lead to defects in the embryo, especially in the neural tube. Deficiency of vitamins B9 and B12 is also associated with some diseases related to the central nervous system such as dementia, depression, or Alzheimer's disease and with vascular diseases such as thrombosis [30-33]. To avoid FA deficits, since the human body is not able to produce FA, adequate levels of vitamin B9 must be achieved through diet or food supplements.

FA derives from three chemical structures: a 6-methylpterin, *p*-aminobenzoic acid, and glutamic acid [30]. There are many different folates depending on the glutamate residues included in the chemical structure [32]. On the other hand, folate exits in different oxidation states. The natural folates have the pterin ring in its reduced state such as 7,8-dihydrofolate or 5,6,7,8-tetrahydrofolate, not being a natural physiological form of this vitamin the pterin ring in its high oxidized state [30, 32, 34].

The solubility of FA in water is low in neutral medium (1.6 mg/L at 25 °C) and depends strongly on the pH of the solution [35]. Under high acidic conditions, the solubility of FA increases significantly, but its stability decreases at pH values below 5. On the other hand, solubility increases significantly with increasing pH (50 mg/mL at 1 M NaOH), being stable solutions when protected from light [30].

In alkaline medium, it presents three well-defined absorption bands in the UV–Vis region, at 256, 283, and 365 nm assigned to the π - π * electronic transitions due to the pterin ring and *p*-aminobenzoic acid moieties of FA [36–38], and its electrochemical properties, both in oxidation and reduction processes, are related to the conjugated pteridine ring [39, 40].

FA has been studied and quantified with different analytical techniques such as photoluminescence [41–43], UV/ Vis absorption spectroscopy [44, 45], electrochemistry [39,

46–49], HPLC [50, 51], or HPLC–MS [52, 53]. Some of these techniques require complex and long sample pretreatments; others imply the modification of the surface used to determine this molecule such as in electrochemistry. Since FA has a characteristic UV/Vis absorption spectrum and its oxidation process is previously described, UV/Vis-SEC can be a suitable instrumental technique to study the electron transfer reaction of vitamin B9.

In this work, a UV/Vis-SEC-based methodology is proposed to quantify FA in a pharmaceutical drug. Specifically, in this work, FA is electrochemically oxidized, while changes in the UV/Vis absorption spectra, which are used to identify and quantify FA, are simultaneously recorded. The determination protocol developed is simple and not timeconsuming and provides optimal results in the identification and quantification of FA. By adapting this protocol to other substances and other matrices, this approach can be used for the analysis of other drugs in real samples.

Experimental

Chemicals and materials

FA (97%, Sigma-Aldrich), KCl (99 + %, ACROS Organics), NaOH (ACROS Organics), glacial acetic acid (VWR), and sodium acetate (>99%, VWR) were of analytical grade and were used as delivered without further purification. Yodocefol® (ItalFarmaco). The protocols used in all the experiments performed in this work ensure safe and careful handling of all reagents.

All solutions were freshly prepared using ultrapure deionized water (18.2 M Ω cm resistivity at 25 °C, Milli-Q Direct 8, Millipore).

Instrumentation

UV/Vis-SEC measurements were performed using a customized UV–VIS SPELEC instrument (Metrohm-DropSens), employing the experimental setup described previously [1, 5]. A deuterium lamp was used because the reagents and products of this electrochemical process absorb electromagnetic radiation in the UV region. Carbon DRP-110 screenprinted electrodes (SPE, Metrohm-Dropsens) were used for UV/Vis-SEC measurements. These C-SPEs consist of a carbon working electrode (C-WE) of 4 mm diameter, a silver pseudo-reference electrode, and a carbon counter electrode.

UV/Vis-SEC experiments in normal configuration were carried out using the Teflon® SEC cell for reflection experiments (DRP-REFLECELL, Metrohm-DropSens) and a reflection probe (DRP-RPROBE, Metrohm-DropSens). UV/Vis-SEC experiments in parallel configuration were carried out by facing and attaching two 100-µm-bare optical fibers (Avantes) to the surface of the C-WE of the SPEs that were placed at the boxed connector for SPEs (DSC, Metrohm-DropSens). The optical pathway was measured for each C-SPE used, which is in all cases between 2 and 3 mm in length. A 50-µL aliquot of the solution studied is used in each experiment.

UV/Vis-SEC study of the effect of pH on FA oxidation

0.1 M acetic/acetate buffer solutions were prepared, being their pH=5. 0.1 M KCl solutions were prepared, being their pH=7. Also, 0.1 M NaOH solutions were prepared, being their pH=12.9.

UV/Vis-SEC experiments in normal configuration were performed to test the influence of the pH on the process of electrochemical oxidation of FA. Solutions of 100 μ M FA in the specific medium tested (buffer acetic/acetate, KCl or NaOH) were prepared, and cyclic voltammetry experiments were carried out. The potential was scanned in the anodic direction at 0.01 V s⁻¹ between 0.00 and +0.90 V (0.1 M NaOH) or between +0.70 and +1.30 V (0.1 M acetic/acetate buffer and 0.1 M KCl), and simultaneously the evolution of the absorbance between 240 and 900 nm is recorded along the whole experiments. The spectrum of the sample at the starting potential was taken as reference, and changes of absorbance respect to this reference spectrum (ΔA) were recorded. The integration time for absorptiometric data in all experiments was 400 ms.

UV/Vis-SEC quantification protocol

UV/Vis-SEC measurements were performed in 0.1 M NaOH (pH = 12.9), changing the concentration of FA between 5 and 100 μ M. SEC experiments were performed collecting the evolution of absorption spectra between 240 and 900 nm during a cyclic voltammetry experiment. The potential was scanned between 0 and +0.90 V at 0.02 V s⁻¹, starting all the experiments in the anodic direction. In all UV/Vis-SEC experiments, the first spectrum of the sample at 0 V was taken as reference, and therefore, the change of absorbance respect to the reference spectrum was recorded, expressed as ΔA in all figures. Time-resolved UV/Vis-SEC measurements were performed using an integration time of 400 ms.

The calibration models were constructed by randomly measuring the different FA solutions, varying its concentration between 5 and 100 μ M. Each point in the calibration models was replicated three times. Before performing the spectroelectrochemical measurements, the SPEs were pretreated by placing 50 μ L of a 0.1 M NaOH solution and performing 10 potential scans between 0 and +0.90 V at 0.02 V s⁻¹. The same C-SPE was used for each calibration

test. The blank solution containing 0.1 M NaOH was measured before and after sampling all the calibration samples to check that the WE does not undergo any modification.

Test sample preparation

Yodocefol® tablets were used as test sample. Each tablet of this pharmaceutical drug contains 400 μ g of FA, 262 μ g of potassium iodide, 2 μ g of vitamin B12, and different excipients (110 mesh lactose monohydrate, microcrystalline cellulose, sodium potato starch glycolate, calcium stearate, trisodium citrate, citric acid, and maltodextrins). To prepare test samples of this drug, first a dispersion of the tablet is prepared in 25 mL of 0.1 M NaOH. After decantation of the non-water-soluble excipients, 1-mL aliquots of the supernatant were centrifuged at 4000 rpm for 28 min in Eppendorf vials. The supernatant obtained was directly used as drug sample to be quantified that considering the starting dilution, the FA concentration in this test sample was 36 μ M.

Data acquisition and analysis software

SPELEC instrument was controlled by DropView SPELEC software (Metrohm-DropSens), performing operando, time-resolved SEC experiments with fully synchronized data acquisition.

Matlab 2018 and R 3.6.1 were used to analyze all UV/Vis-SEC experiments, to generate the calibration models, and to predict the FA concentration of test samples.

Results and discussion

FA oxidation with UV/Vis absorption spectroelectrochemistry

UV/Vis-SEC can be performed using a normal or parallel arrangement. When soluble products of reaction are formed during the electrochemical process, UV/Vis in parallel arrangement is more suitable for quantitative purpose. Nevertheless, UV/Vis-SEC in normal arrangement is very suitable for performing a rapid screening of the best experimental conditions used for quantification. Thus, UV/Vis-SEC in normal arrangement was used to look for the best electrolytic medium to obtain a sensitive and reproducible optical response.

Three different electrolytic media were explored to determine FA using UV/Vis-SEC in normal configuration: (a) 0.1 M NaOH (pH=12.9), (b) 0.1 M KCl (pH=7), and (c) 0.1 M acetic/acetate buffer (pH=5). Figure 1 shows the contour plots for the three different electrolytic media tested, where evolution of absorption spectra during a UV/Vis-SEC experiment was shown. As can be observed, only in alkaline

Fig. 1 Contour plots of the absorbance with time/potential during SEC experiments in a 100 μM FA solutions in (a) 0.1 M NaOH (pH = 12.9), (**b**) 0.1 M KCl (pH=7), and (c) 0.1 M acetic/acetate buffer (pH=5). Potential was scanned between 0.00 and +0.90 V at 0.01 V s⁻¹ in (**a**), between +0.70 V and +1.30 V at 0.01 V s⁻¹ in (**b**), and between +0.70 V and +1.30 V at 0.01 V s⁻¹ in (c). ΔA values in normal configuration were measured taking the starting FA solution as reference spectrum. Integration time was 400 ms



media, well-defined and sufficiently intense UV/Vis absorption signals with suitable spectral resolution were recorded.

Figure 2 shows the SEC responses obtained in parallel configuration with a 100 μ M FA and 0.1 M NaOH solution (pH = 12.9). As can be seen, a small and ill-defined anodic peak can be observed in the cyclic voltammogram between +0.50 and +0.60 V (CV, Fig. 2a). This ill-defined anodic peak must be related to the oxidation of FA since it

Fig. 2 (a) CV of the blank solution (0.1 M NaOH, blue line) and of the FA solution (orange line). (b) Absorption spectra recorded at different potentials during the anodic scan, and (c) CVAs at 260 (blue line), 295 (orange line), 332 (yellow line), and 392 (violet line) nm in a 100 µM FA and 0.1 M NaOH solution. Potential was scanned between 0.00 V and +0.90 V at 0.02 V/s. ΔA values in parallel configuration were measured taking the starting FA solution at 0.00 V as reference spectrum. Integration time for absorbance measurements was 400 ms. Optical path length = 2.3 mm



is the main difference with respect to the CV of the blank solution. However, a clear evolution of the UV/Vis absorption spectra is appreciated during the CV (Fig. 2b). These absorbance changes are related to the oxidation of FA as can be clearly observed in the cyclic voltabsorptograms (CVA, Fig. 2c).

In the evolution of the spectra during the anodic scan (Fig. 2b), four absorption bands were observed, two overlapped positive ones at 332 and 392 nm and two small overlapped negative ones at 260 and 295 nm, showing a single isosbestic point at 312 nm. This isosbestic point indicates that there is an interconversion of the reactant to a single product that absorbs electromagnetic radiation in this spectral range. Negative absorbance values are observed because the reference spectrum is taken in the FA initial solution. At alkaline pH values, FA is found as folate anion [54], showing three bands around 256, 285, and 365 nm according to literature [36, 37, 44, 55, 56]. According to the literature [38], the absorption band decreasing at 260 nm is related to the π - π * transition of the pterin moiety, while the absorption band at 295 nm is related to the π - π * transition of the aminobenzoyl ring. Thus, the two absorption bands with positive values can be ascribed to the formation of a new compound during the oxidation of FA, while the two absorption bands with negative values correspond to the FA that is consumed from the solution adjacent to the C-WE during the oxidation process. CVA at these four absorption bands (Fig. 2c) has shown how the oxidation of FA begins at +0.55 V, since the absorbance increases in the bands associated with the oxidation product of FA and decreases in the bands associated with the consumption of FA. Once the vertex potential (+0.90 V) has been reached, absorbance continues increasing in the cathodic scan until it reaches a maximum value around +0.60 V. From this potential downwards, the absorbance only decreases slightly during the cathodic scan because of the diffusion of the products of the electrochemical process.

As was described in bibliography, FA oxidation reaction is irreversible, since FA is oxidized to an enamine that subsequently suffers a hydrolysis process, breaking this enamine into the pterin moiety as 6-formylpterin and the *p*-aminobenzoyl-L-glutamic acid (PGA) [37, 46]. After oxidation, absorption spectral changes (ΔA) at wavelengths longer than 300 nm should correspond to the 6-formylpterin fragment. This hydrolysis of FA in alkaline medium has been previously reported [40], and according to the literature, the ΔA spectra of derivatives of the 6-formylpterin fragment have shown absorption bands between 300 and 400 nm [55, 57, 58]. In our case, we observe absorption bands at 332 and 392 nm, which confirm this hydrolysis process.

From the electrochemical signal, it is not easy to follow the oxidation of FA due to the ill-defined anodic peak recorded; however, the optical signal clearly confirms that the molecule is oxidized in this alkaline electrolytic medium. This is one of the main advantages of UV/Vis-SEC because the electrochemical oxidation process of molecules can be studied without a clear evidence of the process in the electrical signal.

FA determination with UV/Vis absorption spectroelectrochemistry

Once the optimal conditions for the study of FA were determined, the calibration of FA using the optical signal was performed. In this case, the electrochemical signal cannot be used as no oxidation peak is observed. Different tests and preliminary experiments were carried out to determine the linear range. At concentrations greater than 100 µM, no linear relationship between concentration and absorbance was found. On the other side, concentration values below 5 µM cannot be easily differentiated from the signal noise. Thus, a calibration set of known FA concentration samples between 5 and 100 µM was prepared, with standards of 5, 8, 10, 20, 40, 60, 80, and 100 µM of FA, always in 0.1 M NaOH to ensure that the desired pH is the same in all solutions ($pH = 12.9 \pm 0.1$). A parallel configuration UV/Vis-SEC cell was selected for the measurements because it is more sensitive than normal arrangement for soluble products of the electrochemical reaction due to its longer optical path length.

Figure 3 shows the optical responses obtained during the calibration procedure for some of the tested samples: 0, 8, 40, 80, and 100 µM. Figure 3a shows the UV/Vis absorption spectra of these FA samples recorded at +0.55 V in the reverse scan, in the spectral range of 200-500 nm where absorbance changes are detected. As was expected, the higher the FA concentration, the higher the recorded absorbance related to the oxidation/hydrolysis products. A good reproducibility between measurements was obtained as can be deduced from the two samples shown at 40 µM, which practically are overlapped. The CVAs at 392 nm for the solutions of concentration equal to those shown in Fig. 3a are shown in Fig. 3b. This wavelength is selected because it corresponds to the most sensitive absorption band. Oxidation starts from a potential of +0.55 V onwards, observing an increase of absorbance, reaching a maximum during the reverse scan at +0.55 V.

Figure 4 shows the calibration curves obtained for the concentration range of 5 to 100 μ M using the bands at 332 nm and 392 nm, the two positive absorption bands shown in Fig. 2b and Fig. 3a. Figures of merit of the two calibrations are summarized in Table 1.

The calibration model at 392 nm is more sensitive than the one at 332 nm as deduced from the slope. Very good and similar coefficients of determination (R^2) were obtained for the two calibrations models, which provide similar limits of detection in the two cases. The low values of the standard deviation of residuals (S_{yx}) indicate **Fig. 3** (a) Absorption spectra at +0.55 V obtained during the reverse scan recorded at different concentrations. (b) CVAs at 392 nm obtained at different concentrations. Experimental conditions in the Fig. 2 caption



the low dispersion of data, as can be observed in Fig. 4 for the three replicates of each point, where the error bars are very small. The limits of detection (LOD) are very similar with the two wavelengths.

Repeatability of the UV/Vis-SEC method was evaluated by constructing three regression models for each wavelength (332 and 392 nm) under the same experimental conditions described above and evaluating the repeatability of the slopes. For the calibrations at 332 and 392 nm, a %RSD of 4.36 and 2.20 was obtained, respectively. From these values, we can conclude that the UV/Vis-SEC method has a very good repeatability in the quantification of FA in alkaline medium.

FA determination in a pharmaceutical tablet

Once demonstrated that this UV/Vis-SEC method is reliable, repeatable, and sensitive, the determination of FA in a drug (Yodocefol®) was performed, probing the capability of the method for real samples. Yodocefol® tablet contains 262 µg



Fig. 4 Calibration curves of FA obtained from the CVAs at +0.55 V in the reverse scan at 332 (violet line) band 392 nm (yellow line). Experimental conditions in the caption of Fig. 2. Each point was replicated three times. Optical path length = 2.3 mm

of KI (equivalent to 200 μ g of I₂), 400 μ g of FA, and 2 μ g of vitamin B12 and different excipients. Therefore, the analyte is accompanied with potential interfering compounds. The drug sample was prepared as described in the experimental section.

A new calibration procedure was performed to carry out the determination of a tablet of Yodocefol® using the UV/ Vis-SEC method described above. Since the tablets are dissolved in 25 mL of 0.1 M NaOH solution, and they contain 400 μ g of FA, the real concentration of the test sample was 36 μ M of FA.

Figure 5a shows the UV/Vis absorption spectra registered during the oxidation of the Yodocefol® sample prepared in alkaline medium. Only the four bands that had been observed for the FA standard solutions are detected, although they are slightly noisy due to scattering of light, possibly caused by the presence of small particles of the excipient that have not been completely removed during the centrifugation process. It is noteworthy the robustness of the method, allowing us to perform the determination of the analyte despite the dispersion produced by this complex matrix. Figures of merit for the new calibration models at 332 and 392 nm are very similar to the ones obtained in the calibration shown in Table 1. In this case, the slope is higher $(8.1 \cdot 10^{-4} \ \mu M^{-1}$ at 332 nm and $13.4 \cdot 10^{-4} \ \mu M^{-1}$ at 392 nm) because the optical pathway is longer (2.90 mm). Two replicated experiments are shown in Fig. 5. As can be seen, the two signals (spectra, inset Fig. 5a, and CVAs at 392 nm, Fig. 5b) are very similar indicating the good reproducibility of the method, even in this complex matrix. The same absorption spectrum at +0.55 V in the reverse scan

Table 1 Figures of merit for the linear regression models obtained from the absorbance values at 332 and 392 nm, at+0.55 V in the reverse scan

Wave- length	Slope (µM ⁻¹)	Intercept (a.u.)	R^2	S _{yx}	$LOD \; (\mu M^{-1})$
(nm)	M /				
332	$4.7 \cdot 10^{-4}$	$2.2 \cdot 10^{-3}$	0.998	7.4.10 ⁻⁴	4.7
392	$7.5 \cdot 10^{-4}$	$2.5 \cdot 10^{-3}$	0.998	$1.2.10^{-3}$	4.8

 R^2 coefficient of determination, S_{yx} standard deviation of residuals, *LOD* limit of detection

Fig. 5 (a) Absorption spectra recorded at different potentials during the oxidation of the Yodocefol® sample. Inset shows the spectra of two Yodocefol® samples at +0.55 V in the reverse scan. (b) CVAs at 392 nm for two Yodocefol® samples. Experimental conditions in the Fig. 2 caption. Optical path length = 2.9 mm



is obtained for the Yodocefol® sample and for a standard solution, as well as the same CVAs at 392 nm independently on the sample matrix. These results led us to conclude unequivocally that the only molecule that is oxidized is the FA, which is present in the Yodocefol® sample. The oxidation begins at a potential of +0.55 V, as in the previous cases, increasing the absorbance, until it reaches a maximum absorbance value at +0.55 V in the reverse scan, the potential selected for quantification (Fig. 5b).

A concentration of $37 \pm 4 \,\mu$ M for the linear regression model obtained with ΔA values at +0.55 V in reverse scan at 332 nm and $38 \pm 5 \,\mu$ M for the corresponding calibration at 392 nm was obtained, with the real concentration being 36 μ M. Therefore, a good prediction of the test sample is obtained. Moreover, a low %RSD, 4.8%, was assessed for the regression models at the two characteristic wavelengths, which indicates a high degree of coincidence between repeated determinations.

From these results, we can conclude that the new analytical method based on UV/Vis-SEC allows us the determination of FA in complex matrices with very good figures of merit.

Conclusions

UV/Vis-SEC has been demonstrated to provide very good analytical figures of merit for the determination of FA. It should be noted that electrochemistry is used to generate the oxidation product that, combined with spectroscopy, allows us to obtain a determination of this molecule. In many cases, a double determination of the sample is possible, but sometimes, as in the system shown in this work, the electrochemical signal cannot be used for quantitative purposes, and the determination is made only with the spectroscopic data generated during the oxidation process of the FA. Therefore, the marriage of optical techniques with electrochemistry facilitates the development of analytical methods. It is noteworthy that, in the determination of FA, UV/Vis spectroscopy is used to detect the products of a reaction that are not observed in the electrochemical signal. FA contained in Yodocefol® has been successfully determined. The interfering compounds in this commercial drug do not affect the quantification process, demonstrating the capability of these techniques in the analysis of complex samples. UV/Vis-SEC shows a very good reproducibility, which is remarkable considering the simple pretreatment of the sample prior to its quantification. Moreover, some non-soluble excipient compounds were observed in the test sample, but despite that, highquality analytical figures of merit were obtained. A simple univariate calibration has been used for the determination of test samples, but as has been demonstrated, different absorption bands can be used to carry out the quantification of the sample. Therefore, analytical solutions based on multivariate analysis could be easily implemented to extract the analytical information in case of strong interferences. Additionally, UV/Vis-SEC provides information on the electrochemical oxidation process, allowing to obtain spectra related to the products of the reaction.

Authors' contributions F.O. and A.R. contributed to the acquisition of data. F.O., A.R., A.H, and A.C. contributed to the conception, design, and implementation of the experiments and to the analysis and interpretation of the results. A.H and A.C. contributed to the writing of the manuscript. F.O., A.R., A.H, and A.C. contributed to the revision of the manuscript. This work has been headed by A.H. and A.C.

Funding Open Access funding provided thanks to the CRUE-CSIC agreement with Springer Nature. Authors acknowledge the financial support from Ministerio de Economía y Competitividad (Grant CTQ2017-83935-R-AEI/FEDERUE), Junta de Castilla y León (Grant BU297P18), and Ministerio de Ciencia, Innovación y Universidades (Grant RED2018-102412-T). F.O. is grateful for the contract funded by Junta de Castilla y León, the European Social Fund, and the Youth Employment Initiative.

Data availability Not applicable.

Code availability Not applicable.

Declarations

Competing interests The authors declare no competing interests.

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