#### **REVIEW**



# Prognosis in home dental bleaching: a systematic review

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Received: 3 March 2023 / Accepted: 8 May 2023 / Published online: 5 June 2023 © The Author(s) 2023

#### **Abstract**

**Objectives** The aim of this systematic review was to evaluate the prognosis of at-home dental bleaching using low concentration bleaching products.

**Materials and methods** This review was conducted was performed following the recommendations of the 2020 PRISMA statement and was registered in the International Prospective Register of Systematic Reviews (PROSPERO-CRD42022360530). The PICO question was "What is the prognosis of home teeth whitening treatment?". An advanced electronic search was made in three databases: PubMed, Web of Science, and Embase.

**Results** The database search led to the retrieval of 225 articles. After elimination of duplicate references, the titles and abstracts of the articles were analyzed with respect to the eligibility criteria, and 24 studies were included for the development of the systematic review.

**Conclusions** Most authors state that the color remains stable between 1 and 2.5 years regardless of the type of bleaching agent or the forms of administration, and color stability in cases of severe discolorations presents a higher degree of recurrence. **Clinical relevance** Given the growing demand for dental cosmetic treatments, the following systematic review may aid the clinician's continuing education and evidence-based practice by providing knowledge on the field of at-home dental bleaching agents and their long-term effects.

Keywords Home dental bleaching · Home dental whitening · Prognosis · Systematic review · Clinical trials

# **Background**

Nowadays, esthetic procedures are widely required in daily dental practice. Patients seeking an attractive smile have been increasing significantly, especially in terms of tooth color [1]. Dental bleaching is considered the treatment of choice to improve patient satisfaction in terms of tooth discoloration, due to its non-invasive approach and low cost compared to other cosmetic dentistry procedures [2]. Many types of color problems can affect the appearance of teeth, and the causes of tooth discoloration must be carefully evaluated to establish a correct diagnosis [3, 4].

Discolorations are classified as either extrinsic or intrinsic. Extrinsic discolorations result from the accumulation of chromogenic substances on the external surface of the As for intrinsic discolorations, they occur after a change in the structural composition or thickness of the dental tissues. They can be caused by systemic or local factors. Systemic causes include those related to drugs (e.g., tetracycline), alterations in the structure or thickness of dental tissues [5, 7]. Local causes include pulp necrosis, intrapulpal hemorrhage, remnants of pulp tissue in the chamber after root canal treatment, root canal filling materials, some coronal restorative materials, enamel microcracks, caries, and aging. These discolorations are treated by tooth bleaching techniques [3–6, 8]. Historically, bleaching techniques were introduced to the clinic in 1848 as a treatment for discolorations of non-vital teeth, using chloride of lime [9]. In 1864, Truman introduced a more effective technique for bleaching



tooth [3–5]. They are secondary to the habitual intake of chromogenic dietary sources such as wine, coffee, tea, carrots, oranges, chocolate, tobacco, some mouthwashes, or poor or incorrect oral hygiene habits. These discolorations, most of the times, can be eliminated mechanically by professional prophylaxis treatments [3–7].

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non-vital teeth that consisted in the application of a solution formed by chlorine and acetic acid [10].

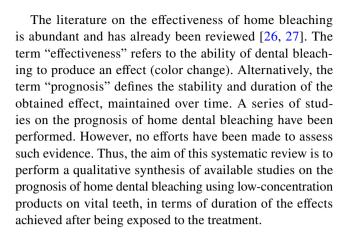
At the end of the nineteenth century, other bleaching agents such as potassium cyanide, oxalic acid, sulfurous acid, aluminum chloride, sodium hypophosphate, pyrozone, hydrogen dioxide, and sodium peroxide were also successfully used in bleaching non-vital teeth [11–16]. Finally, in 1868, whitening techniques for vital teeth were introduced, using oxalic acid, pyrozone, and later hydrogen peroxide (HP) [11–16]. Dental bleaching techniques can be classified into vital and non-vital bleaching techniques. The former are classified as: in-office bleaching, at-home bleaching supervised by the dentist and at-home bleaching without supervision, using over-the-counter bleaching products [17].

Bleaching in the clinic uses bleaching products in high concentrations (25-40% HP). This procedure is carried out after having performed some previous maneuvers such as hard tissue prophylaxis and soft tissue protection through the application of physical barriers. The agents applied can be activated chemically or by light [18]. Alternatively, at-home dental bleaching involves the use of bleaching products with lower concentrations [5]. Originally, these techniques consisted of the application of 10% carbamide peroxide (CP) in individualized splints applied overnight (6–8 h). Currently, gels with concentrations up to 20% are applied [1, 19, 20]. However, the industry has developed products that act more quickly compared to those described above and are more attractive to some patients. These products are presented in the form of gels containing HP in concentrations between 3 and 10%, mainly applied during the day [21, 22].

Supervision by the dentist takes place during check-up appointments [1, 19–23]. The advantages of this technique include: self-administration by the patient, less time in the dental chair, high degree of safety, fewer adverse effects, and low cost [24]. The disadvantages include the need for high patient collaboration since the result is linked to the diligence with which the indicated guidelines are respected. Excessive or prolonged use of the treatment may cause increased tooth sensitivity and soft tissue irritation. Sometimes both techniques can be combined [11, 24].

The third technique, also known as "over-the-counter," involves the use of over-the-counter products containing low concentration bleaching agents. These products are purchased and applied without professional supervision and come in the form of toothpaste, whitening strips, mouthwashes, and prefabricated splints [5, 25].

The literature describes a wide variety of protocols and methods for applying the products described above, although there is still no single accepted protocol [23]. As for the stability of the color obtained after bleaching, this depends to a large extent on diet and habits as they contribute to the development of extrinsic discolorations [5].



## **Materials and methods**

# **Protocol and registration**

This study protocol was registered in the International Prospective Register of Systematic Reviews (PROSPERO-CRD42022360530).

# Search strategy

This systematic review of the literature was performed following the recommendations of the PRISMA statement updated in 2020 [28]. A literature search was conducted in digital databases PubMed, Web of Science, and Embase in May 2022 and was updated on the 29th of March 2023. No language or time restrictions were applied. The search strategy employed three main fields: field #1, keywords regarding prognosis (prognosis or dura\*); field #2, keywords regarding dental bleaching ("tooth bleaching" or "teeth bleaching" or "dental bleaching" or "dental whitening" or bleaching or whitening); and field #3, keywords regarding at-home dental bleaching specifically (home). Keyword selection was based on the descriptors used in previous studies in the field. Whenever possible, both controlled and uncontrolled terms were used. See Supplementary Table 1 for the specific search strategy for each database. Restrictions to publication date and language were not applied. The PICO question guiding the search was: What is the prognosis of home dental bleaching treatment? Which can be subdivided as follows:

P (population): individuals undergoing home dental bleaching; I (intervention): home bleaching using specific low concentration bleaching agent/s with specific application method/s; C (comparison): home bleaching using other specific low concentration bleaching agent/s and/or other specific application method/s; and O (outcomes).

The search strategy, study selection process, data extraction, and quality assessment (risk of bias assessment) were



performed by two independent investigators (R.F. and M.M). In case of doubt, a third investigator was consulted (JL.S).

# **Eligibility criteria**

Studies were selected based on previously established inclusion criteria: (1) RCT; (2) clinical trials; (3) home bleaching protocols using HP in concentrations lower than 10% or CP in concentrations lower than 28%, with post-treatment follow-up times of no less than 2 months; (4) in vivo studies in patients of any age group. Studies in which in-office bleaching protocols were applied with concentrations not dispensable for home use, combined bleaching techniques, home VS in-office bleaching techniques, retrospective studies, in vitro, cohort, clinical cases, and case series were excluded.

# Study selection, data extraction, and synthesis of the evidence

All the selected articles were imported into a citation management software (Mendeley. Elsevier, Amsterdam, Netherlands), and duplicate articles were eliminated. A first screening of the articles according to title and abstract was then performed according to the search strategy described above, and finally a second screening of the full text of the eligible studies was performed.

The following bibliometric data were extracted from each study: author and year of publication. As for the methodological variables, the following were extracted: study type, diagnostic method, number of participants and dropout, bleaching protocol, bleaching product application method, age range, and bleaching method. Lastly, the following outcome variables were extracted: results in terms of DSGU/ $\Delta$ E; results in terms of color change in  $\Delta$ E values (if spectrophotometer was used); and results in terms of color change in DSGU (shade guide unit) values (if a shade guide was used).

After data extraction, a synthesis of the evidence was performed. To do so, the extracted variables were assessed to search for similarities and/or contradictions among the included studies' results. Lastly, a qualitative synthesis of the studies' data regarding the primary outcomes of this review (prognosis of home bleaching, measured over time by analysis of  $\Delta E$  and color change in DSGU dental units) was performed.

#### **Quality assessment**

Quality assessment of the selected studies was carried out using Cochrane Collaboration tools for risk of bias: ROB-2 for randomized controlled trials (RCTs) and ROBINS-I for controlled trials (CTs) [29]. No other analyses could be performed due to the heterogeneity of the data.

#### Results

# **Study selection**

The database search led to the retrieval of 225 articles: 33 in Web of Science, 138 in PubMed, and 54 in Embase. After elimination of duplicate references, the titles and abstracts of the articles were analyzed with respect to the eligibility criteria, and 24 articles were selected for full-text reading. All of the assessed studies were eligible for the qualitative synthesis (Fig. 1).

# Study methodology

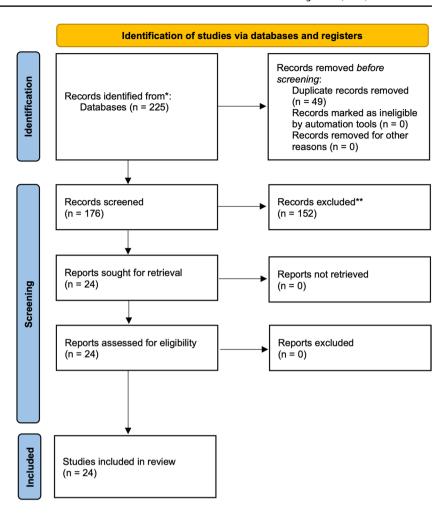
The characteristics of the 24 selected studies are described in Table 1 (methodology) and Table 2 (results). Sixteen randomized clinical trials [1, 2, 19-23, 30-38] and 8 clinical trials [39-46] were selected. In total, 1197 subjects were exposed to home bleaching using different bleaching agents such as HP at concentrations of 3% [35], 5% [20] 5.3% [20] 6% [21, 22, 34, 43], 7.5% [21, 40], and 10% [1, 21] and CP at concentrations of 10% [1, 2, 19, 22, 23, 30–33, 36–39, 42–46], 15% [34, 41, 46], 16% [23, 30, 38], 20% [46], and 28% [37]. A total of 8 studies analyzed the color change combining the use of spectrophotometer and visual color guides (classic vita pan type or similar) [1, 2, 19, 22, 23, 30, 36, 38], while 8 studies used color guides as the only recording method [20, 21, 33, 35, 39, 42, 44, 45]. One study used a spectrophotometer to determine color changes [37], 3 studies combined the use of colorimeter and color guides [40, 41, 46], and 4 studies used only a colorimeter [31, 32, 34, 43]. Some authors used customized splints adapted to the size of the sensor of the spectrophotometer or colorimeter, in order to favor the reproducibility of the measurements [1, 2, 19, 31, 32, 36, 40, 41]. Others simply placed the sensor in the center of the tooth, which is not as precise as the method described above [22, 23, 30, 37, 38, 43, 46].

The study with the lowest number of patients included 20 of them [37] and those with the highest number included 92 [22, 23, 30, 38]. Most of the studies opted for customized bleaching splints with a reservoir as the method of application of the bleaching agent [1, 2, 19–23, 30, 34, 36–38, 41–44, 46]. Six studies applied the agent in customized splints without a reservoir [31–33, 35, 39, 40]. Two studies applied the agent using customized splints with and without a reservoir, following a split-mouth design [

Regarding the bleaching agent application protocols, different patterns were observed. Most of the studies chose short exposure times between 30 min and 3 h [1, 19–21, 23, 30, 34–38, 40, 41, 43, 45]. The remaining studies opted for long exposure times between 6 and 8 h, maintaining a nocturnal pattern



**Fig. 1** Study selection flow diagram



#### Study results

Authors who measured color stability using objective techniques after 2 [43] and 3 months [2, 34, 40, 41] reported homogeneous results between groups. Slight recurrences were also described. However, they did not reach statistically significant values (p < 0.04). The studies that performed a 6-month follow-up [19, 22, 30–33, 35, 44] reported similar results. Authors who measured both  $\Delta E$  and DSGU did not reach statistical significance ((p = 0.3, p = 0.7) [30] and (p > 0.05) [19]). Authors who only measured  $\Delta E$  recorded stable values [31, 32]. The variations recorded indicated a slight regression of the color, but they were not statistically significant (p < 0.05). Three studies only performed DSGU measurements, two of them reported no recurrence [33, 35], and the third study [44] reported a setback of 1 DSGU unit and a recurrence of 12%.

Authors who performed follow-ups after 1 year post bleaching, did not report significant differences between the color change from immediate post bleaching and after 1 year. One study highlighted differences between beach guides, which were clinically significant after  $\Delta E$  calculation [36]. Two studies [23, 38], in the annual and 2 years post-bleaching follow-ups, reported color stability in terms of the  $\Delta E$  values.

In a study in which smokers were compared to non-smokers 2.5 years post bleaching [45], DSGU measurement showed a slight but statistically significant color recurrence in both groups. Two studies assessed patients with intrinsic discolorations secondary to the intake of tetracyclines [42, 46]. Follow-ups were performed at 5 and 7.5 years. The first study,  $\Delta E$  results, showed that the result achieved after the treatment had maintained 68%, 67%, and 66% of the applied concentrations of CP (10%, 15%, and 20%, respectively) [46]. The second study, after 90 months, showed that 60% of the subjects reported no visible changes, 7% reported a slight recurrence, and another 7% showed a moderate recurrence [42].



| Authors         | Year Study tyl | Study type Diagnostic method  | N° of<br>participants<br>(dropout) | Bleaching protocol  | Application<br>method                                | Age range                             | Bleaching agent |
|-----------------|----------------|---|------------------------------------|---|--|---------------------------------------|-----------------|
| Meireles et al. | 2022 RCT       | Vita pan classical,<br>Spectrophotometer                                | 40(1)                              | GI:CP10% (Polanight 10%, SDI) 4 h weeks, preferably at night G2:CP10% (Polanight 10%, SDI) 4 h per day ×2 weeks, preferably at night  | G1:ICWR<br>G2:ICWR                                   | × 18                                  | CP10%           |
| Martini et al.  | 2021 RCT       | Spectrophotometer, Vita<br>pan classical, Vita Bleach<br>guide 3dmaster | 46(10)                             | G1:PC10%(Opalescence, Ultradent) 3 h per day ×3 weeks<br>G2:PC10%(Opalescence, Ultradent) 3 h per day ×3 weeks  | G1:ICWR<br>G2:ICWOR                                  | SN                                    | CP10%           |
| Mailart et al.  | 2021 RCT       | Spectrophotometer, Vita pan<br>classical, Vita Bleach guide<br>3dmaster | (0)09                              | GI: HP10%(Opalescence go, Ultradent) 30 min per day ×2 weeks G2: HP10%(White Class) 30 min per day ×2 weeks G3: CP10% (opalescence) 2 h per day × 2 weeks G4: PC10%(Opalescence) 8 h per day × 2 weeks  | G1 ICWR<br>G2 PCPM<br>G3 ICWR<br>G4:ICWR             | ^ 18                                  | CP10%, HP10%    |
| Darriba et al.  | 2019 RCT       | Spectrophotometer,<br>Vita pan classical                                | 50(0)                              | G1:CP10%(vivastyle vialdent ivoclar) 2 h per day × 2 weeks<br>G2:CP10%(vivastyle vialdent ivoclar 2 h per day × 3 weeks   | G1:ICWR<br>G2:ICWOR                                  | V 18                                  | CP10%           |
| Pinto et al.    | 2017 RCT       | Vita Bleach guide 3dmaster  | 30(NI)                             | G1: HP6% (White Class with calcium – FGM)1 h per day × 7 days;<br>G2, HP7.5%(White Class with calcium—FGM)1 per day × 7 days;<br>G3 HP10% (Oral B 3D White – Oral-B) 30 min 2 times per day × 7 days<br>G4 control group–placebo 1 h per day × 7 days | G1 ICWR<br>G2: ICWOR<br>G3: STRIPS<br>G4: ICWOR      | 12–20                                 | HP6%-7,5%,10%   |
| Botelho et al.  | 2017 RCT       | Colorimeter   | 26(2)                              | G1:CP15%(Nupro White GoldTM, Dentsply r) 2 h per day or all night x3 months G2:HP6%(Crest Whitestrips 30 min 2 times per day x3 months  | G1: ICWR<br>G2: STRIPS                               | 287/304                               | CP15% HP6%      |
| De Geus et al.  | 2017 CT        | Vita clásico, Vita Bleach<br>guide 3dmaster                             | 60(17)                             | G1:Smokers, PC10%Opalescence) 3 h per<br>day×3 weeks<br>G2 NONsmokers PC10%Opalescence) 3 h per<br>day×3 weeks  | G1: PCPM<br>G2: PCPM                                 | 18–54                                 | CP10%           |
| Aka et al.      | 2017 RCT       | Vita pan classical,<br>spectrophotometer                                | 92(2)                              | G1:control G2:PC10% (Opalescence pf)8-10 h per day x 14 days; G3 HP6% (Opalescence Go) 1 h per day x 14 days;   | G1 only color<br>measurement<br>G2: ICWR<br>G3: PCPM | × × × × × × × × × × × × × × × × × × × | CP10% HP6%      |
| Auschill et al. | 2012 RCT       | Vita pan classical  | 30(NI)                             | G1: HP5%(Colgate visible white) 30 min 2 v per day × 14 days G2: HP5,3%s (Whitestrips procter and gamble)   | G1: ICWR<br>G2: STRIPS                               | 18–56                                 | HP5% HP5,3%     |



| Table 1 (continued) | (pan            |   |                                    |  |  |           |                           |
|---------------------|-----------------|---|------------------------------------|--|--|-----------|---------------------------|
| Authors             | Year Study type | Study type Diagnostic method  | N° of<br>participants<br>(dropout) | Bleaching protocol   | Application<br>method  | Age range | Age range Bleaching agent |
| Turkun et al.       | 2010 RCT        | Spectrophotometer   | 20(NI)                             | G1: CP28%(meta trayRemedent) 20 min per<br>day × 10 days<br>G2: PC10%(opalescence PF) 6-8 h per<br>day × 10 days   | G1:PCPM<br>G2: ICWR  | 20–30     | CP28% CP10%               |
| Meireles et al.     | 2010 RCT        | Spectrophotometer, Vita<br>pan classical                                | 92(11)                             | GI: CP10%(cp10) 2 h per day × 3 weeks G2: PC16% (Whiteness Perfect, FGM) 2 h per day × 3 weeks   | G1:ICWR<br>G2:ICWR   | 18–55     | CP10%, CP16%              |
| Meireles et al.     | 2009 RCT        | Spectrophotometer, Vita<br>pan classical                                | 92(3)                              | GI: CP10%(cp10) 2 h per day x 3 weeks G2: PC16% (Whiteness Perfect, FGM) 2 h per day x 3 weeks   | G1:ICWR<br>G2:ICWR   | 18–55     | CP10%, CP16%              |
| Meireles et al.     | 2008 RCT        | Spectrophotometer Vita<br>pan classical                                 | 92(3)                              | GI: CP10%(cp10) 2 h per day × 3 weeks<br>G2: PC16% (Whiteness Perfect, FGM) 2 h per<br>day × 3 weeks   | G1: ICWR<br>G2: ICWR   | 18–55     | CP10%, CP16%              |
| Medeiros et al.     | 2008 CT         | Vitapan classical   | 50(0)                              | G1: placebo G2:PC10% (Opalescence pf)8-10 h per day $\times$ 21 days;  | G1: ICWR<br>G2: ICWR   | 18–25     | CP10%                     |
| Hannig et al.       | 2007 CT         | Colorimeter   | 47(NI)                             | G1:HP6%(Whitestrips) 30 min per day, twice a day × 2 weeks G2:CP10%(vivastyle vialdent ivoclar)1 h per day × 2 weeks   | G1: STRIPS<br>G2: ICWR   | 18–60     | CP10% HP6%                |
| Matis et al.        | 2006 CT         | Colorimeter Vitalescence<br>Esthetic Restorative Masters<br>Shade Guide | 44(?)                              | G1: CP10% 6-8 h per day×6 months<br>G2: PC15% 6-8 h per day×6 months<br>G3: CP20% 6-8 h per day×6 months SPLIT<br>MOUTH DESIGN   | G1: ICWR<br>G2: ICWR<br>G3: ICWR   | > 18      | CP10%,<br>CP15%,CP20%     |
| Leonard et al.      | 2003 CT         | Vita pan classical  | 21(6)                              | G1: CP10% 6-8 h(Opalescence pf) per<br>day×6 months  | G1: ICWR in a quadrant ICWOR on the other quadrant   | > 18      | CP10%                     |
| Myers et al.        | 2003 RCT        | Vita pan classical  | 65(NI)                             | G1: HP3%(ADS Tooth Whitening Gel, Applied Dental Sciences, Inc., Lee, MA)) 30 min per day 3 times per day, tot 90 min)×14 days G2: Placebo 30 min per day 3 times per day, tot 90 min)×14 days | G1: ICWOR<br>G2: ICWOR   | >21       | НР3%                      |
| Matis et al.        | 2002 CT         | Colorimeter, Vita pan classical 26(NI)                                  | 26(NI)                             | G1: CP15%(Rembrandt Xtra-Comfort Non- Sensitizing Bleaching Gel Regular Strength,)2hper day × 14 days  | G1: ICWR on<br>the left anterior<br>quandrant<br>ICWOR on the<br>right anterior<br>quandrant | 23-68     | CP15%                     |



CP10%- placebo CP20% HP 7.5% Bleaching agent CP10% CP10% CP10% Age range +35-35 23-64 20-57 22-79 × 18 G1: ICWOR G2: ICWOR G1: ICWOR G1: ICWOR G1: ICWOR G2: ICWOR G1: ICWOR Application method 32: HP7.5% (daywhite discuss dental) 1 h al 2 G1: CP10% 6–8 h(Opalescence pf)  $\times$  2 weeks G1: CP20%(Opalescence pf)1 h 2 times per G1: CP10% 6-8 h(Opalescence pf) × 5 days G2:PC10% (Opalescence pf) 8–10 h per G1: placebo  $8-10 \text{ h per day} \times 21 \text{ days}$ G1: CP10% 6-8 h per day  $\times$  2 weeks G2: placebo  $6-8 \text{ h} \times 5 \text{ days}$ times per day  $\times$  14 days Bleaching protocol  $day \times 21 days$  $day \times 14 days$ G2:placebo participants (dropout) 52(NI) 24(NI) 50(NI) 29(5) (0)09 Frubyte bioform guide, Lumin Vacuum Shade Diagnostic method Vita pan classical Colorimeter Guide, Vita Colorimeter Colorimeter Study type RCT RCT RCT CI1996 2000 1996 Year Table 1 (continued) Rosenstiel et al. Mokhlish et al. Russel et al. Matis et al. Swift et al.

by the manufacturer; RCT, randomized clinical trial; CT, reservoir; ICWOR, individualized cuvette without reservoir; PCPM, preformed cuvette provided clinical trial; CP, carbamide peroxide, HP, hydrogen peroxide individualized cuvette with

#### Risk of bias

All studies were rated using the Cochrane Collaboration tools for risk of bias: RoB2 was used for rating RCT and ROBINS-I for CT (Figs. 2 and 3).

RCT studies were classified as unclear for the following elements: bias arising from the randomization process [2, 19, 21, 22, 37], bias due to deviations from the intended intervention [2, 19, 22, 23, 30, 32–38], and bias in outcome measurement [2]. Other studies were classified as high risk for the following elements analyzed: bias arising from the randomization process [32] and bias due to deviations from the intended intervention [1, 21, 31]. The remaining studies were classified as low risk for the following elements: bias arising from the randomization process [1, 20, 23, 30, 31, 33–36, 38], bias due to deviations from the intended intervention [20], bias due to missing data on outcomes [1, 2, 19–23, 30–38], bias in outcome measurement [1, 19–23, 33–41], and bias in selection of the reported outcome [1, 2, 19–23, 30–38].

CT studies were classified as low risk for the following elements: bias due to confounding [39–46], bias due to the selection of participants [39–46], bias in classification of interventions [39–46], bias due to deviations from intended interventions [39–46], bias due to missing data [40, 41], bias in measurement of outcomes [40, 44], and bias in the selection of the reported result [39–46]. Other studies were classified as unclear for the following elements: bias due to missing data [39, 43–46] and bias in measurement of outcomes [39, 41–43, 45, 46]. Only one study was classified as high risk for the following element: bias due to missing data [42].

#### **Discussion**

Concern has been expressed about the duration of the effects of dental bleaching. This has been the subject of several studies over the years [3, 4]. Multiple variables such as effectiveness, intra- and post-treatment sensitivity, and long-term color stability have been analyzed [47]. The great heterogeneity in the designs of the research methodologies, the lack of a single clinical and follow-up protocol (or at least a minimum for conducting studies), and the different techniques applied to obtain the data make it difficult to reach a consensus. Nevertheless, the results obtained in the included studies highlight that the color obtained after home dental bleaching remains stable between 1 and 2.5 years regardless of the bleaching agent used or its forms of administration.

The selected studies compared the performance of athome bleaching products by applying different experimental protocols with the aim of monitoring their long-term prognosis. The number of participants was different among



 Table 2
 Summary of the results of the included studies

| Authors         | Year     | Study type | Year Study type Diagnostic method                                      | N°<br>par-<br>ticipants<br>(dropout) | Results in terms of DSGU/ $\Delta E$ | Results: color change in ΔE values (spectrophotometer)  | Results: color change in DSGU (shade guide units) values  | Follow-up |
|-----------------|----------|------------|--|--------------------------------------|--------------------------------------|---|---|-----------|
| Meireles et al. | 2022     | RCT        | Vitapan Classical,<br>Spectrophotometer                                | 40(1)                                | DSGU+ AE                             | G1:2 weeks 11.4(3.3) <sup>a</sup> 1<br>month11.4(2.7) <sup>a</sup><br>3 months 10.5 (2.6)b<br>G2: 2 weeks 7.6(2.4) <sup>a</sup> 1<br>month7.1(2.3) <sup>a</sup><br>3 months 6.4(2.6)b | Used only in initial screening phases   | 3 months  |
| Martini et al.  | 2021 RCT | RCT        | Spectrophotometer, Vitapan<br>Classical, Vita Bleach<br>guide 3dmaster | 46(10)                               | DSGU+ AE                             | G1:baseline vs 1 month 8.4±2.3 baseline vs 1 year: 9.0±2.0 G2 baseline vs 1 month 18.6±2.5 baseline vs 1 year: 8.9±2.5  | Vita classical: G1:baseline vs 1 month 8.1±2.1 baseline vs 1 year: 8.0±2.1 G2:baseline vs 1 year: 8.0±2.1 G2:baseline vs 1 year: 8.1±2.0 VITA Bleachedguide 3D-MAS-TER G1: baseline vs 1 month: 10.4±3.1 baseline vs 1 year: 9.2±2.9 G2:baseline vs 1 month 10.8±2.7 baseline vs 1 year 9.3±2.7 | l year    |
| Mailart et al.  | 2021 RCT | RCT        | Spectrophotometer, Vitapan<br>Classical, Vita Bleach guide<br>3dmaster | (0)09                                | DSGU+∆E                              | G1: 2 weeks 7.03 ± 1.83 1 year 6.03 ± 2.33 G2: 2 weeks 6.75 ± 1.38 1 year 6.48 ± 1.02 G3: 2 weeks 7.13 ± 2.75 1 year 5.67 ± 1.66 G4: 2 weeks 7.49 ± 1.51 1 year 7.75 ± 2.04           | ca: ss5.20±0.99 1 year 30 ss 5.33±1.85 1 year 56 s 4.63±1.58 1 year 33 s 5.63±0.95 1 year 29 chedguide 3D-MAS- chedguide 3D-MAS- cs6.70±1.44 1 year 35 ss 6.86±1.60 1 year 42 ss 7.41±1.40 1 year 72 ss 7.70±1.72 1 year  | l year    |
| Darriba et al.  | 2019     | RCT        | Spectrophotometer + Vitapan<br>Classical                               | 50(0)                                | DSGU+∆E                              | G1:4.74±1.94 4.81 ±2.19<br>G2: 5.77±2.15. 5.56±2.09   | 3.12 -3.45 ± 4.04<br>3.91 – 5.15 ± 4.20   | 6 months  |



| lable 2 (continue | numaca)                           |           |            |                                     |   |           |
|-------------------|-----------------------------------|-----------|------------|-------------------------------------|---|-----------|
| Authors           | Year Study type Diagnostic method | °N        | Results in | Results: color change in $\Delta E$ | Results: color change in DSGU Follow-up | Follow-up |
|                   |                                   | par-      | terms of   | values (spectrophotometer)          | (shade guide units) values              |           |
|                   |                                   | ticipants | DSGU/ AE   |                                     |   |           |
|                   |                                   | (dropout) |            |                                     |   |           |

| Year 2017 2017           | Study type | 7.  | 0                                    |                                    |   |   |                       |
|--------------------------|------------|---|--------------------------------------|------------------------------------|---|---|-----------------------|
| 2017                     |            | Study type Diagnostic method  | N°<br>par-<br>ticipants<br>(dropout) | Results in<br>terms of<br>DSGU/ AE | Results: color change in ΔE values (spectrophotometer)                                  | Results: color change in DSGU<br>(shade guide units) values   | Follow-up             |
|                          | RCT        | Vita Bleach guide 3dmaster  | 30(NI)                               | DSGU                               | 1   | G1: 2M3/1 M<br>G2: 3M1 /1M1<br>G3:3M1/1M1   | 1 year                |
| 7100                     |            | Colorimeter   | 26(2)                                | $\Delta \mathrm{E}$                | G1:7.84 (3.88) 10.11 (3.42)<br>G2:5.53 (4.16) 8.23 (4.69                                | I   | 3 months              |
| De Geus et al. 2017      | CT         | Vita classical, Vita Bleach<br>guide 3dmaster                         | 60(17)                               | DSGU                               | I   | VITA CLASSICAL<br>G1:5.2+-2.1.4.7+-2.2<br>G2:5.7+-2.3 5.1+-2.2<br>Vita 3d master<br>G1:4.1+-1.1 2.6+-1.1<br>G2:4.7+-14 3.3+-1.2 | 30 months (2.5 years) |
| Aka et al. 2017 RCT      |            | Vitapan Classical,<br>spectrophotometer                               | 92(2)                                | DSGU+ ΔE                           | G1:1.1,1,6-1.0,1,3-1.0,1.3<br>G2:5.0,4.8-7.0,6.9-9.8,9.8<br>G3:3.3,3.3-4.8, 4.9-5.7,6.2 | N   | 6 months              |
| Auschill et al. 2012     | RCT        | Vitapan Classical   | 30(NI)                               | DSGU                               | G1;9.63+-1.14 6.35+-3.50<br>G2:9.68+-0.95 6.41+2.65                                     | 1   | 18 months (1.5 years) |
| Turkun et al. 2010       | RCT        | Spectrophotometer   | 20(NI)                               | $\Delta \mathrm{E}$                | G1:4.3 + -1.84 3.2 + -1.25<br>G2:9.3 +2.4. 8.3 +2.73                                    | I   | 1 year                |
| Meireles et al. 2010     | RCT        | Spectrophotometer, Vitapan<br>Classical                               | 92(11)                               | $DSGU + \Delta E$                  | G1:8.5 (6.0–15.0) 2.9 (1.5–11.8)<br>G2:8.8 (6.0–13.5) 2.7 (1.3–5.8)                     | N   | 2 years               |
| Meireles et al. 2009 RCT |            | Spectrophotometer, Vitapan<br>Classical                               | 92(3)                                | DSGU+∆E                            | G1:8.5 (6.0 to 15.0) 2.7 (1.3 to 11.8)<br>G2:8.8 (6.0 to 13.5). 2.7 (1.3 to to 6.0)     | G1::8.7 (3.3 to 14.7), 2.5 (1.1 to 10.5)<br>G2::8.0 (4.3 to 12.7) 2.3 (1.3 to 5.7)  | 1 year                |
| Meireles et al. 2008     | RCT        | Spectrophotometer, Vitapan<br>Classical                               | 92(3)                                | DSGU+∆E                            | G1:8.5 (6.0 to 15.0) 3.0 (1.3 to 10.5)<br>G2:8.8 (6.0 to 13.5). 2.4 (1.2 to 5.8)        | G1:.8.7 (3.3 to 14.7). 22.5 (1.5 to 12.0)<br>G2: 8.0 (4.3 to 12.7) 2.3 (1.0 to 6.2)   | 6 months              |
| Medeiros et al. 2008     | CT         | Vitapan Classical   | 50(0)                                | DSGU                               | 1   | G1:2.04 2.04<br>G2:3.97 2.15  | 6 months              |
| Hannig et al. 2007       | CT         | Colorimeter   | 46(NI)                               | 14                                 | G1: $+1.55 \pm 0.41$ . $73.22 \pm 1.59$<br>G2: $73.55 \pm 1.83$ . $+1.20 \pm 0.37$      |   | 2 months              |
| Matis et al. 2006        | CT         | Colorimeter Vitalescence Esthetic<br>Restorative Masters Shade Guide, | 44(?)                                | DSGU+ AE                           | G1:9./12<br>G2:9.5/12<br>G3:11/12.5   |   | 5 years               |
| Leonard et al. 2003      | CT         | Vitapan Classical   | 21(6)                                | DSGU                               | 1   | G1:C4(C1-C4) baseline<br>B1(B1-C1) post treatment<br>C1(A1-C3) a 90 months  | 90 months (7.5 years) |



| Aumors                     | Year Study type | Year Study type Diagnostic method         | N°<br>par-<br>ticipants<br>(dropout) | Results in terms of DSGU/ AE | Results: color change in ΔE values (spectrophotometer)  | Results: color change in DSGU Follow-up (shade guide units) values | Follow-up |
|----------------------------|-----------------|---|--------------------------------------|------------------------------|---|--|-----------|
| Myers et al.               | 2003 RCT        | Vitapan Classical                         | (NI)                                 | DSGU                         | I   | G1:7.0(D3) 12.0(A2)<br>G2:7.0(D3) 8.0 (A3)                         | 6 months  |
| Matis et al.               | 2002 CT         | Colorimeter, Vitapan Classical            | 26(NI)                               | $DSGU + \Delta E$            | ICRC:4.75 /3,5<br>ICSR: 3,75 / 3  | NI   | 3 months  |
| Mokhlish et al.            | 2000 CT         | Trubyte bioform guide, Colorimeter 24(NI) | 24(NI)                               | DSGU+∆E                      | G1: CENTRAL:6.5/4 LAT-<br>ERAL: 6/3.75 CANINE:8/5<br>G2: CENTRAL:5/3.75<br>LATERAL: 5.5/3.5<br>CANINE:6.5/5.5 |  | 3 months  |
| Swift et al.               | 1999 CT         | Vitapan Classical                         | 29(5)                                | DSGU                         | I   | G1: D3 D2  | 2 years   |
| Matis et al.               | 1998 RCT        | Colorimeter                               | (0)09                                | $\Delta E$                   | G1:1,8, 1,8, 1,5 / 1,8, 1,8, 1,5<br>G2:13.0, 9,6, 8.6 / 6,2, 4,9, 5,0   | I  | 6 months  |
| Rosenstiel et al. 1996 RCT | 1996 RCT        | Colorimeter                               | 52(NI)                               | $\Delta \mathrm{E}$          | G1:4.6, 4,2<br>G2: 1.8,1.8  | I  | 6 months  |
| Russel et al.              | 1996 RCT        | Lumin Vacuum Shade Guide, Vita            | 50(NI)                               | DSGU                         | ı   | G1: A3 A1<br>G2: D3 D3   | 6 months  |

the studies, ranging from 20 to 92. The remaining articles included an average of 51 participants, and only a few studies performed a sample size calculation [1, 19, 22, 23, 30, 36, 38].

The studies included variables such as the presence or absence of a reservoir in the bleaching splints to determine if its presence influenced the result [41, 42]. Color measurements were performed by objective technique, such as the spectrophotometer [37] or colorimeter [31, 32, 34, 43], or subjective operator dependent techniques, using shade guides sorted according to dental brightness [20, 21, 33, 35, 39, 42, 44, 45]. Other studies used a combination of both techniques [1, 2, 19, 22, 23, 30, 36, 38, 40, 41, 46].

The analysis and comparison of the data obtained from the different studies present some limitations since the authors performed the measurements not only in different ways, but also at different times and with different apparatus. In addition, when the measurements were performed with the same apparatus (spectrophotometer or colorimeter), they were not carried out in the same way. In the case of measurements using subjective techniques, it should be noted that the same shade guides were not used in all the studies. This hinders the comparability between studies.

The clinical and follow-up protocols were diverse. The results obtained in the studies regarding the effectiveness of the method of application of the bleaching agent were similar. Therefore, it can be deduced that the form of application of the treatment does not influence the final result. The extent of treatment over time varied between 1 and 3 weeks. In the case of intrinsic staining treatments secondary to drugs (tetracyclines), the treatment was extended up to 6 months [42, 46]. Follow-up times were also heterogenous, ranging from 2 months to 7.5 years.

Authors who measured color stability using objective techniques after 2 [43] and 3 months [2, 34, 40, 41] and who used HP between 6% [34, 43] and 7.5% [40] and CP between 10% [2, 41, 43], 15% [34, 41], and 20% [40] reported homogeneous results between groups. Slight recurrences were also described. However, they did not reach statistically significant values (p < 0.04). Such results could be attributed to the short follow-up time.

The studies that performed 6-month follow-up and applied CP from 10% [19, 22, 30–33, 44] to 16% [30] and HP at 3% [35] and 6% [22] and performed the measurements subjectively [33, 35, 44], objectively [31, 32], and by means of a combination of both [19, 22, 30] and reported similar results. Rosentisel et al. [32] and Matis et al. [31] measured color using a colorimeter. Both applied 10% CP overnight but differed in the extent of treatment: 5 days and 3 weeks, respectively. They agree that after the color measurements and subsequent CIELAB analysis calculating  $\Delta$ E, the color was stable when compared to the post-bleaching values. Also, the variations recorded indicated a



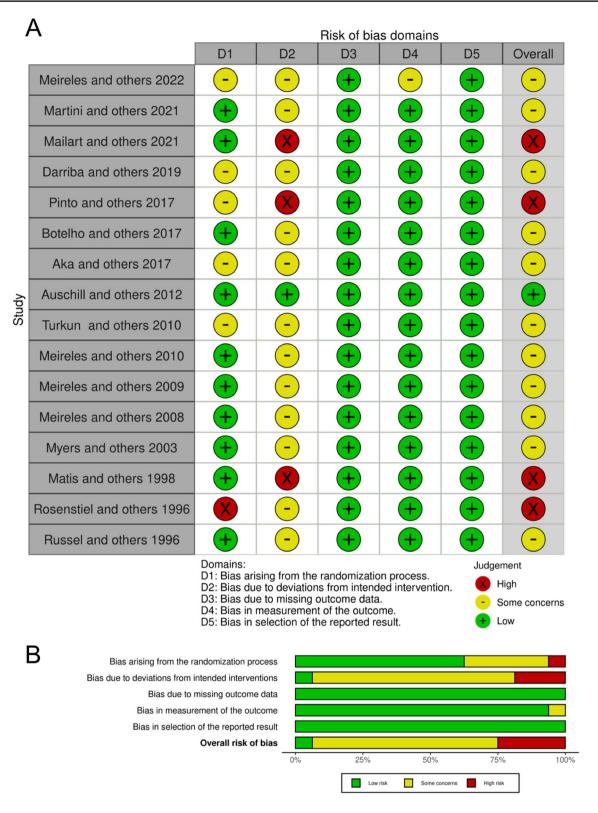
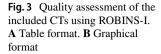
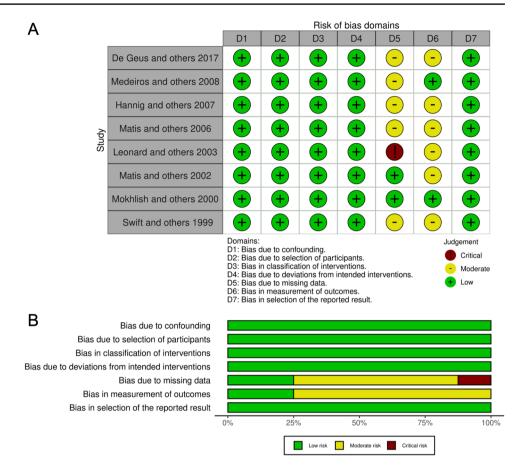


Fig. 2 Quality assessment of the included RCTs using RoB-2. A Table format. B Graphical format





slight regression of the color, but they were not statistically significant (p < 0.05).

In the studies by Meireles et al. [30] and Darriba et al. [19], in which color measurements were made using both techniques, despite having applied different bleaching protocols, the authors reported a slight recurrence in both groups studied. However, again, data did not reach statistical significance ((p=0.3, p=0.7) [30] (p>0.05) [19]). Color measurement by means of color guides confirmed the results obtained after calculation of  $\Delta E$  value. On the other hand, Aka et al. [22] observed a statistically significant (p<0.05) recurrence ( $\Delta E$ ) in the CP group at 10%, and the measurement of color using guides showed better brightness values in the CP group compared to the HP group.

Alternatively, Russell et al. [33], Mayers et al. [35], and Medeiros et al. [44] performed the measurements only by using shade guides. The first two reported no recurrence, while Medeiros et al. [44] reported a setback of 1 DSGU unit and a recurrence of 12%, without attributing this result to a specific cause. The subjectivity of the technique and the difficult comparability of the values derived from it with the values provided by objective techniques, which are capable of recording what the human eye cannot always record, or at least not with such precision, make it difficult to compare the studies with each other.

Other authors performed follow-ups after 1 year post bleaching. Martini et al. [36] performed color measurements using a spectrophotometer and color guides. They reported no significant differences between the color change from immediate post bleaching and after 1 year using the classic vita pan shade guide unit (mean difference =  $0.1 \Delta SGU$ ; 95% central incisor, 0.2 to 0.4; p = 0.53; and at 1 year, mean difference = 0.3 95% central incisor 1.0 to 1.6; p = 0.62). However, significant differences were observed using the VITA Bleachedguide (VITA Zahnfabrik, Bad Säckingen, Germany) (mean difference =  $1.4 \Delta SGU$ ; 95% central incisor 0.7 to 2.1; p < 0.01). After calculation of  $\Delta E$ , these differences were defined as not clinically important, since they were below the 50:50 perception threshold for shade changes in dentistry [36].

Another factor that hinders the comparison of the results of studies using subjective techniques is the diversity between guides. This generates a certain degree of uncertainty about the reliability of the results obtained. Authors such as Mailart et al. [1] and Martini et al. [36] performed follow-up at 1 year using a spectrophotometer and two different color guides and concluded that the color remains stable since no statistically significant differences were observed between the groups after  $\Delta E$  analysis. In addition, no color discrepancies were reported upon comparison with



the two guides. Meireles et al. [23, 38] also performed the color measurements using a spectrophotometer and color guide. In the annual and biennial post-bleaching follow-ups, the results were not significantly different in terms of the  $\Delta E$  values, and the median tooth color did not show statistically significant changes (p > 0.2).

Pinto et al. [21] and Auschill et al. [20] analyzed the shade stability using vita 3DMASTER (VITA Zahnfabrik, Bad Säckingen, Germany) and vita pan classic (VITA Zahnfabrik, Bad Säckingen, Germany) shade guides, respectively. In the study by Pinto et al. [21], all groups showed color stability at 12 months post bleaching. Auschill et al. [20], after 18 months of follow-up, reported recurrence in both groups studied when compared with the results obtained after bleaching, reporting values from  $2.88 \ (p=0.24)$  to  $3.3 \ (p=0.001)$  DSGU.

The origin of discolorations will be decisive in determining both the effect and the prognosis of bleaching. The study conducted by De Geus et al. [45] selected a different sample from the previously mentioned studies, composed of smokers vs. nonsmokers. The aim of their study was to evaluate the stability of whitening at 2.5 years using measurement techniques using classic vita pan and vita bleach 3dmaster shade guides (VITA Zahnfabrik, Bad Säckingen, Germany). At 30 months, a homogeneous high dropout rate (28%) was reported in both groups: 5 participants stopped smoking and 4 reduced the number of cigarettes consumed daily. In the measurements, a slight but statistically significant color recurrence could be detected in both groups. Therefore, the recurrence is not only attributable to extrinsic staining potentially caused by smoking, although in view of the changes in the smoking group, the comparison is not completely fair. The authors whose samples consisted of patients with intrinsic discolorations secondary to the intake of drugs (tetracyclines) were Matis et al. [46] and Leonard et al. [42], who applied similar clinical protocols: nocturnal application of CP for 6 months, at 10%, 15%, and 20% in the case of Matis et al. [46] and only 10% in the case of Leonard et al. [42]. The color measurement techniques and follow-up times varied between the two studies. In the first study [46], they followed up at 5 years using a colorimeter and Vitalescence Esthetic Ultradent shade guides (Vitalescence Esthetic Restorative Masters Shade Guide, Ultradent Products, Inc.), and in the second study [42], they followed up at 7.5 years measuring shade changes using classic vita pan shade guides. Matis et al. [46] reported  $\Delta E$  results, which showed that concentrations of 10%, 15%, and 20% had maintained 68%, 67%, and 66% of the color achieved after bleaching, respectively. The results of Leonard et al. [42] also showed recurrences. After 90 months, 60% of the subjects reported no visible changes, 7% reported a slight recurrence, and another 7% showed a moderate recurrence, but never matching the pretreatment color. In addition, 27%

of the subjects reported having undergone further bleaching sessions before 90 months. Both studies showed higher levels of recurrence than studies whose samples did not include patients with intrinsic staining, which is evidence of a greater degree of difficulty in maintaining the results in these groups of patients.

The main limitation of the present systematic review is based on the great diversity of clinical and follow-up protocols employed in the included studies. In addition, a wide range of measurement methods were used, which make it difficult to compare the results obtained in the studies. Therefore, there is a need to establish a single experimental protocol to facilitate the interpretation of the data obtained.

#### **Conclusions**

Although many studies have demonstrated the general efficacy of bleaching gels, the long-term benefits of the treatment related to color maintenance are not well established in the literature. Most authors state that the color remains stable between 1 and 2.5 years regardless of the type of bleaching agent or the forms of administration such as individualized cuvette with reservoir, individualized cuvette without reservoir, and preformed cuvette provided by the manufacturer. Color stability in cases of severe discolorations such as tetracycline staining presents a higher degree of recurrence.

**Supplementary Information** The online version contains supplementary material available at https://doi.org/10.1007/s00784-023-05069-0.

**Author contribution** M.M. contributed to investigation, conceptualization, methodology, writing-original draft, writing—review and editing.

L.F. contributed to formal analysis, conceptualization writing – review and editing.

J.L.S. contributed to translation, review and editing of the manuscript.

R.F. contributed to investigation, conceptualization, methodology, writing-original draft, writing — review and editing. All authors contributed to preparation and approval of the final version of the manuscript.

Funding Open Access funding provided thanks to the CRUE-CSIC agreement with Springer Nature.

### **Declarations**

Ethical approval Not applicable.

**Informed consent** Not applicable.

**Conflict of interest** The authors declare no competing interests.

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