ORIGINAL PAPER



Assessing the potential of online ICP–MS analysis to optimize Ca/ matrix separation using DGA Resin for subsequent isotopic analysis

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Received: 29 October 2020 / Accepted: 12 March 2021 / Published online: 14 April 2021 © The Author(s) 2021

Abstract

Ca isotopes have gained increasing interest as a diagnostic tool for bone diseases due to the variations in abundances as a consequence of changes in bone-mineral balance. Optimized Ca/matrix separation prior to analysis is a prerequisite for reliable isotope ratio measurements in complex biological matrices such as blood, serum, or urine. The online analysis of analyte/matrix separation by ICP-MS enables direct assessment of elution profiles supporting the optimization process. The integration of transient signals and signal suppression challenge the quantification and interpretation of the elution profiles. Mn and Co remain unretained by the DGA Resin (TrisKem International) from nitric acid. Hence, in the present study, these elements were investigated for their application as standards to monitor signal suppression. Successful analyte quantification was accomplished using a dynamic correction strategy applying a linear gradient of a suppression factor based on Mn and Co intensities. An optimized Ca/matrix separation procedure using DGA Resin is proposed based on the results during online ICP-MS analysis.

Graphic abstract



Keywords Isotopic analysis \cdot Inductively coupled plasma mass spectrometer \cdot Extraction \cdot Bioinorganic chemistry \cdot Metals \cdot (High pressure) liquid chromatography

Introduction

DGA Resin (N,N,N',N'-tetra-n-octyldiglycolamide, TrisKem International, Bruz, France) was developed as an extraction resin in the early 2000s by Horwitz et al. [1]. The primary

advantage of this resin is the high affinity for trivalent REEs and actinides [2, 3], which originated from the need to treat high-level radioactive waste [1, 3]. Therefore, DGA Resin has been applied, first and foremost, for analyte/matrix separation in the nuclear isotopic analysis. Applications include the separation of radioactive isotopes of Sr [4, 5], Y [6], Po [7], Ac [8], Pu [9–11], Am [12], and Cm [13]. The beneficial features and characteristics of DGA Resin are also applicable for analyte/matrix separation of stable isotopes in the fields of geochemical, archaeological, environmental, and biomedical sciences. Table 1 summarizes analyte/matrix separation procedures for isotopic systems using DGA Resin. Future development of analyte/matrix separation procedures might

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so- opic ystem	Sample matrix	Add. resins	RV DGA	Load DGA	Elution DGA	Recovery	Removed (matrix) ele- ments	Co-elution	Detector	References
a	Geological and biological CRMs	I	1 cm ³	2 mol dm ^{-3} HNO ₃	$12 \text{ mol dm}^{-3} \text{HNO}_3$	93% ±2%	Na, Mg, Ti, Rb, Zr, Ba, Nd, Hf, U	n/a	MC ICP-MS	[27]
Ca	GEOLOGICAL CRMs	I	1 cm ³	$3 \text{ mol dm}^{-3} \text{ HNO}_3$	DI water	100%	Mg, Al, Ti, Fe, K	Sr	SMIT	[21]
Ca	Geological and biological CRMs	I	250 mm ³	$4 \text{ mol dm}^{-3} \text{ HNO}_3$	DI water	≥ 95%	Na, Mg, Al, P, K, Ti, Fe, Rb, Sr, Ba	n/a	MC ICP-MS	[20]
Ca	Geological and biological CRMs	AG50W-X12	250 mm ³	5 mol dm^{-3} HNO ₃	DI water	n/a	Na, Mg, Al, K, Ti, Mn, Fe, Cu, Zn, Rb, Sr	n/a	MC ICP-MS	[28]
Sr	Geological and biological CRMs	I	1 cm ³	$2 \text{ mol dm}^{-3} \text{ HNO}_3$	6 mol dm ^{-3} HNO ₃	93%±2%	Na, Mg, Ti, Rb, Zr, Ba, Nd, Hf, U	n/a	MC ICP-MS	[27]
Sr	Geological CRM	I	1 cm ³	$2 \text{ mol dm}^{-3} \text{ HNO}_3$	$0.2 \text{ mol dm}^{-3} \text{ HNO}_3$	> 90%	Mg, Al, Ca, Ti, Fe, Rb, Ba, REEs, Hg	Pb	MC ICP-MS	[22]
Sr	Geological and biological CRMs	I	3 cm^3	$2 \text{ mol dm}^{-3} \text{ HNO}_3$	$0.2 \text{ mol dm}^{-3} \text{HNO}_3$	84–105%	Mg, Al, Ca, Ti, Fe, Rb, Ba, REEs, Hg, Pb	n/a	MC ICP-MS	[23]
Zr	Geological CRMs	I	1 cm ³	7 mol dm ^{-3} HNO ₃	1 mol dm ⁻³ HF	Approx. 95%	Na, Mg, Al, K, Ca, Ti, V, Fe, Ni	Mo	TIMS	[29]
Zr	Geological CRMs	AG1-X8	2 cm^3	12 mol dm ⁻³ HNO ₃	3 mol dm^{-3} HNO ₃ +0.2 mol dm ⁻³ HF	n/a	Mg, Al, Ca, Ti, Fe, Mo	n/a	MC ICP-MS	[30]
Nd	Aerosol	Sr Spec	1 cm^3	$0.01 \text{ mol dm}^{-3} \text{ HCl}$	$0.0075 \text{ mol dm}^{-3} \text{ HNO}_3$	> 70%	La, Ce, Pr, Sm	n/a	MC ICP-MS	[31]
Nd	Geological CRMs	Sr Spec	800 mm ³	3.5 mol dm ⁻³ HNO ₃ +0.23 mol dm ⁻³ H ₃ BO ₃	0.4 mol dm ⁻³ HCl	Approx. 98.2%	Mg, Na, Al, K, Ca, Ti, Fe	n/a	TIMS	[32]
PN	Geological CRM	1	1 cm^3	2 mol dm^{-3} HNO ₃	2 mol dm ⁻³ HCl	> 91%	Mg, Al, Ca, Tï, Fe, Rb, Sr, Ba, Ce, Sm, Hg, Pb	n/a	MC ICP-MS	[22]
ΡN	Meteorites	TRU Spec, LN2	350 mg	3 mol dm ⁻³ HCl	$0.006 \text{ mol dm}^{-3} \text{ HCl}$	71–99%	La, Ce, Pr	n/a	TIMS	[33]
Nd	Geological CRMs	TRU Spec, LN2	950 mm ³	$0.1 \text{ mol dm}^{-3} \text{ HNO}_3$	0.01 mol dm ⁻³ HCl	%06-08	Ce, Pr	n/a	TIMS	[34]

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Table 1	(continued)									
Iso- topic system	Sample matrix	Add. resins	RV DGA	Load DGA	Elution DGA	Recovery	Removed (matrix) ele- ments	Co-elution	Detector	References
PN	Geological CRMs	. 1	1 cm ³	3 mol dm ⁻³ HNO ₃	2 mol dm ⁻³ HCl	%06 <	Na, Mg, Al, K, Ca, Ti, Fe, Ni, Cr, Ba, La, Ce, Pr, Sm, Eu, U	n/a	SMIT	[35]
Ηf	Aerosol	Sr Spec	1 cm ³	$6 \text{ mol dm}^{-3} \text{ HNO}_3$	1 mol dm ⁻³ HNO ₃ +0.2 mol dm ⁻³ HF	> 70%	n/a	n/a	MC ICP-MS	[31]
Ηf	Geological CRMs	Sr Spec	800 mm ³	3.5 mol dm^{-3} $\text{HNO}_3 + 0.23 \text{ mol dm}^{-3}$ H_3BO_3	3.5 mol dm ⁻³ HNO ₃ +1 mol dm ⁻³ HF	Approx. 97.3%	Mg, Na, Al, K, Ca, Ti, Fe	n/a	TIMS	[32]
Pd	Automotive exhaust emis- sions	AGW 50 X-12	0.2 cm ³	3 mol dm ⁻³ HCl	0.5 mol dm ⁻³ HCl	> 50%	Na, Mg, K, Ca, Ni, Cu, Rb, Sr, Zr, Hf, Ir, Hg, Pb	Zn, Ga, Ge, Y, Mo, Cd	ICP-MS (IDMS)	[36]
Pt	Automotive exhaust emis- sions	AGW 50 X-12	0.2 cm ³	4 mol dm ⁻³ HCl	0.04 mol dm ⁻³ HCl	> 70%	Na, Mg, K, Ca, Ni, Cu, Rb, Sr, Zr, Hf, Ir, Hg, Pb	Zn, Ga, Ge, Y, Mo, Cd	ICP-MS (IDMS)	[36]
Pb	Marine sedi- ment	I	1 cm ³	$2 \text{ mol dm}^{-3} \text{ HNO}_3$	5 mol dm ⁻³ HNO ₃	> 93%	Mg, Al, Ca, Ti, Fe, Rb, Sr, Ba, REEs, Hg	n/a	MC ICP-MS	[22]
ЪЪ	Geological and biological CRMs	I	3 cm ³	2 mol dm ⁻³ HNO ₃	5 mol dm ⁻³ HNO ₃	77–96%	Mg, Al, Ca, Ti, Fe, Rb, Sr, Ba, REEs, Hg	n/a	MC ICP-MS	[23]

target further stable isotope systems, which are retained by DGA Resin [1, 14, 15].

Calcium isotope abundance analysis is a growing field of research in biomedicine and can be used as a diagnostic tool for bone diseases like osteoporosis (e.g. [16–19]. Herein, the natural variation of Ca isotopic composition is determined with multicollector thermal ionization mass spectrometry (MC TIMS) or multicollector inductively coupled plasma mass spectrometry (MC ICP-MS). Both analytical techniques require a quantitative Ca/matrix separation for complex matrices such as blood, serum, or urine, to reduce the effects of spectral interferences (e.g. K, Mg, Al, Ti, Sr) and mass fractionation effects (e.g. by Na. Fe, P, Ba) [20]. Since MC TIMS was the preferred method for the reliable determination of Ca isotopic composition during the last decades, the offline Ca/matrix separation protocol from Feng et al. [21] was selected in the present study for further investigation. Feng et al. [21] employed the DGA Resin for Ca purification. Unwanted matrix elements, such as Na, Mg, Al, K, Ti, and Ba, are unretained using nitric acid as eluent [15].

The development and optimization of analyte/matrix separation procedures is a tedious and time-consuming task including the optimization of parameters such as resin volume (RV), eluent composition and acidity, eluent volume, and elution speed. In conventional offline optimization, small defined volumes (e.g. 0.5 cm³ or 1 cm³) are collected during incremental elution and analyzed for their elemental composition, leading to a elution profile (see e.g. [22, 23]). Online analysis of analyte/matrix separation provides a powerful alternative to batch separation. The advantages of online analysis of analyte/matrix separation lie in the continuous observation of a transient signal during the whole separation procedure. The elution profile provides information about the height, sharpness and broadening of the peaks, which enables the determination of the distinct elution volume required. Further, the mixing behavior and equilibration of the eluents can be monitored, as shown in this work. Pooled elution fractions of batch separation do not provide this information, which is crucial for loading and elution of the target element.

Inductively coupled plasma mass spectrometry (ICP-MS) is a preferable method for the measurement of eluted elements in online separation provided by high-performance liquid chromatography (HPLC) or ion chromatography (IC) [24], using micro-columns based on online flow injection [25, 26] or automated systems [22, 23]. The transient signals measured by the ICP-MS must be integrated for the quantification of analytes in the different elution steps to evaluate the separation efficiency and analyte recovery. Variations in the composition of the eluent can cause significant signal suppression, which is a challenge for accurate quantification.

Besides interfering matrix elements, Co and Mn are also unretained by DGA Resin using nitric acid as eluent [15]. These two elements do not influence Ca isotope amount ratio measurements. Therefore, the working hypothesis of the present work was to introduce Co and Mn continuously during the separation procedure to monitor for possible suppression and the mixing behavior of eluents and consequently enable a reliable quantification of the analyte and matrix elements in the transient signal. The aim of our study was to prove this concept and to demonstrate that online analysis (applying monitoring elements) is helpful to optimize offline separation protocols.

Results and discussion

Quantification of the Ca/matrix separation

The first quantification approach investigated for the transient signal (of the elements of interest) employed an external nine-point-calibration [normalized to ⁵⁹Co, prepared in dilute nitric acid (w = 3%)] from offline-analysis. The signals were integrated over a time period corresponding to the incremental time intervals of the transient signal. The transient elemental signal intensities were corrected for the blank by subtracting the average signal intensity of each corresponding fraction for nitric acid ($c = 3 \mod dm^{-3}$) and subboiled water at baseline. Each eluent was doped with Mn and Co (1 ng g^{-1}) to provide a continuous normalization signal. The transient signal was normalized to the signal intensity of ⁵⁹Co (internal standard). ⁵⁵Mn was monitored in parallel. The elution curve was divided into sample load, matrix wash, and Ca elution sections. The calculated total amounts of each of these three sections were summed up and compared to the total amount loaded (see section "Data reduction-internal normalization"). The resulting overall recoveries indicated significant overestimates in the final fraction for Ca as well as for the coeluting elements Sr, In, and Pb (recoveries of 333%, 206%, 177%, and 249%, respectively, see Supplementary Table 2). In contrast, Fe which is also partly co-eluting in the final fraction showed low recoveries (around 21%). It has to be noted, that Fe signals were close to LOD. The gradient from subboiled water to nitric acid ($c = 3 \mod dm^{-3}$) and accompanying suppression effects in the ICP as well as a potential delay of Co (also Mn) elution due to slight retaining by DGA Resin may have hampered this quantification strategy.

Consequently, a second quantification approach for the transient signal (of the elements of interest) was investigated. In this approach, suppression correction was applied prior to quantification using the external nine-point-calibration (mass vs. intensity, prepared in diluted nitric acid (w=3%)) from offline-analysis. Therefore, the suppression factors of Mn and Co were determined in the different eluent matrices first, since both elements were continuously present in

each elution step. Table 2 lists the suppression factors of ⁵⁵Mn as well as ⁵⁹Co in nitric acid ($c = 3 \mod dm^{-3}$), in nitric acid ($c = 3 \mod dm^{-3}$) including a multi-elemental test solution (β (Ca) = 975 ng g⁻¹, see Supplementary Table 2), and in subboiled water relative to diluted nitric acid (w = 2%) during offline ICP-MS measurements. The suppression factors of nitric acid ($c = 3 \mod dm^{-3}$) with and without multielement matrix overlapped within standard deviation (see Table 2). Suppression effects by multi-element matrix are considered negligible. To cope with a variation of suppression factors between elements, a mean suppression factor for each eluent (nitric acid ($c = 3 \mod dm^{-3}$), subboiled water) was calculated from ⁵⁵Mn and ⁵⁹Co intensities. Mean suppression factors of 1.57 ± 0.15 (SD, n = 14) for nitric acid $(c=3 \text{ mol } dm^{-3}; \text{ conditioning, sample load and matrix wash})$ and 0.15 ± 0.05 (SD, n = 18) for subboiled water (column cleaning and Ca elution), respectively, were determined.

After blank subtraction, the transient signal of each elution step was corrected using the mean suppression factors for the corresponding eluent (sample load with nitric acid ($c = 3 \mod dm^{-3}$), matrix wash with nitric acid $(c=3 \text{ mol } \text{dm}^{-3})$, Ca elution with subboiled water). Each complete elution step was corrected with the suppression factor (static model). Following the blank subtraction, a quantification of the transient signal (for each element) was performed using an external nine-point-calibration (mass vs. intensity, see section "Data reduction-dynamic correction of suppression factors"). Then, the calculated total amount of analyte and matrix elements in the sample load step, matrix wash step and Ca elution step were summed up and compared to the amount loaded (see section "Data reductioninternal normalization"). This procedure led to a significant overestimation of all elements which elute-at least partially-in subboiled water: Ca, Fe, Sr, In, and Pb (recoveries of 901%, 449%, 695%, 591%, and 757%, respectively).

Hence, a third quantification approach for the transient signal (of the elements of interest) was investigated. Herein, a concentration gradient from subboiled water to nitric acid $(c=3 \text{ mol } \text{dm}^{-3})$ was assumed when starting the conditioning. Further, a concentration gradient from nitric acid $(c=3 \text{ mol } \text{dm}^{-3})$ to subboiled water was assumed when starting the Ca elution due to on-column mixing. Hence, the transient signal was dynamically corrected using a linear gradient of the mean suppression factors from one eluent to the other (e.g. for Ca elution from nitric acid

 $(c=3 \text{ mol } \text{dm}^{-3}; 1.57 \pm 0.15 \text{ (SD, } n=14))$ to subboiled water $(0.15 \pm 0.05 \text{ (SD, } n=18))$. Consideration of the time required for the gradient of an entire resin volume (200 mm³, 24 s) only accounted for 30% of the overestimate observed for the second quantification approach. Therefore, a time of 141.4 s (280 data points, 1180 mm³=5.9 times RV) required for the gradient was estimated based on the elution profiles of Mn and Co reaching baseline intensities in nitric acid $(c=3 \text{ mol } \text{dm}^{-3};$ covering conditioning, sample load and matrix wash) and in subboiled water (covering Ca elution), respectively (see Figs. 1 and 2). Under the assumption of a linear gradient, dynamic suppression factors ranging from 0.15 ± 0.05 (subboiled water) to 1.57 ± 0.15 (nitric acid, $c=3 \text{ mol } \text{dm}^{-3}$) were applied for the 115.1 s of conditioning



Fig. 1 Elution profiles of ⁵⁵Mn and ⁵⁹Co intensities (*I*) during **a** conditioning (V=1 cm³ nitric acid (c=3 mol dm⁻³); 2 min), sample load (V=0.5 cm³ nitric acid (c=3 mol dm⁻³); 1 min) and matrix wash (V=1 cm³ nitric acid (c=3 mol dm⁻³); 2 min) and **b** Ca elution (V=5 cm³ subboiled water; 10 min) of the blank sample (nitric acid, c=3 mol dm⁻³)

 Table 2
 Suppression factor

 of ⁵⁵Mn and ⁵⁹Co in varying

 matrixes during ICP-MS

 measurements

Matrix	⁵⁵ Mn suppression factor	⁵⁹ Co suppression factor	Mean suppression factor
Nitric acid ($c = 3 \mod dm^{-3}$)	1.73 ± 0.04 (SD, $n=5$)	$1.44 \pm 0.05 \text{ (SD, } n = 5)$	1.57 ± 0.15 (SD, $n = 14$)
Matrix containing nitric acid ($c = 3 \mod \text{dm}^{-3}$)	$1.66 \pm 0.04 \text{ (SD, } n = 2)$	1.39 ± 0.05 (SD, $n=2$)	
Subboiled water	$0.16 \pm 0.06 \text{ (SD, } n = 9)$	0.13 ± 0.04 (SD, $n=9$)	$0.15 \pm 0.05 \text{ (SD, } n = 18)$



Fig. 2 Elution profiles of ⁵⁵Mn and ⁵⁹Co blank-corrected intensities (*I*) during **a** conditioning (V=1 cm³ nitric acid (c=3 mol dm⁻³); 2 min), sample load (V=0.5 cm³ nitric acid (c=3 mol dm⁻³); 1 min) and matrix wash (V=1 cm³ nitric acid (c=3 mol dm⁻³); 2 min) and **b** Ca elution (V=5 cm³ subboiled water; 10 min) of the multi-elemental test solution

and the first 26.3 s sample load. Dynamic suppression factors from 1.57 (nitric acid, $c = 3 \text{ mol } \text{dm}^{-3}$) to 0.15 (subboiled water) were applied for the first 141.4 s of Ca elution.

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Finally, the quantifications of the transient signals for each element were performed using external nine-point-calibration (mass vs. intensity, see section "Data reduction-dynamic correction of suppression factors"). Recoveries were calculated accordingly (see section "Data reduction-internal normalization"). In agreement with Feng et al. [21], Ca recovery was $102\% \pm 15\%$ in the Ca elution step using subboiled water (see Fig. 3 and Supplementary Table 3). Recoveries of Sr, In, and Pb were significantly reduced ($80\% \pm 18\%$, $72\% \pm 30\%$, and $110\% \pm 30\%$, respectively) and the recovery of Fe have increased ($73\% \pm 30\%$) (see Supplementary Table 3). The successful quantification (approach 2) of the online analysis of Ca/matrix separation using DGA resin, with a dynamic suppression correction proved the concept of using Mn and Co signal intensities as indicators for suppression effects.

Optimized Ca/matrix separation

Figure 3 shows the elution profile of Ca, as well as coeluting matrix elements causing spectral (K, Mg, Al, Ti, Sr) and non-spectral interferences (Na. Fe, Ba). Due to the properties of the DGA Resin, most of the matrix-based interferences (Na, Mg, Al, K, and Ti) are removed from Ca within the matrix wash. It should be noted, that a co-elution of Sr is not a concern for Ca isotopic analysis using MC TIMS [21] but a serious issue for Ca isotopic analysis using MC ICP-MS [20] because these elements cannot be separated during analysis in the mass spectrometer when ionized in the plasma source.

Following the results of the successful quantification during online matrix separation, an optimized Ca/matrix separation procedure is proposed in Table 3 based on the following considerations: (1) when re-using the DGA Resin, a precleaning step with diluted hydrochloric acid as applied in

Fig. 3 Elution profiles of several elements of interest during Ca/matrix separation of the multi-elemental test solution, shown as cumulative recoveries (*R*) per 50 mm³ (6 s) after quantification using approach 2, applying dynamic suppression correction and external calibration (mass vs. intensity). Additionally, raw intensities of Co (*I*) are shown as average per 50 mm³ (6 s)



Table 3Proposed elution stepsfor an optimized (online/offline)Ca/matrix separation usingDGA Resin

Step	Eluent	Flow rate/mm ³ min ⁻¹	Time/min	Eluent volume/cm ³
Pre-cleaning	$0.1 \text{ mol dm}^{-3} \text{HCl}$	500 ^a	4	2 (10 RV)
Cleaning	Subboiled water	500^{a}	4	2 (10 RV)
Conditioning	3 mol dm ⁻³ HNO ₃	500	3 ^b	1.5 (7.5 RV)
Sample load	$3 \text{ mol } \text{dm}^{-3} \text{HNO}_3$	500	1	0.5 (2.5 RV)
Matrix wash	$3 \text{ mol } \text{dm}^{-3} \text{HNO}_3$	500	2	1 (5RV)
Ca elution	Subboiled water	500	1.5	0.75 (3.75 RV)

^aCould be probably performed at a higher flow rate (1000 mm³ min⁻¹)

^bA column with 200 mm³ resin volume needs at least six times RV for the concentration gradient between nitric acid ($c = 3 \text{ mol dm}^{-3}$) and subboiled water

previous studies [22, 23, 27] should be considered to remove remaining REEs (e.g. Y and U, see Supplementary Table 3) from the DGA Resin. (2) Since the time required for the concentration gradient from subboiled water to nitric acid $(c=3 \text{ mol } \text{dm}^{-3})$ exceeds the time required for the conditioning step and interferes with the accurate loading acidity of the nitric acid ($c = 3 \mod \text{dm}^{-3}$), an extension of the conditioning step to 1.5 cm³ (3 min at 500 mm³ min⁻¹ pump speed) should be considered. (3) Based on the observed elution profile, the Ca elution volume can be reduced to 750 mm³ (1.5 min at 500 mm³ pump speed, 3.75 RV), while maintaining a recovery of 102% (see Supplementary Table 3: Ca elution (750 mm³), Fig. 3) (Note: Total mass fraction of Ca in the sample load was 721 ng \pm 72 ng, the collected mass fraction of Ca in the Ca elution using 5 cm^3 was 741 ng \pm 111 ng, the collected mass fraction of Ca in Ca elution using 750 mm³ was 740 ng \pm 111 ng).

Conclusion

The quantification of a transient signal during an online separation is challenging because of the dynamic gradient of eluents in an online analysis of analyte/matrix separation following on-column mixing. A modified approach using dynamic suppression correction, based on indicator elements (Mn, Co) that were unretained by the DGA resin, proved to be a robust approach for accurate quantification as a prerequisite for the optimization of analyte/matrix separation procedures.

The online analysis of Ca/matrix separation using DGA Resin enabled insights in the mixing behavior of the eluents (subboiled water, nitric acid ($c = 3 \mod dm^{-3}$)) which was revealed by the elution behavior of indicator elements Mn and Co. Based on the results during online separation, an optimized Ca/matrix separation procedure using DGA Resin is proposed for subsequent Ca isotope ratio analysis by TIMS analysis.

The approach is advantageous for separation procedures with alternating elution solutions and can be applied to any system once suitable monitoring elements (i.e. will not be retained on the resin) are found. This can dramatically increase the efficiency of the method development and allow for rapid checks of ion exchange methods for various different matrices.

Experimental

Type I reagent-grade water (18 M Ω cm) (F+L GmbH, Vienna, Austria) was further purified by sub-boiling distillation (Milestone-MLS GmbH, Leutkirch, Germany). Analytical reagent-grade nitric acid (w=65%; Merck-Millipore, Darmstadt, Germany) was purified by double sub-boiling using a Savillex DST-1000 sub-boiling distillation system (AHF Analysentechnik, Tübingen, Germany). Polyethylene flasks, tubes and pipette tips (VWR International, Radnor, USA) were pre-cleaned in a two-stage washing procedure using diluted nitric acid (w=10% and w=1%, respectively).

Single-element standards were purchased from Inorganic Ventures (Christiansburg, USA: Na, Mg, Al, K, Ca, Fe, Mn, Co, Cu, Zn, Sr, Y, Cd, In, Ba, Pb, and U), Merck (Ti, Cr, Ni, and Mo) and VWR International (Radnor, USA: Tl, Rb (Aristar)).

Two grams of the unbranched DGA Resin (TrisKem International) with a particle size of 50 µm–100 µm was soaked in diluted ethanol ($w \approx 30\%$, Merck) and shaken for 1 h. For the online analysis of Ca/matrix separation, an empty PFA column with a resin volume of 200 mm³ (part. no. CF-0200, ESI) was utilized and self-packed with the soaked DGA Resin following the standard procedure [22].

A multi-elemental test solution containing 10 ng g⁻¹ of Na, Mg, Al, K, Ti, Fe, Cr, Ni, Cu, Zn, Rb, Sr, Y, Mo, Cd, In, Ba, Tl, Pb, and U and 1000 ng g⁻¹ of Ca was gravimetrically prepared in nitric acid ($c=3 \mod \text{dm}^{-3}$) including 1 ng g⁻¹ of Mn and Co spike. The nitric acid ($c=3 \mod \text{dm}^{-3}$) as eluent was prepared gravimetrically and spiked with 1 ng g⁻¹ of Mn and Co. The subboiled water as eluent was spiked with 1 ng g^{-1} of Mn and Co.

For corrections of suppression effects and for the quantification of recoveries, a nine-point-calibration with the same multi-elemental mixture as the test solution was gravimetrically prepared in diluted nitric acid (w = 2%) covering the range of 0.05 ng g⁻¹ and 150 ng g⁻¹. All calibration standards were spiked with 1 ng g⁻¹ of Mn and Co. Furthermore, the performance of the multi-elemental analysis was monitored with in-house quality control solution prepared in nitric acid ($c = 3 \mod \text{dm}^{-3}$), diluted nitric acid (w = 2%) and subboiled water. The quality control solutions were spiked with 1 ng g⁻¹ of Mn and Co.

Offline multi-elemental analysis

To correct suppression effects of the different eluents and for quantification of recoveries, multi-elemental composition of the different eluents and the multi-elemental test solution were determined in standard liquid set-up using the ICP-QMS (NexION 2000B, PerkinElmer) coupled to an ESI SC-2 DX FAST autosampler. General instrumental settings are described in Supplementary Table 1.

Online analysis of Ca/matrix separation

Multi-elemental online analysis of Ca/matrix separation was carried out by connecting an autosampler probe (i.d. 1 µm, ESI, Omaha, USA) using a 10-cm-long PFA Capillary Extension (i.d. 1 µm, ESI) to a pump tubing (i.d. 1.1 µm, red/ red, PerkinElmer, Ontario, Canada), which was clamped into a peristaltic pump (part. no. 3642, Ismatec, Wertheim, Germany). The sample was taken up and transported at a flow rate of 500 mm³ min⁻¹ via a PFA capillary (i.d. 1 µm and ¹/₄-28 nut, ESI) into the PFA column (ESI). The column was filled with 200 mm³ unbranched DGA Resin (part. no. DN-B25-S, TrisKem International). Sampling was performed manually. The column run-off was directly connected to an ICP-QMS (NexION 2000B, PerkinElmer) equipped with a sea-spray glass nebulizer (max. uptake 2 cm³ min⁻¹, PerkinElmer). The multi-element composition was monitored online in a time-resolved sequence using the ICP-QMS (NexION 2000B, PerkinElmer). General instrumental settings are described in Supplementary Table 1.

Procedure of the Ca/matrix separation

Ca/matrix separation was performed following the separation procedure of Feng et al. [21] using unbranched DGA Resin (TrisKem International): A conditioning step using nitric acid ($c = 3 \mod \text{dm}^{-3}$) was followed by a sample loading step using nitric acid ($c = 3 \mod \text{dm}^{-3}$), and a matrix wash step using nitric acid ($c = 3 \mod \text{dm}^{-3}$). Ca was finally eluted using subboiled water. The elution volumes were adapted according to the resin volume used. The corresponding elution steps and parameters are shown in Table 4.

Transient multi-elemental signal was recorded by online analysis for the Ca/matrix separation (transient signal) of a blank sample containing nitric acid ($c=3 \mod dm^{-3}$, 500 mm³). Further, the transient multi-elemental signal was recorded by online analysis for the Ca/matrix separation (transient signal) of a multi-elemental test solution prepared in nitric acid ($c=3 \mod dm^{-3}$, 500 mm³). The multielemental test solution contained approx. 750 ng total Ca as major matrix element and < 10 ng of Na, Mg, Al, K, Ti, Cr, Fe, Ni, Cu, Zn, Rb, Sr, Mo, Cd, In, Ba, Tl, Pb, and U as minor elements.

Data reduction-internal normalization

Elution steps were identified and defined by ⁵⁵Mn and ⁵⁹Co raw intensities. Blank correction of each elution step (loading, matrix wash, Ca elution) was performed by subtracting the average intensities of the corresponding blank elution step. In the case of loading with nitric acid ($c = 3 \mod \text{cm}^{-3}$) and Ca elution with subboiled water, only data points recorded after the signal reached baseline (141.1 s, see "Results and discussion") were used.

After blank subtraction, the intensities of the transient signals (elements of interest) were normalized point-by-point with the corresponding ⁵⁹Co signal intensity, applied as an internal standard. Point-by-point mass fractions

Table 4	Elution steps for online
analysis	of Ca/matrix separation

Step	Eluent	Flow rate/ mm ³ min ⁻¹	Time/min	Eluent volume/cm ³
Cleaning	Subboiled water	500	4 ^a	2.0 ^a (10 RV)
Conditioning	3 mol dm ⁻³ HNO ₃	500	2	1.0 (5 RV)
Sample load	3 mol dm ⁻³ HNO ₃	500	1	0.5 (2.5 RV)
Matrix wash	3 mol dm ⁻³ HNO ₃	500	2	1.0 (5 RV)
Ca elution	Subboiled water	500	10	5.0 (25 RV)

^aThe cleaning procedure was extended to 10 min using 5 cm³ of subboiled water for the blank sample

(ng g^{-1}) during the transient signal were quantified using the nine-point-calibration of the offline multi-elemental measurement, which was also normalized to ⁵⁹Co signal intensity. Cumulative mass fractions for all elements under investigation were calculated as the integral over each elution step. To evaluate the recovery, the overall integrated mass fractions were calculated as the sum of the sample load, the matrix wash and the Ca elution (Note: Only an elution time of 1.5 min corresponding to an elution volume of 750 mm³ for Ca elution was used) and compared to the mass fractions of the loaded sample.

The Limits of Detection and Quantification (LOD, LOQ) was calculated according to DIN ISO 11843-2 [37]. For each elution step, the LOD was defined as $3 \times SD$ of the corresponding baseline blank signal. (Note: In the evaluation of the multi-elemental test solution, all values below LOD were set to zero.) Mass fractions, recoveries, LODs and LOQs, including expanded relative uncertainties (U_{rel} (k=2)) are summarized in Supplementary Table 2.

Data reduction-dynamic correction of suppression factors

Elution steps were identified and defined by ⁵⁵Mn and ⁵⁹Co raw intensities. Blank correction of each elution step (loading, matrix wash, Ca elution) was performed by subtracting the average intensities of the corresponding blank elution step. In the case of loading with nitric acid ($c = 3 \mod \text{dm}^{-3}$) and Ca elution with subboiled water, only data points after reaching the baseline (141.1 s, see "Results and discussion") were used.

After blank subtraction, the intensities of the transient signals (elements of interest) were dynamically corrected for suppression effects in the sample loading and the Ca elution step. Therefore, a linear gradient of the mean suppression factors from one eluent to the other was applied. The time required for the gradient was estimated based on the elution profiles of ⁵⁵Mn and ⁵⁹Co signal intensities reaching baseline intensities (see "Results and discussion"). To enable a quantification without internal normalization, a nine-point-calibration based on total mass (accounted by sample volume (500 mm³) and the density of diluted nitric acid (w = 3%)) plotted against signal intensity was a set-up from the offline multi-elemental measurement. Point-by-point total masses (ng) during the transient signal were quantified using the slope of this nine-point-calibration (mass vs. intensity). Cumulative masses for all elements under investigation were calculated as the integral over each elution step. To evaluate the recovery, the overall integrated total masses were calculated as the sum of the sample load, the matrix wash and the Ca elution (Note: Only an elution time of 1.5 min corresponding to an elution volume of 750 mm³ for Ca elution was used) and compared to the total mass of the loaded

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sample. The mass of the loaded sample was quantified by the nine-point-calibration (accounted by sample volume (500 mm³) of the test solution and density of the nitric acid ($c = 3 \mod dm^{-3}$)). The LOD and LOQ for the total mass were defined as $3 \times SD$ and $10 \times SD$, respectively, of the baseline blank signal from the corresponding eluent. Total masses, recoveries, LODs and LOQs, including expanded relative uncertainties (U_{rel} (k=2)) are summarized in Supplementary Table 3.

Uncertainty calculations

The total combined uncertainty for each elemental total mass $(m_{\rm spl})$ was calculated using a simplified Kragten approach [38] using Eq. (1) as the model equation. The precision of the measured raw intensity of the sample $(I_{\rm spl})$, the estimated correction factor for the suppression effect $(f(I)_{\rm sup})$ and the slope of the calibration curve (k) were identified as the main contributors to the uncertainty.

$$m_{\rm spl} = \frac{I_{\rm spl} - I_{\rm blk}}{f(I)_{\rm sup}} \times \frac{1}{k}$$
(1)

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00706-021-02754-2.

Acknowledgements This work was funded by the Chemical Monthly fellowship (2018) of the Austrian Academy of Sciences (ÖAW) (grant number: Stipendium der Monatshefte für Chemie).

Funding Open access funding provided by Montanuniversität Leoben.

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References

- 1. Horwitz EP, McAlister DR, Bond AH, Barrans RE (2005) Solvent Extr Ion Exch 23:219
- Eichrom Technologies (2020) DGA resins. Eichrom Technologies, LLC. https://www.eichrom.com/eichrom/products/dga-resins/. Accessed 16 Jun 2020
- Bertelsen ER, Jackson JA, Shafer JC (2020) Solvent Extr Ion Exch 38:251
- McLain DR, Tsai Y, Graczyk DG, Canaday JL, Steeb JL (2018) J Radioanal Nucl Chem 317:1439

- Shao Y, Yang G, Tazoe H, Ma L, Yamada M, Xu D (2018) J Environ Radioact 192:321
- Aluicio-Sarduy E, Hernandez R, Valdovinos HF, Kutyreff CJ, Ellison PA, Barnhart TE, Nickles RJ, Engle JW (2018) Appl Radiat Isot 142:28
- Maxwell SL, Culligan BK, Hutchison JB, Utsey RC, McAlister DR (2013) J Radioanal Nucl Chem 298:1977
- 8. McAlister DR, Horwitz EP (2018) Appl Radiat Isot 140:18
- Maxwell SL, Culligan BA, Jones VD, Nichols ST, Noyes GW (2011) J Radioanal Nucl Chem 287:223
- Grate JW, O'Hara MJ, Farawila AF, Douglas M, Haney MM, Petersen SL, Maiti TC, Aardahl CL (2011) Anal Chem 83:9086
- Dolique I, Kabai E, Schuster M (2019) J Radioanal Nucl Chem 322:1423
- 12. Wang Z, Zheng J, Tagami K, Uchida S (2017) J Radioanal Nucl Chem 312:151
- Kazi Z, Guérin N, Christl M, Totland M, Gagné A, Burrell S (2019) J Radioanal Nucl Chem 321:227
- Sasaki Y, Zhu Z-X, Sugo Y, Kimura T (2007) J Nucl Sci Technol 44:405
- 15. Pourmand A, Dauphas N (2010) Talanta 81:741
- Morgan JLL, Skulan JL, Gordon GW, Romaniello SJ, Smith SM, Anbar AD (2012) Proc Natl Acad Sci 109:9989
- Heuser A, Eisenhauer A, Scholz-Ahrens KE, Schrezenmeir J (2016) Isot Environ Health Stud 52:633
- Tanaka Y-K, Mikuni-Takagaki Y, Hidaka K, Wada-Takahashi S, Kawamata R, Hirata T (2019) Anal Sci 35:793
- Eisenhauer A, Müller M, Heuser A, Kolevica A, Glüer CC, Both M, Laue C, Hehn UV, Kloth S, Shroff R, Schrezenmeir J (2019) Bone Rep 10:100200
- 20. Feng L, Zhou L, Yang L, Zhang W, Wang Q, Shuoyun T, Hu Z (2018) J Anal At Spectrom 33:413
- 21. Feng L-P, Zhou L, Yang L, DePaolo DJ, Tong S-Y, Liu Y-S, Owens TL, Gao S (2017) Geostand Geoanal Res 41:93
- Retzmann A, Zimmermann T, Pröfrock D, Prohaska T, Irrgeher J (2017) Anal Bioanal Chem 409:5463

- 23. Zimmermann T, Retzmann A, Schober M, Pröfrock D, Prohaska T, Irrgeher J (2019) Spectrochim Acta Part B 151:54
- 24. Latkoczy C, Prohaska T, Watkins M, Teschler-Nicola M, Stingeder G (2001) J Anal At Spectrom 16:806
- 25. Galler P, Limbeck A, Boulyga SF, Stingeder G, Hirata T, Prohaska T (2007) Anal Chem 79:5023
- Galler P, Limbeck A, Uveges M, Prohaska T (2008) J Anal At Spectrom 23:1388
- Romaniello SJ, Field MP, Smith HB, Gordon GW, Kim MH, Anbar AD (2015) J Anal At Spectrom 30:1906
- Bao Z, Zong C, Chen K, Lv N, Yuan H (2020) Int J Mass Spectrom 448:116268
- Feng L, Hu W, Jiao Y, Zhou L, Zhang W, Hu Z, Liu Y (2020) J Anal At Spectrom 35:736
- Inglis EC, Creech JB, Deng Z, Moynier F (2018) Chem Geol 493:544
- 31. Pourmand A, Prospero JM, Sharifi A (2014) Geology 42:675
- 32. Li C-F, Wang X-C, Guo J-H, Chu Z-Y, Feng L-J (2016) J Anal At Spectrom 31:1150
- 33. Pin C, Gannoun A (2019) J Anal At Spectrom 34:2136
- 34. Pin C, Gannoun A (2019) J Anal At Spectrom 34:310
- Chu Z-Y, Wang M-J, Li C-F, Yang Y-H, Xu J-J, Wang W, Guo J-H (2019) J Anal At Spectrom 34:2053
- Vogl J, Meyer C, Koenig M, Becker D, Noordmann J, Rienitz O, Mamakos A, Riccobono F (2015) J Anal At Spectrom 30:479
- DIN ISO 11843-2:2006-06, Erkennungsfähigkeit—Teil 2: Verfahren im Fall der linearen Kalibrierung (ISO 11843-2:2000; Text Deutsch, Englisch) (2006) Beuth Verlag GmbH
- 38. Kragten J (1994) Analyst 119:2161

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