



Human bocavirus infection in Belgian children with respiratory tract disease

Vanessa Verbeke¹ · Marijke Reynders² · Katelijne Floré² · Wouter Vandewal³ · Sara Debulpaep⁴ · Kate Sauer² · Frederik Cardoen³ · Elizaveta Padalko¹

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Abstract

Human bocavirus (HBoV) has been detected primarily in children with acute lower respiratory tract disease (LRTD), but its occurrence, clinical profile, and role as a causative agent of RTD are not clear. The aim of this study was to investigate the prevalence and the potential clinical relevance of HBoV. Using molecular tests, we tested 1352 nasopharyngeal samples obtained between October 1, 2017 and April 30, 2018 from children up to the age of 16 with RTD for the presence of HBoV DNA and 20 other respiratory pathogens at three different hospitals in Belgium. HBoV was detected in 77 children with a median age of 10.6 months. Consecutive samples were available for 15 HBoV-positive children and showed persistent HBoV positivity in four of them. Mono-infection was observed in six infants. Four of them were born prematurely and were infected during hospitalization at the neonatal intensive care unit (NICU). Only one of these six mono-infected children was diagnosed with recurrent wheezing due to HBoV. This child was carried to term and had a high viral load. Coinfections, most frequently with rhinovirus (52.1%) and adenovirus (49.3%), were observed in 72 patients. In seventeen of them in which HBoV was present at high viral load or higher viral load than its copathogens, bronchi(oli)tis ($n = 8$), recurrent wheezing ($n = 8$) or episodic wheezing ($n = 1$) were diagnosed. Our results suggest that HBoV infection at high viral load in infants is associated with wheezing ($P = 0.013$, Cramer's $V = 0.613$).

Abbreviations

BPD	Bronchopulmonary dysplasia
CLD	Chronic lung disease
CPAP	Continuous positive airway pressure
Cq	Quantitation cycle

CRP	C-reactive protein
EV	Enterovirus
GI	Gastrointestinal
HAdV	Human adenoviruses
HBoV	Human bocavirus
HCoV-229E	Human coronavirus 229E
HCoV-HKU1	Human coronavirus HKU1
HCoV-NL63	Human coronavirus NL63
HCoV-OC43	Human coronavirus OC43
hMPV	Human metapneumovirus

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✉ Elizaveta Padalko
Elizaveta.Padalko@uzgent.be

Vanessa Verbeke
vanessa.verbeke@uzgent.be

Marijke Reynders
marijke.reynders@azsintjan.be

Kateljine Floré
katelijne.flore@azsintjan.be

Wouter Vandewal
wouter.vandewal@stlucas.be

Sara Debulpaep
sara.debulpaep@uzgent.be

Kate Sauer
kate.sauer@azsintjan.be

Frederik Cardoen
frederik.cardoen@stlucas.be

- ¹ Department of Medical Microbiology, Ghent University Hospital, Corneel Heymanslaan 10, 9000 Ghent, Belgium
- ² AZ Sint-Jan Bruges, Ruddershove 10, 8000 Brugge, Belgium
- ³ AZ Sint-Lucas Bruges, Sint-Lucaslaan 29, 8310 Brugge, Belgium
- ⁴ Department of Pediatrics, Ghent University Hospital, Ghent, Belgium

HRV	Human rhinoviruses
HPeV	Human parechovirus
HPIV	Human parainfluenza viruses
HRSV	Human respiratory syncytial virus
IAV	Human influenza A virus
IBV	Human influenza B virus
LRTD	Lower respiratory tract disease
MP	<i>Mycoplasma pneumoniae</i>
ND	Not detected
NICU	Neonatal intensive care unit
NPS	Nasopharyngeal samples
PICU	Pediatric intensive care unit
RTD	Respiratory tract disease
RTI	Respiratory tract infections
RT-PCR	Real-time PCR
SARI	Severe acute respiratory infection
UH	University Hospital
URTD	Upper respiratory tract disease
WBC	White blood cell count

Introduction

Respiratory tract diseases (RTDs) are a leading reason for morbidity in young children and are caused by a broad spectrum of microbial agents. Viruses account for the largest number of respiratory tract infections (RTIs). The so-called respiratory viruses include influenza A and B viruses (IAV, IBV), human parainfluenza viruses (HPIVs), human respiratory syncytial virus (HRSV), human adenoviruses (HAdVs), human rhinoviruses (HRVs), human coronaviruses (HCoV), enteroviruses (EVs) and human parechoviruses (HPeVs). In the 21st century, using large-scale molecular virus screening, several novel viruses have been discovered in patients with respiratory infections. These viruses include human metapneumovirus (hMPV), polyomaviruses KI and WU, several coronaviruses (SARS-CoV, HCoV-NL63, HCoV-HKU1, MERS-CoV) and human bocavirus (HBoV) as described by Allander et al. in 2005 [1–3]. The DNA virus HBoV is a member of the family *Parvoviridae*, genus *Bocaparvovirus*. HBoV is classified into genotypes 1 through 4. HBoV1 is predominantly found in respiratory tract secretions from children with RTD, and HBoV2–4 are found mainly in stool samples from patients with gastroenteritis [3, 4]. HBoV is predominantly present in winter and spring [5]. The average prevalence of HBoV in respiratory tract samples ranges from 1.0% to 56.8%, depending on the country. The worldwide estimate for the total prevalence of HBoV in respiratory infections is 6.3% [6]. HBoV has been reported worldwide in all age groups; however, it has mainly been detected in children who presented at the hospital with RTD. A high HBoV viral load could be an etiologic agent for severe LRTI, and these patients may develop bronchitis and pneumonia with

fever, cough and peribronchial infiltrates detected on a chest X-ray [7, 8]. Low viral loads in coinfection indicate more asymptomatic shedding [9]. Furthermore, HBoV has been suggested as a cause of pediatric gastrointestinal (GI) infection [10, 11] and might even have a causal role in encephalitis [12–15]. However, it has also been found in children with mild infections [16] and in asymptomatic ones, and therefore, the pathogenic role of HBoV is still under discussion [8, 17]. Classically, Koch's postulates have been used to establish a causal relationship between viruses and disease [18]. As there is no animal model so far for HBoV, proving its clinical relevance is challenging. However, HBoV can replicate in human airway epithelium cultures [6, 19], and Deng et al. showed that HBoV1 induces damage to the airway epithelium (loss of cilia, disruption of the tight junction barrier, and a significant decrease in transepithelial electrical resistance) [20, 21]. Furthermore, serological diagnosis of HBoV has recently confirmed significant increases in IgG antibodies in children with pneumonia. These results support the idea that it is a true pathogen in RTI in children [22, 23]. Prolonged viral shedding has been described, about 2.5 months in outpatients and about 4.5 months to 1 year in hospitalized children, which probably explains why HBoV is detected in asymptomatic cases. This prolonged shedding may also explain why the rate of coinfection with other viruses is so high, ranging from 75% to 85% [24].

In this study, we retrospectively analyzed data for 1352 nasopharyngeal samples (NPSs, aspirates and swabs) that were molecularly tested for the presence of HBoV DNA. Our aim was to determine the prevalence of HBoV in children up to 16 years of age who presented at the hospital with RTD and to describe the clinical features of the infected children.

Materials and methods

Specimen collection

We retrospectively analyzed the data of the respiratory molecular screening of 1352 NPSs obtained between October 1, 2017 and April 30, 2018 from children up to the age of 16 who presented with RTD at three different hospitals in Belgium: Ghent University Hospital, AZ Sint-Jan, and AZ Sint-Lucas of Bruges. Since we wanted to describe the whole spectrum of disease attributed to HBoV, we choose to include all patients and use no exclusion criteria. Because of the retrospective nature of this study, there was no asymptomatic control group included. Three hundred two NPSs were collected and analyzed in the Ghent University Hospital, and 572 and 478 samples were collected at AZ Sint-Jan and AZ Sint-Lucas of Bruges, respectively, and analyzed at AZ Sint-Jan Hospital.

Viral DNA extraction and real-time PCR amplification

The NPSs were tested in a routine setting for the presence of HBoV 1-4 DNA among 20 other infectious agents (IAV and IBV; human coronaviruses NL63 [HCoV-NL63], 229E [HCoV-229E], OC43 [HCoV-OC43] and HKU1 [HCoV-HKU1]; HPIV 1, 2, 3 and 4; hMPV A and B, HRV, HRSV A and B, HAdV, EV, HPeV, and *Mycoplasma pneumoniae* [MP]) using real-time PCR (RT-PCR) amplification by TaqMan® technology. The AZ Sint-Jan Hospital also tested for five viruses, 10 bacteria and two fungi. In the Ghent University Hospital, nucleic acid from NPSs was extracted using a NucliSENS EasyMAG (BioMérieux, France) automated extractor followed by a commercial multiplex RT-PCR assay (FTD Respiratory Pathogens 21, Fast Track Diagnostics, Luxembourg) according to manufacturer's instructions. In the AZ Sint-Jan Hospital, nucleic acid extraction on a QIASymphony (QIAGEN, Spain) automated extraction system according to the manufacturer's instructions was followed by a singleplex RT-PCR using an in-house customized TaqMan® Array Card (TAC) (v13.0 premarket version Cambridge-Bruges) [25]. Because a universal primer for HBoV 1-4 was used, distinction between the different genotypes of HBoV was not possible, which is a limitation of our study. The detailed protocol for DNA extraction and RT-PCR amplification is available online as Online Resource 1.

Clinical interpretation of the HBoV PCR test results

HBoV results were categorized in three groups based on the quantitation cycle value (Cq value): $Cq \leq 22$, strongly positive (high viral load); $22 < Cq \leq 30$, positive; $30 < Cq \leq 40$, weakly positive. We were especially interested in patients with HBoV in monoinfection or with HBoV present at a low Cq value (≤ 22) or a lower category of Cq value than its copathogens in cases of coinfection, since the fundamentals of RT-PCR allow us to understand that lower Cq values represent a higher viral load. The PCR results were analyzed together with the white blood cell count (WBC) and differentiation, the value of C-reactive protein (CRP), the clinical characteristics of the patient (URTD vs. LRTD, GI disease, fever), the duration of hospitalization, the need for oxygen therapy, and results of chest X-ray, if available. A history of underlying conditions such as cardiac disease, prematurity, chronic pulmonary disease or previous episodes of wheezing were studied if present. Bronchiolitis or wheezing in young children is a very common clinical problem, and definitions are often confusing. In this study, we define bronchiolitis as a constellation of clinical signs and symptoms occurring in children younger than 2 years, including a viral URT prodrome followed by increased respiratory effort and wheezing [26, 27]. We diagnosed children younger than

two years of age with bronchiolitis if their history showed no previous episodes of wheezing and with "recurrent wheezing" if there had been previous episodes. Children older than two years of age with previous episodes of wheezing were diagnosed with episodic wheezing. Bronchopulmonary dysplasia (BPD) was defined as oxygen dependency beyond 28 days of life [28].

Statistical analysis

Statistical analysis was carried out using IBM SPSS Statistics 25. Categorical variables were compared using the chi-square test, and the continuous variables were compared using an independent *t*-test or one-way ANOVA. *P*-values less than 0.05 were considered significant.

Results

Detection of HBoV

Between October 1, 2017 and April 30, 2018, 1352 NPSs of 1153 infants and children with RTD were received for diagnostic evaluation. HBoV was detected in 87 (6.4%) samples from 77 children. The dataset for these positive patients is available as Online Resource 2. Out of the 87 samples, 13 (14.9%) were monoinfections and 74 (85.1%) were coinfections with one or more other viruses. The 13 single detections belonged to six infants, the 74 samples with coinfections belonged to 72 children. One child had multiple NPSs that were positive for HBoV in mono- and coinfection, so a total of 77 different children were included in this study (Fig. 1). Coinfections were most frequently seen with HRV (38 samples) and HAdV (36 samples). Coinfection of HBoV with bacteria or fungi was not seen.

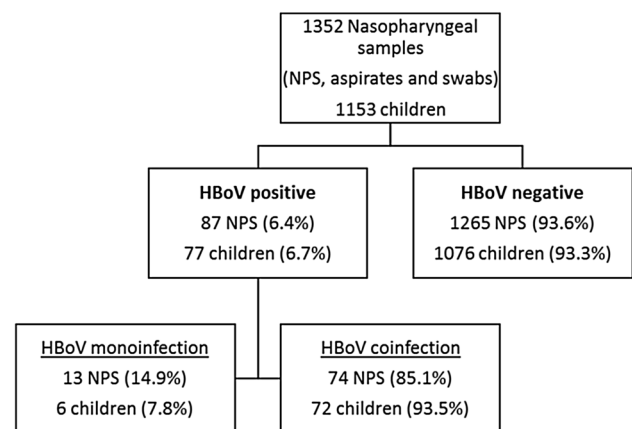


Fig. 1 Flowchart of positivity of nasopharyngeal samples (NPS) for human bocavirus (HBoV) in monoinfection or coinfection in our pediatric study population

The HBoV viral load, expressed as a Cq-value, in children with a mono-infection (mean Cq, 26; range, 10–34) was not different from that in children with a co-infection (mean Cq, 28; range, 13–35) ($P = 0.445$). A high viral load (Cq \leq 22) was observed in four of the 13 (30.8%) samples expressing HBoV alone and in 16 of the 74 (21.6%) samples with co-infection. Patients with a higher viral load were slightly younger, although this was not statistically significant ($P = 0.051$). The mean age of patients in the first (Cq \leq 22), second (22 < Cq \leq 30) and third category (30 < Cq \leq 40) was 11.2, 11.2 and 16.8 months, respectively.

Persistent HBoV positivity

For 15 HBoV-positive children (4 mono-infections and 11 co-infections) various consecutive samples were taken because of a clinical need for them over a period of several weeks to several months (Table 1 and Fig. 2). Four children showed positivity for HBoV in consecutive samples. One child showed persistent HBoV positivity in seven separate determinations spread equally over a period of 97 days. The second child showed HBoV shedding in three determinations over a period of 35 days. The third child was HBoV positive two times within a period of 2 weeks; whereas in the last child, HBoV DNA could be detected in samples collected in October 2017 and in March 2018.

Clinical features associated with HBoV detection

The median age of the HBoV-positive children was 10.6 months (mean age, 13.2 months; range, 19 days–5.4 years). All but one of the children were younger than 3 years, and 62.8% of the children were younger than 12 months. The majority of the children (98.6%) were hospitalized in one of the three hospitals during the course of infection.

Mono-infection

The clinical characteristics of the six patients with respiratory episodes associated with single HBoV detection were analyzed in detail, and the results are shown in Table 2. The children had a median age of 2.6 months (mean age, 3.9 months; range, 19 days–11.5 months).

Nosocomial mono-infection

An important finding was that four out of the six children who were born prematurely at a gestational age of 25, 26, 28 and 33 weeks were still at the NICU of the Ghent University Hospital at the time of infection. Three of them got infected in the same week, so transmission probably occurred via the hospital staff. These premature patients were all treated in incubators and had had no previous contact with other

children. The fact that they were in a protected area is probably the reason there was no co-infection. The patients born at 25, 26 and 28 weeks all developed BPD as a sequela of their premature birth. All three were intubated at birth, received surfactant, and needed long-term ventilator support. Two of them developed chronic oxygen dependency and were discharged with home oxygen therapy. The child born at a gestational age of 25 weeks needed conventional ventilation for 17 days, followed by 10 days of continuous positive airway pressure (CPAP) and long-term oxygen treatment. His respiratory condition was aggravated at an age of 70 days by infection with HBoV (Cq value, 10) with a need for repeated CPAP support and a higher oxygen supply. A chest X-ray at this time point showed peribronchial thickening and atelectasis. This child showed persistent HBoV PCR positivity in seven separate determinations (6 in mono-infection, 1 in co-infection). The other three premature infants expressed HBoV at higher Cq values (30–34) and had only mild deterioration of their respiratory condition. A child born at a gestational age of 26 weeks had severe BPD with a need for invasive ventilation (conventional and high-frequency oscillation) for 33 days followed by 18 days of CPAP. During the infection with HBoV, there was a higher oxygen need. A child born at a gestational age of 28 weeks needed conventional ventilation for 2 days followed by 3 days of oxygen supply by nasal canula. At day 19, infection with HBoV resulted in lower saturation and development of a need for extra oxygen until day 63. A child born at a gestational age of 33 weeks was transferred from another hospital to the NICU because of necrotizing enterocolitis. During her stay, there was an unexplained mild decline in saturation. A respiratory panel showed the presence of HBoV.

Community-acquired mono-infection

The fifth child was born at term and had no important pre-existing medical condition. This infant had been exclusively breastfed since birth. At the age of 4 months, she had a first episode of wheezing, and bronchiolitis was diagnosed. Further she had repetitive mild RTD, a finding that is very common in infants attending daycare. At an age of 4.9, months she presented at the emergency department with severe respiratory distress and a need for non-invasive respiratory support (high-flow O₂ nasal canula). She was admitted to the pediatric intensive care unit (PICU) for three days. Diagnosis of recurrent wheezing, caused by HBoV (Cq value, 16) was made. The sixth child had no LRTD and suffered from an otitis media. No pneumonia or GI disorders were observed in these six mono-infected children. There was no significant elevation of the white blood cell count or CRP value.

Table 1 Overview of results for 15 human bocavirus (HBov)-positive children who were tested multiple times by PCR for respiratory pathogens during the 2017–2018 season

Patient	Pathogen (identification + Cq value + date of PCR run)	Duration HBov positivity (days)	Number of PCR assays performed	Number of samples positive for HBov
Monoinfection				
A	HBov Cq 10 23/12/17 HBov Cq 23 8/01/18 HBov Cq 30 15/01/18 HBov Cq 34 1/01/18 HBov Cq 34 25/12/17 HBov Cq 34 ND 21/02/18 31/03/18	97	7	7
B	HBov Cq 32 15/01/18 HBov Cq 34 5/02/18 HRV 13/04/18 20/04/18	35	3	3
C	HBov Cq 32 5/02/18	14	2	2
D	HBov Cq 33 14/02/18	148	2	2
Coinfection				
P	HBov Cq 32, HAdV, IBV, HPeV, HRV 29/12/17		4	1
X	HBov Cq 33, HAdV 2/03/18		2	2
Y	HBov Cq 33, HAdV 9/10/17		4	1
Z	HBov Cq 34, HAdV, EV, HPeV 14/11/17		2	1
AA	HRSV A, HRV 12/12/17		2	1
T	HCoV-OC43, HRSV A 2/01/18		2	1
AB	HAdV, HRSV B, HRV 1/12/17		2	1
AC	HAdV, HRV 25/02/18		2	1
Q	HRSV A 21/11/17		3	1
AD	ND 28/02/18		3	1
AE	HRV 26/01/18		3	1

Positivity for HBov was reflected in bold

HBov, human bocavirus; HRV, human rhinovirus; ND, not detected; HAdV, human adenovirus; IBV, influenza B virus; HPeV, human parechovirus; EV, enterovirus; HRSV, human respiratory syncytial virus; HCoV-OC43, human coronavirus OC43; hMPV, human metapneumovirus

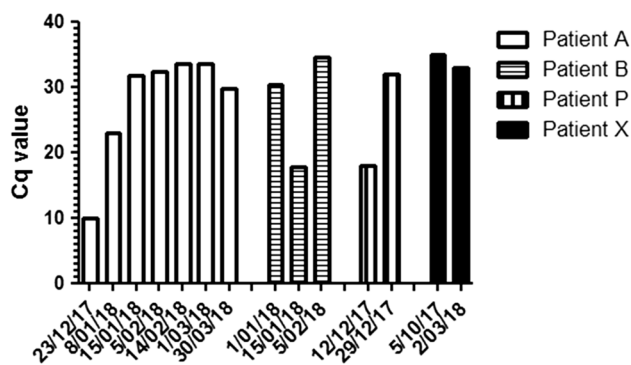


Fig. 2 Persistent positivity for human bocavirus (HBoV) in four patients who were tested multiple times by PCR for respiratory pathogens during the 2017–2018 season

Community-acquired coinfection

The clinical characteristics of patients with HBoV in coinfection were analyzed in detail for a selected subpopulation. Seventeen cases in which HBoV was present at a Cq value ≤ 22 ($n = 15$) or at a lower Cq value than its copathogens ($n = 2$) were analyzed in more detail to assess whether HBoV is a possible respiratory pathogen. An overview of the results can be found in Table 3. The median age of these 17 children was 11.8 months (range, 3.3–32.9 months). Five of these children had been born prematurely, with a gestational age ranging from 31 to 34 weeks. All children were hospitalized briefly in a pediatric ward at one of the three hospitals. Symptoms typically started with rhinitis and cough, which developed to tachypnea, wheezing, crackles, and sometimes the use of accessory muscles and nasal flaring. Bronchi(ol)itis ($n = 8$), recurrent wheezing ($n = 8$) and episodic wheezing ($n = 1$) were the leading diagnoses. Pneumonia was not observed. Chest X-rays were taken of 10 children, nine of which showed abnormalities (peribronchial thickening, atelectasis). Current evidence, however, does not support the routine use of chest radiography in children with bronchiolitis because abnormalities do not correlate well with disease severity. Fever was present in 76.5% of the cases, and hypoxia was present in 41.2% of them (duration of oxygen support varying from 1 to 5 days). Upper respiratory pathology such as conjunctivitis, rhinitis, pharyngitis, stomatitis and otitis media was present in 94.1% of the children. Gastrointestinal disorders, ranging in severity from decreased oral intake to vomiting and diarrhea, was observed in 11 children. There were only slight to moderate elevations in WBC count and CRP, compatible with viral infection.

Discussion

During the respiratory season 2017–2018, 1352 NPSs were received for molecular respiratory screening. HBoV was detected in 6.4% of the samples, which is in accordance with prevalence data described previously [6]. As the two hospitals use slightly different molecular techniques for diagnosis, infection frequencies can be biased. In agreement with previous studies [29], coinfection was common (85.1%), which might reflect the high prevalence of viral infections in young children and the prolonged shedding of HBoV [24]. Higher Cq values dominated. Only 23.0% of the specimens had Cq values ≤ 22 . Competition or interference between viruses in the respiratory tract could explain the low viral loads observed in the majority of coinfections. In our study, HBoV viral loads were comparable between the monoinfected and coinfecting samples, as was also found in previous studies [7, 23, 30]. However, the small number of samples in the monoinfected group makes it difficult to prove any statistical difference between the two.

The median age of the HBoV-positive children was 10.6 months. All but one of the children were younger than 3 years of age. This is in agreement with previous studies in which it was observed that most HBoV infections occur between the ages of 6 months and 3 years [5, 23, 29]. This distribution is compatible with protection from infection by maternal antibodies. The mean age of children who express HBoV in monoinfection (3.9 months) is lower than the mean age of children with HBoV in coinfection (14.0 months) ($P = 0.016$). This makes perfect sense, as older children have had more time to be exposed to different respiratory viruses. Indeed, four 4 prematurely born patients with monoinfection were still hospitalized at the NICU at the time of infection. The HBoV viral load was higher among patients younger than 18 months. This is in concordance with findings of Zhou et al. [7].

Consecutive samples were available for 15 HBoV-positive children. Four children showed persistent HBoV PCR positivity. In one child with seven consecutive samples, a large decrease in viral load was observed after the initial detection, suggesting that a persistent low-level shedding of HBoV in respiratory secretions may follow the HBoV primary infection. This observation was also seen in the case of a second child in which the Cq value increased from 18 to 32 in a two-week period. The Cq values of a third child are indicative of reinfection in mid-January, as the Cq level began January was much higher. In the last child, the observation of similar viral load levels at a 5-month interval could be attributed to protracted viral shedding or to reinfection. In four other children, the Cq value for HBoV was already high at the beginning, ranging

Table 2 Overview of general information and laboratory and clinical parameters for the six children with human bocavirus (HBoV) in mono-infection

General information			Laboratory parameters				Clinical parameters								
Patient	Gender	Age (months)	Prematurity (gestational age)	Medical department	Duration hospitalization (days)	WBC (*10 ⁹ /l)	CRP (mg/L)	First positivity HBoV (date + Cq value)	Fever (°C)	GI disorder	URTD	LRTD	Duration O ₂ therapy (days)	Chest RX	Medical history
A	M	2.2	25 weeks	NICU UH Ghent	206	14.0	<0.6	23/12: 10	No	No	No	Aggravation of CLD	Oxygen dependent at discharge	Peribronchial cuffing, atelectasis	BPD
B	F	0.6	28 weeks	NICU UH Ghent	69	11.7	<0.6	1/01: 30	No	No	No	Aggravation of CLD	63	/	BPD
C	F	1.1	33 weeks	NICU UH Ghent	3	6.9	<0.6	25/12: 34	No	No	No	Lower oxygen saturation	Oxygen dependent at discharge	Normal	Necrotizing enterocolitis
D	M	3.0	26 weeks	NICU UH Ghent	150	10.7	<0.6	21/02: 34	No	No	No	Aggravation of CLD	Oxygen dependent at discharge	/	BPD, pneumothorax
E	F	4.9	No	PICU UH Ghent	4	12.3	6.0 ↑	15/02: 16	No	No	No	Recurrent wheezing	3	Peribronchial cuffing, atelectasis	/
F	M	11.5	No	Pediatric ward AZ Sint-Jan	3	/	/	3/02: 15	Yes (38.4)	No	OMA	No	0	/	Recurrent OMA

WBC, white blood cell count; CRP, C-reactive protein; GI disorder, gastrointestinal disorder; URTD, upper respiratory tract disease; LRTD, lower respiratory tract disease; M, male; F, female; NICU, neonatal intensive care unit; UH Ghent, Ghent University Hospital; CLD, chronic lung disease; BPD, bronchopulmonary dysplasia; PICU, pediatric intensive care unit; OMA, otitis media acuta

Table 3 Overview of general information and laboratory and clinical parameters for the 17 children with human bocavirus (HBov) in coinfection where HBov was present at a low Cq value (≤ 22) or a lower Cq value than its copathogens

General information			Laboratory parameters					Clinical parameters									
Patient	Gender	Age (months)	Prematurity (gestational age)	Duration hospitalization (days)	WBC ($\times 10^9/l$)	CRP (mg/L)	First positivity HBov (date + Cq value)	High viral load CT ≤ 22	Recent virosis 22 < CT ≤ 30	Moderate signal 30 < CT < 40	Fever ($^{\circ}C$)	GI disorder	URTD	LRTD	Duration O ₂ therapy (days)	Chest RX	Medical history
G	M	18.3	31 weeks	5	12.9	16.5 \uparrow	5/01: 17	HBov	-	HAdV	38.6	Gastritis	Rhinitis	Recurrent wheezing	4	Peribronchial cuffing	/
H	F	8.2	34 weeks	5	11.2	9.9 \uparrow	7/04: 16	HBov	-	HCoV-HKU1, HPeV	No	No	Rhinitis, pharyngitis	Recurrent wheezing	3	Peribronchial cuffing	/
I	M	6.7	No	6	19.7 \uparrow	4.2	17/04: 13	HBov	HAdV, HRV	-	No	No	Rhinitis	Recurrent wheezing	2	Peribronchial cuffing	/
J	M	16.6	No	3	12.3	27.0 \uparrow	13/04: 16	HBov	HCoV-NL63	HRV	40.0	No	Conjunctivitis	Bronchitis	0	Peribronchial cuffing	/
K	F	11.9	No	2	12.3	11.0 \uparrow	25/03: 17	HBov	HRSV B, HRV	-	40.0	Gastritis	Rhinitis	Bronchitis	0	/	/
L	F	11.9	34 weeks	6	10.2	7.5 \uparrow	23/03: 14	HBov	HRV	-	No	No	Rhinitis	Bronchitis	1	/	/
M	M	16.8	No	4	6.9	15.0 \uparrow	19/04: 18	HBov , IBV	EV, HRV	-	39.2	No	Stomatitis, conjunctivitis	Recurrent wheezing	0	/	/

Table 3 (continued)

General information			Laboratory parameters					Clinical parameters									
Patient	Gender	Age (months)	Prema-turity (gestational age)	Dura-tion hos-pitali-zation (days)	WBC (*10 ⁹ /l)	CRP (mg/L)	First posi-tivity HBov (date + Cq value)	High viral load CT ≤ 22	Recent virosis 22 < CT ≤ 30	Moder-ate signal 30 < CT < 40	Fever (°C)	GI dis-order	URTD	LRTD	Dura-tion O ₂ therapy (days)	Chest RX	Medical history
N	F	11.8	No	4	21.3 ↑	16.0 ↑	7/03: 14	HBov	-	IBV	No	No	Otitis, rhi-nitis, phar-nyngitis	Recur-rent wheez-ing	0	/	/
O	F	6.0	No	4	16.2	40.0 ↑	10/10: 14	HBov , HAdV	HRV	-	39.4	Gastritis	Con-juncti-vitis	Bronchi-tis	0	/	/
P	M	4.7	No	3	20.5 ↑	26.4 ↑	15/12: 18	HBov	HAdV	-	39.1	Ano-rexia	Rhinitis	Bronchi-olitis	0	Peri-bron-chial cuff-ing	Esopha-geal atresia
Q	M	8.4	No	6	13.7	9.4 ↑	1/02: 22	HBov	hMPV	HAdV, HRV	39.5	Ano-rexia, gastri-tis	Rhinitis	Recur-rent wheez-ing	5	Peri-bron-chial cuff-ing	/
R	M	6.0	No	8	17.0	1.7	26/04: 22	HBov	HRV	-	38.9	Ano-rexia, gas-tritis, enteri-tis	Otitis, rhini-tis	Bronchi-olitis	0	/	/
S	F	16.2	34 weeks	3	13.4	12.7 ↑	3/04: 15	HBov	HCoV-NL63, HRV	-	39.3	Ano-rexia	Rhinitis	Recur-rent wheez-ing	0	Peri-bron-chial cuff-ing	/
T	M	7.4	No	5	18.1 ↑	7.8 ↑	30/04: 17	HBov	HRV	HAdV	39.4	Ano-rexia, gastri-tis	Con-juncti-vitis	Bronchi-olitis	5	Peri-bron-chial cuff-ing	Eczema

Table 3 (continued)

General information			Laboratory parameters					Clinical parameters									
Patient	Gender	Age (months)	Prematurity (gestational age)	Duration hospitalization (days)	WBC (*10 ⁹ /l)	CRP (mg/L)	First positivity HBov (date + Cq value)	High viral load CT ≤ 22	Recent virosis 22 < CT ≤ 30	Moderate signal 30 < CT < 40	Fever (°C)	GI disorder	URTD	LRTD	Duration O ₂ therapy (days)	Chest RX	Medical history
U	M	32.9	No	5	12.7	8.2 ↑	6/04; 16	HBov	-	HRV	38.6	Enteritis	No	Episodic wheezing	4	Normal	/
V	F	16.2	34 weeks	4	10.8	4.7	3/04; 23	-	HBov	HCoV-NL63	38.8	Anorexia	Rhinitis	Recurrent wheezing	0	Peribronchial cuffing	/
W	M	3.3	No	1	12.2	12.5 ↑	25/04; 28	-	HBov	HRV	39.2	Anorexia	Rhinitis	Bronchiolitis	0	/	/

Positivity for HBov was reflected in bold

WBC, white blood cell count; CRP, C-reactive protein; GI disorder, gastrointestinal disorder; URTD, upper respiratory tract disease; LRTD, lower respiratory tract disease; M, male; F, female; HAdV, human adenovirus; HCoV-HKU1, human coronavirus HKU1; HPeV, human parechovirus; HRV, human rhinovirus; ASD, atrial septum defect; HCoV-NL63, human coronavirus NL63; HRSV, human respiratory syncytial virus; IBV, influenza B virus; EV, enterovirus; hMPV, human metapneumovirus

from 33 to 34, and it is therefore not surprising that there was no persistence of HBoV in subsequent runs. Martin et al. found that HBoV sequences in consecutive samples taken from 12 children were not 100% identical and therefore were considered to be HBoV reinfections that contributed to long-term shedding [16]. Since this is a retrospective study, the NPSs are not available anymore in the laboratory, and we are therefore not able to perform sequencing to distinguish between reinfection and persistence. Long-term shedding, which is a feature shared by other human parvoviruses [31], might explain the high frequency of codetection of HBoV with other respiratory pathogens. Therefore, the positivity of HBoV does not necessarily imply respiratory disease, and it is informative for clinicians to have an indication of the viral load.

Proving the clinical relevance and pathogenicity of HBoV is challenging, because fulfillment of Koch's postulates is not possible and because of the high rate of coinfection. One might argue that HBoV is only an aggravating factor of respiratory disease, a persisting virus that is reactivated by the inflammatory process or an innocent bystander that is just detected by chance [32]. Many studies have confirmed the association between HBoV infections in hospitalized children and wheezing episodes [33–35], while other studies fail to find an obvious relationship between HBoV infection and distinct clinical manifestations [24]. In our monoinfected group, three infants were born at a gestational age of 25–28 weeks and already had weak lungs and an immature immune system as a result of their premature birth. All three met the consensus criteria for BPD. As it is known that children with BPD have an increased risk of developing severe disease when acquiring a respiratory viral infection, it is difficult to reliably attribute the observed clinical respiratory signs to the pathogenicity of HBoV. Nevertheless, we saw deterioration of the patients' respiratory status with higher oxygen need and a need for more ventilator support at the time of infection in all three cases. In the monoinfected group, there was only one infant, who was 4.9 months old and without comorbidities, hospitalized at the PICU, for which there was a clear causal relationship between the presence of HBoV at a low Cq value and wheezing. Thus, in our study, only one child out of 77 HBoV-infected children needed hospitalization at the PICU because of severe acute respiratory distress due to HBoV infection. This is lower than the 3.93% observed in a study by Moesker et al. [17].

In order to assess whether HBoV showed characteristics of a respiratory pathogen in the coinfecting group, we chose to focus in detail on cases in which HBoV was expressed at a high viral load or a higher viral load than its copathogens. In these patients, a viral URT prodrome followed by breathing difficulties and wheezing were the most frequently reported clinical signs, and bronchi(oli)tis and recurrent wheezing were the leading diagnoses. In this group, a relationship

was observed between HBoV and LRTD. Some previous studies have found high viral loads to be associated with more-severe symptoms [9], while others have failed to find a clear relationship [5, 7, 23]. In our opinion, it is not possible to establish a clear relationship between viral load and severity of LRTD. In the past, many scoring systems have been developed in an attempt to quantify respiratory distress objectively, but few have demonstrated any predictive validity [36]. Since clinical parameters such as respiratory rates or use of accessory muscles were sometimes lacking in the medical files, it was not possible to make a 100% correct statement about severity. We may assume that all children who needed hospitalization, and certainly those who developed hypoxia with a need for extra oxygen or ventilator support, had rather severe clinical signs.

In conclusion, HBoV is frequently found in NPSs of children with RTD. In our study, we found that HBoV infection at high viral load in infants is associated with bronchi(oli)tis and recurrent or episodic wheezing ($P = 0.013$, Cramer's $V = 0.613$).

Author contributions VV analyzed and interpreted the laboratory and clinical data for all patients of the three participating hospitals included in this study and wrote this manuscript. The microbiologists MR, KF and WV provided the laboratory data from AZ Sint-Jan and AZ Sint-Lucas Hospital of Bruges and helped with the interpretation of the data. The pediatricians SD, KS and FC provided the clinical data from the Ghent University Hospital, the AZ Sint-Jan and AZ Sint-Lucas Hospital of Bruges, respectively, and helped with the interpretation of the data. The study was supervised and coordinated by professor EP. All authors read and approved the final manuscript.

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Data availability The data generated and/or analyzed during the current study are available online: Online Resource 2 Dataset of the 87 nasopharyngeal samples positive for human bocavirus (xls. 447 kB).

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethics approval and consent to participate The study was conducted in accordance with the Declaration of Helsinki and was approved by the Medical Ethics Committee of the three participating hospitals under Belgian Registration number B670201836996.

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