

Pierre Reusser

Current concepts and challenges in the prevention and treatment of viral infections in immunocompromised cancer patients

Presented as an invited lecture
at the 9th International Symposium
Supportive Care in Cancer, St. Gallen,
Switzerland, 26 February–1 March 1997

Abstract Patients with acute leukemia treated with intensive chemotherapy and recipients of bone marrow or peripheral blood stem cell transplants are at high risk of serious viral disease. Recent progress in diagnostic methods and in antiviral drug treatment has permitted the development of efficient management strategies particularly for infections due to herpes simplex virus, varicella-zoster virus, and cytomegalovirus in these patients. By contrast, specific treatment of other virus infections in immunocompromised cancer patients remains an unresolved issue. The emergence of herpesvirus re-

sistance to antiviral drugs is a matter of concern, and its clinical importance among patients with malignancy needs to be elucidated. Future investigations may furthermore help to clarify the role of novel antiviral agents, such as cidofovir, lobucavir, and compound 1263W94, and of the adoptive immunotherapy with virus-specific CTL clones in severely immunodeficient cancer patients.

Key words Bone marrow transplantation · Herpesvirus infection · Antiviral prophylaxis · Antiviral therapy · Adoptive immunotherapy

P. Reusser, M.D.
Department of Medicine,
University Hospital,
Petersgraben 4,
CH-4031 Basel, Switzerland
Tel.: (41) 61-265 25 25
Fax: (41) 61-265 53 53

Introduction

Immunocompromised cancer patients are at elevated risk for serious viral infections. Patients with malignancy who are particularly susceptible to viral disease include those with leukemia or lymphoma, recipients of an allogeneic bone marrow transplant (BMT) or peripheral blood stem cell transplant (PBSCT), and patients treated with high-dose cytoreductive chemotherapy followed by autologous BMT or PBSCT [51, 71]. Both DNA and RNA viruses are important causes of viral disease among profoundly immunodeficient cancer patients (Table 1) [24, 31, 37, 38, 51, 59, 64, 71, 76, 77, 82]. Infections caused by DNA viruses are generally more frequent, which is related to the propensity of these viral agents to establish long-term latency after primary infection and to reactivate during subsequent periods of immunosuppression [51].

A challenging and unresolved issue is the management of infections caused by viruses that do not belong to the family of human herpesviruses. For example, respiratory viruses, such as respiratory syncytial virus and the influenza and parainfluenza viruses, contribute to significant morbidity and mortality in immunocompromised cancer patients [31, 37, 38, 59, 76, 77]. Infections caused by respiratory viruses are characterized by a potential for both community and nosocomial acquisition, high attack rates, and an ability to cause seasonal outbreaks. However, the current diagnostic techniques for these viral agents are of limited clinical value, and there is no specific antiviral therapy of respiratory virus diseases that was proven to be efficacious and safe in severely immunocompromised hosts [59]. At present, no firm treatment recommendations can be made for cancer patients who develop infections caused by respiratory viruses or by other viruses outside the herpesvi-

Table 1 Viruses causing significant morbidity among severely immunodeficient cancer patients

DNA viruses	RNA viruses
Herpes simplex virus types 1 and 2	Respiratory syncytial virus
Varicella-zoster virus	Rhinovirus
Cytomegalovirus	Influenza virus types A, B, C
Epstein-Barr virus	Parainfluenza virus types 1 to 4
Human herpesvirus 6	Measles virus
Hepatitis B virus	Hepatitis A and C viruses
Adenovirus	Rotavirus
Polyomaviruses (JC, BK)	

rus group, and these infections will therefore not be discussed in more detail in this review.

Of the viruses listed in Table 1, herpes simplex virus (HSV) types 1 and 2, varicella-zoster virus (VZV), and cytomegalovirus (CMV) have been particularly well studied, and new strategies for the management of disease caused by these herpesviruses in cancer patients were defined [51, 52, 71, 82]. Progress was made possible by the introduction into clinical use of rapid and sensitive diagnostic methods and of potent antiviral drugs against these viruses in recent years (Fig. 1) [51, 52]. By contrast, the clinical role of other members of the human herpesvirus family (Epstein-Barr virus, human herpesviruses 6, 7, and 8) among patients with malignant disorders has only been partially characterized, and efficient antiviral drug therapy for diseases associated with these viruses is not available at this time [9, 10, 13, 15, 16, 68, 79, 83]. Thus, this review will focus on current concepts and challenges in the prevention and therapy of disease due to HSV, VZV, and CMV among profoundly immunodeficient cancer patients.

Herpes simplex virus infection

HSV infection in adult cancer patients results almost exclusively from reactivation of latent virus [42], and prevention of HSV disease is therefore primarily aimed

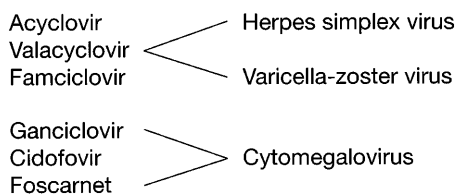


Fig. 1 Antiviral drugs currently available for clinical use against herpesviruses. The drugs are grouped according to their main indications in the prevention and therapy of diseases caused by herpes simplex virus, varicella-zoster virus, and cytomegalovirus. Acyclovir is also used in high doses in the prevention of CMV infection after allogeneic bone marrow transplantation

at HSV seropositive patients. In the absence of antiviral drug prophylaxis, the incidence of HSV reactivation among patients receiving induction chemotherapy for acute leukemia or undergoing allogeneic BMT was reported to be in the range of 60–80% [8, 42]. HSV disease in these patients is most frequently observed at mucocutaneous sites, but esophageal HSV infection is also common and occurs in approximately 10% of patients who have symptoms of upper gastrointestinal disease [8, 66].

Three antiviral agents are currently licensed for treatment of clinical manifestations of HSV infection (Fig. 1). Acyclovir became available for clinical use in the early 1980s and has been extensively evaluated since in the prevention and therapy of HSV disease among both immunocompetent and immunocompromised hosts [78]. Prophylactic acyclovir has become a standard of care at many cancer centers for HSV seropositive patients who are treated for hematologic malignancies or who undergo BMT or PBSCT [71, 78]. Recommended dosages for acyclovir prophylaxis are 250 mg/m² or 5 mg/kg i.v. every 12 h, or between 200 mg three times daily and 800 mg twice daily when the drug is given orally, and the length of treatment is usually 3–5 weeks after the start of chemotherapy [51]. Acyclovir therapy of established mucocutaneous or esophageal HSV disease in cancer patients is given either i.v. at a dose of 250 mg/m² or 5 mg/kg every 8 h, or orally at a dose of 200–400 mg five times daily for a duration of 7–10 days [51, 72, 78]. For therapy of HSV pneumonia or HSV encephalitis, acyclovir i.v. at 500 mg/m² or 10 mg/kg given every 8 h for 10–14 days is recommended by some experts, but antiviral treatment of these conditions has not been studied systematically in cancer patients.

More recently, valacyclovir, a prodrug of acyclovir with 3–5 times its oral bioavailability, and famciclovir, the oral prodrug of penciclovir, were introduced and were shown to exert potent inhibitory activity against HSV types 1 and 2, and VZV [4, 60, 69, 70, 75]. A major advantage of these new agents over acyclovir is their high oral bioavailability, which permits less frequent dosing when treating HSV or VZV disease [4, 49, 60, 69, 75]. However, current results documenting the therapeutic efficacy of valacyclovir and famciclovir for these indications are derived from trials among immunocompetent patients, and firm conclusions regarding the antiviral activity of these drugs in immunocompromised cancer patients cannot be inferred from these studies.

Varicella-zoster virus infection

The clinical manifestations of VZV infection are chickenpox and herpes zoster. Chickenpox results from pri-

primary VZV infection and occurs in most cases in children under the age of 10 years. Children with acute leukemia who develop chickenpox are at particularly high risk for VZV pneumonia, which had an incidence of 32% and a fatality rate of 7% in one study [23]. Herpes zoster is due to reactivation of latent VZV and, among cancer patients, is most frequently observed in those with leukemia or lymphoma and in recipients of autologous or allogeneic BMT [12, 39, 40].

For therapy of established chickenpox or herpes zoster in immunodeficient cancer patients, the use of acyclovir i.v. at a dose of 500 mg/m² or 10 mg/kg given every 8 h for 7–10 days is the currently recommended treatment of choice [3, 48, 51, 67, 78]. Valaciclovir and famciclovir are being evaluated for therapy of herpes zoster after BMT, but results from these trials have not been reported yet.

Prevention of chickenpox among immunocompromised hosts requires strict isolation from infectious persons. However, isolation procedures may be too late in some instances, because patients with chickenpox can be contagious up to 2 days before the skin rash appears [67]. VZV seronegative cancer patients who have been exposed to infectious individuals may benefit from infusions of VZV hyperimmune globulins if this treatment is initiated within 96 h after the exposure [67]. Moreover, immunization with a live attenuated VZV vaccine in seronegative children with leukemia was associated with high seroconversion rates and with a reduction of breakthrough varicella infections and of subsequent herpes zoster [26, 30].

Allogeneic BMT recipients are at risk of VZV reactivation for a prolonged period of time after the transplant [29, 39]. The efficacy of long-term acyclovir prophylaxis was therefore assessed in these patients, but the results obtained indicate that such prophylactic treatment is not advisable, since it only delays the occurrence of herpes zoster and carries the potential for induction of VZV resistance [29, 33, 36, 46].

Cytomegalovirus infection

Patients with acute leukemia who are treated with intensive induction chemotherapy and recipients of an allogeneic or autologous BMT or PBSCT are at elevated risk of developing CMV infection and serious CMV disease, such as interstitial pneumonia. In the absence of efficient antiviral prophylaxis, the incidence of CMV infection among these cancer patients is in the range of 30–50%, and approximately one-third of allograft recipients and 10–20% of autograft recipients with documented CMV infection develop CMV pneumonia, which without specific therapy is fatal in over 80% of cases [21, 43, 54, 71, 80]. A strong independent predic-

tor of CMV disease after BMT is CMV viremia, which was associated with a 5.5-fold increased risk for CMV pneumonia in one series [45]. Furthermore, failure to restore a CD8⁺ cytotoxic T-lymphocyte (CTL) response specific for CMV after BMT or PBSCT was also recognized to be an important predisposing factor for CMV infection and CMV pneumonia during the first 3 months after transplant [35, 55, 57].

Therapy for established CMV pneumonia after BMT or PBSCT remains a major challenge despite the introduction into clinical use of potent antiviral drugs against CMV in recent years. Single-agent therapy of CMV pneumonia, including intravenous ganciclovir and foscarnet, proved to be generally ineffective in BMT recipients [1, 50]. However, several nonrandomized studies evaluating a combination of intravenous ganciclovir plus high doses of intravenous immune globulins for therapy of CMV pneumonia suggested lower fatality rates than in historical controls [19, 50, 62]. Updated results from these studies showed an average survival rate of about 65% early after completion of this combination therapy, but survival 6 months later was only between 30–40% [82]. Thus, with the best presently available therapy the long-term outcome of CMV pneumonia after BMT remains severe, and emphasis must be placed on the prevention of CMV disease in these patients.

Among CMV seronegative allograft recipients, CMV infection can efficiently be prevented by the exclusive use of seronegative or leukocyte-depleted blood products if the BMT or PBSCT donor is also seronegative [6, 7]. Among CMV seropositive patients or seronegative patients with a seropositive organ donor, one approach to prevention of CMV disease is based on suppressing CMV reactivation by the prophylactic use of antiviral drugs. High-dose intravenous acyclovir was evaluated for this indication after allogeneic BMT and was associated with improved survival compared with that in untreated or placebo-treated patients, but was only partially effective in preventing CMV disease [44, 47]. Ganciclovir is considerably more potent than acyclovir against CMV [22]. Prophylactic intravenous ganciclovir in two placebo-controlled studies of allogeneic BMT recipients resulted in significant reduction of CMV disease compared with controls, but there was no survival advantage for ganciclovir-treated patients, which may be related in part to a higher number of deaths from nonviral infections in these patients [28, 81].

An additional strategy for the prevention of CMV disease in transplant recipients, termed “preemptive treatment”, was recently introduced; it consists in administration of antiviral drugs only to patients with virologically documented CMV infection [18, 27, 63]. This strategy permits the restriction of antiviral treatment to the patients most at risk for CMV disease,

which is particularly important in view of the toxicity associated with the potent anti-CMV agents currently available (Fig. 1). Preemptive treatment with intravenous ganciclovir based on positive CMV cultures was investigated in two controlled trials after allogeneic BMT, and resulted in decreased rates of CMV disease and improved survival [27, 63]. However, 12–13% of patients screened by surveillance cultures in these studies developed CMV disease without prior evidence of CMV infection. The proportion of patients missed by culture-based screening for CMV might be reduced by the use of more sensitive and rapid detection methods, such as the antigenemia assay or the polymerase chain reaction (PCR) in peripheral blood specimens [2, 5, 17, 25]. A first randomized comparison of culture-based and PCR-based initiation of preemptive ganciclovir treatment after allogeneic BMT yielded encouraging results; PCR-monitored patients had a lower incidence of CMV disease and better survival by day 100 after transplant [18].

Because of the toxicity and low oral bioavailability of licensed antiviral agents against CMV, there is a need for alternative drugs for the treatment of CMV disease in immunodeficient cancer patients. Several novel antiviral compounds and modified formulations of known drugs have been developed recently and are being investigated for possible clinical use (Table 2). These new agents share a potent inhibitory activity against CMV and an oral bioavailability that in most cases is superior to that of the presently available anti-CMV drugs.

Immunotherapy

Passive immunization by the administration of (hyper) immune globulin infusions to prevent CMV infection and disease after BMT remains a controversial issue. In view of the unclear benefit and substantial costs of this approach, passive immunization cannot be advocated for routine use after BMT or PBSCT without further studies (for more extended discussion see [41, 51]).

The CMV-specific CD8⁺ CTL response plays a crucial role in the protection of allograft and autograft recipients from CMV infection and CMV pneumonia during the posttransplant course [35, 55, 57]. Restora-

Table 2 New compounds and formulations with antiviral activity against cytomegalovirus that can be administered orally

Ganciclovir prodrug
Cidofovir prodrug
Lobucavir (BMS-180194)
Adefovir dipivoxil
1263 W94

tion of this cellular immune response in BMT recipients by the adoptive transfer of CTL clones specific for CMV has been shown to be safe [58, 74]. Ongoing studies are aimed at determining the efficacy of this new strategy in the prevention of CMV infection and CMV disease after allogeneic BMT (S.R. Riddell, personal communication).

Herpesvirus resistance to antiviral drugs

The emergence of herpesvirus resistance to antiviral agents in immunocompromised hosts is being reported with increasing frequency [53]. Infections due to drug-resistant HSV, VZV, and CMV have become recognized as a clinically significant problem particularly in patients with the acquired immunodeficiency syndrome, whereas data on the importance of herpesvirus resistance in immunocompromised cancer patients remain limited [14, 20, 32, 34, 56, 65, 73]. A survey among centers affiliated to the European Group for Blood and Marrow Transplantation (EBMT) indicated that antiviral drug resistance of herpesviruses is proven or suspected in a substantial number of EBMT centers (Table 3) [56]. Moreover, most patients with acyclovir-resistant HSV disease or ganciclovir-resistant CMV infection appeared to respond to foscarnet treatment [56]. However, broader use of susceptibility testing to antiviral agents is required for better definition of the incidence and clinical importance of disease caused by resistant herpesviruses in BMT recipients. Future investigations should be facilitated by the development of more rapid and efficient methods for detection of herpesvirus resistance [11, 61]. Furthermore, antiviral drugs with mechanisms of activation that differ from those of acyclovir or ganciclovir need to be investigated further to establish alternative treatment options in BMT or PBSCT recipients with herpesvirus disease resistant to these agents.

Acknowledgements The author thanks M. Marti for assistance in manuscript preparation.

Table 3 Herpesvirus resistance to antiviral drugs among 68 centers of the European Group for Blood and Marrow Transplantation. (Data from [55])

Resistant virus	Centers reporting resistance ^a n (%)
Herpes simplex virus	17 (25)
Varicella-zoster virus	3 (4)
Cytomegalovirus	19 (28)

^a 39 (57%) of the 68 centers observed no herpesvirus resistance

References

1. Aschan J, Ringdén O, Ljungman P, et al (1992) Foscarnet for treatment of cytomegalovirus infections in bone marrow transplant recipients. *Scand J Infect Dis* 24:143-150
2. Aspin MM, Gallez-Hawkins GM, Giugni TD, et al (1994) Comparison of plasma PCR and bronchoalveolar lavage fluid culture for detection of cytomegalovirus infection in adult bone marrow transplant recipients. *J Clin Microb* 32:2266-2269
3. Balfour HH Jr, Bean B, Laskin OL, et al (1983) Acyclovir halts progression of herpes zoster in immunocompromised patients. *N Engl J Med* 308:1448-1453
4. Beutner KR, Friedmann DJ, Forszpaniak C, Andersen PL, Wood MJ (1995) Valaciclovir compared with acyclovir for improved therapy for herpes zoster in immunocompetent adults. *Antimicrob Agents Chemother* 39:1546-1553
5. Boeckh M, Bowden RA, Goodrich JM, et al (1992) Cytomegalovirus antigen detection in peripheral blood leukocytes after allogeneic marrow transplantation. *Blood* 80:1358-1364
6. Bowden RA, Sayers M, Flournoy N, et al (1986) Cytomegalovirus immune globulin and seronegative blood products to prevent primary cytomegalovirus infection after marrow transplantation. *N Engl J Med* 314:1006-1010
7. Bowden RA, Slichter SJ, Sayers M, et al (1995) A comparison of filtered leukocyte-reduced and cytomegalovirus (CMV) seronegative blood products for the prevention of transfusion-associated CVM infection after marrow transplantation. *Blood* 86:3598-3603
8. Bustamante CI, Wade JC (1991) Herpes simplex virus infection in the immunocompromised cancer patient. *J Clin Oncol* 9:1903-1915
9. Carrigan DR, Knox KK (1994) Human herpesvirus 6 (HHV-6) isolation from bone marrow: HHV-6-associated bone marrow suppression in bone marrow transplant patients. *Blood* 84:3307-3310
10. Carrigan DR, Drobyski WR, Russler SK, Tapper MA, Knox KK, Ash RC (1991) Interstitial pneumonitis associated with human herpesvirus-6 infection after marrow transplantation. *Lancet* 338:147-149
11. Chou S, Erice A, Jordan MC, et al (1995) Analysis of the UL97 phosphotransferase coding sequence in clinical cytomegalovirus isolates and identification of mutations conferring ganciclovir resistance. *J Infect Dis* 171:576-583
12. Christiansen NP, Haake RJ, Hurd DD (1991) Early herpes zoster infection in adult patients with Hodgkin's disease undergoing autologous bone marrow transplant. *Bone Marrow Transplant* 7:435-437
13. Cone RW, Hackman RC, Huang M-LW, et al (1993) Human herpesvirus 6 in lung tissue from patients with pneumonitis after bone marrow transplantation. *N Engl J Med* 329:156-161
14. Drew WL, Miner RC, Busch DF, et al (1991) Prevalence of resistance in patients receiving ganciclovir for serious cytomegalovirus infection. *J Infect Dis* 163:716-719
15. Drobyski WR, Dunne WM, Burd EM, et al (1993) Human herpesvirus-6 (HHV-6) infection in allogeneic bone marrow transplant recipients: evidence of a marrow-suppressive role for HHV-6 in vivo. *J Infect Dis* 167:35-39
16. Drobyski WR, Knox KK, Majewski D, Carrigan DR (1994) Brief report: fatal encephalitis due to variant B human herpesvirus-6 infection in a bone marrow-transplant recipient. *N Engl J Med* 330:1356-1360
17. Einsele H, Steidle M, Vallbracht A, et al (1991) Early occurrence of human cytomegalovirus infection after bone marrow transplantation as demonstrated by the polymerase chain reaction technique. *Blood* 77:1104-1110
18. Einsele H, Ehninger G, Hebart H, et al (1995) Polymerase chain reaction monitoring reduces the incidence of cytomegalovirus disease and the duration and side effects of antiviral therapy after bone marrow transplantation. *Blood* 86:2815-2820
19. Emanuel D, Cunningham I, Jules-Elysee K, et al (1988) Cytomegalovirus pneumonia after bone marrow transplantation successfully treated with the combination of ganciclovir and high-dose intravenous immune globulin. *Ann Intern Med* 109:777-782
20. Englund JA, Zimmermann ME, Swierkosz EM, Goodmann JL, Scholl DR, Balfour HH Jr (1990) Herpes simplex virus resistant to acyclovir. A study in a tertiary care center. *Ann Intern Med* 112:416-422
21. Enright H, Haake R, Weisdorf D, et al (1993) Cytomegalovirus pneumonia after bone marrow transplantation. Risk factors and response to therapy. *Transplantation* 55:1339-1346
22. Faulds D, Heel RC (1990) Ganciclovir. A review of its antiviral activity, pharmacokinetic properties and therapeutic efficacy in cytomegalovirus infections. *Drugs* 39:597-638
23. Feldman S, Lott L (1987) Varicella in children with cancer. Impact of antiviral therapy and prophylaxis. *Pediatrics* 80:465-472
24. Flomenberg P, Babbitt J, Drobyski WR, et al (1994) Increasing incidence of adenovirus disease in bone marrow transplant recipients. *J Infect Dis* 169:775-781
25. Gerna G, Furione M, Baldanti F, et al (1995) Quantitation of human cytomegalovirus DNA in bone marrow transplant recipients. *Br J Haematol* 91:674-683
26. Gershon AA, Steinberg SP, and Varicella Vaccine Collaborative Study Group of the National Institute of Allergy and Infectious Diseases (1989) Persistence of immunity to varicella in children with leukemia immunized with live attenuated varicella vaccine. *N Engl J Med* 320:892-897
27. Goodrich JM, Mori M, Gleaves CA, et al (1991) Prevention of cytomegalovirus disease after allogeneic marrow transplantation by early treatment with ganciclovir. *N Engl J Med* 325:1601-1607
28. Goodrich JM, Bowden RA, Fisher L, et al (1993) Ganciclovir prophylaxis to prevent cytomegalovirus disease after allogeneic marrow transplant. *Ann Intern Med* 118:173-178
29. Han CS, Miller W, Haake R, Weisdorf D (1994) Varicella zoster infection after bone marrow transplantation: incidence, risk factors and complications. *Bone Marrow Transplant* 13:277-283
30. Hardy I, Gershon AA, Steinberg SP, et al (1991) The incidence of zoster after immunization with live attenuated varicella vaccine. A study in children with leukemia. *N Engl J Med* 325:1545-1550

31. Harrington RD, Hooton TM, Hackman RC, et al (1992) An outbreak of respiratory syncytial virus in a bone marrow transplant center. *J Infect Dis* 165:987-993
32. Hill EL, Hunter GA, Ellis MN (1991) In vitro and in vivo characterization of herpes simplex virus clinical isolates recovered from patients infected with human immunodeficiency virus. *Antimicrob Agents Chemother* 35:2322-2328
33. Jacobson MA, Berger TG, Fikrig S, et al (1990) Acyclovir-resistant varicella zoster virus infection after chronic oral acyclovir therapy in patients with the acquired immunodeficiency syndrome (AIDS). *Ann Intern Med* 112:187-191
34. Knox KK, Drobyski WR, Carrigan DR (1991) Cytomegalovirus isolate resistant to ganciclovir and foscarnet from a marrow transplant patient (letter). *Lancet* I:1292-1293
35. Li C-R, Greenberg PD, Gilbert MJ, et al (1994) Recovery of HLA-restricted cytomegalovirus (CMV)-specific T-cell responses after allogeneic bone marrow transplant: correlation with CMV disease and effect of ganciclovir prophylaxis. *Blood* 83:1971-1979
36. Ljungman P, Wilczek H, Gahrton G, et al (1986) Long term acyclovir prophylaxis in bone marrow transplant recipients and lymphocyte proliferation responses to herpes virus antigens in vitro. *Bone Marrow Transplant* 1:185-192
37. Ljungman P, Gleaves CA, Meyers JD (1989) Respiratory virus infection in immunocompromised patients. *Bone Marrow Transplant* 4:35-40
38. Ljungman P, Andersson J, Aschan J, et al (1993) Influenza A in immunocompromised patients. *Clin Infect Dis* 17:244-247
39. Locksley RM, Flournoy N, Sullivan KM, Meyers JD (1985) Infection with varicella-zoster virus after marrow transplantation. *J Infect Dis* 152:1172-1181
40. Mazur MH, Dolin R (1978) Herpes zoster at the NIH: a 20 year experience. *Am J Med* 65:738-744
41. Meyers JD (1989) Prevention of cytomegalovirus infection after marrow transplantation. *Rev Infect Dis* 11 [Suppl 7]:S1691-S1705
42. Meyers JD, Flournoy N, Thomas ED (1980) Infection with herpes simplex virus and cell-mediated immunity after marrow transplant. *J Infect Dis* 142:338-346
43. Meyers JD, Flournoy N, Thomas ED (1986) Risk factors for cytomegalovirus infection after human marrow transplantation. *J Infect Dis* 153:478-488
44. Meyers JD, Reed EC, Shepp DH, et al (1988) Acyclovir for prevention of cytomegalovirus infection and disease after allogeneic marrow transplantation. *N Engl J Med* 318:70-75
45. Meyers JD, Ljungman P, Fisher LD (1990) Cytomegalovirus excretion as a predictor of cytomegalovirus disease after marrow transplantation: importance of cytomegalovirus viremia. *J Infect Dis* 162:373-380
46. Perren TJ, Powles RL, Easten D, et al (1988) Prevention of herpes zoster in patients by long-term oral acyclovir after allogeneic bone marrow transplantation. *Am J Med* 85 [Suppl 2A]:99-101
47. Prentice HG, Gluckman E, Powles RL, et al (1994) Impact of long-term acyclovir on cytomegalovirus infection and survival after allogeneic bone marrow transplantation. *Lancet* 343:749-753
48. Prober CG, Kirk LE, Keeney RE (1982) Acyclovir therapy of chickenpox in immunosuppressed children - a collaborative study. *J Pediatr* 101:622-625
49. Pue M, Benet LZ (1993) Pharmacokinetics of famciclovir in man. *Antiviral Chem Chemother* 4 [Suppl 1]:47-55
50. Reed EC, Bowden RA, Dandliker PS, et al (1988) Treatment of cytomegalovirus pneumonia with ganciclovir and intravenous cytomegalovirus immunoglobulin in patients with bone marrow transplants. *Ann Intern Med* 109:783-788
51. Reusser P (1994) Prophylaxis and treatment of herpes virus infections in immunocompromised cancer patients. *Baillière's Clin Infect Dis* 1:523-544
52. Reusser P (1996) Human cytomegalovirus infection and disease after bone marrow and solid organ transplantation. *Baillière's Clin Infect Dis* 3:357-371
53. Reusser P (1996) Herpesvirus resistance to antiviral drugs: a review of the mechanisms, clinical importance and therapeutic options. *J Hosp Infect* 33:235-248
54. Reusser P, Fisher LD, Buckner CD, Thomas ED, Meyers JD (1990) Cytomegalovirus infection after autologous bone marrow transplantation: occurrence of cytomegalovirus disease and effect on engraftment. *Blood* 75:1888-1894
55. Reusser P, Riddell SR, Meyers JD, et al (1991) Cytotoxic T lymphocyte response to cytomegalovirus following human allogeneic bone marrow transplantation: pattern of recovery and correlation with cytomegalovirus infection and disease. *Blood* 78:1373-1380
56. Reusser P, Cordonnier C, Einsele H, et al, for the Infectious Diseases Working Party of the European Group for Blood and Marrow Transplantation (EBMT) (1996) European survey of herpesvirus resistance to antiviral drugs in bone marrow transplant recipients. *Bone Marrow Transplant* 17:813-817
57. Reusser P, Attenhofer R, Hebart H, Helg C, Chapuis B, Einsele H (1997) Cytomegalovirus-specific T-cell immunity in recipients of autologous peripheral blood stem cell or bone marrow transplants. *Blood* 89:3873-3879
58. Riddell SR, Watanabe KS, Goodrich JM, et al (1992) Restoration of viral immunity in immunodeficient humans by the adoptive transfer of T cell clones. *Science* 257:238-241
59. Sable CA, Hayden FG (1995) Orthomyxoviral and paramyxoviral infections in transplant patients. *Infect Dis Clin North Am* 9:987-1003
60. Sacks SL, Aoki FY, Diaz-Mitoma F, Sellors J, Shafran SD, for the Canadian Famciclovir Study Group (1996) Patient-initiated, twice-daily oral famciclovir for early recurrent genital herpes. A randomized, double-blind multicenter trial. *JAMA* 276:44-49
61. Safrin S, Elbeik T, Mills J (1994) A rapid screen test for in vitro susceptibility of clinical herpes simplex virus isolates. *J Infect Dis* 169:879-882
62. Schmidt GM, Kovacs A, Zaia JA, et al (1988) Ganciclovir/immunoglobulin combination therapy for the treatment of human cytomegalovirus-associated interstitial pneumonia in bone marrow allograft recipients. *Transplantation* 46:905-907
63. Schmidt GM, Horak DA, Niland JC, et al (1991) A randomized, controlled trial of prophylactic ganciclovir for cytomegalovirus pulmonary infection in recipients of allogeneic bone marrow transplants. *N Engl J Med* 324:1005-1011
64. Shields AF, Hackman RC, Fife KH, Corey L, Meyers JD (1985) Adenovirus infections in patients undergoing bone-marrow transplantation. *N Engl J Med* 312:529-533

-
65. Slavin MA, Bindra RR, Gleaves CA, Pettinger MB, Bowden RA (1993) Ganciclovir sensitivity of cytomegalovirus at diagnosis and during treatment of cytomegalovirus pneumonia in marrow transplant recipients. *Antimicrob Agents Chemother* 37:1360-1363
 66. Spencer GD, Hackman RC, McDonald GB, et al (1986) A prospective study of unexplained nausea and vomiting after marrow transplantation. *Transplantation* 42:602-607
 67. Straus SE, Ostrove JM, Inchauspé G, et al (1988) Varicella-zoster virus infections. Biology, natural history, treatment, and prevention. *Ann Intern Med* 108:221-237
 68. Straus SE, Cohen JI, Tosato G, Meier J (1993) Epstein-Barr virus infections: biology, pathogenesis, and management. *Ann Intern Med* 118:45-58
 69. Tyring S, Barbarash RA, Nahlik JE, et al (1995) Famciclovir for the treatment of acute herpes zoster: effects on acute disease and postherpetic neuralgia. A randomized, double-blind, placebo-controlled trial. *Ann Intern Med* 123:89-96
 70. Vere Hodge RA, Sutton D, Boyd MR, et al (1989) Selection of an oral prodrug (BRL 42810; famciclovir) for the antiherpes virus agent BRL 39123 [9-(4-hydroxy-3-hydroxymethyl-1-yl)guanine; penciclovir]. *Antimicrob Agents Chemother* 33:1765-1773
 71. Wade JC (1993) Management of infection in patients with acute leukemia. *Hematol Oncol Clin North Am* 7:293-315
 72. Wade JC, Newton B, McLaren C, et al (1982) Intravenous acyclovir to treat mucocutaneous herpes simplex virus infection after marrow transplantation. A double-blind trial. *Ann Intern Med* 96:265-269
 73. Wade JC, McLaren C, Meyers JD (1983) Frequency and significance of acyclovir-resistant herpes simplex virus isolated from marrow transplant patients receiving multiple courses of treatment with acyclovir. *J Infect Dis* 148:1077-1082
 74. Walter EA, Greenberg PD, Gilbert MJ, et al (1995) Reconstitution of cellular immunity against cytomegalovirus in recipients of allogeneic bone marrow by transfer of T-cell clones from the donor. *N Engl J Med* 333:1038-1044
 75. Weller S, Blum MR, Doucette M, et al (1993) Pharmacokinetics of the acyclovir pro-drug valaciclovir after escalating single- and multiple-dose administration to normal volunteers. *Clin Pharmacol Ther* 54:595-605
 76. Wendt CH, Weisdorf DJ, Jordan MC, Balfour HH Jr, Hertz MI (1992) Parainfluenza virus respiratory infection after bone marrow transplantation. *N Engl J Med* 326:921-926
 77. Whimbey E, Elting LS, Couch RB, et al (1994) Influenza A virus infections among hospitalized adult bone marrow transplant recipients. *Bone Marrow Transplant* 13:437-440
 78. Whitley RJ, Gnann JW Jr (1992) Acyclovir: a decade later. *N Engl J Med* 327:782-789
 79. Wilborn F, Brinkmann V, Schmidt CA, Neipel F, Gelderblom H, Siegert W (1994) Herpesvirus type 6 in patients undergoing bone marrow transplantation: serologic features and detection by polymerase chain reaction. *Blood* 83:3052-3058
 80. Wingard JR, Piantadosi S, Burns WH, et al (1990) Cytomegalovirus infections in bone marrow transplant recipients given intensive cytoreductive therapy. *Rev Infect Dis* 12 [Suppl 7]: S793-S804
 81. Winston DJ, Ho WG, Bartoni K, et al (1993) Ganciclovir prophylaxis of cytomegalovirus infection and disease in allogeneic bone marrow transplant recipients. Results of a placebo-controlled, double-blind trial. *Ann Intern Med* 118:179-184
 82. Zaia JA, Forman SJ (1995) Cytomegalovirus infection in the bone marrow transplant recipient. *Infect Dis Clin North Am* 9:879-900
 83. Zutter MM, Martin PJ, Sale GE, et al (1988) Epstein-Barr virus lymphoproliferation after bone marrow transplantation. *Blood* 72:520-529