



Simulation of fracture in vascular tissue: coupling a continuum damage formulation with an embedded representation of fracture

Christopher Miller¹ · T. Christian Gasser^{1,2}

Received: 1 August 2023 / Accepted: 27 October 2023
© The Author(s) 2023

Abstract

The fracture of vascular tissue, and load-bearing soft tissue in general, is relevant to various biomechanical and clinical applications, from the study of traumatic injury and disease to the design of medical devices and the optimisation of patient treatment outcomes. The fundamental mechanisms associated with the inception and development of damage, leading to tissue failure, have yet to be wholly understood. We present the novel coupling of a microstructurally motivated continuum damage model that incorporates the time-dependent interfibrillar failure of the collagenous matrix with an embedded phenomenological representation of the fracture surface. Tissue separation is therefore accounted for through the integration of the cohesive crack concept within the partition of unity finite element method. A transversely isotropic cohesive potential per unit undeformed area is introduced that comprises a rate-dependent evolution of damage and accounts for mixed-mode failure. Importantly, a novel crack initialisation procedure is detailed that identifies the occurrence of localised deformation in the continuum material and the orientation of the inserted discontinuity. Proof of principle is demonstrated by the application of the computational framework to two representative numerical simulations, illustrating the robustness and versatility of the formulation.

Keywords Vascular tissue · Soft-biological tissue · Microstructure · Fracture · Cohesive zone model · Continuum damage model · Coupled formulation · Embedded discontinuity · Partition of unity finite element method

1 Introduction

Cardiovascular diseases are the leading cause of mortality worldwide, representing an estimated 45% of all deaths in Europe alone [1]. Biomechanical factors are known to influence the inception and development of cardiovascular pathologies [2], such as atherosclerosis [3, 4] and aneurysms [5, 6]. The accrual of microdamage and the onset of vascular tissue failure are of particular importance, with their influence on clinical treatment and management strategies becoming increasingly evident. Nevertheless, there remains a pronounced deficit in the knowledge and understanding

regarding specific microstructural damage and failure mechanisms, and how this ultimately manifests at the macroscale.

Much of this information has been gleaned from the study of bone, where intrinsic and extrinsic mechanisms are known to contribute toward its fracture toughness [7–10]. Additionally, skin has been seen to exhibit mostly intrinsic toughening processes, where crack propagation is described by crack blunting, accompanied by interfibrillar sliding, fibril straightening, fibrillar reorientation, and elastic stretching [11, 12]. However, such mechanisms undoubtedly emanate from the extracellular matrix (ECM) architecture and the underlying structure-function relationships that govern the mechanical functionality of the tissue in question. Furthermore, with only a limited number of load cases, such as dissection [13–15] and penetration [16, 17], having been investigated towards identifying crack tip mechanisms in vascular tissue, there is evidently still much more to discern concerning influential mechanisms in the healthy, and significantly, the diseased arterial wall.

The complexity and inherent challenges associated with the experimental investigation of failure have prompted the pursuance of computational methods to model damage [18–

✉ T. Christian Gasser
gasser@kth.se

Christopher Miller
chrismi@kth.se

¹ Solid Mechanics, Department of Engineering Mechanics, KTH Royal Institute of Technology, Stockholm, Sweden

² Faculty of Health Sciences, University of Southern Denmark, Odense, Denmark

24] and fracture [25–31] in soft biological tissues. The computational modelling of fracture, in particular, constitutes an indispensable tool not only to predict the failure of cracking structures but also to shed insights into fracture processes. Various continuum-oriented methods have been proposed to tackle large deformation non-linear fracture mechanical problems and, specifically, regularise the non-polar continuum by introducing a length-scale associated with the damage mechanism of consequence [32–36]. This gives rise to mesh-independent results but at the cost of requiring an extensively refined mesh within the fracture process zone that invariably necessitates lengthy simulation times, which then prevents the analysis of most clinically relevant problems.

An alternate means of modelling material failure is to instead characterise the fracture surface discontinuously. Embedded formulations such as the embedded finite element method (EFEM) [37], the extended finite element method (XFEM) [38], and the partition of unity finite element method (PUFEM) [39], allow for a cohesive zone to be inserted directly within the element. A traction separation law (TSL) consequently defines the materials' inelastic failure properties and determines the opening of the fracture surface. Approaches of this kind are particularly well-suited to clearly-defined problems where for example there is a single crack and no branching. However, fracture mechanical concepts alone cannot fully quantify soft tissue rupture that exhibits a distinct toughening mechanism [40, 41]. In the context of computational modelling using embedded discontinuities, the continuum material should therefore ideally consider the development of damage. The inclusion of such behaviour would then necessitate a crack initialisation criteria that accounts for the occurrence of material instabilities and localisation in the continuum [42, 43].

In our present work, we use the PUFEM, previously employed to model arterial dissection [25], to numerically investigate the fracture mechanical properties of the arterial wall. The computational framework provides a discrete representation of a discontinuity by enhancing the nodal degrees of freedom, giving rise to a two-field variational formulation with embedded strong discontinuities. The utilised crack tracking algorithm is detailed extensively elsewhere [44]. All constitutive descriptions are taken from our previous study that concerns collagen-related damage in soft biological tissues [45]. The continuous material model is microstructurally motivated and incorporates an interfibrillar failure mechanism that brings about collagen fiber damage. A transversely isotropic cohesive material is defined whereby a phenomenological TSL provides a surface-based description of fracture, with parameters derived from a bottom-up approach that qualitatively replicates a single-element mixed-mode fracture representation (detailed in [45]). A novel crack initialisation criterion and associated

definition of the normal to the fracture surface is introduced, indicating the instance of numerical material instability and the subsequent placement of a discontinuity.

To effectively demonstrate the robustness and versatility of the computational modelling framework, we simulate the uniaxial tensile rupture and symmetry-constrained compact tension (symconCT) testing of the healthy porcine aorta. The latter is an experimental setup that results in the stable propagation of fracture at loading along either the circumferential or axial directions [46].

2 Continuum mechanical framework

In this section, we introduce the fundamental continuum mechanical features associated with the numerical implementation of the PUFEM. The kinematics of strong discontinuities and preliminary information concerning the continuum stress and cohesive traction responses are therefore detailed. A derivation of the spatial variational formulation is additionally presented.

2.1 Kinematics

We consider the reference configuration of a body to contain an embedded discontinuity $\partial\Omega_{0d}$, that separates the body into the subdomains Ω_{0+} and Ω_{0-} , see Fig. 1. A deformation $\chi(\mathbf{X})$ then maps Ω_{0+} and Ω_{0-} into the spatial configurations Ω_+ and Ω_- , with \mathbf{X} denoting the referential position of a material point. The kinematics of the separation is represented by means of a strong discontinuity in the displacement field $\mathbf{u} = \mathbf{u}_c(\mathbf{X}) + \mathcal{H}(\mathbf{X})\mathbf{u}_e(\mathbf{X})$, where the Heavside func-

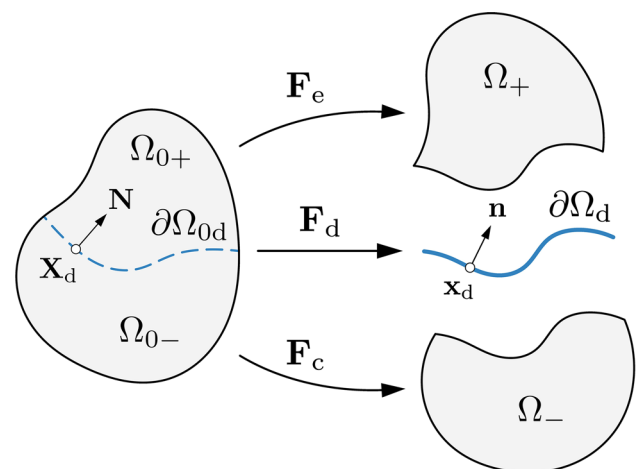


Fig. 1 Discontinuous kinematics representing the reference configuration $\Omega_0 = \Omega_{0+} \cup \partial\Omega_{0d} \cup \Omega_{0-}$, and the current configuration $\Omega = \Omega_+ \cup \partial\Omega_d \cup \Omega_-$, of a body separated by a strong discontinuity: the related deformation gradients are $\mathbf{F}_e = \mathbf{I} + \text{Grad}\mathbf{u}_e + \text{Grad}\mathbf{u}_c$, $\mathbf{F}_d = \mathbf{I} + \text{Grad}\mathbf{u}_c + \mathbf{u}_e \otimes \mathbf{N}/2$ and $\mathbf{F}_c = \mathbf{I} + \text{Grad}\mathbf{u}_c$

tion, $\mathcal{H}(\mathbf{X})$, holds a value of 0 for $\mathbf{X} \in \Omega_{0-}$ and a value of 1 for $\mathbf{X} \in \Omega_{0+}$ [47]. The additive decomposition of \mathbf{u} ultimately introduces the smooth displacement fields \mathbf{u}_c and \mathbf{u}_e , which characterise the compatible and enhanced displacements, respectively.

Following the definition of the displacement field, the deformation gradient, $\mathbf{F}(\mathbf{X}) = \mathbf{I} + \text{Grad}\mathbf{u}(\mathbf{X})$, of the body is defined as

$$\mathbf{F}(\mathbf{X}) = \mathbf{I} + \text{Grad}\mathbf{u}_c(\mathbf{X}) + \mathcal{H}(\mathbf{X})\text{Grad}\mathbf{u}_e(\mathbf{X}) + \delta_d(\mathbf{X})\mathbf{u}_e(\mathbf{X}) \otimes \mathbf{N}(\mathbf{X}_d), \tag{1}$$

where \mathbf{I} is the identity tensor and $\text{Grad}(\bullet) = \partial(\bullet)/\mathbf{X}$ is the material gradient operator. The Dirac-delta function, δ_d , holds a value of 0 in $\mathbf{X} \in \Omega_{0-} \cup \Omega_{0+}$ and a value of ∞ at $\mathbf{X} \in \partial\Omega_{0d}$. The relation $\text{Grad}\mathcal{H}(\mathbf{X}) = \delta_d\mathbf{N}(\mathbf{X}_d)$ is applied, with $\mathbf{N}(\mathbf{X}_d)$ being the unit normal vector defining the orientation of the discontinuity at an arbitrary point $\mathbf{X}_d \in \Omega_{0d}$.

A constitutive representation for the separation of a material body necessitates the assumption of a fictitious discontinuity $\partial\Omega_d$, which is a bijective mapping of $\partial\Omega_{0d}$ into the current configuration by means of the average deformation gradient

$$\mathbf{F}_d(\mathbf{X}_d) = \mathbf{I} + \text{Grad}\left(\mathbf{u}_c + \frac{1}{2}\mathbf{u}_e\right) = \mathbf{I} + \text{Grad}\mathbf{u}_c + \frac{1}{2}\mathbf{u}_e \otimes \mathbf{N}. \tag{2}$$

The factor 1/2 enforces that $\partial\Omega_d$ is the middle between the two surfaces defining the crack [48]. The related unit normal vector of the fictitious discontinuity $\partial\Omega_d$ is then found through the weighted push-forward operation of the covariant vector \mathbf{N}

$$\mathbf{n} = \frac{\mathbf{N}\mathbf{F}_d^{-1}}{|\mathbf{N}\mathbf{F}_d^{-1}|}. \tag{3}$$

We note that the constrained (incompressible) deformation of the continuous material, and so the determination of $\bar{\sigma}$ and \bar{c} in the subsequent section, requires the multiplicative decomposition of the deformation into volume-changing and volume-preserving parts [49]. As such, the modified-deformation gradient $\bar{\mathbf{F}} = J^{-1/3}\mathbf{F}$, with the Jacobian $J = \det\mathbf{F}$, is introduced, completing the kinematic description.

2.2 Continuum response

The continuum response is determined subject to a classical uncoupled formulation [50], in order that incompressibility, a focal feature of soft biological tissues, is enforced. The Helmholtz free energy is therefore $\Psi = \bar{\Psi} + U$, with $\bar{\Psi}$ denoting a purely isochoric contribution and $U = K(\ln J)^2/2$, with K being the bulk modulus, denoting a

purely volumetric contribution. Consequently, the Cauchy stress and the associated spatial stiffness are given by

$$\boldsymbol{\sigma} = \bar{\boldsymbol{\sigma}} + \boldsymbol{\sigma}_{\text{vol}}, \quad \mathbb{C} = \bar{\mathbb{C}} + \mathbb{C}_{\text{vol}}, \tag{4}$$

where $\bar{\boldsymbol{\sigma}}$ and $\bar{\mathbb{C}}$ are constitutive isochoric contributions, whose full derivation is detailed in Sect. 3.1. The volumetric contributions are defined subject to

$$\boldsymbol{\sigma}_{\text{vol}} = p\mathbf{I}, \quad \mathbb{C}_{\text{vol}} = \tilde{p}\mathbf{I} \otimes \mathbf{I} - 2p\mathbb{I}, \tag{5}$$

where $p = dU/dJ$, is the hydrostatic pressure which acts as a Lagrangian multiplier, set to enforce incompressibility. For simplicity, we also use the scalar function $\tilde{p} = p + J\partial p/\partial J$, and \mathbb{I} is the fourth-order identity tensor.

2.3 Cohesive traction response

The coalescence of underlying soft tissue failure mechanisms is lumped into a representative discrete surface [51]. The cohesive response assumes the existence of a transversely isotropic potential per unit undeformed area, $\partial\Omega_{0d}$, of the discontinuity,

$$\psi = \psi(\mathbf{u}, \mathbf{n}, \delta). \tag{6}$$

As such, the cohesive zone properties depend upon; the gap displacement, $\mathbf{u} = \mathbf{u}_e(\mathbf{X}_d)$, the current normal of the discontinuity, \mathbf{n} , and the scalar internal damage variable $\delta \in \{0, \infty\}$ that records the state of damage. The cohesive potential has to obey objectivity requirements, i.e. $\psi = \psi(\mathbf{u}, \mathbf{n}, \delta) = \psi(\mathbf{Q}\mathbf{u}, \mathbf{Q}^{-T}\mathbf{n}, \delta)$, where \mathbf{Q} is an arbitrary proper orthogonal tensor.

The damage surface $\phi(\mathbf{u}, \delta)$ is introduced, and the *Karush–Kuhn–Tucker* loading and unloading conditions, $\dot{\delta} \geq 0, \phi \leq 0, \dot{\delta}\phi = 0$, and the consistency condition, $\dot{\delta}\dot{\phi} = 0$, have to be satisfied.

In accordance with the Coleman–Noll procedure [52], the referential first Piola–Kirchhoff traction vector, \mathbf{T} , and the internal dissipation, \mathcal{D}_{int} , are provided by

$$\mathbf{T} = \frac{\partial\psi}{\partial\mathbf{u}}, \quad \mathcal{D}_{\text{int}} = -\frac{\partial\psi}{\partial\delta}\dot{\delta} \geq 0. \tag{7}$$

Furthermore, to ensure a consistent linearisation within the PUFEM, we introduce the stiffness tensors

$$\mathbf{C}_u = \frac{\partial\mathbf{T}}{\partial\mathbf{u}}, \quad \mathbf{C}_n = \frac{\partial\mathbf{T}}{\partial\mathbf{n}}, \quad \mathbf{C}_\delta = \frac{\partial\mathbf{T}}{\partial\delta}, \tag{8}$$

that describe the stiffness of the cohesive zone with respect to all the dependent variables.

2.4 Variational formulation

Beginning from the standard single-field variational principle, $\int_{\Omega_0} \text{Grad} \delta \mathbf{u} : \mathbf{P}(\mathbf{F}) dV - \delta \Pi^{\text{ext}}(\delta \mathbf{u}) = 0$, where dV , $\mathbf{P}(\mathbf{F})$ and $\delta \mathbf{u}$ denote the referential volume element, the first Piola–Kirchoff stress tensor and the admissible variation of the displacement field, respectively; the variational foundation for the PUFEM formulation may be derived [53]. Given the additive decomposition of the displacement field $\delta \mathbf{u} = \delta \mathbf{u}_c + \mathcal{H} \delta \mathbf{u}_e$, application of a push-forward operation leads to the two variational statements

$$\left. \begin{aligned} & \int_{\Omega_-} \text{sym}(\text{grad}_c \delta \mathbf{u}_c) : \boldsymbol{\sigma}_c dv \\ & + \int_{\Omega_+} \text{sym}(\text{grad}_e \delta \mathbf{u}_c) : \boldsymbol{\sigma}_e dv - \delta \Pi_c^{\text{ext}}(\delta \mathbf{u}_c) = 0, \\ & \int_{\Omega_+} \text{sym}(\text{grad}_e \delta \mathbf{u}_e) : \boldsymbol{\sigma}_e dv \\ & + \int_{\partial \Omega_d} \mathbf{t} \cdot \delta \mathbf{u}_e ds - \delta \Pi_e^{\text{ext}}(\delta \mathbf{u}_e) = 0, \end{aligned} \right\} \quad (9)$$

where dv is the spatial volume element in the current configuration and ds is the surface element on the fictitious discontinuity $\partial \Omega_d$. The quantities $\boldsymbol{\sigma}_c = J_c^{-1} \mathbf{P}(\mathbf{F}_c) \mathbf{F}_c^T$ and $\boldsymbol{\sigma}_e = J_e^{-1} \mathbf{P}(\mathbf{F}_e) \mathbf{F}_e^T$ are the compatible and enhanced Cauchy stress tensors, whilst $\mathbf{t} = \mathbf{T} dS/ds$ is the Cauchy traction vector acting on $\partial \Omega_d$. The virtual energies $\delta \Pi_c^{\text{ext}}$ and $\delta \Pi_e^{\text{ext}}$ refer to the referential domains Ω_{0-} and Ω_{0+} , respectively. Finally, $\text{sym}(\bullet) = ((\bullet) + (\bullet)^T)/2$ gives the symmetric part of (\bullet) , whilst $\text{grad}_c(\bullet) = \text{Grad}(\bullet) \mathbf{F}_c^{-1}$ and $\text{grad}_e(\bullet) = \text{Grad}(\bullet) \mathbf{F}_e^{-1}$ are spatial gradient operators.

The consistent linearisation of Eq. (9) is extensively detailed in [39] and the outcome with respect to the finite element implementation is later elaborated upon in Sect. 4.1.

3 Constitutive descriptions

Here we outline the fundamental aspects concerning the motivation and mathematical necessities for the continuum microstructural damage model and the transversely isotropic cohesive zone model. We also establish the criteria governing crack initialisation and the orientation of the fracture surface.

3.1 Continuum microstructural damage model

The key features of the continuum microstructural damage model, first introduced in [45], are henceforth summarised to provide a generalised overview. Further details are provided in the appendices for the interested reader, which are taken directly from the original publication.

3.1.1 Histomechanical assumptions

We consider vascular tissue as a fiber-reinforced material at finite deformations, whereby collagen fibers with varying orientations are embedded in an isotropic ground substance. Each fiber is comprised of numerous CFPG-complexes, mechanical sub-units containing undulated collagen fibrils adjacently adjoined by a proteoglycan-rich interfibrillar matrix, see Fig. 2a. In the event that soft tissue is exposed to tension, fibrils begin to straighten and carry load. Two time-dependent proteoglycan (PG) deformations are considered: (i) their ability to slide when the CFPG-complex is exposed to heightened stresses, reducing the stress back down towards a homeostatic target stress, and (ii) their capacity to recover back towards a referential minimal energy state at low stresses [54]. Finally, it is assumed that interfibrillar failure will occur at some limiting PG deformation, see Fig. 2b, leading to the cessation of the CFPG-complexes load-carrying ability. Furthermore, this process is postulated as being stress-dependent; upon exposure to excessive supra-physiological stress, there is assumed to be a significant decline in the structural integrity of the interfibrillar material.

3.1.2 Collagen associated kinematics

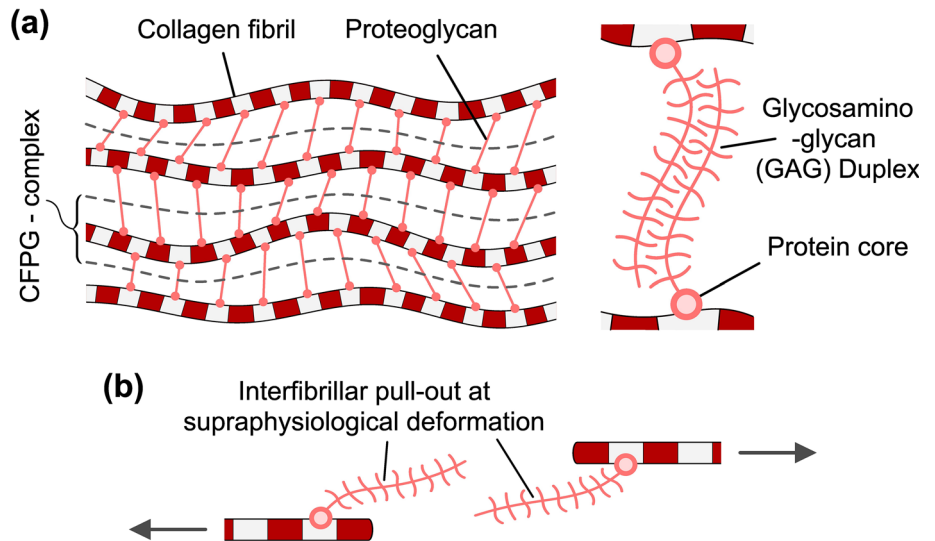
Collagen fiber are assumed to have a referential orientation, in Ω_0 , defined by the unit vector \mathbf{M} . An orientation density function $\rho(\mathbf{M})$ then defines the referential alignment distribution of collagen fibers. If we consider an affine deformation, then the spatial orientation of a fiber, in Ω , is $\mathbf{m} = \bar{\mathbf{F}} \mathbf{M}$, and $\lambda = |\mathbf{m}|$ denotes its stretch.

The i th CFPG complex within a fiber is therefore subjected to the one-dimensional stretch λ , which can be multiplicatively decomposed into three parts, see Figure 2 in [45]. Firstly, the straightening stretch $\lambda_{st i}$, maps the CFPG-complex from Ω into $\Omega_{st i}$, and acts to straighten the initially undulated fibril, where $\lambda_{st i}$ is drawn from the probability density function $f_{\lambda_{st}}(\lambda_{st})$. A triangular distribution is used for convenience, yet the model remains general and is equally applicable to any probability density function.

Secondly, the interfibrillar deformation $\lambda_{pg i}$, maps the CFPG-complex from $\Omega_{st i}$, into $\Omega_{pg i}$, and represents the collective deformation of all PGs within the CFPG complex. However, if the threshold PG deformation $\lambda_{pg \text{ fail}}$, is surpassed, then interfibrillar failure occurs and the CFPG complex alternatively maps into the other intermediate configuration, $\Omega_{pg \text{ fail } i}$. Finally, the elastic fibril deformation $\lambda_{f i}$, maps the CFPG-complex into Ω , such that

$$\lambda = \lambda_{f i} \bar{\lambda}_i ; \bar{\lambda}_i = \lambda_{pg i} \lambda_{st i} \quad (\text{no summation}), \quad (10)$$

Fig. 2 a A collagen fiber is composed of multiple CFPG-complexes, themselves consisting of collagen fibrils and a PG-dominated interfibrillar matrix that connects adjacent fibrils. PGs are made up of a GAG duplex and are attached to fibrils via a protein core, **b** following supraphysiological deformations interfibrillar failure occurs in the PG-rich matrix, leading to collagen fibril pull-out



encapsulates the multiplicative kinematics of the i th CFPG-complex belonging to a collagen fiber with the referential orientation \mathbf{M} .

3.1.3 CFPG-complex constitutive properties

A CFPG-complex is considered linear elastic in a first Piola–Kirchoff stress vs stretch setting, and so

$$P_{fi} = \begin{cases} k_f \left(\frac{\lambda}{\bar{\lambda}_i} - 1 \right) & ; \lambda > \bar{\lambda}_i \\ 0 & ; \text{otherwise} \end{cases} \quad (11)$$

defines the 1D stress of the i th CFPG-complex, where k_f is a stiffness parameter i.e., the fibrils stiffness. The corresponding referential stiffness of the CFPG-complex is then defined as

$$C_{fi} = 2 \frac{\partial S_{fi}}{\partial \lambda^2} = \begin{cases} \frac{k_f}{\bar{\lambda}_i^3} & ; \lambda > \bar{\lambda}_i \\ 0 & ; \text{otherwise} \end{cases} \quad (12)$$

where with the assumption of incompressibility, $S_{fi} = P_{fi}/\lambda$ denotes the second Piola–Kirchhoff stress.

Following on from the discussion of the different interfibrillar PG deformations in Sect. 3.1.1, λ_{pgi} evolves subject to

$$\frac{\partial \lambda_{pgi}}{\partial t} = \begin{cases} \frac{P_{fi}/P_0 - 1}{\eta_{slid}} & ; P_{fi} \geq P_0 \\ \frac{1 - \lambda_{pgi}}{\eta_{rec}} & ; P_{fi} < P_0 \end{cases} \quad (13)$$

where P_0 , is the desired homeostatic CFPG-complex stress and η_{slid} , and η_{rec} , are the rate parameters that govern the aforementioned mechanisms.

Lastly, the limiting interfibrillar PG deformation, $\lambda_{pg\ fail}$, likewise eluded to in Sect. 3.1.1, is defined by

$$\lambda_{pg\ fail} = \frac{\alpha}{P_0} (P_f - P_0) + \beta, \quad (14)$$

where α and β are phenomenological damage-related parameters that determine the dependence of interfibrillar failure on the stress of a CFPG-complex, and additionally, as per Eq. (13), the rate of interfibrillar PG deformation. Specifically, α determines the impact of the stress state when the CFPG-complex is stressed beyond P_0 , whilst β is independent of the stress.

3.1.4 Collagen fiber properties

The relation Eq. (13) results in a highly non-linear development of λ_{pgi} . Expressly, the imposition that only straightened fibrils are permitted to slide necessitates a discrete implementation of the model. As such, when determining the overall collagen fiber properties, a finite number n_f of CFPG-complexes are considered. Inverse transform sampling is used to generate a straightening stretch λ_{sti} for the i th CFPG-complex, with relevant equations pertaining to a triangular distribution provided in “Appendix A”.

We present here a general summary of the integral method for collagen fiber stress and stiffness determination. Upon inspection of Eq. (12), we observe that the referential stiffness of a CFPG-complex is solely dependent upon the overall stretch λ , of the fiber. As such, the referential stiffness of a collagen fiber is found through the referential stiffness of a single CFPG-complex C_f , multiplied by the proportion of those CFPG-complexes that are engaged in load-bearing, which we term $p_{rec} \in \{0, 1\}$. To identify said proportion, we linearly interpolate the cumulative density with respect to the squared fiber stretch, over the n_f discrete CFPG-complexes.

It is also necessary to compute the collagen fiber damage variable $\delta \in \{0, 1\}$, which represents the proportion of those CFPG-complexes that have failed based on the limiting interfibrillar deformation expressed by Eq. (14). It should be noted that this differs from the damage variable used in the cohesive traction response. The determination of δ is detailed in ‘‘Appendix B’’. The contribution of failed CFPG-complexes is then neglected from the overall fiber stiffness, and the referential collagen fiber stiffness reads, generally, as

$$C = \max\{0, C_f(p_{rec} - \delta)\} . \tag{15}$$

The corresponding second Piola–Kirchoff stress is then ascertained through the integration of the stiffness with respect to the squared fiber stretch, across all those CFPG-complexes in tension. It is defined by the general expression

$$S = \int_{\bar{\lambda}_d^2}^{\lambda^2} C d\lambda^2 , \tag{16}$$

where the lower bound, $\bar{\lambda}_d$, is a function of δ , i.e., the bounds of integration will alter based on the proportion of failed CFPG-complexes. The dependency of C and S upon the loading history, and as such, the development of internal variables such as λ_{pgi} and δ , leads to an expansive set of piecewise analytical expressions that are additionally detailed in ‘‘Appendix B’’.

Finally, the Piola transform, i.e., the push-forward operation into the spatial configuration, determines the corresponding Cauchy stress $\sigma = \lambda^2 S$, with incompressible fiber deformation having been assumed.

3.1.5 Macroscopic collagen contribution

In line with the theory of fibrous tissues [55], the total collagenous stress is found by integration of the fiber stress over the solid angle ω , i.e., by a spherical integration over the unit sphere, and thus is given by the relation

$$\bar{\sigma}_{coll} = \int_{\omega} \rho(\mathbf{M}) \frac{\sigma(\lambda(\mathbf{M}))}{\lambda^2} \text{dev}(\mathbf{m} \otimes \mathbf{m}) d\omega . \tag{17}$$

There is precedent for the use of the Bingham distribution to define the referential collagen fiber orientation density in the computational modelling of vascular tissue [56, 57]. The orientation density function $\rho(\mathbf{M})$, was therefore defined according to

$$\rho(\mathbf{M}(\phi, \theta)) = \frac{1}{2c} \exp[\kappa_1 (\cos \phi \cos \theta)^2 + \kappa_2 (\cos \phi \sin \theta)^2] , \tag{18}$$

with ϕ , and θ , denoting the azimuthal and elevation angles, respectively. With specific regard to the morphology of the

arterial wall, values of $\phi = \theta = 0$ relate to the circumferential direction, values of $\phi = \pi/2$ and $\theta = 0$ relate to the axial (longitudinal) direction, whilst $\theta = \pi/2$ relates to the out-of-plane radial direction. Additionally, the parameters κ_1 , and κ_2 , determine the shape of the distribution, whilst c , is a normalisation parameter.

3.1.6 Non-collagenous contribution

The isotropic ground substance, consisting of non-collagenous material, is described by the (classical) Neo-Hookean strain energy density function

$$\psi_{nH} = \mu(\bar{I}_1 - 3)/2 , \tag{19}$$

where μ denotes the referential shear stiffness and $\bar{I}_1 = \text{tr} \bar{\mathbf{b}}$ is the modified first invariant, with $\bar{\mathbf{b}} = \bar{\mathbf{F}}\mathbf{F}^T$ being the modified left Cauchy–Green tensor. The relation $\bar{\sigma}_{nH} = \mu \text{dev}(\bar{\mathbf{b}})$, then provides the deviatoric contribution of the Cauchy stress.

3.1.7 Overall soft tissue properties

The additive decomposition of the Cauchy stress, by means of the rule of mixtures, gives the total deviatoric stress as

$$\bar{\sigma} = \bar{\sigma}_{coll} + \bar{\sigma}_{nH} , \tag{20}$$

The total stress for load-bearing soft-biological tissue is then obtained through the further addition of the isochoric stress σ_{vol} , as per the treatment discussed in Sect. 2.2.

3.2 Transversely isotropic cohesive zone model

In order to particularize the transversely isotropic cohesive model introduced in Sect. 2.3, we restrict our attention to the cohesive potential of the form $\psi = \psi(\mathbf{u} \otimes \mathbf{u}, \mathbf{n} \otimes \mathbf{n}, \delta)$. Through application of the theory of invariants, with i_1, \dots, i_5 denoting deformation dependent variables [58], the potential may be alternatively expressed as $\psi = \psi(i_1, i_2, i_3, i_4, i_5, \delta)$, which following particularization reads

$$\psi = \psi(i_1, i_4, \delta) = \frac{\kappa(\delta)}{2} [i_1 + (1 - \beta)i_4] , \tag{21}$$

with $i_1 = \mathbf{I} : (\mathbf{u} \otimes \mathbf{u})$, and $i_4 = (\mathbf{u} \otimes \mathbf{u}) : (\mathbf{n} \otimes \mathbf{n})$, defining the invariants used in this constitutive description. The expression

$$\kappa = \frac{T_0}{\delta} \left[a \exp\left(-b\left(\frac{\delta}{\alpha}\right)^c\right) + (1 - a) \exp\left(-d\left(\frac{\delta}{\alpha}\right)^e\right) \right] , \tag{22}$$

describes the evolution of the interface stiffness under normal loading (mode I) along with the accumulation of damage δ . The non-dimensional parameters a, b, c, d , and e define the shape of the traction separation law as motivated by the response of the continuum damage model introduced in Sect. 3.1.

The parameter T_0 is the respective peak normal traction, and the parameter α normalizes the displacement and is defined by

$$\alpha = \frac{G}{T_0 \zeta}, \tag{23}$$

with G denoting the respective mode I fracture energy. The energy needed to form a unit of undeformed fracture surface, whilst ζ is the numerical approximation of the integral $\int_0^\infty (a \cdot \exp(-bx^c) + (1 - a) \cdot \exp(-dx^e)) dx$.

In addition, the normal and shear responses at the interface are coupled by the parameter β . It represents the ratio between the elastic stiffness and peak traction in the normal and shear directions, such that $\beta\kappa$ and βT_0 yield the interfaces' shear stiffness and the peak shear traction, respectively. As with the parameters a, \dots, e , also κ is motivated by the response of the continuum damage model introduced in Sect. 3.1.

Towards finalising the constitutive description, the development of the state of interface damage needs to be specified. The damage surface $\phi(\mathbf{u}, \delta) = |\mathbf{u}| - \delta$, and the expression

$$\dot{\delta} = \frac{|\mathbf{u}| - \delta}{\eta_{cz}}, \tag{24}$$

therefore determine the evolution of the internal scalar damage variable δ (given that $\phi > 0$). Here, the rate constant η_{cz} specifies how fast damage in the cohesive zone is able to develop. Interface damage is linked to volume deformation, and the time needed for interstitial fluid to flow into the failure domain thus determines the time scale set by η_{cz} .

Figure 3 displays the normal and shear traction responses of the proposed cohesive zone model, with the accumulation of interface damage for η_{cz} being much smaller than the loading rate applied to the interface for the provided example.

The first Piola–Kirchoff traction vector is determined through Eq. (7)₁, and so is defined as

$$\mathbf{T} = \kappa \mathbf{u}_n + \beta \kappa \mathbf{u}_s, \tag{25}$$

where $\mathbf{u}_n = (\mathbf{u} \cdot \mathbf{n})\mathbf{n}$ and $\mathbf{u}_s = \mathbf{u} - \mathbf{u}_n$ denote the normal and shear displacement at the interface, respectively. The stiffnesses associated with the cohesive zone model are found

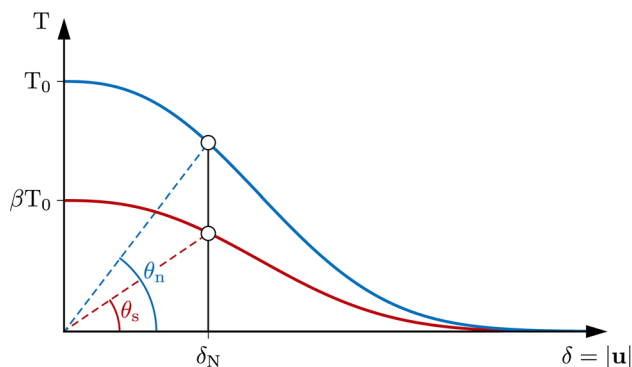


Fig. 3 Demonstration of the normal (blue) and shear (red) traction responses of the proposed transversely isotropic cohesive zone model with the accumulation of interface damage $\delta = |\mathbf{u}|$, where $\theta_n = \tan^{-1}(\kappa)$ and $\theta_s = \tan^{-1}(\beta\kappa)$ for illustrative purposes. (Color figure online)

through the application of Eq. (8) to Eq. (25), leading to

$$\left. \begin{aligned} \mathbf{C}_u &= \kappa(1 - \beta)\mathbf{n} \otimes \mathbf{n} + \beta\kappa \mathbf{I} \\ \mathbf{C}_n &= \kappa(1 - \beta)\mathbf{n} \otimes \mathbf{u} \\ \mathbf{C}_\delta &= \gamma(\mathbf{u}_n + \beta\mathbf{u}_s) \end{aligned} \right\} \tag{26}$$

where the term $\gamma = \partial\kappa/\partial\delta$ is obtained by appropriate treatment of Eq. (22).

Finally, the internal dissipation, following Eq. (7)₂ reads

$$\mathcal{D}_{int} = -\frac{1}{2} \frac{\partial\kappa}{\partial\delta} \dot{\delta}(i_1 + (1 - \beta)i_4) > 0, \tag{27}$$

and imposes a constraint on the model parameters to be used in the traction separation law. Notably, the proposed approach of defining the interface properties through the stiffness factor κ naturally accounts for mixed-mode conditions at the interface.

3.3 Cohesive zone initialisation

The nature of the crack initialisation criteria in approaches utilising embedded strong discontinuities is critical because it determines the evolution of the fracture surface and thus crack propagation in the failed material. As the continuum microstructural model in the presented framework incorporates an internal damage mechanism and no associated method of regularisation, the initialisation of the cohesive zone must employ a bifurcation analysis and account for the emergence of material instability.

To this end, following the sufficient development of collagen fiber damage, the first Piola–Kirchhoff-related elasticity tensor

$$\mathcal{A}_{IJKL} = J \mathbf{F}_{iI}^{-1} \mathbf{F}_{kK}^{-1} c_{ijkl}, \tag{28}$$

becomes non-positive definite, and localization in the non-polar continuum leads to material instability [42, 43]. This therefore marks the state of deformation and the point in space \mathbf{X} in which the cohesive zone model is introduced. In contrast to the description of the bulk material, the incompressibility of tissue within the fracture process zone is not enforced. Given the transversely isotropic nature of our cohesive zone description, it is necessary to determine whether a normal or shear deformation initialises the localization. We, therefore, consider the eigenvalue problem

$$(\mathcal{A} - \lambda \mathbf{I}) : \mathbf{v} = \mathbf{0}, \quad (29)$$

and use the first negative (lowest) eigenvalue $\lambda_i; i = 1, \dots, 9$ [59] to inform the crack initialisation criteria. In the event that the lowest eigenvalue falls in the range $1 \leq i \leq 3$, the interface begins at the peak normal traction T_0 , whilst for $4 \leq i \leq 9$, shear failure triggers the fracture and the cohesive zone model begins at the peak shear traction βT_0 .

It is also necessary to define the referential normal vector \mathbf{N} of the discontinuity upon the activation of the cohesive zone model. Our symconCT experiments [46] showed fracture propagation along directions that were in between the maximum principal stress direction and the weakest material direction. Therefore, the condition

$$\tilde{\mathbf{n}} = w_\sigma \mathbf{n}_\sigma + w_{cf} \mathbf{n}_{cf}, \quad (30)$$

with the weights $w_\sigma = w_{cf} = 0.5$, was used to set the normal to the interface. Here, \mathbf{n}_σ is a principal stress direction, the push forward of \mathbf{N}_σ , which relates to the eigenvector \mathbf{v}_i associated with the lowest eigenvalue. Its determination subject to the solution of Eq. 29 is detailed later in Sect. 4.2.3. The vector \mathbf{n}_{cf} denotes the direction along which the in-plane orientation density of collagen fibers is lowest according to Eq. 18, i.e., the deformed axial direction. Both vectors refer to the point of loss of positive definiteness of \mathcal{A} in the deformed configuration Ω , and so the pull-back and normalization operation

$$\mathbf{N} = \frac{\tilde{\mathbf{n}}\mathbf{F}}{|\tilde{\mathbf{n}}\mathbf{F}|}, \quad (31)$$

provides the orientation of the discontinuity in the reference configuration Ω_0 . Application of Cauchy's stress theorem to the first Piola–Kirchhoff stress of the continuum, $\mathbf{P} = J\boldsymbol{\sigma}\mathbf{F}^{-T}$, then provides the the peak normal traction T_0 .

4 Finite element implementation

This section details the key aspects of the framework's finite element implementation. A brief elaboration of the PUFEM

formulation's cornerstone features is followed by a discussion of the specifics for the numerical implementation of the continuum microstructural damage model and the transversely isotropic cohesive zone model.

4.1 Overview of PUFEM formulation implementation

The computational framework is implemented in the multi-purpose finite element analysis program FEAP [60], using an in-house code. Within the PUFEM, the standard polynomial interpolation functions are enriched by the Heaviside function to attain good local approximations for the underlying problem. The displacement field \mathbf{u} is therefore expressed as

$$\mathbf{u} = \sum_{i=1}^{n_{\text{elem}}} N^I \mathbf{u}_{Ic} + \mathcal{H} \sum_{i=1}^{n_{\text{elem}}} N^I \mathbf{u}_{Ie}, \quad (32)$$

with N^I representing the standard interpolation functions, where I is an index running between 1 and n_{elem} , the total number of nodes per element. Furthermore, the regular and enhanced nodal displacements are denoted by \mathbf{u}_{Ic} and \mathbf{u}_{Ie} , respectively.

With the use of the displacement field in Eq. (32), further treatment of the variational statements provided in Eq. (9) leads to a linearised set of equations for a single finite element

$$\begin{bmatrix} \mathbf{K}_{\mathbf{u}_c \mathbf{u}_c} & \mathbf{K}_{\mathbf{u}_c \mathbf{u}_e} \\ \mathbf{K}_{\mathbf{u}_e \mathbf{u}_c} & \mathbf{K}_{\mathbf{u}_e \mathbf{u}_e} \end{bmatrix}_{i-1}^e \begin{bmatrix} \Delta \mathbf{u}_c \\ \Delta \mathbf{u}_e \end{bmatrix}_i^e = \begin{bmatrix} \mathbf{f}_{\mathbf{u}_c}^{\text{ext}} \\ \mathbf{f}_{\mathbf{u}_e}^{\text{ext}} \end{bmatrix}_{i-1}^e - \begin{bmatrix} \mathbf{f}_{\mathbf{u}_c}^{\text{int}} \\ \mathbf{f}_{\mathbf{u}_e}^{\text{int}} \end{bmatrix}_{i-1}^e. \quad (33)$$

The indices i and $i - 1$ relate to the iteration steps of the global Newton iteration procedure, whilst $\Delta \mathbf{u}_c$ and $\Delta \mathbf{u}_e$ are increments in the compatible and enhanced displacements, respectively.

The vectors $\mathbf{f}_{\mathbf{u}_c}^{\text{ext}}, \mathbf{f}_{\mathbf{u}_e}^{\text{ext}}$ and $\mathbf{f}_{\mathbf{u}_c}^{\text{int}}, \mathbf{f}_{\mathbf{u}_e}^{\text{int}}$ are the nodal force vectors due to external and internal loading. With the subscripts $(\bullet)_{\mathbf{u}_c}$ and $(\bullet)_{\mathbf{u}_e}$ denoting an association with compatible and enhanced degrees of freedom, respectively. The stiffness matrices $\mathbf{K}_{\mathbf{u}_c \mathbf{u}_c}, \mathbf{K}_{\mathbf{u}_c \mathbf{u}_e}, \mathbf{K}_{\mathbf{u}_e \mathbf{u}_c}$ and $\mathbf{K}_{\mathbf{u}_e \mathbf{u}_e}$ emerge from the consistent linearisation of the internal loading vectors with respect to the compatible and enhanced displacements. For details concerning the construction of the nodal force vectors and stiffness matrices, based on the output quantities from the continuum (Sect. 3.1) and discontinuous (Sect. 3.2) constitutive descriptions, see [39].

The solution of the linearised system (33) is often termed a monolithic approach. In the case of fracture mechanical problems however, a staggered approach is frequently utilised as it oftentimes presents as a more robust option. In this case,

the system may alternatively be split into the two subsystems

$$[\mathbf{K}_{\mathbf{u}_c \mathbf{u}_c}]_{i-1}^e [\Delta \mathbf{u}_c]_i^e = [\mathbf{f}_{\mathbf{u}_c}^{\text{ext}}]_{i-1}^e - [\mathbf{f}_{\mathbf{u}_c}^{\text{int}}]_{i-1}^e ; \tag{34}$$

$$[\mathbf{K}_{\mathbf{u}_c \mathbf{u}_c}]_{i-1}^e [\Delta \mathbf{u}_c]_i^e = [\mathbf{f}_{\mathbf{u}_c}^{\text{ext}}]_{i-1}^e - [\mathbf{f}_{\mathbf{u}_c}^{\text{int}}]_{i-1}^e . \tag{35}$$

that are solved separately, one after the other. As such the contributions from $\mathbf{K}_{\mathbf{u}_c \mathbf{u}_c}$ and $\mathbf{K}_{\mathbf{u}_c \mathbf{u}_c}$ are neglected, resulting in the increments $\Delta \mathbf{u}_c$ and $\Delta \mathbf{u}_c$ being independent from one another.

A critical feature of numerical approaches that use embedded strong discontinuities, such as the PUFEM, is the usage of an effective method for the tracking of 3D crack surfaces. This work uses a two-step algorithm, first introduced in [44]. Here, the discontinuity normal \mathbf{N} , defined in Sect. 3.3, serves as a trial state, which is then corrected to meet the morphology (geometry) of the already existent crack. The method employs non-local smoothing to circumvent topological difficulties, thus providing a closed crack surface.

4.2 Numerical implementation of the continuum microstructural damage model

The finite element implementation of the continuum microstructural damage model requires the numerical treatment of some of those relations established in Sect. 3.1. Furthermore, Sequential details concerning the full numerical implementation of the continuum microstructural damage model are provided in Table 1. It conveys the ordering and usage of the various equations used to compute the overall soft-tissue stress and stiffness.

4.2.1 Temporal update of the proteoglycan stretch

A first-order accurate backward Euler discretization of Eq. (13) informs the evolution of the interfibrillar PG deformation based on the stress-state of a CFPG-complex. The i th collective PG stretch is initialized as $\lambda_{\text{pg}i0} \leftarrow 1.0$, after which it evolves subject to

$$\lambda_{\text{pg}iN} = \begin{cases} \lambda_{\text{pg}iN-1} + \frac{\Delta t}{\eta_{\text{slid}}} (P_{fi}/P_0 - 1) ; & P_{fi} > P_0 \\ \lambda_{\text{pg}iN-1} + \frac{\Delta t}{\eta_{\text{rec}}} (1 - \lambda_{\text{pg}iN-1}) ; & P_{fi} \leq P_0, \end{cases} \tag{36}$$

where the subscripts $(\bullet)_N$ and $(\bullet)_{N-1}$ relate to the current and previous time points, respectively. Here, $\Delta t = t_N - t_{N-1}$ denotes the size of the time step and the updated $\lambda_{\text{pg}iN}$ is stored in the history $\mathcal{H}_{i+1N}^{\text{con}} \leftarrow \lambda_{\text{pg}iN}$ and serves as input for the next iteration.

4.2.2 Macroscopic collagen contribution—numerical integration

The determination of the macroscopic collagen contribution occurring at a Gauss point of a finite element, according to Eq. (17), requires the spherical integration of the collagen fiber response over the unit sphere. Spherical t-designs [61]

$$\int_{\omega} (\bullet) d\omega \approx \left(\frac{4\pi}{l_{\text{int}}} \right) \sum_{j=1}^{l_{\text{int}}} (\bullet)_j , \tag{37}$$

have been used to numerically perform said integration and calculate the 3D collagen-related stress, where l_{int} denotes the total number of spherical integration points.

4.2.3 Cessation of damage development in localised finite elements

Upon the initiation of a localisation in the continuum bulk material, arising from the significant accrual of collagen fiber damage, fracture formation begins, and all subsequent irreversible deformations are henceforth to be entirely represented by the cohesive traction model. Accordingly, once an eigenvalue λ_i of \mathcal{A} becomes negative, any further development of damage in the continuum model for the localised element is prevented, ultimately forcing the accumulation of damage in the cohesive zone and thus the propagation of the fracture surface. All algorithmic details concerning the identification and specification of localization are listed in Table 2.

However, in limited circumstances, there may also exist finite elements whose Gauss points meet the aforementioned localisation criterion, but they are not located ahead of the crack tip, and therefore the problem inhibits the initialisation of the cohesive zone model—the crack-tracking algorithm only permits the placement of cohesive zones in those elements situated at the crack tip. Therefore, to prevent the formation of a localised deformation, the further development of damage in the continuum model is also prevented in these elements.

4.3 Numerical implementation of the transversely isotropic cohesive zone model

The cohesive model is implemented in a user material routine, where the input data consists of the gap displacement $\mathbf{u}(\mathbf{x}_d^l)$ and normal vector $\mathbf{n}(\mathbf{x}_d^l)$ at the current time point. Additionally, the peak traction \mathbf{T}_0 as well as the damage variable $\delta(\mathbf{x}_d^l)$ from the previous time point are available, with \mathbf{x}_d^l denoting the spatial coordinate of the l th integration point of the discontinuity.

Table 1 Numerical implementation of the continuum damage model, detailing the determination of the Cauchy stress and the spatial stiffness at the Gauss point level of a partition of unity finite element*Initialization at time $t=0$*

Define the integration weight $w = 4\pi/l_{int}$ of a spherical integration point
 DO $j = 1, l_{int}$ % Loop over spherical integration points
 Define referential fiber direction \mathbf{M}_j and collagen fiber density $\rho_j = \rho(\mathbf{M}_j)$
 Define initial value of damage variable $\delta_j = 0$ and initialize history vector $\mathcal{H}_{j,1}^{con} \leftarrow \delta_j$
 DO $i = 1, n_f$ % Loop over CFPG-complexes
 Define straightening stretch λ_{st_i} according to Eq. (A3)
 Define initial PG stretch $\lambda_{pg_i} = 1$ and initialize history vector $\mathcal{H}_{j,i+1}^{con} \leftarrow \lambda_{pg_i}$
 ENDDO
 ENDDO

Algorithm at time t

Given: isochoric part of the deformation gradient $\bar{\mathbf{F}}$
 Initialize stress and elasticity tensors $\bar{\sigma}_{coll} \leftarrow 0, \bar{c}_{coll} \leftarrow 0$
 DO $j = 1, l_{int}$ % Loop over spherical integration points
 Compute deformed fiber direction $\mathbf{m}_j = \bar{\mathbf{F}}\mathbf{M}_j$ and fiber stretch $\lambda_{jN} = |\mathbf{m}_j|$
 Read damage variable at previous time point from history vector $\delta_{jN-1} \leftarrow \mathcal{H}_{j,1N-1}^{con}$
 Initialize number of fibrils recruited $n_r = 0$
 DO $i = 1, n_f$ % Loop over CFPG-complexes
 Read PG stretch at previous time point from history vector $\lambda_{pg_{iN-1}} \leftarrow \mathcal{H}_{j,i+1N-1}^{con}$
 Compute CFPG-complex stress P_{f_i} according to Eq. (11)
 IF ($P_{f_i} > 0$) $n_r = i$
 Compute $\lambda_{pg_{iN}}$ according to Eq. (36) and $\lambda_{pg_{fail_i}}$ according to Eq. (14)
 Initialize the damage variable $\delta_{jN} = \delta_{jN-1}$
 IF ($i > 2$)
 IF ($\lambda_{pg_{i-1N}} > \lambda_{pg_{fail_{i-1}}}$) AND ($\lambda_{pg_{iN}} \leq \lambda_{pg_{fail_i}}$)
 Compute the damage variable δ_{jN} at fiber stretch λ_{jN} according to Eq's. (B4,B5)
 Account for prior damage according to $\delta_{jN} = \max\{\delta_{jN}, \delta_{jN-1}\}$
 ENDIF
 Update integration constant A according to Eq's. (B7,B8)
 ENDIF
 Store PG stretch in history vector $\mathcal{H}_{j,i+1N}^{con} \leftarrow \lambda_{pg_{iN}}$
 ENDDO
 Compute the second Piola–Kirchhoff stress S_j according to Eq's. (B9,B10)
 Compute stress factor $a = S_j$ and stiffness factor $b = (S_j + \lambda^2 \partial S_j / \partial \lambda^2) / \lambda^4$
 Map collagen stress and elasticity to continuum metric

$$\bar{\sigma}_{coll j} = \rho_j a \text{dev}(\mathbf{m} \otimes \mathbf{m})$$

$$\bar{c}_{coll j} = \rho_j b \text{dev}(\mathbf{m} \otimes \mathbf{m}) \otimes \text{dev}(\mathbf{m} \otimes \mathbf{m}) - 1/6(\mathbf{I} \otimes \bar{\sigma}_{coll j} + \bar{\sigma}_{coll j} \otimes \mathbf{I})$$
 Update collagen stress/elasticity tensors $\bar{\sigma}_{coll} \leftarrow \bar{\sigma}_{coll} + \bar{\sigma}_{coll j}, \bar{c}_{coll} \leftarrow \bar{c}_{coll} + \bar{c}_{coll j}$
 Store damage variable in history vector $\mathcal{H}_{j,i+1N}^{con} \leftarrow \delta_{jN}$
 ENDDO
 Multiply collagen stress and elasticity tensors by w : $\bar{\sigma}_{coll} \leftarrow w \bar{\sigma}_{coll}, \bar{c}_{coll} \leftarrow w \bar{c}_{coll}$
 Compute non collagenous and volumetric stress/elasticity contributions $\bar{\sigma}_{nH}, \bar{c}_{nH}, \sigma_{vol}, c_{vol}$
 Compute total tissue stress/elasticity σ, c according to Eq. (20)

The indices j and i relate to a fiber and CFPG-complex, respectively. The subscripts $(\bullet)_N$ and $(\bullet)_{N-1}$ relate to the current and previous time points, respectively

Table 2 Algorithm for identification and specification of localization conditions at the Gauss-point level of a finite element

```

(1) Initialize location mode  $loc_{mode} = null$ 
(2) Solve the eigenvalue problem  $(\mathcal{A} - \lambda \mathbf{I}) : \mathbf{v}$ , which gives  $i = 1, \dots, 9$  eigenvalues  $\lambda_i$  and
    eigenvectors  $\mathbf{v}_i$  of dimension 9
(3) Find the lowest eigenvalue  $\lambda_m$  with the corresponding eigenvector  $\mathbf{v}_m; m \in \{1, \dots, 9\}$ 
(4) IF ( $\lambda_m < 0$ ) % Localisation criteria satisfied
    (a) Load index table
         $m = 1 : i = 1; j = 1$ 
         $m = 2 : i = 2; j = 2$ 
         $m = 3 : i = 3; j = 3$ 
         $m = 4 : i = 1; j = 2$ 
         $m = 5 : i = 2; j = 3$ 
         $m = 6 : i = 1; j = 3$ 
         $m = 7 : i = 2; j = 1$ 
         $m = 8 : i = 3; j = 2$ 
         $m = 9 : i = 3; j = 1$ 
    (b) Set failure mode
        IF  $m \in \{1, 2, 3\}$ 
             $loc_{mode} = normal$ 
        ELSEIF  $m \in \{4, \dots, 9\}$ 
             $loc_{mode} = shear$ 
        ENDIF
    (c) Set referential normal  $\mathbf{N}_\sigma$  and shear  $\mathbf{M}_\sigma$  direction
         $N_{\sigma k} = v_{ik}; M_{\sigma k} = v_{jk}; k = 1, 2, 3$ 
    (d) Compute referential peak normal traction
        IF ( $loc_{mode} = normal$ )
             $T_0 = \mathbf{N}_\sigma \cdot \mathbf{P}\mathbf{N}_\sigma$ 
        ELSEIF  $loc_{mode} = shear$ 
             $T_0 = (\mathbf{M}_\sigma \cdot \mathbf{P}\mathbf{N}_\sigma) / \beta$ 
        ENDIF
    ENDIF
ENDIF
    
```

The backward Euler discretization of Eq. (24) yields the iteration $\delta_N = (\delta_{N-1} + |\mathbf{u}|\xi_N)/(1 + \xi_N)$ with $\xi_N = \Delta t / \eta_{cz}$ for the damage variable, where δ_{N-1} and Δt denote the damage state at the previous time point and the time increment, respectively. The stiffness factor κ may then be computed by virtue of Eq. (22), and the model’s cohesive traction vector \mathbf{T} and the stiffness quantities $\mathbf{C}_u, \mathbf{C}_n, \mathbf{C}_\delta$ determined according to Eqs. (25, 26), respectively. The detailed algorithm is listed in Table 3.

In order to prevent the occurrence of an infinite scalar elastic stiffness κ that would result in the case of $\delta = 0$, as per Eq. (22), the damage variable is initialised with a value of $1/\epsilon$, where ϵ is an arbitrarily large number. The elastic stiffness-based implementation ensures the compatibility of the traction field in the cohesive zone with the stress field in the continuum (bulk) material.

5 Application of framework to numerical examples

In order to demonstrate the ability of the proposed constitutive formulation to study soft-tissue failure, we investigate fracture in the medial layer of the aortic vessel wall. The tissue is described by the set of parameters listed in Table 4, which represents porcine aortic wall tissue previously tested in our lab, with the cohesive zone model parameters numerically derived directly from the softening behaviour of the continuum via the bottom-up approach laid out in [45].

We first apply the discussed computational framework to the simulation of the uniaxial tensile testing of a notched specimen loaded along the arterial wall’s axial direction until rupture. We then move our attention to the simulation of the symconCT fracture test [46], an experimental setup

Table 3 Numerical implementation of the transversely isotropic cohesive zone model, detailing the determination of the traction vector and associated stiffness entities at the Gauss integration point level of a PUFEM element

Initialization at time $t = 0$

Define initial damage $\delta = \epsilon$, with $\epsilon = 0.001$ mm

Initialise cohesive zone history vector $\mathcal{H}^{\text{dis}} \leftarrow \delta$

Algorithm at time t

Given: gap displacement \mathbf{u} and normal of discontinuity \mathbf{n}

Compute normal displacement $\mathbf{u}_n = (\mathbf{u} \cdot \mathbf{n})\mathbf{n}$ and shear displacement $\mathbf{u}_s = \mathbf{u} - \mathbf{u}_n$

Import prior damage from history $\delta_{j_{N-1}} \leftarrow \mathcal{H}^{\text{dis}}$

Compute damage criterion $\phi = |\mathbf{u}| - \delta_{N-1}$

IF $\phi < 0$ % No further damage accumulates

Current damage is equivalent to prior damage $\delta_N = \delta_{N-1}$

Compute κ according to Eq. (22) and set the damage-related stiffness $\gamma = 0$

ELSEIF $\phi > 0$ % Further damage accumulates

Update damage state $\delta_N = (\delta_{N-1} + |\mathbf{u}|\xi_N)/(1 + \xi_N)$ with $\xi_N = \Delta t/\eta_{cz}$

Compute κ according to Eq. (22) and the scalar damage-related stiffness $\gamma = \partial\kappa/\partial\delta$

ENDIF

Store damage variable in history vector $\mathcal{H}^{\text{dis}} \leftarrow \delta_N$

Compute traction vector \mathbf{T} according to Eq. (25)

Compute stiffness quantities \mathbf{C}_u , \mathbf{C}_n , \mathbf{C}_δ according to Eq. (26)

The subscripts $(\bullet)_N$ and $(\bullet)_{N-1}$ relate to the current and previous time points, respectively

conceived such that a more controlled fracture may take place, and in doing so, microstructural failure and damage mechanisms be more readily investigated. The quasi-static monolithic solution of the system (33) was computed, and all simulations were carried out on a DELL Precision 7560 Notepad with an i9 Intel processor. The fracture is forced to initiate in the middle of the notch and then propagates according to the method described previously.

5.1 Uniaxial tensile rupture test

A $30 \times 24 \times 1.0$ mm³ sized patch of tissue was discretised into 560 linear tetrahedral elements, see Fig. 4 (left). As the prescribed traction separation law properties result in a cohesive length of approximately 2 mm, the model represents an overly coarse discretisation of the geometry, with the primary aim of evaluating the numerical robustness and capability of the PUFEM model. Dirichlet boundary conditions were applied perpendicular to the notch, in the axial direction, at the two outer edges, and a displacement rate of 10.0 mm s^{-1} was prescribed. The monolithic solution of the system (33) was computed and differing magnitudes of load increment were investigated. The resulting impact on the mechanical response is demonstrated in Fig. 4 (right).

The large element size paired with the small cohesive length yields a somewhat scattered force response, which did not pose any issues concerning numerical stability. As the first principal stress direction and the direction of the least in-plane collagen fiber coincide for this problem, the fracture

propagates along a straight line from the notch towards the opposite edge of the specimen.

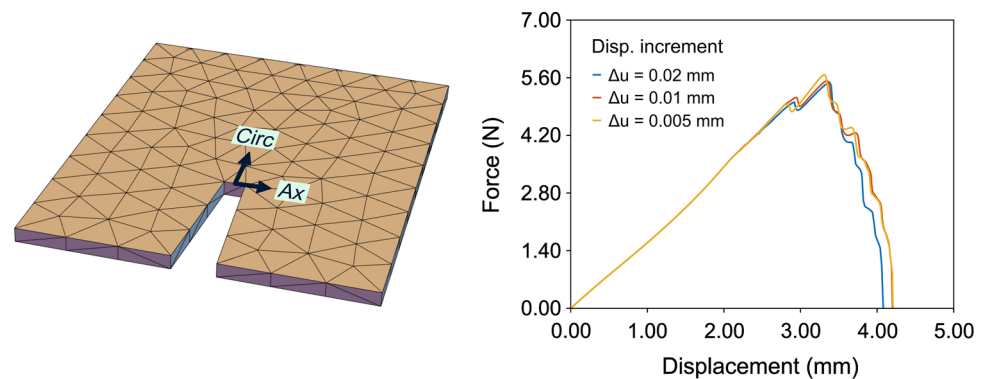
5.2 Symmetry-constrained compact tension test

The same 3D domain used in Sect. 5.1 was subsequently discretized into approximately 11k linear tetrahedral elements and then tightly linked to two rigid bodies, mimicking the clamps of the symconCT ex-vivo tissue test. As in the symconCT laboratory experiment, the specimen was first pre-stretched, undergoing an initial loading sequence where the distance between the hinge points of the clamps was increased. Subsequently, during the primary loading phase, these points were kept fixed in space, and the displacement of the loading points were increased at a rate of 3 mm/min.

Tissue fracture was studied for the loading of the tissue specimen in both the axial and circumferential directions. Whilst the specimen in the case of axial loading was completely fractured following 123 load steps, when exposed to loading in the circumferential direction, the crack ran into the clamps (rigid body) after 85 loading steps. With the computer system used, the simulation of this problem took approximately 20 and 15 min, respectively. Figure 5 displays a contour plot of the von Mises stress for the axial loading case approximately halfway through the fractured symconCT test, whilst Fig. 6 (left) and (right) compare the simulated load–displacement curves and the alternate fracture paths resulting for specimens loaded in the axial and circumferential directions, respectively.

Table 4 Material and structural parameters representing the medial layer of the porcine aorta

Matrix material		
Neo-Hookean parameter	μ	30 kPa
Collagen fibers		
<i>CFPG-complex structural parameters</i>		
Collagen fibril stiffness	k_f	100 MPa
Collagen fibril target homeostatic stress	T_0	0.5 MPa
Rate of PG sliding	η_{slid}	$8e2 \text{ s}^{-1}$
Rate of PG recovery	η_{rec}	$2e1 \text{ s}^{-1}$
<i>CFPG-complex failure parameters</i>		
Damage parameter 1	α	-0.09
Damage parameter 2	β	1.1
<i>Recruitment parameters</i>		
Lower limit	λ_A	1.05
Mode	λ_B	1.25
Upper limit	λ_C	1.45
<i>Discretization parameters</i>		
Number of CFPG-complexes	n_f	15
Collagen fiber orientation distribution		
<i>Distribution parameters</i>		
Bingham concentration parameter 1	κ_1	11.7
Bingham concentration parameter 2	κ_2	9.6
<i>Numerical spherical integration parameters</i>		
Number of spherical integration points	l_{int}	120
Cohesive zone model parameters		
Shape parameter	a	0.7435
Shape parameter	b	9.9064
Shape parameter	c	1.8404
Shape parameter	d	1.6935
Shape parameter	e	1.2540
Integral shape parameter	ζ	0.34683
Mode I fracture energy	G	0.50958 Nmm/mm^2
Normal and shear coupling parameter	β	0.8
Rate parameter	η_{cz}	0.01 or 1.0 s^{-1}

Fig. 4 Results for the uniaxial rupture test. A monotonically increasing axial displacement was applied at a rate of 10.0 ms^{-1} , (left) coarse discretisation of the tissue sample, (right) influence of the displacement increment on the mechanical response, with convergence realised at sufficiently small increments


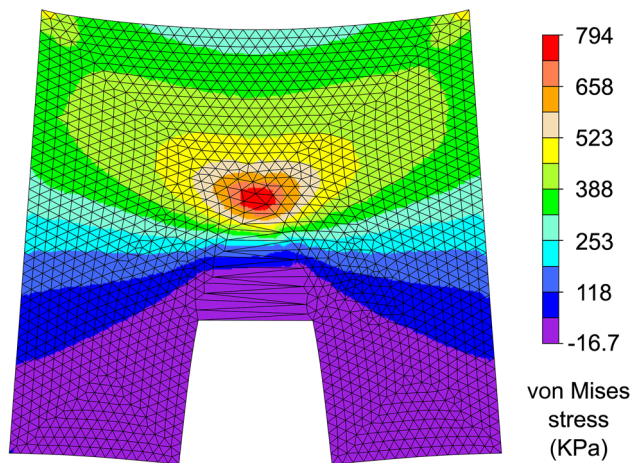


Fig. 5 von Mises stress contour plot for the sample loaded in the axial direction once the crack has propagated approximately halfway through the length of the specimen. A stress concentration ahead of the crack tip is clearly visible, as is the significant decline in stress for those PUFEM elements that have localised and a cohesive zone is present

6 Conclusion

The failure of vascular tissue, occurring from traumatic injury or the presentation of disease, is an issue of undoubted clinical relevance. For instance, the rupture or ulceration of the fibrous cap that characterises atherosclerotic lesions exposes flowing blood to highly thrombogenic plaque contents and the potential incidence of an acute thromboembolic event. Likewise, the rupture of abdominal aortic aneurysms (AAA) carries a demonstrable risk of mortality due to internal bleeding. However, the underlying structure-function relationships that govern both normal and pathological mechanical functionality, and ultimately the fracture properties of vascular tissue, are as yet unknown. To date, limited efforts have been taken to divulge the histo-mechanical mechanisms associ-

ated with damage development and rupture of the arterial wall, whether it be experimentally or computationally. The biomechanical simulation of fracture in load-bearing soft biological tissues, in particular, represents a uniquely challenging endeavour from a numerical standpoint.

The present study combines non-linear continuum mechanical and discontinuous cohesive zone frameworks within a robust and efficient PUFEM formulation that can effectively characterise 3D crack propagation. The continuum model incorporates time-dependent interfibrillar damage phenomena, motivated by the microstructural architecture of the collagenous ECM, whilst the response of the (phenomenological) transversely isotropic cohesive zone model is drawn directly from that of the continuum model. Significantly, the outlined traction separation law can effectively characterise mixed-mode fracture. Shear loading may influence failure processes in soft biological tissues, so it is imperative that such features are incorporated when seeking to model rupture and its associated processes. Given our cohesive description, the parameter β determines the coupling between normal and shear loading at the interface. As indicated by the single finite element response at softening, a constant β has been used. It implies that the structures that support normal and shear load transitioning in the failure process zone diminish homogeneously with the accumulation of damage δ .

To the best of the authors' knowledge, this is the first instance in which a 3D finite-strain continuum damage formulation has been coupled with an embedded representation of the fracture surface. Namely, when the development of collagen fiber damage in the continuum material gives rise to a localised deformation, a discontinuous cohesive zone is embedded within the finite element of interest and all further evolution of damage in the continuum material is halted. All subsequent irreversible deformations are thus occupied by the cohesive zone, leading to the separation of the crack face

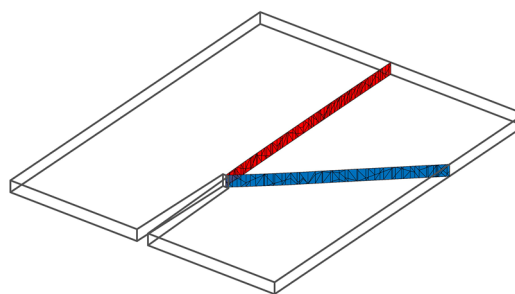
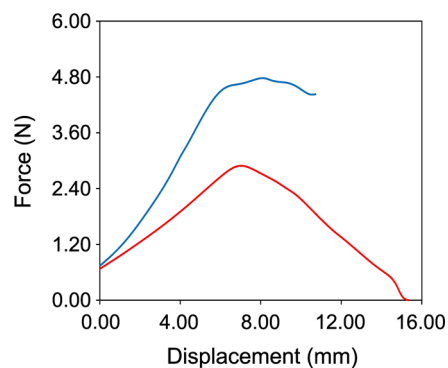


Fig. 6 Results for symconCT testing, demonstrating the impact of sample orientation relative to loading direction and notch placement, (left) the mechanical response following the pre-stretching of the specimen, with red and blue denoting loading in the axial and circumferential

directions, respectively, (right) the associated fracture paths, also displaying the facets formed when a discontinuity presents in a PUFEM element. Fracture paths are shown in the unloaded reference configuration. (Color figure online)

in the PUFEM element. The localisation criteria, and specifically the definition of the normal to the fracture surface, is of paramount importance to numerical simulations that employ embedded discontinuities. A myriad of contributing factors likely influence the formation of fracture in load-bearing soft biological tissues. For instance, the orientation of the discontinuity may be a function of the stress state, the rate of stress, and the material's microstructural organisation, with the true origin and impact of such considerations difficult to fully establish. Equation (30) may therefore be seen as the first attempt to address this problem, and the amassment of experimental data from individual tissues would likely result in more adequate definitions.

The discussed computational framework was able to qualitatively capture fracture mechanical behaviour for the explored representative numerical examples, with stable failure propagation negating the need for the introduction of inertial effects through transient solution methods. The application of the model to the uniaxial tensile rupture test aptly showcased its versatility, with the reliable simulation of tissue failure for a course mesh size and relatively large load increments. The simulation of the symconCT example well-replicated the general experimental observations for samples loaded in the axial direction, with the successful demonstration of a centralised crack trajectory and the stable/controlled decline of the force following the inception of fracture. Furthermore, as our framework is generic, it is likely that fracture in tissue types other than the vascular wall could also be analyzed.

However, in the case of samples loaded in the circumferential direction, disagreement between the simulation and experimental observations was evident. Whilst the numerical simulation could effectively reproduce the initial orientation of the crack (a consequence of the initialised normal definition), it was unable to recreate the 'zig-zag' pattern witnessed experimentally, see [46]. From a histological and microstructural perspective, this may stem from the repeated branching of the crack and the subsequent arrest of the remaining mother crack due to unknown failure mechanisms. The numerical formulation, as it currently stands, is limited to the presence of a single cohesive zone within a given PUFEM element, and so cannot suitably mimic the postulated phenomena. It is also worth noting that in the case of the symconCT testing of aneurysmatic aorta specimens loaded in the circumferential direction, the crack frequently propagates into the clamps, see Fig. 7. This observation stems from our ongoing experimental study concerning human thoracic aortas and matches with the PUFEM simulations predicted in this work. The diseased human aorta is particularly fibrous, histology that then promotes fracture in between collagen fibers.

Many of the recent modelling strategies used to characterise fracture in soft biological tissues have focussed on the phase field method [15, 28, 34]. It can handle topologi-

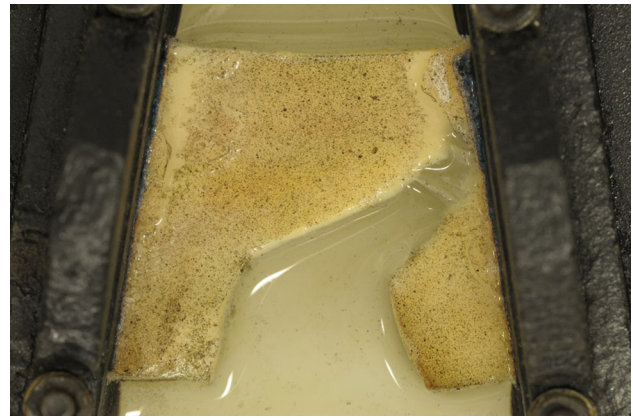


Fig. 7 Symmetry-constrained compact tension (symconCT) test of the aneurysmatic thoracic human aorta loaded along the circumferential direction. The experimentally provoked fracture path matches that predicted by the PUFEM method, as shown in Fig. 6

cally complex fractures, such as branching and intersecting cracks, without the need to explicitly trace the fracture surface, circumventing many of the numerical complexities of crack tracking. However, it does not allow full access to the constitutive description of fracture, and it is therefore not necessarily ideal with respect to designing and validating fracture laws. Additionally, the explicit integration of physical aspects, such as nonlinearities and the time dependencies associated with fracture processes, are not well understood.

Acknowledgements This research has been supported by the project Grant 2015-04476 from the Swedish Research Council (VR). We also thank Marta Alloisio, KTH Royal Institute of Technology, who provided the image shown in Fig. 7.

Funding Open access funding provided by Royal Institute of Technology.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

Appendix A: Straightening stretch probability distribution

Fibril straightening is characterised by the triangular probability density function. As such, the piecewise expression

$$F_{\lambda_{st}}(\lambda_{st}) = \begin{cases} \frac{(\lambda_{st} - \lambda_A)^2}{(\lambda_C - \lambda_A)(\lambda_B - \lambda_A)} ; & \lambda_A \leq \lambda_{st} \leq \lambda_B \\ 1 - \frac{(\lambda_C - \lambda_{st})^2}{(\lambda_C - \lambda_A)(\lambda_C - \lambda_B)} ; & \lambda_B < \lambda_{st} \leq \lambda_C \end{cases} \tag{A1}$$

defines the cumulative density function for the triangular distribution, with λ_A and λ_C corresponding to the collagen fiber stretches where the first and last fibril are initially recruited to load bearing, respectively, and λ_B is the mode of the distribution. Consequently, the piecewise expression

$$F_{\lambda_{st}}^{-1}(p) = \begin{cases} \lambda_A + \sqrt{(\lambda_C - \lambda_A)(\lambda_B - \lambda_A)p} ; & 0 \leq p \leq F_{\lambda_{st}}(\lambda_B) \\ \lambda_C - \sqrt{(\lambda_C - \lambda_A)(\lambda_B - \lambda_A)(1 - p)} ; & F_{\lambda_{st}}(\lambda_B) < p \leq 1 \end{cases} \tag{A2}$$

denotes the associated inverse cumulative density function for the triangular distribution. Finally, the relation

$$\lambda_{sti} = F_{\lambda_{st}}^{-1}\left(\frac{i - 1}{n_f - 1}\right) ; 1 \leq i \leq n_f \tag{A3}$$

then defines the generated straightening stretch λ_{sti} for the i th CFPG-complex via inverse transform sampling, with n_f denoting the total number discrete CFPG-complexes considered.

Appendix B: Collagen fiber stress and stiffness determination

A full accounting of the collagen fiber stress and stiffness determination is provided, with the inclusion of details that were omitted from the main text. For a thorough and exhaustive explanation of the constitutive descriptions intricate details, the reader is referred to the original publication [45].

B.1 Proportion of broken CFPG-complexes

As a finite number of CFPG-complexes are considered, in order to determine δ , it is necessary to interpolate the space between neighboring discrete CFPG-complexes. To achieve this, we linearly approximate the relationship between λ_{pg} and λ_{st} across the interpolated space, through the set of expressions

$$\lambda_{pg} = m\lambda_{st} + c, \quad m = \frac{\lambda_{sti} - \lambda_{sti-1}}{\lambda_{pgi} - \lambda_{pgi-1}}, \quad c = \lambda_{pgi} - m\lambda_{sti} \tag{B4}$$

If the limiting CFPG-complex at which failure occurs is present within the interpolated space i.e. $\lambda_{pgi-1} > \lambda_{pgfaili-1}$ and $\lambda_{pgi} \leq \lambda_{pgfaili}$, then through application of Eq's (10,11,B4) to Eq (14), we arrive at the function

$$g(\lambda_{st}) = \frac{\alpha}{P_0} \left[k_f \left(\frac{\lambda}{\lambda_{st}(m\lambda_{st} + c)} - 1 \right) - P_0 \right] - \beta - (m\lambda_{st} + c) = 0. \tag{B5}$$

It can be solved by means of the Newton method to provide the straightening stretch corresponding to the limiting CFPG-complex at which failure has occurred. The expression, $\delta = F_{\lambda_{st}}(\lambda_{st})$, then provides the proportion of failed CFPG-complexes, i.e. the damage of the collagen fiber.

B.2 Stress of a collagen fiber—integration over all CFPG-complexes

A linear interpolation among the n_f discrete CFPG-complexes, with respect to the squared fiber stretch, may be used to approximate the percentage of load-bearing CFPG-complexes, with the damage variable δ being used to omit those CFPG-complexes that have failed and therefore no longer contribute towards load-bearing. The collagen fiber stiffness can be approximated through analytical functions, and following subsequent integration, the stress can be obtained. The general expressions for the stiffness and stress of a collagen fiber are derived subsequently.

Given $n_r \in \{0, n_f\} \cap \mathbb{Z}$ number of discrete CFPG-complexes currently recruited to load bearing, i.e. those where $P_f > 0$, the fiber's referential stiffness reads

$$C = \begin{cases} 0 ; & n_r = 0 \\ \max \left\{ \frac{k_f}{\lambda^3} \left[\gamma_{n_r} + \frac{\lambda^2 - \bar{\lambda}_{n_r}^2}{\bar{\lambda}_{n_r+1}^2 - \bar{\lambda}_{n_r}^2} - \delta \right], 0 \right\} ; & 0 < n_r < n_f \\ \frac{k_f}{\lambda^3} [1 - \delta] ; & n_r = n_f \end{cases} \tag{B6}$$

where $\gamma_{n_r} = F_{\lambda_{st}}(\lambda_{st n_r})$, $\bar{\lambda}_{n_r} = \lambda_{st n_r} \lambda_{pg n_r}$ and $\bar{\lambda}_{n_r+1} = \lambda_{st n_r+1} \lambda_{pg n_r+1}$ have been used. This expression can then be integrated over all CFPG-complexes currently recruited to load-bearing according to

$$A_i = \begin{cases} 0 ; n_r = 0 \\ \frac{2k_f(\bar{\lambda}_i - \bar{\lambda}_{i-1})}{\bar{\lambda}_i \bar{\lambda}_{i-1}} \left[(\gamma_{i-1} - \delta) + \frac{\bar{\lambda}_i \bar{\lambda}_{i-1} - \bar{\lambda}_{i-1}^2}{\bar{\lambda}_i^2 - \bar{\lambda}_{i-1}^2} \right] ; & \delta \leq \gamma_{i-1} \\ \frac{2k_f(\bar{\lambda}_i - \bar{\lambda}_d)}{\bar{\lambda}_i \bar{\lambda}_d} \left[(\gamma_{i-1} - \delta) + \frac{\bar{\lambda}_i \bar{\lambda}_d - \bar{\lambda}_{i-1}^2}{\bar{\lambda}_i^2 - \bar{\lambda}_{i-1}^2} \right] ; & \gamma_{i-1} < \delta < \gamma_i \\ 0 ; \delta > \gamma_i \end{cases} \tag{B7}$$

where $\gamma_{i-1} = F_{\lambda_{st}}(\lambda_{st i-1})$, $\bar{\lambda}_i = \lambda_{st i} \lambda_{pg i}$ and $\bar{\lambda}_{i-1} = \lambda_{st i-1} \lambda_{pg i-1}$ have been used. Furthermore, $\bar{\lambda}_d$, relates to the altered lower bound of integration due to damage and is given by

$$\bar{\lambda}_d = \left[\bar{\lambda}_{i-1}^2 + (\delta - \gamma_{i-1})(\bar{\lambda}_i^2 - \bar{\lambda}_{i-1}^2) \right]^{1/2}. \tag{B8}$$

The total integration constant is then defined by $A = \sum_{i=2}^{n_r} A_i$. The second Piola–Kirchhoff stress S of a collagen fiber is determined through the subsequent addition of A to the integral over the remaining portion of the interpolated stiffness function, leading to the piecewise expression

$$S = \begin{cases} 0; & n_r = 0 \\ A + \frac{2k_f(\lambda - \bar{\lambda}_{n_r})}{\lambda \bar{\lambda}_{n_r}} \left[(\gamma_{i-1} - \delta) + \frac{\lambda \bar{\lambda}_{n_r} - \bar{\lambda}_{n_r}^2}{\bar{\lambda}_{n_r+1}^2 - \bar{\lambda}_{n_r}^2} \right]; & 0 > n_r > n_f, \delta \leq \gamma_{n_r} \\ A + \frac{2k_f(\lambda - \bar{\lambda}_d)}{\lambda \bar{\lambda}_d} \left[(\gamma_{i-1} - \delta) + \frac{\lambda \bar{\lambda}_d - \bar{\lambda}_{n_r}^2}{\bar{\lambda}_{n_r+1}^2 - \bar{\lambda}_{n_r}^2} \right]; & 0 > n_r > n_f, \gamma_{n_r} < \delta \leq \gamma_{n_r+1}, \lambda \leq \bar{\lambda}_d \\ 0; & 0 > n_r > n_f, \gamma_{n_r} < \delta \leq \gamma_{n_r+1}, \lambda > \bar{\lambda}_d \\ 0; & 0 > n_r > n_f, \delta > \gamma_{n_r+1} \\ A + 2k_f(1 - \delta) \left(\frac{1}{\bar{\lambda}_{n_f}} - \frac{1}{\lambda} \right); & n_r = n_f, \delta < 1 \\ 0; & \delta = 1, \end{cases} \tag{B9}$$

where $\bar{\lambda}_d$ once again relates to the altered lower bound of integration due to damage, and in this case is given by

$$\bar{\lambda}_d = \left[\bar{\lambda}_{n_r}^2 + (\delta - \gamma_{i-1})(\bar{\lambda}_{n_r+1}^2 - \bar{\lambda}_{n_r}^2) \right]^{1/2}. \tag{B10}$$

References

1. Wilkins E, Wilson L, Wickramasinghe K, Bhatnagar P, Leal J, Luengo-Fernandez R, Burns R, Rayner M, Townsend N (2017) European cardiovascular disease statistics 2017. European Heart Network, Belgium
2. Bäck M, Gasser TC, Michel J-B, Caligiuri G (2013) Review. Biomechanical factors in the biology of aortic wall and aortic valve diseases. *Cardiovasc Res* 99:232–241
3. Ohayon J, Finet G, Gharib AM, Herzka DA, Tracqui P, Heroux J, Rioufol G, Kotys MS, Elagha A, Pettigrew RI (2008) Necrotic core thickness and positive arterial remodeling index: emergent biomechanical factors for evaluating the risk of plaque rupture. *Am J Physiol Heart Circ Physiol* 295:717–727
4. Karlöf E, Seime T, Dias N, Lengquist M, Witasp A, Almqvist H, Kronqvist M, Gådin JR, Odeberg J, Maegdefessel L, Stenvinkel P, Matic LP, Hedin U (2019) Correlation of computed tomography with carotid plaque transcriptomes associates calcification with lesion-stabilization. *Atherosclerosis* 288:175–185
5. Maier A, Essler M, Gee MW, Eckstein HH, Wall WA, Reeps C (2012) Correlation of biomechanics to tissue reac-

- tion in aortic aneurysms assessed by finite elements and [18f]-fluorodeoxyglucose-PET/CT. *Int J Numer Methods Biomed Eng* 28:456–471
6. Erhart P, Grond-Ginsbach C, Hakimi M, Lasitschka F, Dihlmann S, Böckler D, Hyhlik-Dürr A (2014) Finite element analysis of abdominal aortic aneurysms: predicted rupture risk correlates with aortic wall histology in individual patients. *J Endovasc Ther* 21:556–564
7. Vashishth D, Tanner KE, Bonfield W (2003) Experimental validation of a microcracking-based toughening mechanism for cortical bone. *J Biomech* 36(1):121–124
8. Yang QD, Cox BN, Nalla RK, Ritchie RO (2006) Re-evaluating the toughness of human cortical bone. *Bone* 38(6):878–887
9. Li S, Abdel-Wahab A, Silberschmidt VV (2013) Analysis of fracture processes in cortical bone tissue. *Eng Fract Mech* 110:448–458
10. Kataruka A, Mendu K, Okeoghene O, Puthuvelil J, Akono AT (2017) Microscopic assessment of bone toughness using scratch tests. *Bone Rep* 6:17–25

11. Yang W, Sherman VR, Gludovatz B, Schaible E, Stewart P, Ritchie RO, Meyers MA (2015) On the tear resistance of skin. *Nat Commun* 6:6649
12. Pissarenko A, Yang W, Quan H, Poyer B, Williams A, Brown KA, Meyers MA (2020) The toughness of porcine skin: quantitative measurements and microstructural characterization. *J Mech Behav Biomed Mater* 109:103848
13. Roach MR, He JC, Kratky RG (1999) Tear propagation in isolated, pressurized porcine thoracic aortas. *Can J Cardiol* 15:569–575
14. Sommer G, Gasser TC, Regitnig P, Auer M, Holzapfel GA (2008) Dissection of the human aortic media: an experimental study. *J Biomech Eng* 130:12
15. Ban E, Cavinato C, Humphrey JD (2022) Critical pressure of intramural delamination in aortic dissection. *Ann Biomed Eng* 50:183–194
16. Gasser TC, Gudmundson P, Dohr G (2009) Failure mechanisms of ventricular tissue due to deep penetration. *J Biomech* 42:626–633
17. Forsell C, Gasser TC (2011) Numerical simulation of the failure of ventricular tissue due to deep penetration: the impact of constitutive properties. *J Biomech* 44:45–51
18. Gasser TC (2011) An irreversible constitutive model for fibrous soft biological tissue: a 3d microfiber approach with demonstrative application to abdominal aortic aneurysms. *Acta Biomater* 7:2457–2466
19. Balzani D, Brinkhues S, Holzapfel GA (2012) Constitutive framework for the modeling of damage in collagenous soft tissues with application to arterial walls. *Comput Methods Appl Mech Eng* 213–216:139–151

20. Holzapfel GA, Ogden RW (2020) An arterial constitutive model accounting for collagen content and cross-linking. *J Mech Phys Solids* 136:103682
21. Linka K, Hillgärtner M, Itskov M (2018) Fatigue of soft fibrous tissues: multi-scale mechanics and constitutive modeling. *Acta Biomater* 71:398–410
22. Hamedzadeh A, Gasser TC, Federico S (2018) On the constitutive modelling of recruitment and damage of collagen fibres in soft biological tissues. *Eur J Mech A/Solids* 72:483–496
23. Zitnay JL, Jung GS, Lin AH, Qin Z, Li Y, Yu SM, Buehler MJ, Weiss JA (2020) Accumulation of collagen molecular unfolding is the mechanism of cyclic fatigue damage and failure in collagenous tissues. *Sci Adv* 6(35):2795
24. Gregory J, Hazel AL, Shearer T (2021) A microstructural model of tendon failure. *J Mech Behav Biomed Mater* 122:104665
25. Gasser TC, Holzapfel GA (2006) Modeling the propagation of arterial dissection. *Eur J Mech A/Solids* 25:617–633
26. Gasser TC, Holzapfel GA (2007) Modeling dissection failure during balloon angioplasty. *Ann Biomed Eng* 35:711–723
27. Ferrara A, Pandolfi A (2010) A numerical study of arterial media dissection processes. *Int J Fract* 8:435–540
28. Gültekin O, Dal H, Holzapfel GA (2016) A phase-field approach to model fracture of arterial walls: theory and finite element analysis. *Comput Methods Appl Mech Eng* 312:542–566
29. Raina A, Miehe C (2016) A phase-field model for fracture in biological tissues. *Biomech Model Mechanobiol* 15:479–496
30. Nagaraja S, Leichsenring K, Ambati M, De Lorenzis L, Boel M (2021) On a phase-field approach to model fracture of small intestine walls. *Acta Biomater* 130:317–331
31. Alloisio M, Gasser TC (2023) Fracture properties of the porcine aorta. Part 2: Fem modeling and inverse parameter identification. *Acta Biomater*
32. Peerlings RHJ, Borst RD, Brekelmans WAM, Vree JHPD (1996) Gradient enhanced damage for quasi-brittle materials. *Int J Numer Methods Eng* 39(19):3391–3403
33. Bažant ZP, Jirásek M (2002) Nonlocal integral formulations of plasticity and damage: survey of progress. *J Eng Mech* 128(11):1119–1149
34. Miehe C, Hofacker M, Welschinger F (2010) A phase field model for rate-independent crack propagation: robust algorithmic implementation based on operator splits. *Comput Methods Appl Mech Eng* 199(45):2765–2778
35. Jirásek M, Bauer M (2012) Numerical aspects of the crack band approach. *Comput Struct* 110–111:60–78
36. Bažant ZP, Pijaudier-Cabot G (1988) Nonlocal continuum damage, localization instability and convergence. *J Appl Mech* 55:287–293
37. Ortiz M, Leroy Y, Needleman A (1987) A finite element method for localized failure analysis. *Comput Methods Appl Mech Eng* 61:189–214
38. Moës N, Dolbow J, Belytschko T (1999) A finite element method for crack growth without remeshing. *Int J Numer Methods Eng* 46(1):131–150
39. Gasser TC, Holzapfel GA (2005) Modeling 3D crack propagation in unreinforced concrete using PUFEM. *Comput Methods Appl Mech Eng* 194:2859–2896
40. Chittajallu SNSH, Richhariya A, Tse KM, Chinthapenta V (2022) A review on damage and rupture modelling for soft tissues. *Bio-engineering* 9(1):26
41. Bircher K, Zündel M, Pensalfini M, Ehret AE, Mazza E (2019) Tear resistance of soft collagenous tissues. *Nat Commun* 12(1):792
42. Ogden RW (2000) Elastic and pseudo-elastic instability and bifurcation. In: Petryk H (ed) *Material instabilities in elastic and plastic solids*. Springer, Vienna, pp 209–259
43. Bigoni D (2012) Cambridge University Press
44. Gasser TC, Holzapfel GA (2006) 3D crack propagation in unreinforced concrete. A new smoothing algorithm for tracking 3D crack surfaces. *Comput Methods Appl Mech Eng* 195:5198–5219
45. Miller C, Gasser TC (2022) A bottom-up approach to model collagen fiber damage and failure in soft biological tissues. *J Mech Phys Solids* 169:105086
46. Alloisio M, Chatziefraimidou M, Roy J, Gasser TC (2023) Fracture of porcine aorta-part 1: symconct fracture testing and dic. *Acta Biomater*
47. Wells GN, de Borst R, Sluys LJ (2002) A consistent geometrically non-linear approach for delamination. *Int J Numer Methods Eng* 54:1333–1355
48. Wells GN, Sluys LJ (2001) Three-dimensional embedded discontinuity model for brittle fracture. *Int J Solids Struct* 38:897–913
49. Ogden RW (1978) Nearly isochoric elastic deformations: application to rubberlike solids. *J Mech Phys Solids* 26:37–57
50. Simo JC, Taylor RL (1991) Quasi-incompressible finite elasticity in principal stretches. Continuum basis and numerical algorithms. *Comput Methods Appl Mech Eng* 85:273–310
51. Ortiz M, Pandolfi A (1999) Finite-deformation irreversible cohesive elements for three-dimensional crack-propagation analysis. *Int J Numer Methods Eng* 44:1267–1282
52. Coleman BD, Noll W (1963) The thermodynamics of elastic materials with heat conduction and viscosity. *Arch Ration Mech Anal* 13:167–178
53. Melenk JM, Babuška I (1996) The partition of unity finite element method: basic theory and applications. *Comput Methods Appl Mech Eng* 139:289–314
54. Miller C, Gasser TC (2021) A microstructurally motivated constitutive description of collagenous soft biological tissue towards the description of their non-linear and time-dependent properties. *J Mech Phys Solids* 154:104500
55. Lanir Y (1983) Constitutive equations for fibrous connective tissues. *J Biomech* 16:1–12
56. Alastrué V, Saez P, Martínez MA, Doblaré M (2010) On the use of the Bingham statistical distribution in microsphere-based constitutive models for arterial tissue. *Mech Res Commun* 37:700–706
57. Gasser TC, Gallinetti S, Xing X, Forsell C, Swedenborg J, Roy J (2012) Spatial orientation of collagen fibers in the abdominal aortic aneurysm wall and its relation to wall mechanics. *Acta Biomater* 8:3091–3103
58. Spencer AJM (1984) Constitutive theory for strongly anisotropic solids. In: Spencer AJM (ed) *Continuum theory of the mechanics of fibre-reinforced composites*. CISM Courses and Lectures No. 282, International Centre for Mechanical Sciences. Springer, Wien, pp 1–32
59. Itskov M (2000) On the theory of fourth-order tensors and their applications in computational mechanics. *Comput Methods Appl Mech Eng* 189(2):419–438
60. Taylor RL (2007) FEAP—a finite element analysis program, version 8.2 user manual. University of California at Berkeley, Berkeley, California
61. Hardin RH, Sloane NJA (1996) McLaren’s improved snub cube and other new spherical designs in three dimensions. *Discrete Comput Geom* 15:429–441

Publisher’s Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.