



The microbial RNA metagenome of *Aedes albopictus* (Diptera: Culicidae) from Germany

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Received: 11 May 2022 / Accepted: 10 June 2022 / Published online: 20 July 2022
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Abstract

Aedes albopictus is a highly invasive mosquito species that has become widespread across the globe. In addition, it is an efficient vector of numerous pathogens of medical and veterinary importance, including dengue, chikungunya and Zika viruses. Among others, the vector potential of mosquitoes is influenced by their microbiome. However, this influence is very dynamic and can vary between individuals and life stages. To obtain a rough overview on the microbiome of *Ae. albopictus* populations in Germany, pooled female and pooled male individuals from seven German locations were investigated by total RNA sequencing. The mosquito specimens had been collected as larvae in the field and processed immediately after adult emergence, i.e. without females having fed on blood. RNA fragments with high degrees of identity to a large number of viruses and microorganisms were identified, including, for example, *Wolbachia pipientis* and *Acinetobacter baumannii*, with differences between male and female mosquitoes. Knowledge about the natural occurrence of microorganisms in mosquitoes may be translated into new approaches to vector control, for example *W. pipientis* can be exploited to manipulate mosquito reproduction and vector competence. The study results show how diverse the microbiome of *Ae. albopictus* can be, and the more so needs to be adequately analysed and interpreted.

Keywords *Aedes albopictus* · Microbiome · Germany · Vector control

Introduction

Aedes albopictus is a thermophilic mosquito species native to the Asian-Pacific region. Due to globalisation and its high ecological and physiological plasticity, it has become widespread in many regions in the world. Presently, *Ae. albopictus* is considered the most invasive mosquito species of the world (Benedict et al. 2007; Bonizzoni et al. 2013). Climate warming and the resulting mild winters favour the establishment, reproduction and spread of *Ae. albopictus* in temperate climes, such as Central Europe (e.g., Walther et al. 2017; Fălcută et al. 2020).

Aedes albopictus is highly vector-competent for numerous arboviruses, including dengue, chikungunya, yellow fever, Zika, West Nile and various encephalitis viruses (Paupy et al. 2009; Martinet et al. 2019). It thus has a major impact on human and veterinary health. The vector competence, i.e. the ability of a haematophagous arthropod to transmit a pathogen, can be influenced by the arthropod's microbiome (Engel and Moran 2013; Jupatanakul et al. 2014) which is defined by Berg et al. (2020) as 'a characteristic microbial community occupying a reasonable well-defined habitat which has distinct physio-chemical properties'.

It has been shown that the microbiome may have a general impact on the development, reproduction and physiology of an invertebrate (Minard et al. 2013; Coon et al. 2014, 2016, 2017). For example, the endosymbiont *Wolbachia pipientis* is known to be widely distributed in invertebrates (Yang 2000; Hilgenboecker et al. 2008). Bourtzis and O'Neill (1998) and Ahmad et al. (2017) have demonstrated that *W. pipientis* can affect both the reproduction of insects and the replication and dissemination of pathogenic viruses in an insect vector. These effects are major reasons why the study of the microbiome of mosquitoes has become so popular in

Handling Editor: Una Ryan

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recent years. However, the microbiome is not static, but may change during development and can be influenced by many factors such as sex, age and life stage of the host, geographic location, breeding habitat characteristics and food supply (Wang et al. 2011; Zouache et al. 2011; Boissière et al. 2012; Terenius et al. 2012; Jupatanakul et al. 2014; Chen et al. 2020). Water temperature and nutrient content of the breeding habitat, for example, can strongly influence its bacterial community and thus have an impact on the microbiome of developing mosquito larvae (Hörtnagl et al. 2010; Onchuru et al. 2016). In turn, the microbiome ingested from the breeding habitat may considerably influence larval growth and development (Coon et al. 2014, 2016, 2017).

Insects take up a variety of microorganisms from their environment (Strand 2018). In the case of mosquitoes, this mainly occurs in the larval stage, when individuals are confronted with large numbers of microorganisms in their aquatic habitats during feeding. Larval nutrition can therefore have a major impact on the composition of the microbiome (Wang et al. 2011; Boissiere et al. 2012; Coon et al. 2016). By contrast, occasions to take up microorganisms in the adult stage are limited: both sexes feed on sugary plant juices and only females feed on blood, with the latter occasionally facilitating the uptake of disease agents. There is evidence that the insect host can exert some control over its microbiome via the innate immune response (Douglas 2015; Smith et al. 2015).

In recent years, the microbiome of several mosquito species has been studied, among them *Ae. albopictus*, *Aedes japonicus*, *Anopheles gambiae* and *Culex pipiens*, with the focus of most studies being on the midgut microbiota of adult mosquitoes (Wang et al. 2011; Gimonneau et al. 2014). It turned out that the microbiome of some species is

extremely diverse, and a variety of bacterial phyla such as Actinobacteria, Proteobacteria, Bacteroidetes or Firmicutes could be detected (Moro et al. 2013; Zotzmann et al. 2017; Wang et al. 2018).

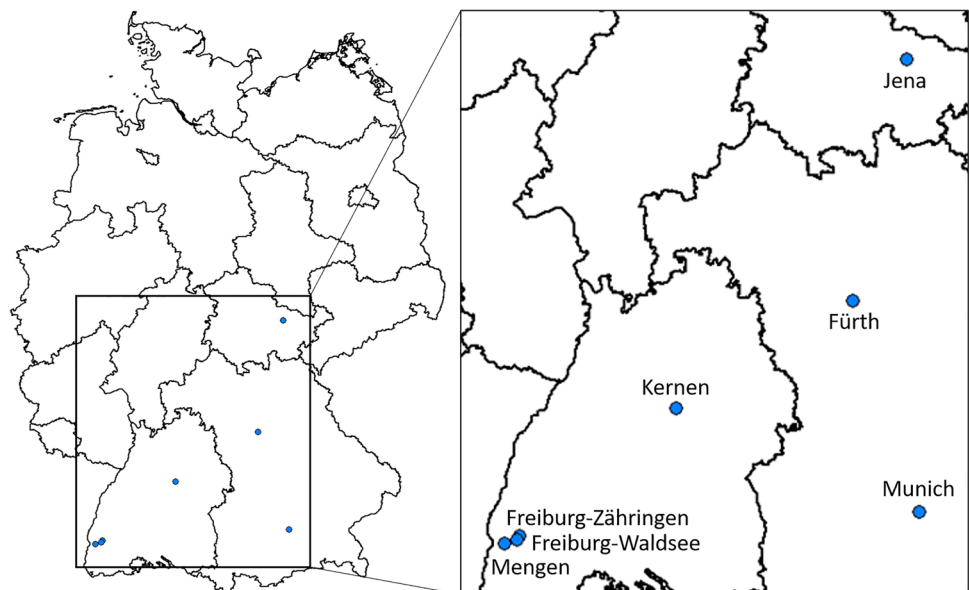
The extent to which the microbiome of a species differs between populations and individuals is largely unexplored. Furthermore, nothing is known about microorganisms naturally occurring in *Ae. albopictus* in Germany, the influence they have on their host and whether they pose a threat to humans or may be exploited to their benefit. This study presents the first preliminary insights into the microbial RNA metagenome of *Ae. albopictus* from Germany which can be considered to represent the mosquito's microbiome. When interpreting the results, however, it is essential to keep in mind that RNA reads similar to a certain microbial species vary considerably in number, and contigs generated from them are of various lengths and have various degrees of probability to be identical to a certain microbial species.

Materials and methods

Mosquito origin

The *Ae. albopictus* specimens investigated in this study had been collected as larvae in the field at seven sites in Germany in 2020 (Fig. 1): Mengen, Freiburg-Waldsee, Freiburg-Zähringen, Kernen, Munich, Fürth and Jena. These were the locations successfully checked for the presence of *Ae. albopictus* aquatic stages from all German cities known to possess established populations at the time of the study. Individuals were obtained by sieving potential breeding containers in cemeteries (Mengen, Freiburg-Waldsee,

Fig. 1 Collection sites in Germany of *Ae. albopictus* specimens examined (blue dots)



Freiburg-Zähringen, Munich, Jena), in gardens of a settlement (Kernen) and an allotment garden complex (Fürth).

Collected larvae were kept in water from their breeding habitat until adult emergence while being fed ad libitum with ground TabiMin fish food pellets (Tetra, Melle, Germany). Shortly after hatching, adults were killed by freezing at $-20\text{ }^{\circ}\text{C}$, without being offered blood or a sugar solution. They were morphologically identified on a chilling table using a stereomicroscope according to the determination key by Becker et al. (2010) and stored in 75% ethanol until further processing. Except for the Freiburg-Zähringen site, from where only one female and no male were available, two females and two males per site were analysed.

Nucleic acid extraction and sequencing

For nucleic acid extraction, the mosquitoes were removed from the ethanol and dried for about 1 min at room temperature for the alcohol to evaporate. Subsequently, 13 female and 12 male mosquitoes were pooled by sex and then completely homogenised in 500 μl serum-free ZB5d medium (FLI-intern cell culture medium = Eagle's minimal essential medium with Earle's and Hank's salts plus non-essential amino acids) containing 5 μl of a ready-to-use mixture of penicillin–streptomycin and 1 μl of a ready-to-use mixture of gentamicin–amphotericin (Thermo Fisher Scientific, Dreieich, Germany). Three steel beads with a diameter of 3 mm (TIS GmbH, Gauting, Germany) were added and the samples agitated for 2 min at 30 Hz in a TissueLyser II (Qiagen, Hilden, Germany). Nucleic acid was then extracted from 200 μl of the supernatant using the NucleoMag VET kit (Macherey–Nagel, Düren, Germany) according to the manufacturer's instructions, but without the addition of carrier RNA. The concentration of extracted RNA (12.2 ng/ μl for the female sample, 11.3 ng/ μl for the male sample) was measured using a NanoDrop Lite (Thermo Fisher Scientific).

Further processing of the sample for total RNA sequencing with Ion Torrent technology, including manual library preparation, was performed following the protocol described by Wylezich et al. (2018). Briefly, the extracted RNA was transcribed into double-stranded cDNA using the cDNA Synthesis System Kit (Roche, Mannheim, Germany), then fragmented by an M220 Focused-ultrasonicator (Covaris Ltd., Brighton, UK) and prepared for Ion Torrent-compatible library generation by means of the GeneRead L Core Kit (Qiagen) and Ion Xpress barcode adapters (Thermo Fisher Scientific). The resulting library was subjected to quality control in a 2100 Bioanalyzer (Agilent Technologies, Santa Clara, USA), using the High Sensitivity DNA Kit (Agilent Technologies) and quantification with a KAPA Library Quantification Kit (Roche). The library was then sequenced on an Ion Torrent S5 XL System (Thermo Fisher Scientific) according to the manufacturer's instructions.

Data analysis

Sequencing results were edited and analysed using Geneious Prime version 2021.0.1 (Biomatters, Auckland, New Zealand). For this, sequences trimmed to a minimum length of 25 bp by the Geneious BBDuk tool were merged by the BBMerge tool, using a merge rate set to 'high'. Both merged data and data that could not be merged were assembled de novo to cluster all closely related sequences into contigs, based on a 'custom sensitivity' setting. The obtained consensus sequences were sorted according to their length, resulting in groups of contigs with similar base pair length. Each contig of these groups was subsequently aligned with GenBank entries (www.ncbi.nlm.nih.gov), with equal results being summarised.

Results

A total of 5,000,504 RNA reads were generated for the female *Ae. albopictus* pool and 4,653,856 reads for the male pool. In the microbial RNA metagenome of female mosquitoes, RNA fragments with high identities to 42 different microorganismal species from 37 different families were detected, whereas in males RNAs with high identities to a total of 38 different species from 36 families were found (Table 1, Supplementary Tables 1 and 2). In the pool of female mosquitoes, there were a total of 213 contigs (in the range of 69.91–100% percent identity (p.i.), 91–100% query cover and 32–1674 bp), 20 (9.35%) of which matched with eukaryotic, 136 (64.02%) with bacterial and 57 (26.63%) with viral species. In the male mosquito pool, a total of 1380 contigs (82.99–100% p.i., 96–100% query cover, 28–1702 bp) was analysed, 19 (1.87%) of which could be assigned to eukaryotes, 338 (24.29%) to bacteria and 1023 (73.83%) to viruses (Fig. 2). Often, an identification at the species level was not possible and only the genus could be determined.

Contigs with high identities to 13 species and 12 genera of viruses/microorganisms were identified in mosquitoes of both sexes (Fig. 3). Contigs arguing for seven microbial species formerly described in *Ae. albopictus* were found in both female and male mosquitoes. Four additional microbial species suggested by the contigs had been found in other mosquito species previously. Another two species of bacterial contigs female *Ae. albopictus* were suggestive of had also been found in other mosquito species, but were not present in our pool of male mosquitoes. Instead, a bacterial species indicated by contigs found in the male pool, but not in the female one of this study, had previously been detected in other mosquito species.

Table 1 shows all viruses and microorganisms whose RNA was identified in *Ae. albopictus* with a p.i. of at least

Table 1 Species of viruses/microorganisms whose RNA matched that isolated from *Ae. albopictus* to at least 97% (see Supplementary Tables 1 and 2 for more details and species with RNA identity lower than 97%)

Species	Phylum	Found in sex	Remark
Viruses			
<i>Aedes albopictus</i> anphevirus	Riboviria	♂♀	Insect-specific virus; previously detected in <i>Ae. albopictus</i> (Manni and Zdobnov 2020)
<i>Aedes phasmavirus</i>	Riboviria	♂♀	Previously detected in <i>Ae. albopictus</i> (Shi et al. 2020)
Barstukas virus	Riboviria	♂♀	Previously detected in various <i>Aedes</i> mosquitoes (Batson et al. 2021)
Guapiaçu virus	Riboviria	♂♀	Insect-specific virus; previously detected in <i>Ae. terrens</i> and <i>Ae. scapularis</i> (Batson et al. 2021; Oliveira Ribeiro et al. 2021)
High Island virus	Riboviria	♂♀	Previously detected in mosquitoes and other invertebrates (Sadeghi et al. 2017)
Usinis virus	Riboviria	♂♀	Previously detected in <i>Ae. albopictus</i> (Batson et al. 2021)
Wenzhou sobemo-like virus	Riboviria	♂♀	Previously detected in <i>Ae. albopictus</i> (Kubacki et al. 2020)
Bacteria			
<i>Acidovorax avenae</i>	Proteobacteria	♀	Plant pathogen (Walcott and Gitaitis 2000)
<i>Acinetobacter baumannii</i>	Proteobacteria	♂♀	Human pathogen; previously detected in <i>Ae. albopictus</i> (Minard et al. 2013)
<i>Acinetobacter dispersus</i>	Proteobacteria	♂	Previously detected on human skin and human wounds, in water and soil (Nemec et al. 2016)
<i>Acinetobacter johnsonii</i>	Proteobacteria	♂♀	Human pathogen; previously detected in <i>Ae. albopictus</i> and other mosquito species (Seifert et al. 1993; Minard et al. 2013)
<i>Acinetobacter oleivorans</i>	Proteobacteria	♂	Previously detected in soil (Uniyal et al. 2016)
<i>Acinetobacter tandooii</i>	Proteobacteria	♀	Previously detected in termites (van Dexter and Boopathy 2019)
<i>Aeromonas hydrophila</i>	Proteobacteria	♀	Pathogenic to many different vertebrates and humans; lives in water habitats (Emerson and Norris 1905; Wohlgenut et al. 1970; Hazen et al. 1978; Agger et al. 1985)
<i>Arthrobacter woluwensis</i>	Actinobacteria	♀	Potential human pathogen (Bernasconi et al. 2004; Li et al. 2021)
<i>Chryseobacterium aureum</i>	Bacteroidetes	♂	Previously detected in river water in Korea (Lee et al. 2019)
<i>Chryseobacterium indoltheticum</i>	Bacteroidetes	♂	Potential human pathogen; previously detected in marine mud (Calderón et al. 2011)
<i>Chryseobacterium scopthalmum</i>	Bacteroidetes	♂	Fish pathogen (Shahi et al. 2018)
<i>Elizabethkingia anophelis</i>	Bacteroidetes	♂♀	Human pathogen: previously detected in <i>An. gambiae</i> (Kämpfer et al. 2011)
<i>Escherichia coli</i>	Proteobacteria	♂♀	Intestinal bacterium; previously detected in <i>An. funestus</i> (Straif et al. 1989)
<i>Hydrogenophaga pseudoflava</i>	Proteobacteria	♀	Previously detected in the midgut of <i>An. gambiae</i> (Straif et al. 1989)
<i>Leclercia adecarboxylata</i>	Proteobacteria	♂	Potential human pathogen (Hess et al. 2008)
<i>Limnobacter humi</i>	Proteobacteria	♂	Previously detected in humus soil (Nguyen and Kim 2017)
<i>Micrococcus luteus</i>	Actinobacteria	♀	Potential human pathogen (Fosse et al. 1985)
<i>Paracoccus yeei</i>	Proteobacteria	♀	Human pathogen; previously detected in the salivary glands of <i>Ae. aegypti</i> (Arias and Clark 2019; Balaji et al. 2021)
<i>Pseudomonas luteola</i>	Proteobacteria	♀	Human pathogen; previously detected in humid environments (Kostmann et al. 1990; Altinok et al. 2007)
<i>Serratia marcescens</i>	Proteobacteria	♂	Human pathogen; previously detected in <i>Anopheles</i> mosquitoes (Hejazi and Falkiner 1997; Bai et al. 2019)
<i>Wolbachia pipientis</i>	Proteobacteria	♂♀	Previously detected in <i>Ae. albopictus</i> (Wiwatanaratanabutr 2013)
<i>Zooglea resiniphila</i>	Proteobacteria	♂	Previously detected in activated sludge (An et al. 2016)
Eukaryota			
<i>Candida sake</i>	Ascomycota	♂♀	Previously detected in oral cavity of HIV-positive people (Hoegl et al. 1998)
<i>Conidiobolus coronatus</i>	Zoopagomycota	♀	Potential human pathogen; previously detected on dead leaf (Fischer et al. 2008)

97% and a query cover of 100%, representing a minimum contig length of 56 bp. RNAs with less p.i. and query cover, shorter lengths or RNAs that could not be assigned to a species, but only to a genus or higher systematic level, are listed

in Supplementary Tables 1 and 2. Only the first group of RNAs is discussed in the following.

RNAs with identity to *Aedes albopictus* anphevirus (89.23–100% p.i., 42–370 bp, 281 contigs) and *Aedes*

Fig. 2 Percentage of contigs from RNA reads from *Ae. albopictus* females and males according to viral, bacterial and eukaryotic origin

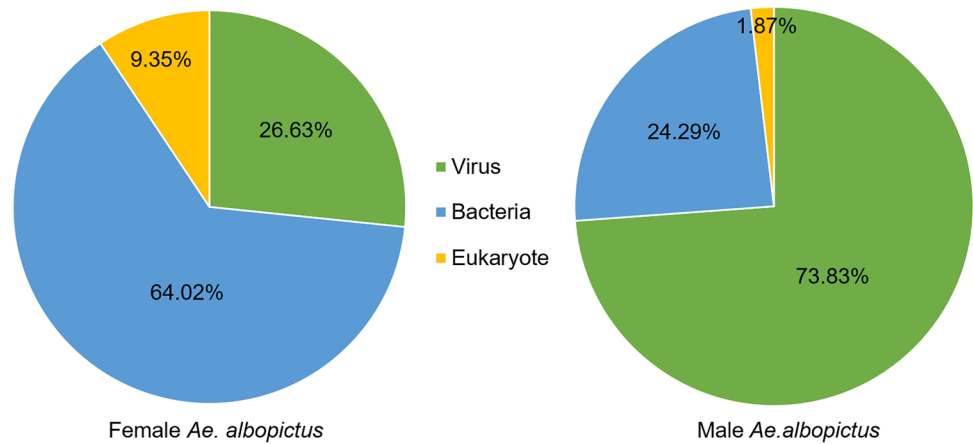
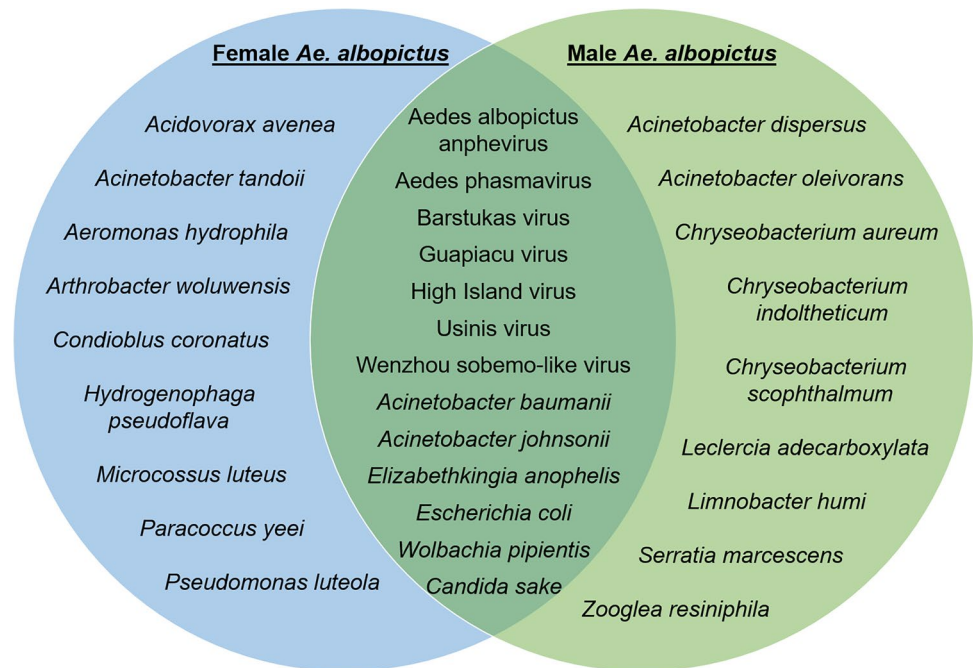


Fig. 3 Assignment of RNA contigs obtained from *Ae. albopictus* to viral, bacterial and eukaryotic species, according to mosquito sex



phasmavirus (96.57–100% p.i., 30–350 bp, 13 contigs) were detected in both female and male *Ae. albopictus* (Table 1). Both viruses are considered insect-specific and had been isolated from adult *Ae. albopictus* before, with *Aedes phasmavirus* being detected in all life stages of the mosquito (Manni and Zdobnov 2020; Shi et al. 2020). In addition, both *Ae. albopictus* sexes harboured RNA similar to Barstukas virus (97.91–100% p.i., 83–524 bp, 5 contigs) and Guapiacu virus (98.50–98.83% p.i., 133–170 bp, 2 contigs) which had previously been identified in adult *Aedes* mosquitoes (Batson et al. 2021). RNA suggesting High Island virus (97.69–98.70% p.i., 120–386 bp, 3 contigs), Usinis virus (99.74–100%, p.i., 118–390 bp, 6 contigs) and Wenzhou sobemo-like virus (96.47–100% p.i., 31–1677 bp, 750 contigs) was also found in both female and male *Ae. albopictus*. High Island virus had formerly been detected in adult

mosquitoes of the species *Psorophora ciliata* (Sadeghi et al. 2017), Usinis virus in adult *Ae. aegypti* and *Ae. albopictus* (Batson et al. 2021) and Wenzhou sobemo-like virus in adult *Ae. albopictus* (Kubacki et al. 2020).

In addition to the RNA of these viruses, RNA with high identities to various bacteria was identified. Among others, RNA largely matching *Acinetobacter baumannii* (97.50–100% p.i., 40–170 bp, 3 contigs), *A. johnsonii* (98.76–100% p.i., 150–188 bp, 7 contigs), *Elizabethkingia anophelis* (99.12–100% p.i., 90–480 bp, 7 contigs), *Escherichia coli* (97.37–100% p.i., 53–167 bp; 14 contigs) and *W. pipientis* (96.91–100% p.i., 33–1052 bp, 204 contigs) was found in both female and male *Ae. albopictus*. *Acinetobacter baumannii* and *A. johnsonii* are human pathogens and had previously been reported from adult *Ae. albopictus* (Seifert et al. 1993; Minard et al. 2013), *A.*

baumannii also from lice and fleas (Kempf et al. 2012). The ubiquitous *E. coli* is a potentially pathogenic (Kaper et al. 2004), widely spread intestinal bacterium of humans and other vertebrates, which had been recognised in adult *Anopheles funestus* and *An. gambiae* before (Straif et al. 1989). *Elizabethkingia anophelis* has emerged as a human pathogen in Africa and Asia (Lau et al. 2016) and had previously been detected in adult *An. gambiae* (Kämpfer et al. 2011). *Wolbachia pipientis* is a widely distributed essential bacterial symbiont of mosquitoes, which had frequently been described from *Ae. albopictus* (e.g. Wiwatanaratnabutr 2013; Park et al. 2016) and other mosquito species (e.g. Kittayapong et al. 2000).

Finally, RNA corresponding to that of *Candida sake* (100% p.i., 104–189 bp, 3 contigs) was detected in both *Ae. albopictus* females and males in this study. The fungus belongs to a genus widely distributed in arthropods but has also been extracted from the oral cavity of HIV-positive humans (Hoegl et al. 1998).

In addition to RNA fragments suggesting the same microbial species to occur in both *Ae. albopictus* females and males, RNA fragments referring to some bacteria were found in one mosquito sex only (Supplementary Tables 1 and 2). These include those of the bacteria *A. dispersus* (99.72% p.i., 422 bp, 1 contig), *A. oleivorans* (100% p.i., 1378 bp, 2 contigs), *Chryseobacterium aureum* (99.11% p.i., 676 bp, 1 contig), *C. indoltheticum* (100% p.i., 602–686 bp, 3 contigs), *C. scophthalmum* (99.79% p.i., 470 bp, 1 contig), *Leclercia adecarboxylata* (100% p.i., 96–546 bp, 6 contigs), *Limnobacter humi* (100% p.i., 106 bp, 1 contig), *Serratia marcescens* (100% p.i., 80 bp, 1 contig) and *Zooglea resiniphila* (100% p.i., 56 bp, 1 contig) in the males (Table 1). *Acinetobacter dispersus* can be frequently found on human skin and in water and soil (Kang et al. 2011; Nemeč et al. 2016). *Acinetobacter oleivorans* had been detected in soil (Kang et al. 2011) and *C. aureum* in river water in Korea (Lee et al. 2019). *Chryseobacterium indoltheticum* is a widespread bacterium occurring in soil and water which may be pathogenic to humans (Calderón et al. 2011), and *C. scophthalmum* is a fish pathogen (Shahi et al. 2018). *Leclercia adecarboxylata* had previously been documented in other insects such as the potato beetle *Leptinotarsa decemlineata* (Muratoglu et al. 2009) and is also considered potentially pathogenic for humans (Hess et al. 2008). By contrast, *L. humi* had been recognised from humus soil (Nguyen and Kim 2017). Another human pathogen similar to RNA which was found in male *Ae. albopictus* was *S. marcescens* (Hejazi and Falkiner 1997). This bacterium had been detected in adult *An. sinensis* mosquitoes previously (Bai et al. 2019) and might become a problem in mosquito laboratory colonies (Seitz et al. 1987). *Zooglea resiniphila* had been found in activated sludge (Gao et al. 2018).

RNAs with similarity to some bacterial species were identified in the female *Ae. albopictus* of this study but not in the males (Table 1). These include *Acidovorax avena* (100% p.i., 121 bp, 1 contig), *Acinetobacter tandoii* (100% p.i., 558 bp, 1 contig), *Aeromonas hydrophila* (100% p.i., 33–1012 bp, 5 contigs), *Arthrobacter woluwensis* (100% p.i., 120–411 bp, 4 contigs), *Hydrogenophaga pseudoflava* (100% p.i., 176–189 bp, 2 contigs), *Micrococcus luteus* (100% p.i., 118–140 bp, 2 contigs), *Paracoccus yeei* (100% p.i., 126–168 bp, 2 contigs) and *Pseudomonas luteola* (100% p.i., 137–198 bp, 2 contigs). *Acidovorax avena* is a plant-pathogenic bacterium (Walcott and Gitaitis 2000), whereas *A. tandoii* had been detected in termites (van Dexter and Boopathy 2019). *Aeromonas hydrophila* is a pathogen of many different vertebrates including humans (Emerson and Norris 1905; Wohlgemut et al. 1970; Agger et al. 1985), which can naturally be found in water habitats (Hazen et al. 1978). *Arthrobacter woluwensis* is a potential human pathogen, which can cause endocarditis, among other symptoms (Bernasconi et al. 2004; Li et al. 2021). *Hydrogenophaga pseudoflava* had previously been detected in the midgut of adult *An. gambiae* (Straif et al. 1989). RNA fragments suggesting another potential human pathogen, which had led to human meningitis in the past, is *M. luteus* (Fosse et al. 1985). *Paracoccus yeei*, on the other hand, is a human bacterial pathogen, which had formerly been isolated from the salivary glands of adult *Ae. aegypti* (Balaji et al. 2021) and can lead to human dialysis-related peritonitis (Arias and Clark 2019). *Pseudomonas luteola* is another fish pathogen, which can cause, for example, meningitis and wound infection in immunocompromised humans (Kostmann et al. 1990; Altinok et al. 2007).

In addition to RNAs with high identities to the above bacteria found in only one sex of *Ae. albopictus*, RNA with a high identity to the fungus *Conidiobolus coronatus*, which has a human-pathogenic potential (Fischer et al. 2008), could be detected in the female mosquitoes.

Discussion

The mosquitoes in this study were pooled from seven sites within Germany known to be populated by *Ae. albopictus*. Since the tiger mosquito is controlled in Germany by Bti (*Bacillus thuringiensis israelensis*) larvicide as soon as local reproduction is detected (Becker et al. 2017, 2022), the finding of larvae is difficult and was limited in the framework of this study. Due to the pooling of the collected samples, no statement can be made about the geographical origin of the microorganisms or the individual colonisation of *Ae. albopictus* specimens. Moreover, all viruses and microorganisms referred to in this study were identified exclusively by their RNAs and the alignment of those with sequences in the used

databases. Therefore, it is not known whether exactly these species were present or other (unknown) species with closely related RNA sequences, whether they were viable viruses, living symbionts or similar and able to replicate/multiply in *Ae. albopictus*, and whether they were arbitrarily taken up from the environment and had no additional correlation to the mosquitoes.

As adult mosquitoes emerged from collected larvae were tested here, and no food sources whatsoever had been offered to the investigated adults, the detected RNAs, or the microbes characterised by them, must be supposed to have been transmitted transstadially from mosquito larva to pupa and through metamorphosis to adult. During metamorphosis, the midgut of mosquitoes is transformed and the digestive cells are histolysed (Fernandes et al. 2014). The survival of microorganisms or the persistence of RNA, respectively, must therefore be supposed to be possible only intracellularly or with certain mechanisms of adapted symbionts. To clarify this and check for the viability of microorganisms, instead of mere RNA, cultivation attempts are necessary, but were not carried out in this study.

As we studied the microbial RNA metagenome of complete mosquitoes, it cannot be determined in which organ or tissue the found RNAs had been localised. Principally, the composition of the bacterial fauna in different organs of a mosquito can be very variable, and some bacteria colonise several organs in the mosquito at the same time (Gao et al. 2020). The tropism of microbes might give information about their migration paths in the mosquito, or about transmissibility from mosquito parent to offspring or mosquito female to blood host. For the latter, the emergence of the microorganism in the mosquito salivary glands would be requisite (Anderson et al. 2010). Whether this was the case for the potential human pathogens RNAs found were suggestive of, such as *A. baumannii*, *E. coli* or *A. hydrophila*, cannot be determined retrospectively. In addition to the localisation of the microorganisms in the mosquito, the pathogen load which was also not determined in this study might be decisive for a mosquito to become a vector. Thus, the mere presence of a pathogen in the mosquito might not be sufficient for transmission (Beerntsen et al. 2000).

The presence in the microbiome of certain bacteria is beneficial to the mosquito. For example, *A. baumannii* and *A. johnsonii* improve blood digestion and nectar assimilation in *Ae. albopictus* (Minard et al. 2013). However, the influence on mosquito development, reproduction and physiology of most microorganisms found in the microbiome is largely unknown.

The most common phyla ever found in the microbiome of adult *Ae. albopictus* include Proteobacteria, Bacteroidetes, Firmicutes and Actinobacteria (Mancini et al. 2018). The most common bacterial genera found in *Aedes*, *Anopheles* and *Culex* species are *Enterobacter*,

Escherichia, *Klebsiella*, *Pseudomonas* and *Serratia* as detected in mosquitoes from the USA, England and India (Demaio et al. 1996; Touré et al. 2000; Pidiyar 2002). RNA with identities to all four bacterial phyla as well as to all five genera were found in the *Ae. albopictus* samples from Germany. In addition, RNA fragments indicating viruses and fungi, such as Riboviria and Ascomycota, were identified. In summary, RNAs with identities to a high number of microorganisms were detected in the German *Ae. albopictus* some of which represent microorganisms already described from this mosquito species previously, such as *W. pipientis*, *A. baumannii* or Usinis virus (Minard et al. 2013; Wiwatanaratanabutr 2013; Batson et al. 2021). Some other viruses or microbial species suggested by the RNA analysis in this study, such as High Island virus, Guapiacu virus and *E. anophelis*, have not been detected in *Ae. albopictus* before, but in other mosquito species and other invertebrates (Kämpfer et al. 2011; Sadeghi et al. 2017; Batson et al. 2021; Oliveira Ribeiro et al. 2021). Furthermore, RNA fragments suggesting microorganisms previously not described from mosquitoes at all, such as *L. humi*, *Z. resiniphila* and *C. aureum*, were found.

It has also been shown that the midgut microbiome of adult mosquitoes may reduce a mosquito's susceptibility to pathogens (Dong et al. 2009; Bahia et al. 2014) and have a general influence on its vector competence (Dodson et al. 2014; Jupatanakul et al. 2014). It can thus be harnessed by manipulating its microorganisms to artificially reduce vector competence. In culture, for example, *E. coli* was genetically modified in order to express two surface molecules that suppress the development of *Plasmodium berghei*. Unfortunately, *E. coli* had difficulties in colonising the mosquitoes and disappeared from their midgut shortly after infection (Riehle et al. 2007). In addition, there are studies that show that an infection of mosquitoes with *Wolbachia* leads to a strong inhibition of the development of potential pathogens. The infection with *Wolbachia* of the wMel strain, for example, leads to a highly reduced replication of dengue virus in *Ae. aegypti*. The reason for this seems to be the *Wolbachia*-linked upregulation of the immune system of the mosquito (Blagrove et al. 2012). Such a way of influencing vector competence is certainly also possible with the help of other organisms from the microbiome.

Since insecticides and physical measures are often inefficient tools for mosquito control (Bourtzis et al. 2016; Pang et al. 2017; Flores and O'Neill 2018), *W. pipientis* is also exploited for innovative biological control by manipulating mosquito reproduction. *Wolbachia* infection can lead to the feminisation of genetically male insects, to the killing of male siblings by females or cytoplasmic incompatibility, which ensures that females can successfully mate only with males harbouring the same *Wolbachia* strain (Werren et al. 2008).

Efficient control tools are particularly important in areas where mosquitoes serve as vectors of human disease agents such as Zika virus, yellow fever virus and malaria parasites. Knowledge about the natural occurrence of microorganisms in mosquitoes can therefore contribute to developing and designing new forms of vector control.

It is difficult to explain the differences in the RNA presence between female and male *Ae. albopictus* in this study. Individuals of both genders were collected from the same breeding sites of the seven locations, so differences between the microbial RNA cannot be attributed to developmental conditions. However, RNA with high identities to 25 species of microorganisms were found in the males but not in the females, and RNA with high identities to 29 species of microorganisms were found in the females but not in the males, with clear differences in the distribution of microorganism to kingdoms (in females, most contigs were assigned to bacteria, whereas in males most contigs were assigned to viruses). For example, RNA fragments arguing for plants were found in the pool of males, but not in the pool of females. That sex has an influence on the microbiome has been shown in previous studies (Chen et al. 2020). Also, Rani et al. (2009) detected *Chryseobacterium*, *Pseudomonas* and *Serratia* species only in females of *Anopheles stephensi*. One possible explanation for the differences might be owing to the limited number of individuals examined, with such differences becoming smaller the more individuals are studied per site. Another explanation might be that female and male larvae in fact have different food preferences and therefore take up different microorganisms with their food, but this is mere speculation and cannot be substantiated. Although it is known that larvae consume a wide range of food (Gimnig et al. 2002; Ye-Ebiyo et al. 2003) and have different feeding preferences depending on species (Merritt et al. 1992), no data exist about different feeding preferences of female and male larvae of the same species.

Especially in cases of a low percent identity and a low query coverage of the found contig sequences with sequences in GenBank, results have to be considered carefully with regard to the occurrence of the respective microorganisms in German *Ae. albopictus*. This applies, for example, to *Kocuria rhizophila*. RNA found in female *Ae. albopictus* matched this soil bacterium with a percent identity of 85.43% only and was registered with only one read. Due to the low percent identity, it is unlikely that the detected RNA belonged to exactly this species. Also, in the case of very short sequence lengths, the linked microorganisms must be viewed critically. This was the case for *Wuchereria bancrofti*-RNA where the sequence length was 45 bp only. Although percent identity and query coverage were both close to 100%, the probability of the RNA belonging to another organism, possibly a filarial species not described so far, is very high. Hitherto, *W. bancrofti* has

not been reported from *Ae. albopictus*, and its geographic distribution range is restricted to subtropical and tropical regions (Service 2001).

Possible contamination must also be considered and, in fact, has already been described in similar studies. Genera such as *Flavobacterium*, *Micrococcus*, *Microbacterium*, *Chryseobacterium*, *Neskia* and *Acidovorax* have been found contaminating laboratory reagents like DNA extraction kits (Salter et al. 2014). In this study, *Flavobacterium*, *Micrococcus*, *Chryseobacterium* and *Acidovorax* could be detected in both sexes of German *Ae. albopictus*, *Neskia* only in females.

Conclusion

The microbiome of vector species such as *Ae. albopictus* holds much potential for the development of efficient control measures and the reduction of vector competence. However, the influence of many microorganisms on the mosquito is still largely unexplored.

Studying the composition of a mosquito's microbiome is difficult, as it is influenced by many factors and can vary considerably both within a species and between species. It is necessary to investigate the influences of the microbiome in more detail and to examine its diversity in a large number of mosquito microbiomes.

To clarify the question of which influence the microorganisms of the microbiome have on the mosquito, it would be helpful to cultivate the microorganisms. However, cultivation is only possible with a small number of species of the microorganisms detected so far. Only with detailed knowledge about the composition and influence of the microbiome on the mosquito can these microorganisms be used for innovative approaches to vector and disease management. The study demonstrates that the microbiome of German *Ae. albopictus* is comprehensive and might be worth further investigations.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s00436-022-07576-7>.

Author contribution JR, DW, MB and HK designed the study; DW and HK collected and identified the mosquitoes; JR and DH carried out the molecular work; JR analysed and interpreted the data; JR and HK wrote the main manuscript text; JR prepared the figure; and all authors reviewed and edited the manuscript. All authors approved the final manuscript version.

Funding Open Access funding enabled and organized by Projekt DEAL.

Data availability Data supporting the conclusions of this article are included within the article and its supplementary tables.

Declarations

Competing interests The authors declare no competing interests.

Ethics approval Not applicable.

Consent to participate Not applicable.

Consent for publication Not applicable.

Conflict of interest The authors declare no competing interests.

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References

- Agger WA, McCormick JD, Gurwith MJ (1985) Clinical and microbiological features of *Aeromonas hydrophila*-associated diarrhea. *J Clin Microbiol* 21:909–913. <https://doi.org/10.1128/jcm.21.6.909-913.1985>
- Ahmad NA, Vythilingam I, Lim YAL, Zabari NZAM, Lee HL (2017) Detection of *Wolbachia* in *Aedes albopictus* and their effects on chikungunya virus. *Am J Trop Med Hyg* 96:148–156. <https://doi.org/10.4269/ajtmh.16-0516>
- Allen TD, Lawson PA, Collins MD, Falsen E, Tanner RS (2006) *Cloacibacterium normanense* gen. nov., sp. nov., a novel bacterium in the family Flavobacteriaceae isolated from municipal wastewater. *Int J Syst Evol Microbiol* 56:1311–1316. <https://doi.org/10.1099/ijs.0.64218-0>
- Altinok I, Balta F, Capkin E, Kayis S (2007) Disease of rainbow trout caused by *Pseudomonas luteola*. *Aquaculture* 273:393–397. <https://doi.org/10.1016/j.aquaculture.2007.10.025>
- An W, Guo F, Song Y, Gao N, Bai S, Dai J, Wei J, Zhang L, Yu D, Xia M, Yu Y, Qi M, Tian C, Chen H, Wu Z, Zhang T, Qui D (2016) Comparative genomics analyses on EPS biosynthesis genes required for floc formation of *Zoogloea resiniphila* and other activated sludge bacteria. *Water Res* 102:494–504. <https://doi.org/10.1016/j.watres.2016.06.058>
- Anderson SL, Richards SL, Smartt CT (2010) A simple method for determining arbovirus transmission in mosquitoes. *J Am Mosq Control Assoc* 26:108–111. <https://doi.org/10.2987/09-5935.1>
- Arias MA, Clark J (2019) *Paracoccus yeei* as a cause of peritoneal dialysis peritonitis in the United Kingdom. *Idcases* 15:e00486. <https://doi.org/10.1016/j.idcr.2019.e00486>
- Bahia AC, Dong Y, Blumberg BJ, Mlambo G, Tripathi A, BenMarzouk-Hidalgo OJ, Chandra R, Dimopoulos G (2014) Exploring *Anopheles* gut bacteria for *Plasmodium* blocking activity. *Environ Microbiol* 16:2980–2994. <https://doi.org/10.1111/1462-2920.12381>
- Bai L, Wang L, Vega-Rodríguez J, Wang G, Wang S (2019) A gut symbiotic bacterium *Serratia marcescens* renders mosquito resistance to *Plasmodium* infection through activation of mosquito immune responses. *Front Microbiol* 10:1580. <https://doi.org/10.3389/fmicb.2019.01580>
- Balaji S, Shekaran SG, Prabakaran SR (2021) Cultivable bacterial communities associated with the salivary gland of *Aedes aegypti*. *Int J Trop Sci* 41:1203–1211. <https://doi.org/10.1007/s42690-020-00310-9>
- Batson J, Dudas G, Haas-Stapleton E, Kistler AL, Li LM, Logan P, Ratnasiri K, Retallack H (2021) Single mosquito metatranscriptomics identifies vectors, emerging pathogens and reservoirs in one assay. *eLife* 10:e68353. <https://doi.org/10.7554/eLife.68353>
- Becher PG, Jensen RE, Natsopoulou ME, Verschut V, De Fine Licht HH (2018) Infection of *Drosophila suzukii* with the obligate insect-pathogenic fungus *Entomophthora muscae*. *J Pest Sci* 91:781–787. <https://doi.org/10.1007/s10340-017-0915-3>
- Becker N, Petric D, Zgomba M, Boase C, Madon M, Dahl C, Kaiser A (2010) Mosquitoes and their Control, 2nd edn. Springer, Berlin Heidelberg
- Becker N, Schön S, Klein AM, Ferstl I, Kizgin A, Tannich E, Kuhn C, Pluskota B, Jöst A (2017) First mass development of *Aedes albopictus* (Diptera: Culicidae) – its surveillance and control in Germany. *Parasitol Res* 116:847–858. <https://doi.org/10.1007/s00436-016-5356-z>
- Becker N, Langentepe-Kong SM, Tokatlian Rodriguez A, Oo TT, Reichle D, Lühken R, Schmidt-Chanasit J, Lüthy P, Puggioli A, Bellini R (2022) Integrated control of *Aedes albopictus* in Southwest Germany supported by the Sterile Insect Technique. *Parasit Vectors* 15:9. <https://doi.org/10.1186/s13071-021-05112-7>
- Berentsen BT, James AA, Christensen BM (2000) Genetics of mosquito vector competence. *Microbiol Mol Biol Rev* 64:115–137. <https://doi.org/10.1128/MMBR.64.1.115-137.2000>
- Benedict MQ, Levine RS, Hawley WA, Lounibos LP (2007) Spread of the tiger: global risk of invasion by the mosquito *Aedes albopictus*. *Vector Borne Zoonot Dis* 7:76–85. <https://doi.org/10.1089/vbz.2006.0562>
- Bennett AJ, Bushmaker T, Cameron K, Ondzie A, Niama FR, Parra HJ, Mombouli JV, Olson SH, Munster VJ, Goldberg TJ (2019) Diverse RNA viruses of arthropod origin in the blood of fruit bats suggest a link between bat and arthropod viromes. *Virology* 528:64–72. <https://doi.org/10.1016/j.virol.2018.12.009>
- Berg G, Rybakova D, Fischer D, Cernava T, Champomier Vergès MC, Charles T, Chen X, Cocolin L, Eversole K, Herrero Corral G, Kazou M, Kinkel L, Lange L, Lima N, Loy A, Macklin JA, Maguin E, Mauchline T, McClure R, Mitter B, Ryan M, Sarand I, Smidt H, Schelkle B, Roume H, Kiran SG, Selvin J, Correa S, de Souza R, van Overbeek L, Singh BK, Wagner M, Walsh A, Sessitsch A, Schlöter M (2020) Microbiome definition re-visited: old concepts and new challenges. *Microbiome* 8:103. <https://doi.org/10.1186/s40168-020-00875-0>
- Bernasconi E, Valsangiacomo C, Peduzzi R, Carota A, Moccetti T, Funke G (2004) *Arthrobacter woluwensis* subacute infective endocarditis: case report and review of the literature. *Clin Infect Dis* 38:27–31. <https://doi.org/10.1086/381436>
- Blagrove MSC, Arias-Goeta C, Failloux A-B, Sinkins SP (2012) *Wolbachia* strain wMel induces cytoplasmic incompatibility and blocks dengue transmission in *Aedes albopictus*. *Proc Natl Acad Sci USA* 109:255–260. <https://doi.org/10.1073/pnas.1112021108>
- Boissière A, Tchioffo MT, Bachar D, Abate L, Marie A, Nsango SE, Shahbazkia HR, Awono-Ambene PH, Levashina EA, Christen R, Morlais I (2012) Midgut microbiota of the malaria mosquito vector *Anopheles gambiae* and interactions with *Plasmodium falciparum* infection. *PLoS Pathog* 8:e1002742. <https://doi.org/10.1371/journal.ppat.1002742>
- Bonizzoni M, Gasperi G, Chen X, James AA (2013) The invasive mosquito species *Aedes albopictus*: current knowledge and future

- perspectives. *Trends Parasitol* 29:460–468. <https://doi.org/10.1016/j.pt.2013.07.003>
- Bourtzis K, O'Neill S (1998) *Wolbachia* infections and arthropod reproduction. *Bioscience* 48:287–293. <https://doi.org/10.2307/1313355>
- Bourtzis K, Lees RS, Hendrichs J, Vreysen MJB (2016) More than one rabbit out of the hat: radiation, transgenic and symbiont-based approaches for sustainable management of mosquito and tsetse fly populations. *Acta Trop* 157:115–130. <https://doi.org/10.1016/j.actatropica.2016.01.009>
- Calderón G, García E, Rojas P, García E, Rosso M, Losada A (2011) *Chryseobacterium indologenes* infection in a newborn: a case report. *J Med Case Rep* 5:10. <https://doi.org/10.1186/1752-1947-5-10>
- Carr EL, Kämpfer P, Patel BKC, Gürtler V, Seviour RJ (2003) Seven novel species of *Acinetobacter* isolated from activated sludge. *Int J Syst Evol Microbiol* 53:953–963. <https://doi.org/10.1099/ijs.0.02486-0>
- Cerezales M, Xanthopoulou K, Ertel J, Nemeč A, Bustamante Z, Seifert H, Gallego L, Higgins PG (2018) Identification of *Acinetobacter seifertii* isolated from Bolivian hospitals. *J Med Microbiol* 67:834–837. <https://doi.org/10.1099/jmm.0.000751>
- Chen WM, Cho NT, Huang WC, Young CC, Sheu SY (2013) Description of *Gemmobacter fontiphilus* sp. nov., isolated from a freshwater spring, reclassification of *Catellibacterium nectariphilum* as *Gemmobacter nectariphilum* comb. nov., *Catellibacterium changlense* as *Gemmobacter changlense* comb. nov., *Catellibacterium aquatile* as *Gemmobacter aquaticus* nom. nov., *Catellibacterium caeni* as *Gemmobacter caeni* comb. nov., *Catellibacterium nanjingense* as *Gemmobacter nanjingensis* comb. nov., and emended description of the genus *Gemmobacter* and of *Gemmobacter aquatilis*. *Int J Syst Evol Microbiol* 63:470–478. <https://doi.org/10.1099/ijs.0.042051-0>
- Chen S, Zhang D, Augustinos A, Doudoumis V, Bel Mokhtar N, Maiga H, Tsiamis G, Bourtzis K (2020) Multiple factors determine the structure of bacterial communities associated with *Aedes albopictus* under artificial rearing conditions. *Front Microbiol* 11:605. <https://doi.org/10.3389/fmicb.2020.00605>
- Coon KL, Vogel KJ, Brown MR, Strand MR (2014) Mosquitoes rely on their gut microbiota for development. *Mol Ecol* 23:2727–2739. <https://doi.org/10.1111/mec.12771>
- Coon KL, Brown MR, Strand MR (2016) Mosquitoes host communities of bacteria that are essential for development but vary greatly between local habitats. *Mol Ecol* 25:5806–5826. <https://doi.org/10.1111/mec.13877>
- Coon KL, Valzania L, McKinney DA, Vogel KJ, Brown MR, Strand MR (2017) Bacteria-mediated hypoxia functions as a signal for mosquito development. *Proc Natl Acad Sci USA* 114:5362–5369. <https://doi.org/10.1073/pnas.1702983114>
- Dahal RH, Kim J (2018) *Fluviicola kyonggii* sp. nov., a bacterium isolated from forest soil and emended description of the genus *Fluviicola*. *Int J Syst Evol Microbiol* 68:1885–1889. <https://doi.org/10.1099/ijsem.0.002759>
- Demaio J, Pumpuni CB, Kent M, Beier JC (1996) The midgut bacterial flora of wild *Aedes triseriatus*, *Culex pipiens* and *Psorophora columbiae* mosquitoes. *Am J Trop Med Hyg* 54:219–223. <https://doi.org/10.4269/ajtmh.1996.54.219>
- Derelle E, Monier A, Cooke R, Worden AZ, Grimsley NH, Moreau H (2015) Diversity of viruses infecting the green microalga *Ostreococcus lucimarinus*. *J Virol* 89:5812–5821. <https://doi.org/10.1128/JVI.00246-15>
- Dodson BL, Hughes GL, Paul O, Matarachiero AC, Kramer LD, Rasgon JL (2014) *Wolbachia* enhances West Nile virus (WNV) infection in the mosquito *Culex tarsalis*. *PLoS Negl Trop Dis* 8:2965. <https://doi.org/10.1371/journal.pntd.0002965>
- Dong Y, Manfredini F, Dimopoulos G (2009) Implication of the mosquito midgut microbiota in the defense against malaria parasites. *PLoS Pathog* 5:e1000423. <https://doi.org/10.1371/journal.ppat.1000423>
- Douglas AE (2015) Multiorganismal insects: diversity and function of resident microorganisms. *Annu Rev Entomol* 60:17–34. <https://doi.org/10.1146/annurev-ento-010814-020822>
- Emerson H, Norris C (1905) “Red-Leg” – an infectious disease of frogs. *J Exp Medicine* 7:32–57. <https://doi.org/10.1084/jem.7.1.32>
- Engel P, Moran NA (2013) The gut microbiota of insects – diversity in structure and function. *FEMS Microbiol Rev* 37:699–735. <https://doi.org/10.1111/1574-6976.12025>
- Fălcuță E, Prioteasa LF, Horváth C, Păstrav IR, Schaffner F, Mihalca AD (2020) The invasive Asian tiger mosquito *Aedes albopictus* in Romania: towards a country-wide colonization? *Parasitol Res* 119:841–845. <https://doi.org/10.1007/s00436-020-06620-8>
- Fernandes MK, Neves CA, Serrão JE, Martins GF (2014) *Aedes aegypti* midgut remodeling during metamorphosis. *Parasitol Int* 63:506–512. <https://doi.org/10.1016/j.parint.2014.01.004>
- Fischer N, Ruef C, Ebnöther C, Bächli EB (2008) Rhinofacial *Conidiobolus coronatus* infection presenting with nasal enlargement. *Infection* 36:594–596. <https://doi.org/10.1007/s15010-008-8056-5>
- Flores HA, O'Neill SL (2018) Controlling vector-borne diseases by releasing modified mosquitoes. *Nat Rev Microbiol* 16:508–518. <https://doi.org/10.1038/s41579-018-0025-0>
- Fosse T, Peloux Y, Granthil C, Toga B, Bertrando J, Sethian M (1985) Meningitis due to *Micrococcus luteus*. *Infection* 13:280–281. <https://doi.org/10.1007/BF01645439>
- Gao N, Xia M, Dai J, Yu D, An W, Li S, Liu S, He P, Zhang L, Wu Z, Bi X, Chen S, Haft DH, Qiu D (2018) Both widespread PEP-CTERM proteins and exopolysaccharides are required for floc formation of *Zoogloea resiniphila* and other activated sludge bacteria. *Environ Microbiol* 20:1677–1692. <https://doi.org/10.1111/1462-2920.14080>
- Gao H, Cui C, Wang L, Jacobs-Lorena M, Wang S (2020) Mosquito microbiota and implications for disease control. *Trends Parasitol* 36:98–111. <https://doi.org/10.1016/j.pt.2019.12.001>
- Gimnig JE, Ombok M, Otieno S, Kaufman MG, Vulule JM, Walker ED (2002) Density-dependent development of *Anopheles gambiae* (Diptera: Culicidae) larvae in artificial habitats. *J Med Entomol* 39:162–172. <https://doi.org/10.1603/0022-2585-39.1.162>
- Gimonneau G, Tchioffo MT, Abate L, Boissière A, Awono-Ambéné PH, Nsango SE, Christen R, Morlais I (2014) Composition of *Anopheles coluzzii* and *Anopheles gambiae* microbiota from larval to adult stages. *Infect Genet Evol* 28:715–724. <https://doi.org/10.1016/j.meegid.2014.09.029>
- Hazen TC, Fliermans CB, Hirsch RP, Esch GW (1978) Prevalence and distribution of *Aeromonas hydrophila* in the United States. *Appl Environ Microbiol* 36:731–738. <https://doi.org/10.1128/aem.36.5.731-738.1978>
- Hejazi S, Falkiner FR (1997) *Serratia marcescens*. *J Med Microbiol* 46:903–912. <https://doi.org/10.32388/HXQR6V>
- Hess B, Burchett A, Huntington MK (2008) *Leclercia adecarboxylata* in an immunocompetent patient. *J Med Microbiol* 57:896–898. <https://doi.org/10.1099/jmm.0.47673-0>
- Hilgenboecker K, Hammerstein P, Schlattmann P, Telschow A, Werren JH (2008) How many species are infected with *Wolbachia*? A statistical analysis of current data. *FEMS Microbiol Letters* 281:215–220. <https://doi.org/10.1111/j.1574-6968.2008.01110.x>
- Hoegl L, Schönián G, Ollert M, Korting HC (1998) *Candida sakei*: a relevant species in the context of HIV-associated oropharyngeal candidosis? *J Mol Med* 76:70–73. <https://doi.org/10.1007/s001090050192>

- Holmes B, Steigerwalt AG, Nicholson AC (2013) DNA-DNA hybridization study of strains of *Chryseobacterium*, *Elizabethkingia* and *Empedobacter* and of other usually indole-producing non-fermenters of CDC groups IIc, IIe, IIh and IIIi, mostly from human clinical sources, and proposals of *Chryseobacterium bernardetii* sp. nov., *Chryseobacterium carnis* sp. nov., *Chryseobacterium lactis* sp. nov., *Chryseobacterium nakagawai* sp. nov. and *Chryseobacterium taklimakanense* comb. nov. *Int J Syst Evol Microbiol* 63:4639–4662. <https://doi.org/10.1099/ijs.0.054353-0>
- Hörtnagl P, Pérez MT, Zeder M, Sommaruga R (2010) The bacterial community composition of the surface microlayer in a high mountain lake. *FEMS Microbiol Ecol* 73:458–467. <https://doi.org/10.1111/j.1574-6941.2010.00904.x>
- Hunter WJ, Kuykendall LD, Manter DK (2007) *Rhizobium selenireducens* sp. nov.: a selenite-reducing alpha-Proteobacteria isolated from a bioreactor. *Curr Microbiol* 55:455–460. <https://doi.org/10.1007/s00284-007-9020-9>
- Joseph SW, Carnahan AM, Brayton PR, Fanning GR, Almazan R, Drabick C, Trudo EWJR, Colwell RR (1991) *Aeromonas jandaei* and *Aeromonas veronii* dual infection of a human wound following aquatic exposure. *J Clin Microbiol* 29:565–569. <https://doi.org/10.1128/jcm.29.3.565-569.1991>
- Jupatanakul N, Sim S, Dimopoulos G (2014) The insect microbiome modulates vector competence for arboviruses. *Viruses* 6:4294–4313. <https://doi.org/10.3390/v6114294>
- Kämpfer P, Matthews H, Glaeser SP, Martin K, Lodders N, Faye I (2011) *Elizabethkingia anophelis* sp. nov., isolated from the midgut of the mosquito *Anopheles gambiae*. *Int J Syst Evol Microbiol* 61:2670–2675. <https://doi.org/10.1099/ijs.0.026393-0>
- Kang YS, Jung J, Jeon CO, Park W (2011) *Acinetobacter oleivorans* sp. nov. is capable of adhering to and growing on diesel-oil. *J Microbiol* 49:29–34. <https://doi.org/10.1007/s12275-011-0315-y>
- Kanyuka K, Ward E, Adams MJ (2003) *Polymyxa graminis* and the cereal viruses it transmits: a research challenge. *Mol Plant Pathol* 4:393–406. <https://doi.org/10.1046/j.1364-3703.2003.00177.x>
- Kaper JB, Nataro JP, Mobley HL (2004) Pathogenic *Escherichia coli*. *Nat Rev Microbiol* 2:123–140. <https://doi.org/10.1038/nrmicr0818>
- Kempf M, Abdissa A, Diatta G, Trape J-F, Angelakis E, Mediannikov O, La Scola B, Raoult D (2012) Detection of *Acinetobacter baumannii* in human head and body lice from Ethiopia and identification of new genotypes. *Int J Infect Dis* 16:e680–e683. <https://doi.org/10.1016/j.ijid.2012.05.1024>
- Kim D, Baik KS, Kim MS, Park SC, Kim SS, Rhee MS, Kwak YS, Seon CN (2008) *Acinetobacter soli* sp. nov., isolated from forest soil. *J Microbiol* 46:396–401. <https://doi.org/10.1007/s12275-008-0118-y>
- Kittayapong P, Baisley KJ, Baimai V, O'Neill SL (2000) Distribution and diversity of *Wolbachia* infections in Southeast Asian mosquitoes (Diptera: Culicidae). *J Med Entomol* 37:340–345. <https://doi.org/10.1093/jmedent/37.3.340>
- Kostmann JR, Solomon F, Fekete T (1990) Infections with *Chryseomonas luteola* (CDC Group Ve-1) and *Flavimonas oryzae* (CDC Group Ve-2) in neurosurgical patients. *Rev Infect Dis* 13:233–236. <https://doi.org/10.1093/clinids/13.2.233>
- Kubacki J, Flacio E, Qi W, Guidi V, Tonolla M, Fraefel C (2020) Viral metagenomic analysis of *Aedes albopictus* mosquitoes from southern Switzerland. *Viruses* 12:929. <https://doi.org/10.3390/v12090929>
- Larkin JM, Williams PM (1978) *Runella slithyformis* gen. nov., sp. nov., a curved, nonflexible, pink bacterium. *Int J Syst Bacteriol* 28:32–36. <https://doi.org/10.1099/00207713-28-1-32>
- Lau SKP, Chow WN, Foo CH, Curreem SOT, Lo GCS, Teng JLL, Chen JHK, Ng RHY, Wu AKL, Cheung IYY, Chau SKY, Lung DC, Lee RA, Tse CWS, Fung KSC, Que TL, Woo PCY (2016) *Elizabethkingia anophelis* bacteremia is associated with clinically significant infections and high mortality. *Sci Rep* 6:26045. <https://doi.org/10.1038/srep26045>
- Lee JE, Hwang EM, Cha CJ, Kim GB (2019) *Chryseobacterium aureum* sp. nov., isolated from the Han River, Republic of Korea. *Int J Syst Evol Microbiol* 69:1628–1633. <https://doi.org/10.1099/ijs.0.003370>
- Li SY, Kao CC, Hu YC, Lai CH, Jiang YP, Mao YC, Huang YT, Liu PY (2021) *Arthrobacter woluwensis* bacteremia: a clinical and genomic report. *Pathogens* 10:443. <https://doi.org/10.3390/pathogens10040443>
- Ling LL, Schneider T, Peoples AJ, Spoering AL, Engels I, Conlon BP, Mueller A, Schäberle TF, Hughes DE, Epstein S, Jones M, Lazarides L, Steadman VA, Cohen DR, Felix CR, Fetterman KA, Millett WP, Nitti AG, Zullo AM, Chen C, Lewis K (2015) A new antibiotic kills pathogens without detectable resistance. *Nature* 517:455–459. <https://doi.org/10.1038/nature14098>
- Mancini MV, Damiani C, Accoti A, Tallarita M, Nunzi E, Cappelli A, Bozic J, Catanzani R, Rossi P, Valzano M, Serrao A, Ricci I, Spaccapelo R, Favia G (2018) Estimating bacteria diversity in different organs of nine species of mosquito by next generation sequencing. *BMC Microbiol* 18:126. <https://doi.org/10.1186/s12866-018-1266-9>
- Manni M, Zdobnov EM (2020) A novel anphevirus in *Aedes albopictus* mosquitoes is distributed worldwide and interacts with the host RNA interference pathway. *Viruses* 12:1264. <https://doi.org/10.3390/v12111264>
- Martinet JP, Ferté H, Failloux AB, Schaffner F, Depaquit J (2019) Mosquitoes of north-western Europe as potential vectors of arboviruses: a review. *Viruses* 11:1059. <https://doi.org/10.3390/v11111059>
- Merritt RW, Dadd RH, Walker ED (1992) Feeding behavior, natural food, and nutritional relationships of larval mosquitoes. *Annu Rev Entomol* 37:349–376. <https://doi.org/10.1146/annurev.en.37.010192.002025>
- Minard G, Tran FH, Raharimalala FN, Hellard E, Ravelonandro P, Mavingui P, Valiente Moro C (2013) Prevalence, genomic and metabolic profiles of *Acinetobacter* and *Asaia* associated with field-caught *Aedes albopictus* from Madagascar. *FEMS Microbiol Ecol* 83:63–73. <https://doi.org/10.1111/j.1574-6941.2012.01455.x>
- Moissenet D, Becker K, Mérens A, Ferroni A, Dubern B, Vu-Thien H (2012) Persistent bloodstream infection with *Kocuria rhizophila* related to a damaged central catheter. *J Clin Microbiol* 50:1495–1498. <https://doi.org/10.1128/JCM.06038-11>
- Moreira JS, Riccetto AGL, Da Silva MTN, Vilela MMDS (2015) Endocarditis by *Kocuria rosea* in an immunocompetent child. *Braz J Infect Dis* 19:82–84. <https://doi.org/10.1016/j.bjid.2014.09.007>
- Moro CV, Tran FH, Raharimalala FN, Ravelonandro P, Mavingui P (2013) Diversity of culturable bacteria including *Pantoea* in wild mosquito *Aedes albopictus*. *BMC Microbiol* 13:70. <http://www.biomedcentral.com/1471-2180/13/70>
- Muratoglu H, Kati H, Demirbag Z, Sezen K (2009) High insecticidal activity of *Leclercia adecarboxylata* isolated from *Leptinotarsa decemlineata* (Col.: Chrysomelidae). *Afr J Biotechnol* 24:7111–7115. <http://www.academicjournals.org/AJB>
- Nemec A, Radolfova-Krizova L, Maixnerova M, Vrestiakova E, Jezek P, Sedo O (2016) Taxonomy of haemolytic and/or proteolytic strains of the genus *Acinetobacter* with the proposal of *Acinetobacter courvalinii* sp. nov. (genomic species 14 sensu Bouvet & Jeanjean), *Acinetobacter dispersus* sp. nov. (genomic species 17), *Acinetobacter modestus* sp. nov., *Acinetobacter proteolyticus* sp. nov. and *Acinetobacter vivianii* sp. nov. *Int J Syst Evol Microbiol* 66:1673–1685. <https://doi.org/10.1099/ijs.0.000932>
- Nguyen TM, Kim J (2017) *Limnobacter humi* sp. nov., a thiosulfate-oxidizing, heterotrophic bacterium isolated from humus soil, and emended description of the genus *Limnobacter* Spring

- et al. 2001. *J Microbiol* 55:508–513. <https://doi.org/10.1007/s12275-017-6645-7>
- Nogueira MM, Neves E, Johnsson R (2021) Effects of habitat structure on the mollusc assemblage in *Mussismilia* corals: evaluation of the influence of different coral growth morphology. *J Mar Biol Assoc UK* 101:61–69. <https://doi.org/10.1017/S0025315421000023>
- Oliveira Ribeiro Gd, Da Costa AC, Gill DE, Ribeiro ESD'A, Da Rego MOS, Monteiro FJC, Villanova F, Nogueira JS, Maeda AY, de Preira Souza R, Tahmasebi R, Morais VS, Pandey RP, Raj VS, Scandar SAS, da Silva Vasami FG, D'Agostino LG, Maiorka PC, Deng X, Nogueira ML, Sabino EC, Delwart ED, Leal EL, Cunha MS (2021) Guapiáçu virus, a new insect-specific flavivirus isolated from two species of *Aedes* mosquitoes from Brazil. *Sci Rep* 11:4674. <https://doi.org/10.1038/s41598-021-83879-6>
- Onchuru TO, Ajamma YU, Burugu M, Kaltenpoth M, Masiga D, Villingier J (2016) Chemical parameters and bacterial communities associated with larval habitats of *Anopheles*, *Culex* and *Aedes* mosquitoes (Diptera: Culicidae) in western Kenya. *Int J Trop Insect Sci* 36:146–160. <https://doi.org/10.1017/S1742758416000096>
- Pang T, Mak TK, Gubler DJ (2017) Prevention and control of dengue – the light at the end of the tunnel. *Lancet Infect Dis* 17:e79–e87. [https://doi.org/10.1016/S1473-3099\(16\)30471-6](https://doi.org/10.1016/S1473-3099(16)30471-6)
- Park CH, Lim HW, Kim HW, Lee WGL, Roh JY, Park MY, Shin EH (2016) High prevalence of *Wolbachia* infection in Korean populations of *Aedes albopictus* (Diptera: Culicidae). *J Asia-Pacific Entomol* 19:191–194. <https://doi.org/10.1016/j.aspen.2015.12.014>
- Paupy C, Delatte H, Bagny L, Corbel V, Fontenille D (2009) *Aedes albopictus*, an arbovirus vector: from the darkness to the light. *Microbes Infect* 11:1177–1185. <https://doi.org/10.1016/j.micinf.2009.05.005>
- Pełkala A, Paździor E, Antychowicz J, Bernad A, Głowacka H, Więcek B, Niemczuk W (2018) *Kocuria rhizophila* and *Micrococcus luteus* as emerging opportunist pathogens in brown trout (*Salmo trutta* Linnaeus, 1758) and rainbow trout (*Oncorhynchus mykiss* Walbaum, 1792). *Aquaculture* 486:285–289. <https://doi.org/10.1016/j.aquaculture.2017.12.028>
- Pidiyar V (2002) *Aeromonas culicicola* sp. nov., from the midgut of *Culex quinquefasciatus*. *Int J Syst Evol Microbiol* 52:1723–1728. <https://doi.org/10.1099/ijs.0.02019-0>
- Pitt A, Schmidt J, Koll U, Hahn MW (2019) *Aquirufa antheringensis* gen. nov., sp. nov. and *Aquirufa nivalisilvae* sp. nov., representing a new genus of widespread freshwater bacteria. *Int J Syst Evol Microbiol* 69:2739–2749. <https://doi.org/10.1099/ijsem.0.003554>
- Pladdies T, Babenzien HD, Cypionka H (2004) Distribution of *Nevskia ramosa* and other rosette-forming neustonic bacteria. *Microb Ecol* 47:218–223. <https://doi.org/10.1007/s00248-003-1070-3>
- Ponte R, Jenkis SG (1992) Fatal *Chromobacterium violaceum* infections associated with exposure to stagnant waters. *Pediatr Infect Dis J* 11:583–586
- Rani A, Sharma A, Rajagopal R, Adak T, Bhatnagar RK (2009) Bacterial diversity analysis of larvae and adult midgut microflora using culture-dependent and culture-independent methods in lab-reared and field-collected *Anopheles stephensi* – an Asian malarial vector. *BMC Microbiol* 9:96. <https://doi.org/10.1186/1471-2180-9-96>
- Reiss I, Borkhardt A, Füssle R, Sziegoleit A, Gortner L (2000) Disinfectant contaminated with *Klebsiella oxytoca* as a source of sepsis in babies. *Lancet* 356:310. [https://doi.org/10.1016/S0140-6736\(00\)02509-5](https://doi.org/10.1016/S0140-6736(00)02509-5)
- Riehle MA, Moreira CK, Lampe D, Lauzon C, Jacobs-Lorena M (2007) Using bacteria to express and display anti-*Plasmodium* molecules in the mosquito midgut. *Int J Parasitol* 37:595–603. <https://doi.org/10.1016/j.ijpara.2006.12.002>
- Romano-Bertrand S, Bourdier A, Aujoulat F, Michon AL, Masnou A, Parer S, Marchand H, Jumas-Bilak E (2016) Skin microbiota is the main reservoir of *Roseomonas mucosa*, an emerging opportunistic pathogen so far assumed to be environmental. *Clin Microbiol Infect* 22:e1-7. <https://doi.org/10.1016/j.cmi.2016.05.024>
- Sadeghi M, Popov V, Guzman H, Phan TG, Vasilakis N, Tesh R, Delwart E (2017) Genomes of viral isolates derived from different mosquito species. *Virus Res* 242:49–57. <https://doi.org/10.1016/j.virusres.2017.08.012>
- Salter SJ, Cox MJ, Turek EM, Caslus ST, Cookson WO, Moffatt MF, Turner P, Parkhill J, Lohman NJ, Walker AW (2014) Reagent and laboratory contamination can critically impact sequences-based microbiome analyses. *BMC Biol* 12:87. <https://doi.org/10.5194/gi-2016-11-RC2>
- Schmidt R, Battaglia V, Scow K, Kane S, Hristova KR (2008) Involvement of a novel enzyme, MdpA, in methyl tert-butyl ether degradation in *Methylbium petroleiphilum* PM1. *Appl Environ Microbiol* 74:6631–6638. <https://doi.org/10.1128/AEM.01192-08>
- Seifert H, Strate A, Schulze A, Pulverer G (1993) Vascular catheter-related bloodstream infection due to *Acinetobacter johnsonii* (formerly *Acinetobacter calcoaceticus* var. *lwoffii*): report of 13 cases. *Clin Infect Dis* 17:632–636. <https://doi.org/10.1093/clinfid/17.4.632>
- Seitz HM, Maier WA, Rottok M, Becker-Feldmann H (1987) Concomitant infections of *Anopheles stephensi* with *Plasmodium berghei* and *Serratia marcescens*: additive detrimental effects. *Zentralbl Bakt Mikrobiol Hyg A* 266:155–166. [https://doi.org/10.1016/s0176-6724\(87\)80029-9](https://doi.org/10.1016/s0176-6724(87)80029-9)
- Service MW (2001) The encyclopedia of arthropod-transmitted infections. CABI Publishing, Liverpool School of Tropical Medicine, UK
- Shahi N, Sharma P, Pandey J, Bisht I, Mallik SK (2018) Characterization and pathogenicity study of *Chryseobacterium scophthalmum* recovered from gill lesions of diseased golden mahseer, *Tor putitora* (Hamilton, 1822) in India. *Aquaculture* 485:81–92. <https://doi.org/10.1016/j.aquaculture.2017.11.018>
- Shi C, Zhao L, Atoni E, Zeng W, Hu X, Matthijssens J, Yuan Z, Xia H (2020) Stability of the virome in lab- and field-collected *Aedes albopictus* mosquitoes across different developmental stages and possible core viruses in the publicly available virome data of *Aedes* mosquitoes. *mSystems* 5:e00640-20. <https://doi.org/10.1128/mSystems.00640-20>
- Shil RK, Mojumder S, Sadida FF, Uddin M, Sikdar D (2014) Isolation and identification of cellulolytic bacteria from the gut of three phytophagous insect species. *Braz Arch Biol Technol* 57:927–932. <https://doi.org/10.1590/S1516-8913201402620>
- Smith AH, Łukasik P, O'Connor MP, Lee A, Mayo G, Drott MT, Doll S, Tuttle R, Disciullo RA, Messina A, Oliver KM, Russel JA (2015) Patterns, causes and consequences of defensive microbiome dynamics across multiple scales. *Mol Ecol* 24:1135–1149. <https://doi.org/10.1111/mec.13095>
- Straif SC, Mbogo CNM, Toure AM, Walker ED, Kaufman M, Toure YT, Beier JC (1989) Midgut bacteria in *Anopheles gambiae* and *An. funestus* (Diptera: Culicidae) from Kenya and Mali. *J Med Entomol* 35:222–226. <https://doi.org/10.1093/jmedent/35.3.222>
- Strand MR (2018) Composition and functional roles of the gut microbiota in mosquitoes. *Curr Opin Insect Sci* 28:59–65. <https://doi.org/10.1016/j.cois.2018.05.008>
- Terenius O, Lindh JM, Eriksson-Gonzales K, Bussièrè L, Laugen AT, Bergquist H, Titanji K, Faye I (2012) Midgut bacterial dynamics in *Aedes aegypti*. *FEMS Microbiol Ecol* 80:556–565. <https://doi.org/10.1111/j.1574-6941.2012.01317.x>
- Touré AM, Mackey AJ, Wang ZX, Beier JC (2000) Bactericidal effects of sugar-fed antibiotics on resident midgut bacteria of newly

- emerged anopheline mosquitoes (Diptera: Culicidae). *J Med Entomol* 37:246–249. <https://doi.org/10.1093/jmedent/37.2.246>
- Uniyal S, Paliwal R, Verma M, Sharma RK, Rai JPN (2016) Isolation and characterization of fipronil degrading *Acinetobacter calcoaceticus* and *Acinetobacter oleivorans* from rhizospheric zone of *Zea mays*. *Bull Environ Contam Toxicol* 96:833–838. <https://doi.org/10.1007/s00128-016-1795-6>
- van Dexter S, Boopathy R (2019) Biodegradation of phenol by *Acinetobacter tandoii* isolated from the gut of the termite. *Environ Sci Pollut Res Int* 26:34067–34072. <https://doi.org/10.1007/s11356-018-3292-4>
- van Pham HT, Jeong SW, Kim J (2015) *Aquabacterium olei* sp. nov., an oil-degrading bacterium isolated from oil-contaminated soil. *Int J Syst Evol Microbiol* 65:3597–3602. <https://doi.org/10.1099/ijsem.0.000458>
- Walcott RR, Gitaitis RD (2000) Detection of *Acidovorax avenae* subsp. *citrulli* in watermelon seed using immunomagnetic separation and the polymerase chain reaction. *Plant Dis* 84:470–474. <https://doi.org/10.1094/PDIS.2000.84.4.470>
- Walther D, Scheuch DE, Kampen H (2017) The invasive Asian tiger mosquito *Aedes albopictus* (Diptera: Culicidae) in Germany: local reproduction and overwintering. *Acta Trop* 166:186–192. <https://doi.org/10.1016/j.actatropica.2016.11.024>
- Wang Y, Gilbreath TM, Kukutla P, Yan G, Xu J (2011) Dynamic gut microbiome across life history of the malaria mosquito *Anopheles gambiae* in Kenya. *PLoS One* 6:e24767. <https://doi.org/10.1371/journal.pone.0024767>
- Wang X, Liu T, Wu Y, Zhong D, Zhou G, Su X, Xu J, Sotero CF, Sadduddin AA, Wu K, Chen X-G, Yan G (2018) Bacterial microbiota assemblage in *Aedes albopictus* mosquitoes and its impacts on larval development. *Mol Ecol* 27:2972–2985. <https://doi.org/10.1111/mec.14732>
- Watson DW, Mullens BA, Petersen JJ (1993) Behavioral fever response of *Musca domestica* (Diptera: Muscidae) to infection by *Entomophthora muscae* (Zygomycetes: Entomophthorales). *J Invert Pathol* 61:10–16. <https://doi.org/10.1006/jipa.1993.1003>
- Werren JH, Baldo L, Clark ME (2008) *Wolbachia*: master manipulators of invertebrate biology. *Nat Rev Microbiol* 6:741–751. <https://doi.org/10.1038/nrmicro1969>
- Wiklund T, Bylund G (1990) *Pseudomonas anguilliseptica* as a pathogen of salmonid fish in Finland. *Dis Aquat Org* 8:13–19
- Wiwatanaratnabutr I (2013) Geographic distribution of wolbachial infections in mosquitoes from Thailand. *J Invert Pathol* 114:337–340. <https://doi.org/10.1016/j.jip.2013.04.011>
- Wohlgemut K, Pierce RL, Kirkbride CA (1970) Bovine abortion associated with *Aeromonas hydrophila*. *J Am Med Assoc* 160:1001–1002
- Wylezich C, Papa A, Beer M, Höper D (2018) A versatile sample processing workflow for metagenomic pathogen detection. *Sci Rep* 8:13108. <https://doi.org/10.1038/s41598-018-31496-1>
- Yang M (2000) Long PCR improves *Wolbachia* DNA amplification: wsp sequences found in 76% of sixty-three arthropod species. *Insect Mol Biol* 9:393–405. <https://doi.org/10.1046/j.1365-2583.2000.00203.x>
- Yang CH, Li YH (2011) *Chromobacterium violaceum* infection: a clinical review of an important but neglected infection. *J Chin Med Assoc* 74:435–441. <https://doi.org/10.1016/j.jcma.2011.08.013>
- Ye-Ebiyo Y, Pollack RJ, Kiszewski A, Spielman A (2003) Enhancement of development of larval *Anopheles arabiensis* by proximity to flowering maize (*Zea Mays*) in turbid water and when crowded. *Am J Trop Med Hyg* 68:748–752. <https://doi.org/10.4269/ajtmh.2003.68.748>
- Yoon JH, Oh TK, Park YH (2004) *Kangiella koreensis* gen. nov., sp. nov. and *Kangiella aquimarina* sp. nov., isolated from a tidal flat of the Yellow Sea in Korea. *Int J Syst Evol Microbiol* 54:1829–1835. <https://doi.org/10.1099/ijms.0.63156-0>
- Zamora L, Vela AI, Palacios MA, Sánchez-Porro C, Svensson-Stadler LA, Domínguez L, Moore ERB, Ventosa A, Fernández-Garayzábal JF (2012) *Chryseobacterium viscerum* sp. nov., isolated from diseased fish. *Int J Syst Evol Microbiol* 62:2934–2940. <https://doi.org/10.1099/ijms.0.036699-0>
- Zhu D, Xie C, Huang Y, Sun J, Zhang W (2014) Description of *Comamonas serinivorans* sp. nov., isolated from wheat straw compost. *Int J Syst Evol Microbiol* 64:4141–4146. <https://doi.org/10.1099/ijms.0.066688-0>
- Zotzmann S, Steinbrink A, Schleich K, Frantzmann F, Xoumpholphakdy C, Spaeth M, Moro CV, Mavingui P, Klimpel S (2017) Bacterial diversity of cosmopolitan *Culex pipiens* and invasive *Aedes japonicus* from Germany. *Parasitol Res* 116:1899–1906. <https://doi.org/10.1007/s00436-017-5466-2>
- Zouache K, Raharimalala FN, Raquin V, van Tran-Van R, Lala HR, Ravelonandro P, Mavingui P (2011) Bacterial diversity of field-caught mosquitoes, *Aedes albopictus* and *Aedes aegypti*, from different geographic regions of Madagascar. *FEMS Microbiol Ecol* 75:377–389. <https://doi.org/10.1111/j.1574-6941.2010.01012.x>

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