#### **ORIGINAL ARTICLE**



# Expression patterns of HNF4α, TTF-1, and SMARCA4 in lung adenocarcinomas: impacts on clinicopathological and genetic features

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#### Abstract

**Introduction** HNF4 $\alpha$  expression and SMARCA4 loss were thought to be features of non-terminal respiratory unit (TRU)-type lung adenocarcinomas, but their relationships remained unclear.

**Materials and methods** HNF4 $\alpha$ -positive cases among 241 lung adenocarcinomas were stratified based on TTF-1 and SMARCA4 expressions, histological subtypes, and driver mutations. Immunohistochemical analysis was performed using xenograft tumors of lung adenocarcinoma cell lines with high *HNF4A* expression.

**Result** HNF4 $\alpha$ -positive adenocarcinomas(n = 33) were divided into two groups: the variant group(15 mucinous, 2 enteric, and 1 colloid), where SMARCA4 was retained in all cases, and the conventional non-mucinous group(6 papillary, 5 solid, and 4 acinar), where SMARCA4 was lost in 3/15 cases(20%). All variant cases were negative for TTF-1 and showed wild-type *EGFR* and frequent *KRAS* mutations(10/18, 56%). The non-mucinous group was further divided into two groups: TRU-type(n = 7), which was positive for TTF-1 and showed predominantly papillary histology(6/7, 86%) and *EGFR* mutations(3/7, 43%), and non-TRU-type(n = 8), which was negative for TTF-1, showed frequent loss of SMARCA4(2/8, 25%) and predominantly solid histology(4/8, 50%), and never harbored *EGFR* mutations. Survival analysis of 230 cases based on histological grading and HNF4 $\alpha$  expression revealed that HNF4 $\alpha$ -positive poorly differentiated (grade 3) adenocarcinoma showed the worst prognosis. Among 39 cell lines, A549 showed the highest level of *HNF4A*, immunohistochemically HNF4 $\alpha$  expression positive and SMARCA4 lost, and exhibited non-mucinous, high-grade morphology in xenograft tumors. **Conclusion** HNF4 $\alpha$ -positive non-mucinous adenocarcinomas included TRU-type and non-TRU-type cases; the latter tended to exhibit the high-grade phenotype with frequent loss of SMARCA4, and A549 was a representative cell line.

Keywords Lung adenocarcinoma · HNF4a · TTF-1 · SMARCA4 · KRAS · A549

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#### Introduction

Lung cancer is the leading cause of cancer-related death in many developed countries, including the United States and Japan [1, 2]. Adenocarcinoma is the most common histological subtype of lung cancer [3].

The existence of a distinct subset of lung adenocarcinomas arising from the terminal respiratory unit (TRU) was previously proposed by Yatabe et al. [4–6]. TRU-type lung adenocarcinomas, which are estimated to account for 75%–80% of primary lung adenocarcinomas, show histologically non-mucinous lepidic growth or papillary components and frequently express thyroid transcription factor-1 (TTF-1), which is the master regulator of lung differentiation at high levels [4–6]. The genetic backgrounds of TRU-type adenocarcinomas have been investigated in detail. *Epidermal growth factor receptor (EGFR)* mutations and *anaplastic lymphoma kinase (ALK)* fusions were found to be specific to TRU-type adenocarcinomas [7, 8]. However, limited information is currently available on non-TRU-type lung adenocarcinomas.

Non-TRU-type lung adenocarcinomas are not a single entity but include various histological and molecular subtypes [9–11]. Kim et al. reported that mucinous adenocarcinomas without TTF-1 expression can be regarded as non-TRU-type lung adenocarcinomas [12], and Yatabe et al. reported that the representative non-TRU-type lung adenocarcinomas were poorly differentiated and exhibited solid morphology [4]. Our previous report revealed that the main group of non-TRUtype lung adenocarcinomas were hepatocyte nuclear factor 4 $\alpha$  (HNF4 $\alpha$ )-positive adenocarcinomas with gastrointestinal features that frequently harbored *KRAS* mutations and *TTF-1* inactivating mutations/hypermethylation [11].

HNF4 $\alpha$  is one of the ligand-dependent transcription factors and specifically expressed in the liver and gastrointestinal organs (stomach, small intestine, and pancreas) but not in normal human lung tissue [13, 14]. HNF4 $\alpha$ regulates epithelial cell polarity and morphogenesis and plays an important role in gastrointestinal and hepatic cell differentiation [15–18]. HNF4 $\alpha$  also has a role as an oncoprotein and is involved in carcinogenesis, cancer growth, and invasion in various cancers such as hepatocellular cancer, colorectal cancer, gastric cancer, and Barret's esophageal cancer [19–22]. In the field of lung adenocarcinoma, HNF4 $\alpha$  was first reported as a characteristic marker for invasive mucinous adenocarcinomas (IMA) [23], which were regarded as non-TRU-type lung adenocarcinomas. However, the frequency of HNF4a expression in adenocarcinomas other than IMA is not well recognized, especially in non-mucinous lung adenocarcinomas.

SMARCA4 is one of the catalytic subunits in SWI/SNIF chromatin remodeling complexes and has recently been

suggested as a tumor suppressor [24–28]. We previously reported that the inactivating mutations of *SMARCA4* were correlated with the epithelial-mesenchymal transition (EMT) phenotype of lung adenocarcinoma cell lines, and loss of SMARCA4 expression was frequent in poorly differentiated non-TRU-type adenocarcinomas, showing a lack of lepidic growth, low expressions of TTF-1 and wild-type *EGFR* [28]. Both the expression of HNF4 $\alpha$  and the loss of SMARCA4 are considered characteristics of non-TRU-type adenocarcinomas, but their relationship remains unclear.

This is the first report focusing on the relationships among immunohistochemical expression patterns of HNF4 $\alpha$ , TTF-1, and SMARCA4, histological subtypes, and driver mutations. The whole sections of 241 primary lung adenocarcinomas were used in this study. HNF4 $\alpha$  expression was found not only in mucinous, enteric, and colloid adenocarcinomas but also in morphologically conventional non-mucinous adenocarcinomas. Some of them heterogeneously expressed HNF4 $\alpha$  and TTF-1, which were mutually exclusive within the same tumor. These cases were considered TRU-type adenocarcinomas and frequently harbored *EGFR* mutations. Moreover, TTF-1-negative and HNF4 $\alpha$ positive non-mucinous adenocarcinomas showed wild-type *EGFR* and frequent SMARCA4 loss, and tended to show a high-grade solid morphology and very poor prognosis.

We also examined the histological and immunohistochemical features of xenograft tumors derived from lung adenocarcinoma cell lines. The HNF4 $\alpha$ -positive lung adenocarcinoma cell lines (A549, Calu3, H1651, and H2405) all showed non-mucinous and high-grade morphology, and the A549 cell line showed a marked loss of SMARCA4, indicating that it was a representative cell line of HNF4 $\alpha$ positive, non-mucinous lung adenocarcinoma with highgrade morphology.

#### **Materials and methods**

#### **Case selections**

Details are shown in Online Resource 1.

#### **Histological analysis**

Details are shown in Online Resource 1.

#### Immunohistochemical analysis

Detailed staining and evaluation protocols are shown in Online Resource 2.

#### Sequencing using a next-generation sequencer

Mutations of primary lung tumors were investigated using the MINtS system, employing a MiSeq sequencer (Illumina K.K.), as previously reported [29]. The protocol of RNA extraction is shown in Online Resource 3.

#### **Cell lines and medium**

We used 39 non-squamous non-small-cell lung cancer cell lines. Detailed information is available in our previous reports [9, 28, 30–34].

## Mutational analysis of the 39 cell lines

Gene mutations in the 39 cell lines were based on our previous reports [9, 28, 30–34] and data from the Cancer Cell Line Encyclopedia (https://portals.broadinstitute.org/ccle/).

# Gene expression profile and single nucleotide polymorphism array analyses of 39 lung adenocarcinoma cell lines

A comprehensive gene expression analysis was undertaken using an oligonucleotide microarray (GeneChip Human Genome U133A; Affymetrix), as previously described [35–37]. Analysis with a single nucleotide polymorphism array (Human Mappings 50 K Xbal array; Affymetrix) was performed using the Genome Imbalance Map algorithm, as previously described [38].

# Xenograft tissues of lung adenocarcinoma cell lines

Details are shown in Online resource 4.

# **Statistical analysis**

For all statistical analyses, SPSS 26 (SPSS, Chicago, IL, USA) was used. Correlations between clinicopathological features and HNF4 $\alpha$  expression were analyzed using the  $\chi^2$  test. The Kaplan–Meier method was used for the calculation of survival curves, and the Wilcoxon method was used for comparisons. Multivariate analysis was performed using the Cox proportional hazards model. Differences were considered significant for *p*-values < 0.05.

# Results

# Clinicopathological features of HNF4 $\alpha$ -positive adenocarcinomas

We conducted an immunohistochemical analysis of HNF4α using 241 primary lung adenocarcinoma samples surgically resected at Jichi Medical University Hospital and found that 33 samples (14%) were positive for HNF4 $\alpha$ . Table 1 shows the relationships between HNF4 $\alpha$ expression and the clinicopathological features of 238 patients (241 samples). A total of 6 lung adenocarcinoma samples from the 3 patients with double primary lung adenocarcinomas were all positive for TTF-1 and negative for HNF4 $\alpha$ . All samples of mucinous (15/15, 100%), enteric (2/2, 100%), and colloid (1/1, 100%) adenocarcinoma exhibited HNF4 $\alpha$  expression. HNF4 $\alpha$  expression was detected in a proportion of acinar (4/24, 17%), papillary (6/123, 5%), and solid (5/43, 12%) adenocarcinomas. Representative figures of HNF4α-positive lung adenocarcinomas are shown in Fig. 1. None of the HNF4 $\alpha$ -positive lung adenocarcinomas showed hepatoid differentiation. None of the in-situ non-mucinous, minimally invasive, or lepidic adenocarcinoma samples (WHO grade 1), representing TRU-type adenocarcinomas, exhibited HNF4a expression.

Table 1 also shows the correlations among HNF4 $\alpha$  expression levels and driver mutations, clinicopathological factors and immunohistochemical patterns. In HNF4 $\alpha$ -positive samples, the frequency of *KRAS* mutations was significantly high (20/33, 61%) (p < 0.001), and the frequency of *EGFR* mutations was significantly low (3/33, 9%) (p < 0.001), whereas no common drive mutations other than *KRAS* and *EGFR* (e.g., *ALK*, *HER2*, *MET*, *BRAF*, *RET*, or *ROS1*) were found. *EGFR* and *KRAS* mutations were mutually exclusive.

HNF4 $\alpha$  expression was correlated with the advanced pT stage (pT2-pT4) (p = 0.001) and STAS (p = 0.001), but not correlated with pleural invasion, lymphatic or vessel invasion, intrapulmonary metastasis, or nodal involvement.

Immunohistochemically, HNF4 $\alpha$  expression was correlated with a loss of SMARCA4 (p = 0.035) and MUC5AC expression (p < 0.001), and inversely correlated with the expression of TTF-1 (p < 0.001) (Table 1), but seven samples were double-positive for TTF-1 and HNF4 $\alpha$ , including six papillary adenocarcinomas and one solid adenocarcinoma. Although the loss of SMARCA2 was not significantly more frequent in HNF4 $\alpha$ -positive adenocarcinomas, two of the four HNF4 $\alpha$ -positive Grade 3 adenocarcinomas that expressed SMARCA4 showed the loss of SMARCA2. **Table 1** Relationships among HNF4 $\alpha$  expression and clinicopathologic factors, including expression patterns of TTF-1 and SMARCA4, and genetic status of EGFR, KRAS, ALK, HER2, MET, BRAF, RET, and ROS1 in 241 primary lung adenocarcinomas

	HNF4α expression			
	Positive	Negative	<i>p</i> -value <sup>g</sup>	
Age <sup>a</sup>				
60 y/o over	29	170	0.476	
60 y/o less	4	35		
Sex <sup>a</sup>				
Male	21	111	0.309	
Female	12	94		
Smoking status <sup>b</sup>				
Never	11	87	0.263	
Current/Ex-smoker	22	112		
Hitsology				
AIS/MIA	0	9		
Lepidic adenocarcinoma	0	23		
Papillary adenocarcinoma	6	117		
Acinar adenocarcinoma	4	20		
Micropapillary adenocarcinoma	0	1		
Solid adenocarcinoma	5	38		
Mucinous adenocarcinoma	15	0		
Colloid adenocarcinoma	2	0		
	1	0		
Pathological I stage		102	0.001	
	6 27	103	<0.001	
	21	105		
Pathological Stage				
0-II	27	150	0.529	
III-1V	6	45		
Nodal involvement <sup>e</sup>				
Positive	8	56	0.631	
Negative	25	139		
Pleural invasion				
Positive	15	80	0.282	
Negative	18	128		
Pulmonary metastasis <sup>d</sup>				
Positive	1	13	0.419	
Negative	31	195		
Lymphatic invasion				
Positive	14	96	0.689	
Negative	19	112		
Vessel invasion				
Positive	16	100	0.556	
Negative	17	108		
STAS <sup>e</sup>				
G1-G2	12	190	0.001	
G3	6	18	0.001	
TTF-1		-		
Positive	7	187	< 0.001	
Negative	26	21	<u>001</u>	
MUCSAC				
Desitive	21	11	<0.001	
rositive	21	11	< 0.001	

	HNF4α e	xpression	
	Positive	Negative	<i>p</i> -value <sup>g</sup>
SMARCA4			
Retained	30	205	<u>0.035</u>
Lost	3	3	
SMARCA2			
Retained	30	197	0.184
Lost	3	8	
EGFR mutation <sup>f</sup>			
Positive	3	107	< 0.001
Negative	30	98	
KRAS mutation <sup>f</sup>			
Positive	13	19	< 0.001
Negative	20	186	
ALK fusion <sup>f</sup>			
Positive	0	2	0.569
Negative	33	203	
HER2 mutation <sup>f</sup>			
Positive	0	2	0.569
Negative	33	203	
MET mutation <sup>f</sup>			
Positive	0	9	0.22
Negative	33	196	
BRAF mutation <sup>f</sup>			
Positive	0	4	0.418
Negative	33	201	
RET fusion <sup>f</sup>			
Positive	0	2	0.569
Negative	33	203	
ROS1 fusion <sup>f</sup>			
Positive	0	3	0.484
Negative	33	202	

 $a_n = 238$  because three patients underwent double cancer

 ${}^{b}n = 232$  because smoking status was unknown in nine samples

 $c_n = 228$  because we excluded seven samples whose nodal involvement unknown and six double cancer samples

 $^{d}n = 240$  because pulmonary metastasis in one sample was unknown

 $e_n = 226$  because invasive mucinous adenocarcinomas (n = 15) were excluded

fn = 238 because we were not able to conduct the gene mutation analysis for three samples

<sup>g</sup>Underlined values indicate p < 0.05

## TTF-1 and SMARCA4 expression and gene mutation patterns differed in HNF4α-positive lung adenocarcinomas according to histology

Based on the 2021 WHO classification of thoracic tumors [39], we divided HNF4 $\alpha$ -positive adenocarcinoma cases (n = 33) into two groups: the variant group (mucinous, enteric, and colloid adenocarcinomas) (n = 18) and the conventional



Fig.1 HE (top) and HNF4 $\alpha$  staining (bottom) sections from five representative samples of HNF4 $\alpha$ -positive lung adenocarcinomas. Scale bar: 50  $\mu$ m

non-mucinous group (acinar, papillary, and solid adenocarcinomas) (n = 15) (Fig. 2a). All variant group cases were diffusely HNF4 $\alpha$ -positive and completely TTF-1-negative. None of them harbored *EGFR* mutations, but more than half of the cases harbored the *KRAS* mutation (10/18, 55.6%). In contrast, almost half of the cases in the non-mucinous group were double-positive for TTF-1 and HNF4 $\alpha$  (7/15, 46.7%), and their expression patterns were heterogenous and mutually exclusive within the same tumor (Online Resource 5a).

The three *EGFR*-mutated cases in the non-mucinous group were all double-positive for TTF-1 and HNF4 $\alpha$ . Given the high frequency of *EGFR* mutations in these double-positive cases (3/7, 43%), we speculated that the double-positive adenocarcinomas were of the TRU-type and that TTF-1-positive TRU-type adenocarcinomas were induced to express HNF4 $\alpha$ through the local loss of TTF-1 (e.g., by epigenetic silencing). In addition, all cases in the variant group retained SMARCA4 expression, but in the non-mucinous group, loss of SMARCA4 was detected in 3 of the 15 cases (20%), much more frequently than in HNF4 $\alpha$ -negative non-mucinous adenocarcinomas (3/208, 1.4%) (Fig. 2a and Table 1). Figure 2b shows histological images of two representative cases of HNF4 $\alpha$ -positive non-mucinous adenocarcinoma with the loss of SMARCA4.

The loss of SMARCA2, a paralog of SMARCA4, did not correlate with the expression of HNF4 $\alpha$  (Table 1) and was detected among HNF $\alpha$ -positive cases in both the variant group (5.6%, 1/18) and conventional non-mucinous group (13.3%, 2/15) (Fig. 2a and Online Resource 5b). MUC5AC expression was frequently positive in HNF4 $\alpha$ -positive cases (in both the variant and conventional groups), but was almost negative in TTF-1-positive cases (6/7, 85.7%) (Fig. 2a and Online Resource 5b).

# HNF4α-positive non-mucinous adenocarcinomas with high-grade morphology (WHO grade 3) showed the worst prognosis

The three-tiered grading system is the common prognostic indicator of non-mucinous lung adenocarcinomas [39]. In the present study, the 5-year survival rates of grade 1 (n = 29), grade 2 (n = 128), and grade 3 (n = 56) groups were 100%, 86.0%, and 61.4% respectively, and the survival rates differed significantly (grade 1 vs. grade 2: p = 0.032, grade 2 vs. grade 3: p = 0.002) (Fig. 3a). Next, for survival analysis, we re-classified non-mucinous adenocarcinoma cases of each grade group into HNF4 $\alpha$ -positive and HNF4 $\alpha$ -negative groups: HNF4 $\alpha$ -positive grade 3 group (n = 6), HNF4 $\alpha$ negative grade 3 group (n = 50), HNF4 $\alpha$ -positive grade 2 group (n = 9), HNF4 $\alpha$ -negative grade 2 group (n = 119), and HNF4 $\alpha$ -negative grade 1 group (n = 29), as well as the variant group (n = 17). Notably, the HNF4 $\alpha$ -positive grade 3 group showed worse prognosis than the HNF4 $\alpha$ -negative grade 3 group (3-year survival rates of 51.4% and 69.3%, respectively) (p = 0.024), showing the worst prognosis among the six groups (Fig. 3b).

We found that in grade 3 non-mucinous adenocarcinomas (n = 56), sex, pleural invasion, pStage, HNF4 $\alpha$  expression and MUC5AC expressions, were poor prognostic factors (Online Resource 6a). We performed a multivariate analysis, excluding the expression of MUC5AC, which correlated with the expression of HNF4 $\alpha$ , and found that the expression of HNF4 $\alpha$  and the pStage remained significant in the multivariate analysis (HR, 3.318; CI, 1.344–8.188 for HNF4 $\alpha$ expression and HR, 9.019; CI, 4.107–19.804 for pStage)



**Fig. 2 a** The histological subtypes (mucinous, enteric, colloid, papillary, acinar, and solid adenocarcinomas), histological grades, immunohistochemical expression of HNF4 $\alpha$ , TTF-1, SMARCA4, SMARCA2 and MUC5AC and genetic mutations of *EGFR* and *KRAS* in 33 HNF4 $\alpha$ -positive lung adenocarcinoma cases, with division into the variant and non-mucinous groups. **b** HE, SMARCA4, HNF4 $\alpha$ ,

(Online Resource 6b). Although HNF4 $\alpha$ -positive grade 3 non-mucinous adenocarcinomas frequently showed the loss of SMARCA4 (2/6, 33%), it was not identified as a poor prognostic factor (Online Resource 6a).

We also compared clinicopathological factors among the six groups (Online Resource 7) and found that advanced pT factor, advanced pStage, lymph node metastasis, vessel invasion, pleural invasion, and pulmonary metastasis were most frequently observed in the HNF4 $\alpha$ -positive grade 3 group, indicating that this group was the aggressive phenotype.

# Xenograft tumors of HNF4α-positive lung adenocarcinoma cell lines showed high-grade, non-mucinous morphology

Finally, we examined whether  $HNF4\alpha$ -positive grade 3 adenocarcinoma cell lines were present among the 39 non-squamous non-small cell lung cancer cell lines. Online Resource and TTF-1 staining of representative cases of HNF4 $\alpha$ -positive nonmucinous adenocarcinomas with loss of SMARCA4 (Cases 22 and 26). Both cases were grade 3 adenocarcinomas, SMARCA4 lost, HNF4 $\alpha$ -positive, and TTF-1-negative. Note that lymphoid cells within the tumor were SMARCA4-positive (100×magnification, Scale bar: 100 µm)

8 shows the gene-level expressions of *HNF4A* and *TTF-1* in the 39 cell lines. The four cell lines with the highest expression of *HNF4A* were A549, H2405, Calu-3, and H1651, in that order. Online Resource 8 also shows the common driver mutations of the 39 cell lines, and among the four *HNF4A*-high cell lines, *SMARCA4* and *KRAS* mutations were found in A549, *HER2* amplification was found in Calu-3, and no common driver mutations were found in H2405 or H1651.

Figure 4a summarizes (i) the genetic status of *EGFR*, *MET*, *HER2*, *KRAS*, and *SMARCA4* (upper panel), (ii) genelevel expressions of *HNF4A*, *TTF-1*, and *SMARCA4* (middle panel), and (iii) protein-level expressions of HNF4 $\alpha$ , TTF-1, SMARCA4, and ACTB (lower panel) for the four *HNF4A*high cell lines (A549, H2405, H1651, and Calu3), compared with the four representative TRU-type cell lines with *TTF-1*high expressions (HCC827, PC3, H1648, and H2009). The four *HNF4A*-high cell lines showed high HNF4 $\alpha$  expression and low level of TTF-1, except for H1651, at both the gene



Fig. 2 (continued)

and protein levels. A marked decrease in the expression level of SMARCA4 was only observed in *SMARCA4*-mutated A549, whereas the other three *HNF4A*-high cell lines exhibited SMARCA4 expression. An aberrant band of SMARCA4 was detected in H2405 by western blot analysis (Fig. 4a).

Next, using xenograft tumors of the four *HNF4A*-high cell lines, we examined the histological growth patterns in HE staining and performed immunohistochemical analysis for HNF4 $\alpha$ , TTF-1, and SMARCA4 (Fig. 4b). A549 and H1651 showed solid growth patterns, H2405 showed solid growth patterns with focal cribriform patterns, and Calu-3 showed fused glandular and papillary growth patterns (Fig. 4b, the top row). All of these growth patterns are features of grade 3 primary lung adenocarcinoma, and notably, none of the cell lines showed morphological features of mucinous adenocarcinoma. Immunohistochemically, all of the four *HNF4A*high cell lines were HNF4 $\alpha$ -positive and TTF-1-negative in the nucleus (Fig. 4b, the second and third row), but H1651, which showed high TTF-1 expression at both the gene and protein levels, exhibited intracytoplasmic TTF-1 expression. SMARCA4 expression was diffusely lost in the A549 xenograft tumor but retained in the other three cell lines (Fig. 4b, the bottom row).

#### Discussion

Here, we have shown that HNF4 $\alpha$  expression was not limited to mucinous, enteric, or colloid adenocarcinomas, which showed gastrointestinal morphology, but also appeared in morphologically conventional non-mucinous adenocarcinomas such as acinar, papillary, and solid adenocarcinomas.

In the present study (mostly Asian cases), the frequency of *KRAS* mutations was significantly higher in HNF4 $\alpha$ -positive adenocarcinomas (39.4%, 13/33 cases) than in HNF4 $\alpha$ -negative adenocarcinomas (9.2%, 19/207 cases). Based on The Cancer Genome Atlas data of 456 primary lung adenocarcinomas (mostly Caucasian cases), the frequency of *KRAS* mutations was not significantly Fig. 3 a Overall survival among 213 cases of non-mucinous adenocarcinomas categorized according to the WHO grading system. b The prognoses of 230 lung adenocarcinomas were analyzed in 6 groups; HNF4 $\alpha$  + G3: HNF4 $\alpha$ -positive grade 3 (n = 6), HNF4 $\alpha$ -G3: HNF4 $\alpha$ -negative grade 3 (n = 50), HNF4 $\alpha$  + G2: HNF4 $\alpha$ -positive grade 2 (n = 9), HNF4 $\alpha$ -G2: HNF4 $\alpha$ -negative grade 2 (n = 119), HNF4 $\alpha$ -G1:HNF4α-negative grade 1 (n = 29), and the variant group (n = 17). The samples with unknown prognoses (n = 5) and double carcinoma cases (n = 3)were excluded



higher in *HNF4A*-high cases (35.9%, 42/117 cases) than in *HNF4A*-low cases (29.5%, 100/339 cases) (p = 0.121) (Online resource 9). *KRAS* mutations in lung adenocarcinoma are more frequent in Caucasians than in Asians. We speculate that this is not because of the higher frequency of HNF4 $\alpha$ -positive cases in Caucasians, but because of the higher frequency of *KRAS* mutations in HNF4 $\alpha$ -negative adenocarcinomas (mainly TRU-type lung adenocarcinomas) in Caucasians.

The results obtained herein revealed the absence of common driver mutations other than *KRAS* mutations in mucinous adenocarcinomas (60%, 9/15 cases). *NRG1* gene fusion, which has been reported in *KRAS* wild-type mucinous adenocarcinomas, was not examined in the present study [40]. CD74-NRG1 fusion genes have been identified as driver oncogenes and ERBB2/ERBB3 receptors may be the target of these fusion genes [41]. We previously noted that the knockdown of HNF4A in lung adenocarcinoma cell lines suppressed the expression and phosphorylation of ERBB3 (data not shown). The relationship among ERBB3, HNF4A, and NRG1 fusion genes is a topic for future studies. Although they were not identified in the present study, ALK fusion genes have been detected in mucinous adenocarcinomas [42], but are TTF-1-positive TRU-type adenocarcinomas with a mucinous morphology, a distinct entity from HNF4 $\alpha$ -positive mucinous lung adenocarcinomas, exhibiting gastrointestinal features.

In the present study, the frequency of TTF-1 expression, loss of SMARCA4, and *EGFR* mutations differed according



**Fig. 4** a Genetic status of *EGFR*, *MET*, *HER2*, *KRAS*, and *SMARCA4* (upper panel), gene-level expressions of *HNF4A*, *TTF-1*, and *SMARCA4* (middle panel), and protein expression levels of HNF4 $\alpha$ , TTF-1, SMARCA4, and ACTB (lower panel) for 8 cell lines, including the four cell lines that highly express HNF4A (A549, H2405, H1651, and Calu-3) and the four cell lines that highly express TTF-1 (HCC827, PC3, H1648, and H2009). In the upper panel, the gray box indicates the presence of genetic abnormalities and the white box

indicates the absence of genetic abnormalities. In the middle lane, red means more than or equal to the average of each gene expression, orange means under the average but more than or equal to one-quarter of the average, and green means under one-quarter of the average. **b** The histological features and immunohistochemical expression patterns of HNF4 $\alpha$ , TTF-1, and SMARCA4 for the xenograft tumors of A549, H2405, H1651, and Calu-3

to histology. All cases of mucinous, enteric, and colloid adenocarcinoma were HNF4 $\alpha$ -positive, TTF-1-negative, and SMARCA4 retained, and showed a high frequency of *KRAS* mutations (10/18, 55.6%) and no *EGFR* mutations (0/18, 0%). A recent study showed that 16 cases of enteric and mucinous adenocarcinoma lacked common driver mutations except for *KRAS* mutations [43], indicating that mucinous, enteric, and colloid adenocarcinomas might form a single spectrum of HNF4 $\alpha$ -positive non-TRU-type adenocarcinomas showing gastrointestinal differentiation. However, enteric adenocarcinomas occasionally show focal TTF-1 expression and *EGFR* mutations [44], suggesting that some enteric adenocarcinomas. The etiology of enteric adenocarcinoma is controversial and requires further investigation.

Approximately half of the HNF4 $\alpha$ -positive non-mucinous adenocarcinomas were TTF-1-positive (7/15, 47%), and they frequently showing papillary predominant histology (6/7, 86%). They were often accompanied by a non-mucinous lepidic component (5/7, 71%) and often lacked MUC5AC expression (1/7, 14.3%). Furthermore, approximately half

of the cases harbored *EGFR* mutations (3/7, 43%), suggesting that the double-positive cases for HNF4 $\alpha$  and TTF-1 were derived from TRU-type lung adenocarcinomas. In these cases, HNF4 $\alpha$  and TTF-1 were expressed heterogeneously and were mutually exclusive within the same tumor. We speculate that focal loss of TTF-1 expression may be partially due to *TTF-1* gene hypermethylation, as previously reported [11].

The remaining half of the HNF4 $\alpha$ -positive non-mucinous adenocarcinomas were totally TTF-1-negative (8/8, 100%) and never harbored *EGFR* mutations (0/8, 0%), suggesting that they were the non-TRU-type lung adenocarcinomas. Half of them were poorly differentiated solid adenocarcinomas (4/8, 50%), potentially differing from mucinous, enteric, and colloid adenocarcinomas, which showed histologically gastrointestinal differentiation. We previously reported that HNF4 $\alpha$  was not a significant prognostic factor in lung adenocarcinomas at any stage [10], but confirmed that the expression of HNF4 $\alpha$  and a solid morphology were independent poor prognostic factors in advanced stage samples. In this study, HNF4 $\alpha$ -positive poorly differentiated (grade 3) non-mucinous adenocarcinomas were aggressive phenotypes and showed the worst prognosis, and HNF4 $\alpha$  expression was an independent prognostic factor in grade 3 non-mucinous lung adenocarcinomas. These results suggest that the expression of HNF4 $\alpha$  plays a distinctive role in the progression of lung adenocarcinoma and a poor prognosis.

HNF4α was recently shown to be involved in the growth and invasion of various cancers as oncoprotein [19–22]. A previous study that examined the expression of HNF4α and mucin profiles in lung mucinous adenocarcinomas [45] reported that HNF4α induced the expression of MUC3 in *KRAS*-mutated mucinous adenocarcinomas, which is a poor prognostic factor for mucinous adenocarcinomas of the breast and appendix [46, 47]. Chen et al. demonstrated that the HNF4α-BC200-FMR-positive feedback loop promoted cell growth and metastasis in *KRAS*-mutated, HNF4α-positive cell lines (A549) [48]. Therefore, the expression of HNF4α has potential as a therapeutic target in lung adenocarcinomas, particularly *KRAS*-mutated lung adenocarcinomas.

Herein, we found that HNF4 $\alpha$ -positive non-mucinous adenocarcinomas frequently showed loss of SMARCA4 (3/15, 20%), much more frequently than in mucinous, enteric, and colloid adenocarcinomas (0/18, 0%), and HNF4 $\alpha$ -negative non-mucinous adenocarcinomas (3/208, 1.4%). Additionally, loss of SMARCA4 was more frequently observed in HNF4 $\alpha$ -positive grade 3 adenocarcinomas (2/6, 33%). The function of SMARCA4 varies among different organs and diseases, and SMARCA4 inactivation in lung cancer is related to the loss of lung lineage transcription and early metastasis [49]. We speculated the HNF4 $\alpha$  expression in grade 3 adenocarcinoma may imply dedifferentiation associated with the inactivated SMARCA4 function, resulting in high-grade morphology and poor prognosis.

In the present study, loss of SMARCA4 was only found in 2.5% (6/241) of the lung adenocarcinoma samples. *SMARCA4* mutation rates were reported to account for approximately 8% of non-small cell lung cancers, but not all mutations resulted in loss of SMARCA4 expression [49]. Some variants of *SMARCA4* mutation may show intact SMARCA4 expression despite the loss of its function [50]. Note that, unlike HNF4 $\alpha$ , loss of SMARCA4 was not an independent prognostic factor in grade 3 adenocarcinomas in our study (Online Resource 6), but we did not investigate the mutational status of *SMARCA4* in SMARCA4-retained adenocarcinomas. Further studies are needed to elucidate the relationship between HNF4 $\alpha$  expression and the function of SMARCA4.

We also demonstrated that four lung adenocarcinoma cell lines (A549, H1651, H2405, and Calu-3) had high HNF4 $\alpha$ expression at both the gene and protein levels. All four cell lines tended to show high expression levels of *Vimentin* and *ZEB1* compared with the *TTF-1*-high cell lines, and relatively low expression of *CDH1* (Online Resource 10), indicating dedifferentiation or EMT. All four cell lines may be regarded as representatives of non-mucinous HNF4 $\alpha$ -positive lung adenocarcinomas with grade 3 morphology, but their immunohistochemical and genetic features varied. We propose that A549 is not a mucinous adenocarcinoma cell line [48], but A549 may be a representative cell line of HNF4 $\alpha$ -positive grade 3 lung adenocarcinomas with aggressive pathological features.

In conclusion, a subset of HNF4α-positive adenocarcinomas, such as mucinous adenocarcinomas with gastrointestinal differentiation, are TTF-1-negative and SMARCA4 retained, often showing KRAS mutations. In addition, some conventional non-mucinous adenocarcinomas are HNF4αpositive, which include not only TRU-type adenocarcinomas that are double-positive for TTF-1 and HNF4 $\alpha$  but also non-TRU-type poorly differentiated (grade 3) adenocarcinomas with frequent loss of SMARCA4 expression. HNF4αpositive grade 3 adenocarcinoma shows a very poor prognosis, and HNF4a expression is an independent prognostic factor in grade 3 lung adenocarcinomas. Thus, examining the status of HNF4 $\alpha$  expression is important for not only assuming the etiology and gene mutational status but also predicting the prognosis in non-mucinous adenocarcinomas. The A549 cell line may be considered a representative cell line of HNF4 $\alpha$ -positive grade 3 adenocarcinomas.

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**Data availability** Raw data can be obtained from the corresponding author upon reasonable request.

Code availability N/A.

#### Declarations

**Ethics approval** The study was approved by the Institutional Ethics Review Committee at Jichi Medical University, Tochigi, Japan.

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#### References

- Siegel RL, Miller KD, Jemal A (2019) Cancer statistics, 2019. CA Cancer J Clin 69(1):7–34. https://doi.org/10.3322/caac.21551
- Katanoda K, Hori M, Saito E et al (2021) Updated trends in cancer in Japan: incidence in 1985–2015 and mortality in 1958–2018-A sign of decrease in cancer incidence. J Epidermiol 31(7):426–450. https://doi.org/10.2188/jea.JE20200416
- Barta JA, Powell CA, Wisnivesky JP (2019) Global epidemiology of lung cancer. Ann Glob Health 85(1):8. https://doi.org/10.5334/ aogh.2419
- Yatabe Y, Mitsudomi T, Takahashi T (2002) TTF-1 expression in pulmonary adenocarcinomas. Am J Surg Pathol 26(6):767–773. https://doi.org/10.1097/00000478-200206000-00010
- Yatabe Y, Mitsudomi T (2007) Epidermal growth factor receptor mutations in lung cancers. Pathol Int 57(5):233–244. https://doi. org/10.1111/j.1440-1827.2007.02098.x
- Yatabe Y, Kosaka T, Takahashi T, Mitsudomi T (2005) EGFR mutation is specific for terminal respiratory unit type adenocarcinoma. Am J Surg Pathol 29(5):633–639. https://doi.org/10.1097/ 01.pas.0000157935.28066.35
- Yoshida A, Tsuta K, Nakamura H et al (2005) Comprehensive histologic analysis of ALK-rearranged lung carcinomas. Am J Surg Pathol 35(8):1226–1234. https://doi.org/10.1097/PAS.0b013 e3182233e06
- Takeuchi K, Soda M, Togashi Y et al (2012) RET, ROS1 and ALK fusions in lung cancer. Nat Med 18(3):378–381. https://doi.org/ 10.1038/nm.2658
- Matsubara D, Ishikawa S, Oguni S, Aburatani H, Fukayama M, Niki T (2010) Co-activation of epidermal growth factor receptor and c-MET defines a distinct subset of lung adenocarcinomas. Am J Pathol 177(5):2191–2204. https://doi.org/10.2353/ajpath.2010. 100217
- Matsubara D, Yoshimoto T, Soda M et al (2020) Reciprocal expression of trefoil factor-1 and thyroid transcription factor-1 in lung adenocarcinomas. Cancer sci 111(6):2183–2195. https://doi. org/10.1111/cas.14403
- 11. Matsubara D, Soda M, Yoshimoto T et al (2017) Inactivating mutations and hypermethylation of the NKX2-1/TTF-1 gene in non-terminal respiratory unit-type lung adenocarcinomas. Cancer Sci 108(9):1888–1896. https://doi.org/10.1111/cas.13313
- Kim YK, Shin DH, Kim KB et al (2015) MUC5AC and MUC5B enhance the characterization of mucinous adenocarcinomas of the lung and predict poor prognosis. Histopathology 67(4):520–528. https://doi.org/10.1111/his.12693
- Yeh MM, Bosch DE, Daoud SS (2019) Role of hepatocyte nuclear factor 4-alpha in gastrointestinal and liver diseases. World J of Gastroenterol 25(30):4074–4091. https://doi.org/10.3748/wjg. v25.i30.4074

- 14. Tanaka T, Jiang S, Hotta H et al (2006) Dysregulated expression of P1 and P2 promoter-driven hepatocyte nuclear factor-4alpha in the pathogenesis of human cancer. J Pathol 208(5):662–672. https://doi.org/10.1002/path.1928
- Babeu JP, Darsigny M, Lussier CR, Boudreau F (2009) Hepatocyte nuclear factor 4alpha contributes to an intestinal epithelial phenotype in vitro and plays a partial role in mouse intestinal epithelium differentiation. Am J Physiol Gastrointest Liver Physiol 297(1):G124-134. https://doi.org/10.1152/ajpgi.90690.2008
- Tuncer S, Sade-Memisoglu A, Keskus AG et al (2020) Enhanced expression of HNF4α during intestinal epithelial differentiation is involved in the activation of ER stress. FEBS J 287(12):2504– 2523. https://doi.org/10.1111/febs.15152
- Moore BD, Khurana SS, Huh WJ, Mills JC (2016) Hepatocyte nuclear factor 4α is required for cell differentiation and homeostasis in the adult mouse gastric epithelium. Am J Physiol Gastrointest Liver Physiol 311(2):G267–G275. https://doi.org/10.1152/ ajpgi.00195.2016
- Li J, Nign G, Duncan SA (2000) Mammalian hepatocyte differentiation requires the transcription factor HNF-4alpha. Genes Dev 14(4):464–474
- Chang HR, Nam S, Kook MC et al (2016) HNF4α is a therapeutic target that links AMPK to WNT signalling in earlystage gastric cancer. Gut 65(1):19–32. https://doi.org/10.1136/ gutjnl-2014-307918
- Huang Y, Xian L, Liu Z et al (2022) AMPKα2/HNF4A/BORIS/ GLUT4 pathway promotes hepatocellular carcinoma cell invasion and metastasis in low glucose microinviroment. Biochem Pharmacol 203:115198. https://doi.org/10.1016/j.bcp.2022.115198
- Darsigny M, Babeu JP, Seidman EG et al (2010) Hepatocyte nuclear factor-4alpha promotes gut neoplasia in mice and protects against the production of reactive oxygen species. Cancer Res 70(22):9423–9433. https://doi.org/10.1158/0008-5472. CAN-10-1697
- Nowicki-Osuch K, Zhuang L, Jammula S et al (2021) Molecular phenotyping reveals the identity of Barrett's esophagus and its malignant transition. Science 373(6556):760–767. https://doi.org/ 10.1126/science.abd1449
- Sugano M, Nagasaka T, Sasaki E et al (2013) HNF4α as a marker for invasive mucinous adenocarcinoma of the lung. Am J Surg Pathol 37(2):211–218. https://doi.org/10.1097/PAS.0b013e3182 6be303
- Medina PP, Romero OA, Kohno T et al (2008) Frequent BRG1/ SMARCA4-inactivating mutations in human lung cancer cell lines. Hum Mutat 29(5):617–622. https://doi.org/10.1002/humu. 20730
- Glaros S, Cirrincione GM, Palanca A, Metzger D, Reisman D (2008) Targeted knockout of BRG1 potentiates lung cancer development. Cancer Res 68(10):3689–3696. https://doi.org/10.1158/ 0008-5472.CAN-07-6652
- Romero OA, Setien F, John S et al (2012) The tumour suppressor and chromatin-remodelling factor BRG1 antagonizes Myc activity and promotes cell differentiation in human cancer. EMBO Mol Med 4(7):603–616. https://doi.org/10.1002/emmm.201200236
- Fukuoka J, Fujii T, Shih JH et al (2004) Chromatin remodeling factors and BRM/BRG1 expression as prognostic indicators in non-small cell lung cancer. Clin Cancer Res 10(13):4314–4324. https://doi.org/10.1158/1078-0432.CCR-03-0489
- Matsubara D, Kishaba Y, Ishikawa S et al (2013) Lung cancer with loss of BRG1/BRM, shows epithelial mesenchymal transition phenotype and distinct histologic and genetic features. Cancer Sci 104(2):266–273. https://doi.org/10.1111/cas.12065
- 29. Inoue Y, Shiihara J, Miyazawa H et al (2017) A highly specific and sensitive massive parallel sequencer-based test for somatic mutations in non-small cell lung cancer. PLoS ONE 12(4):e0176525. https://doi.org/10.1371/journal.pone.0176525

- Matsubara D, Kanai Y, Ishikawa S et al (2012) Identification of CCDC6-RET fusion in the human lung adenocarcinoma cell line, LC-2/ad. J Thorac Oncol 7(12):1872–1876. https://doi.org/10. 1097/JTO.0b013e3182721ed1
- Matsubara D, Ishikawa S, Oguni S, Aburatani H, Fulayama M, Niki T (2010) Molecular predictors of sensitivity to the MET inhibitor PHA665752 in lung carcinoma cells. J Thorac Oncol 5(9):1317–1324. https://doi.org/10.1097/JTO.0b013e3181e2a409
- 32. Ito T, Matsubara D, Tanaka I et al (2016) Loss of YAP1 defines neuroendocrine differentiation of lung tumors. Cancer Sci 107(10):1527–1538. https://doi.org/10.1111/cas.13013
- 33. Matsubara D, Yoshimoto T, Akolekar N et al (2023) Genetic and phenotypic determinants of morphologies in 3D cultures and xenografts of lung tumor cell lines. Cancer Sci 114(4):1757–1770. https://doi.org/10.1111/cas.15702
- Ibrahim R, Matsubara D, Osman W et al (2014) Expression of PRMT5 in lung adenocarcinoma and its significance in epithelialmesenchymal transition. Hum Pathol 45(7):1397–1405. https:// doi.org/10.1016/j.humpath.2014.02.013
- Ishii M, Hashimoto S, Tsutsumi S et al (2000) Direct comparison of GeneChip and SAGE on the quantitative accuracy in transcript profiling analysis. Genomics 68(2):136–143. https://doi.org/10. 1006/geno.2000.6284
- Midorikawa Y, Yamamoto S, Ishikawa S et al (2006) Molecular karyotyping of human hepatocellular carcinoma using singlenucleotide polymorphism arrays. Oncogene 25(40):5581–5590. https://doi.org/10.1038/sj.onc.1209537
- 37. Wang T, Niki T, Goto A et al (2007) Hypoxia increases the motility of lung adenocarcinoma cell line A549 via activation of the epidermal growth factor receptor pathway. Cancer sci 98(4):506– 511. https://doi.org/10.1111/j.1349-7006.2007.00428.x
- Ishikawa S, Komura D, Tsuji S et al (2005) Allelic dosage analysis with genotyping microarrays. Biochem Biophys Res Commun 333(4):1309–1314. https://doi.org/10.1016/j.bbrc.2005.06.040
- WHO Classification of Tumours Editorial Board (2021) Thoracic Tumours. Thoracic Tumours. 5th ed. Lyon, France: International Agency for Research on Cancer
- Shim HS, Kenudson M, Zheng Z et al (2015) Unique Genetic and Survival Characteristics of Invasive Mucinous Adenocarcinoma of the Lung. J Thorac Oncol 10(8):1156–1162. https://doi.org/10. 1097/JTO.000000000000579
- 41. Werr L, Plenker D, Dammert MA et al (2022) CD74-NRG1 Fusions Are Oncogenic In Vivo and Induce Therapeutically Tractable ERBB2:ERBB3 Heterodimerization. Mol Cancer Ther 21(5):821–830. https://doi.org/10.1158/1535-7163

- Duruisseaux M, Antoine M, Rabbe N et al (2017) Lepidic predominant adenocarcinoma and invasive mucinous adenocarcinoma of the lung exhibit specific mucin expression in relation with oncogenic drivers. Lung Cancer 109:92–100. https://doi.org/ 10.1016/j.lungcan.2017.05.007
- 43. Palmirotta R, Lovero D, D'Oronzo S et al (2020) Pulmonary enteric adenocarcinoma: an overview. Expert Rev Mol Med 22:e1. https://doi.org/10.1017/erm.2020.2
- Okada F, Takeda M, Fujii T et al (2024) Clinicopathological and genetic analyses of pulmonary enteric adenocarcinoma. J Clin Pathol 77(2):111–115. https://doi.org/10.1136/jcp-2022-208583
- 45. Guo M, Tomoshige K, Meister M et al (2017) Gene signature driving invasive mucinous adenocarcinoma of the lung. EMBO Mol Med 9(4):462–481. https://doi.org/10.15252/emmm.20160 6711
- 46. Rakha EA, Boyce RW, Abd El-Rehim D et al (2005) Expression of mucins (MUC1, MUC2, MUC3, MUC4, MUC5AC and MUC6) and their prognostic significance in human breast cancer. Mod Pathol 18(10):1295–1304. https://doi.org/10.1038/modpathol. 3800445
- 47. Shibahara H, Higashi M, Yokoyama S et al (2014) A comprehensive expression analysis of mucins in appendiceal carcinoma in a multicenter study: MUC3 is a novel prognostic factor. PLoS ONE 9(12):e115613. https://doi.org/10.1371/journal.pone.0115613
- Chen X, Zhao Y, Wang D et al (2021) The HNF4α-BC200-FMR1– Positive Feedback Loop Promotes Growth and Metastasis in Invasive Mucinous Lung Adenocarcinoma. Cancer Res 81(23):5904– 5918. https://doi.org/10.1158/0008-5472.CAN-21-0980
- Concepcion CP, Ma S, LaFave LM et al (2022) Smarca4 inactivation promotes lineage-specific transformation and early metastatic features in the lung. Cancer Disdov 12(2):562–585. https://doi. org/10.1158/2159-8290.CD-21-0248
- Schoenfeld AJ, Bandlamudi C, Lavery JA et al (2020) The genomic landscape of SMARCA4 alterations and associations with outcomes in patients with lung cancer. Clin Cancer Res 26(21):5701–5708. https://doi.org/10.1158/1078-0432. CCR-20-1825

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