COMMENTARY



Henning Morawietz¹

Received: 17 June 2023 / Revised: 17 June 2023 / Accepted: 19 June 2023 / Published online: 23 June 2023 © The Author(s) 2023

Keywords Cardiac function · Hydrogen peroxide · Isocitrate dehydrogenase · Metabolism · Redox signaling

Commentary on: Nanadikar MS, Vergel Leon AM, Guo J, van Belle GJ, Jatho A, Philip ES, Brandner AF, Böckmann RA, Shi R, Zieseniss A, Siemssen CM, Dettmer K, Brodesser S, Schmidtendorf M, Lee J, Wu H, Furdui CM, Brandenburg S, Burgoyne JR, Bogeski I, Riemer J, Chowdhury A, Rehling P, Bruegmann T, Belousov VV, Katschinski DM (2023) IDH3 γ functions as a redox switch regulating mitochondrial energy metabolism and contractility in the heart. Nat Commun 14:2123.

Metabolic disorders are important risk factors for cardiovascular diseases. The underlying molecular mechanisms are not well-understood. Therefore, the search for potential links between metabolism and cardiovascular system is a hot area of research in physiology and pathophysiology. Especially the role of reactive oxygen species (ROS) in this context is controversially discussed.

ROS play a major role in the regulation of cardiovascular function and the development of cardiovascular diseases [1]. ROS include reactive free oxygen radicals like superoxide anions ($O_2^{-\bullet}$) and stable non-radical oxidants like hydrogen peroxide (H_2O_2). Free oxygen radicals can have pathophysiological effects by reducing nitric oxide (NO) availability and increasing peroxynitrate formation and oxidative modification of different biomolecules [7]. On the other hand, H_2O_2 is considered as an important physiological signaling molecule due to its rather long half-life and its ability to pass membranes. Depending on its intracellular location and concentration, H_2O_2 could be protective or deleterious in



The type of transgenic models is important for the analysis of ROS in the cardiovascular system [9]. Transgenic models can differ in the type of affected ROS, their intracellular or tissue-specific ROS localization, and their effect on ROS concentration. High or low ROS concentrations can mediate different, sometimes opposite effects. Low H₂O₂ concentrations in the nanomolar range might be protective, while high H_2O_2 concentration above 100 µm can induce cytotoxic effects and cell death. The sensitivity to ROS differs between cells and tissues due to their antioxidative capacity. Furthermore, the assays used to measure ROS needs to be critically discussed [2]. Taking these points into account, the development of appropriate transgenic models is crucial to test the role of H_2O_2 as a potential molecular switch between metabolic and cardiovascular functions in vivo (Fig. 1).

This important scientific question has been addressed by Dörthe Katschinski and her colleagues [6]. The paper has been recently published in *Nature Communications* and is highlighted as the Paper of the Month by the German Physiological Society. In this excellent study, Nanadikar et al. developed a novel chemogenetic transgenic mouse model of inducible overexpression of the hydrogen peroxide sensor HyPer using D-amino acid oxidase (DAO) (HyPer-DAO mice) in cardiomyocytes. The primarily nuclear localization

🙆 Springer



Henning Morawietz Henning.Morawietz@tu-dresden.de

¹ Division of Vascular Endothelium and Microcirculation, Department of Medicine III, Faculty of Medicine and University Hospital Carl Gustav Carus, TUD Dresden University of Technology, Fetscherstr. 74, 01307 Dresden, Germany



Fig. 1 Impact of hydrogen peroxide activation in the cardiomyocytes of HyPer mice on cardiac contractility, mitochondrial isocitrate dehydrogenase 3 γ activity, ATP production, and metabolism. Parts of the figure are adapted from SMART – Servier Medical Art, Servier: https://smart.servier.com. Abbreviations: ATP, adenosine triphosphate; Cys148, Cysteine 148; D-ala, D-Alanine; DAO, D-amino acid oxidase; H₂O₂, hydrogen peroxide; HyPer, Hydrogen Peroxide sensor; IDH3 γ , isocitrate dehydrogenase 3 γ ; TCA, tricarboxylic acid

of HyPer in this model resulted in a better fine tuning of intracellular H₂O₂ formation in comparison to classical overexpression or knockout of endogenous H2O2 sources like Nox4. The HyPer probe was reversibly inducible in response to the DAO-specific substrate D-Alanine (D-ala). Using this model, increased endogenous H₂O₂ formation in cardiomyocytes was leading to impaired cardiac contractility. The authors identified novel redox-sensitive target proteins by proteomics. Hydrogen peroxide could reversibly oxidize protein cysteine thiols (-SH) to sulfenic acid (-SOH) finally resulting in disulfide-bridge formation. Such a reversible oxidation of a protein cysteine thiol by H_2O_2 is considered as a redox switch. Most of the redox-sensitive proteins identified in this screen had cysteine residues. The authors focused on proteins involved in mitochondrial metabolism and discovered the γ -subunit of the TCA cycle enzyme isocitrate dehydrogenase (IDH) 3 as a novel redox switch linking metabolism and cardiac function. HyPer-DAO overexpressing HEK cells exhibited a reversible redox modification and activity of IDH3 after activation of DAO in vitro. Endogenously produced H_2O_2 impaired ATP generation in the mitochondria. In a series of elegant experiments using microsecond molecular dynamics simulations and cysteine-gene-edited cells, they could prove that IDH3 γ cysteine (Cys) 148 and 284 are critically involved in the H_2O_2 -dependent regulation of IDH3 activity. Finally, they provide evidence that the redox modification of IDH3 γ Cys148 and Cys284 is responsible for ATP production in the mitochondria. In summary, these data shed light into a novel mechanism how mitochondrial metabolism could be linked via redox-sensitive mechanisms to cardiac function.

What are the implications of these exciting findings for cardiovascular physiology and pathophysiology? First, redox-sensitive modifications of these and other proteins or cysteine residues could provide novel regulatory mechanisms to fine tune the cross-talk between metabolism and cardiac function. Second, this novel transgenic mouse model might be used to study the role of H_2O_2 in important processes like myocardial ischemia and reperfusion. Especially interesting would be a detailed molecular analysis of the most probably redox-sensitive mechanisms of stunning [3].

In conclusion, the use of HyPer power might provide stunning novel views into redox-sensitive processes of cardiovascular physiology and pathophysiology.

Author contribution The author wrote the manuscript.

Funding Open Access funding enabled and organized by Projekt DEAL. This work was supported by research grants of Deutsche Forschungsgemeinschaft (DFG) (Grants MO 1695/4–1, MO 1695/5–1 and IRTG 2251), German Centre for Cardiovascular Research (DZHK) (Grant 81X2800207) and by funding of the Excellence Initiative by the German Federal and State Governments (Institutional Strategy, measure 'support the best', 3–25 2, Grant F-03661–553-41B-1250000).

Data availability Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Human and animal ethics Not applicable.

Consent for publication The author gives consent for the publication of identifiable details, which can include images and details within the text ("Material") to be published in the above journal and article.

Competing interests The author declares no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are

included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Amponsah-Offeh M, Diaba-Nuhoho P, Speier S, Morawietz H (2023) Oxidative stress, antioxidants and hypertension. Antioxidants (Basel) 12:281. https://doi.org/10.3390/antiox12020281
- Brandes RP, Rezende F, Schroder K (2018) Redox regulation beyond ROS: why ROS should not be measured as often. Circ Res 123:326–328. https://doi.org/10.1161/CIRCRESAHA.118. 313146
- Heusch G (2021) Myocardial stunning and hibernation revisited. Nat Rev Cardiol 18:522–536. https://doi.org/10.1038/ s41569-021-00506-7
- Langbein H, Brunssen C, Hofmann A, Cimalla P, Brux M, Bornstein SR, Deussen A, Koch E, Morawietz H (2016) NADPH oxidase 4 protects against development of endothelial dysfunction and atherosclerosis in LDL receptor deficient mice. Eur Heart J 37:1753–1761. https://doi.org/10.1093/eurheartj/ehv564
- Morawietz H (2018) Cardiovascular protection by Nox4. Cardiovasc Res 114:353–355. https://doi.org/10.1093/cvr/cvx252

- 6. Nanadikar MS, Vergel Leon AM, Guo J, van Belle GJ, Jatho A, Philip ES, Brandner AF, Bockmann RA, Shi R, Zieseniss A, Siemssen CM, Dettmer K, Brodesser S, Schmidtendorf M, Lee J, Wu H, Furdui CM, Brandenburg S, Burgoyne JR, Bogeski I, Riemer J, Chowdhury A, Rehling P, Bruegmann T, Belousov VV, Katschinski DM (2023) IDH3gamma functions as a redox switch regulating mitochondrial energy metabolism and contractility in the heart. Nat Commun 14:2123. https://doi.org/10.1038/s41467-023-37744-x
- Schroder K (2020) NADPH oxidases: current aspects and tools. Redox Biol 34:101512. https://doi.org/10.1016/j.redox.2020. 101512
- Schurmann C, Rezende F, Kruse C, Yasar Y, Lowe O, Fork C, van de Sluis B, Bremer R, Weissmann N, Shah AM, Jo H, Brandes RP, Schroder K (2015) The NADPH oxidase Nox4 has anti-atherosclerotic functions. Eur Heart J 36:3447–3456. https://doi.org/ 10.1093/eurheartj/ehv460
- Swain L, Nanadikar MS, Borowik S, Zieseniss A, Katschinski DM (2018) Transgenic organisms meet redox bioimaging: one step closer to physiology. Antioxid Redox Signal 29:603–612. https:// doi.org/10.1089/ars.2017.7469
- Zhao QD, Viswanadhapalli S, Williams P, Shi Q, Tan C, Yi X, Bhandari B, Abboud HE (2015) NADPH oxidase 4 induces cardiac fibrosis and hypertrophy through activating Akt/mTOR and NFkappaB signaling pathways. Circulation 131:643–655. https:// doi.org/10.1161/CIRCULATIONAHA.114.011079

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.