REVIEW ARTICLE

Gastric carcinogenesis

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Abstract

Introduction In most patients, gastric cancer is diagnosed in advanced stage. Curative treatment options are limited and the mortality is high. The process of gastric carcinogenesis is triggered by *Helicobacter pylori*-driven gastritis and is further characterized by its complexity of interaction with other risk factors. Health care systems are challenged for the improvement of prevention, early diagnosis, and effective treatments.

Methods An extensive literature research has been performed to elucidate the interplay between etiological factors involved in gastric carcinogenesis.

Results H. pylori is the most important carcinogen for gastric adenocarcinoma. Evidence is provided by experiments including animal studies as well as clinical observational and interventional studies in humans. Eradication has the potential to prevent gastric cancer and offers the greatest benefit if performed before premalignant changes of the gastric mucosa have occurred. Bacterial virulence factors are essential players in modulating the immune response involved in the initiation of the carcinogenesis in the stomach. Host genetic factors contribute to the regulation of the inflammatory response and in the aggravation of mucosal damage. The harmful role of environmental factors is restricted to salt intake and smoking of tobacco. The ingestion of fruit and vegetables has some protective effect. Conclusion Infection with H. pylori is the major risk factor for gastric cancer development, and thus, eradication of the Helicobacter offers a promising best option for prevention

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of the disease. Bacterial virulence, host genetic factors, and environmental influences are interacting in the multifactorial process of gastric carcinogenesis.

Keywords Gastric cancer \cdot *Helicobacter pylori* \cdot Intestinal metaplasia \cdot CagA \cdot Interleukin-1 beta

Introduction and background

The incidence of gastric cancer shows an impressive fall in certain areas in the world and still on a global scale represents the second most common cause of cancer-related death with about 700,000 patients dying each year [1]. In more than two thirds of patients, gastric cancer is advanced at initial presentation, and only palliative therapy can be offered at this stage. The clinical history of tumor-related symptoms is usually of recent date with alarm symptoms, weight loss, decreasing appetite, anemia, nausea, and vomiting. About 40% of patients never report dyspeptic symptoms within their recent or remote medical history [2]. The 1-year and 5-year survival rates are low with only 42% and 24%, respectively [3].

Therefore, the challenge is to combat gastric cancer with adequate and cost-effective screening programs that allow an early diagnosis. General population screening facilitates the detection of early gastric cancer but is implemented in large scales only in Japan and Korea [4, 5]. It is questionable if these programs are transferable to other regions in the world and particularly to areas where the incidence of gastric cancer is considerably lower. A retrospective calculation from Singapore estimated the cost-effectiveness of mass screening on the basis of a 24year period. According to this estimation, endoscopic screening for stomach cancer is cost-effective in moderate to high-risk populations but not in low-incidence regions as in most parts of Europe or North America [6]. There is high demand for cheaper and non-invasive tools of gastric cancer screening with the primary aim to detect premalignant lesions.

A further approach to fight gastric cancer is the assessment of changes in gene systems (genomics) and in protein expression (proteomics). With the help of these and other new technologies, new information can be gained about biological behavior and prognosis of the tumor concerning the response to current therapy modalities as well as for the development of new therapies.

Gastric cancer of the sporadic type has three major contributing factors. The current concept recognizes the trigger of *Helicobacter pylori* infection, specific host susceptibility, including familiary factors, and environmental influences. Completely distinct is the development of hereditary gastric cancer with familial association where a single gene mutation is responsible.

Hereditary gastric cancer

Hereditary genetic alterations are rare and reported in the literature to be associated with 5% of all gastric cancer cases with the majority of these being of the diffuse type according to Barber et al. [7]. The causative germ line mutation has been identified to be *CDH1*, the *e-cadherin* gene. Endoscopic surveillance has not been successful in sufficiently protected subjects with the *CDH1* gene mutation, and therefore the option of prophylactic gastrectomy has been identified in 76.5% to 100% of the resected specimens after prophylactic gastrectomy [8–10].

Mutation of the e-cadherin gene represents an initial step in the process of downstream gene activation leading towards further enhancement of proliferation and cancer formation. A malfunction of the e-cadherin/\beta-catenin complex frees β -catenin from the cell membrane and provides a higher cytoplasmatic pool, activating the Wntdependent signaling pathways that play a major role in gastric carcinogenesis [11]. By this and the β -cateninrelated activation of T cell factor-dependent transcription, a cascade is orchestrated including several target genes like cyclins and matrix metalloproteinases that are relevant for the initiation of invasive gastric cancer [12]. The mutation of the CDH 1 is of the recessive type with one allele being altered and the other subsequently being inactivated directly in the gastric tissue either by hypermethylation, somatic mutation, and loss of heterozygosity, intragenic deletion, or specific polymporphisms. However, alterations of the CDH 1 gene are reported not only for hereditary gastric cancer. Hypermethylation of the CDH 1

promoter can be detected in 40-80% of all primary gastric cancer [13].

Sporadic gastric cancer

Sporadic cancer accounts for 95% of all cases. It has a complex pathogenesis, in which several factors are involved including a familial risk and environmental factors. The key player is the *H. pylori* infection.

H. pylori

H. pylori-the epidemiological evidence

Infection with *H. pylori* is the major risk factor for the development of gastric cancer, and medical intervention represents the best option for prevention of the disease [14]. In 1994, the WHO classified *H. pylori* as class I carcinogen based mainly on epidemiological evidence for its role in the pathogenesis of gastric adenocarcinoma and has been reinforced in the more recent WHO contribution by the IARC in 2010 [15]. The prevalence of *H. pylori* infection varies among populations with rates up to 80% and is still high (40–50%) in people over 50 years of age in many Western countries.

First substantial evidence was obtained from animal studies on Mongolian gerbils, where *H. pylori* is a complete carcinogen that is capable of inducing gastric adenocarcinomas without the influence of any cocarcinogens [16, 17]. There were also hints for the multifactorial etiology of gastric cancer, when supplementation of the animals with nitrosamines lead to higher rates of cancer incidence and a more rapid carcinogenesis [18, 19].

Numerous studies attempted to assess the attributable risk of H. pylori infection for gastric carcinogenesis in the human system. In 2003, the Helicobacter and Cancer Collaborative Group combined data from all available case-control studies nested with prospective cohorts to assess more reliably the relative risk for gastric cancer. In this study, 1,228 patients were included and a clear association of H. pylori infection to non-cardia gastric cancer (OR 3.0; 95% CI 2.3-3.8) was reported. This association was even stronger when blood samples for H. pylori serology were obtained 10 years or longer before cancer diagnosis (OR 5.9; 95% CI 3.4-10.3) [20]. A possible explanation is the loss of H. pylori colonization in the presence of atrophic gastritis and IM, so that gastric cancer patients have a loss of anti-H. pylori antibodies at the time of disease manifestation.

Another meta-analysis from Asia analyzed 19 studies with approximately 2,500 gastric cancer patients and almost 4,000 matched controls resulting in an OR of 1.92 (95% CI

1.32–2.78) for the development of non-cardia gastric cancer in *H. pylori* positive patients [21] which was in concordance with a previous similar analysis [22].

For a more meticulous assessment of the H. pyloriattributable risk for gastric carcinogenesis, bacterial virulence factors had been taken into consideration for risk analysis. Several studies have shown that the risk of gastric cancer is influenced by the presence of the CagA, and moreover, antibodies to CagA persist longer in the serum. Ekström reported an increase of the H. pylori-attributable OR for non-cardia cancer from 2.2 to 21.0 if the CagA status was co-evaluated by immunoblot analysis [23]. In this analysis, 71% to 91% of gastric cancer in the studied population was attributable to H. pylori infection. A metaanalysis by Huang and colleagues revealed a further 1.64fold increase of gastric cancer risk for CagA-positive strains compared to CagA-negative ones (16 studies, n=5,054) [24]. Again, the lower risk is due to the loss of antibodies in advanced stage gastric cancer. In fact, in patients with early gastric cancer, the prevalence of H. pylori antibodies is much higher compared to individuals with advanced gastric cancer resulting in a different attributable risk [21]. Further relevance is given by the time of serum sampling. The associated OR for gastric cancer development in case of H. pylori infection is significantly higher if serum samples are taken within 90 days after tumor gastrectomy [25].

While most studies claim that *H. pylori* infection is only related to distal gastric cancer, we could recently demonstrate a high prevalence also in patients with proximally located adenocarcinomas. Approximately 80% of patients with proximal gastric cancer revealed positive evidence for an actual or past infection with *H. pylori* if correct allocation of the primary tumor is performed and adenocarcinomas of the distal esophagus are strictly excluded [26]. Finally, the risk for gastric carcinogenesis by *H. pylori* infection is equal in intestinal and diffuse type gastric cancer [26, 27].

H. pylori and bacterial virulence factors

Several bacterial virulence factors define the malignant potential of each *H. pylori* strain. The best investigated factor is the cag pathogenicity island (PAI) type IV secretion system that can induce a more severe inflammatory response and can increase the risk for gastric carcinogenesis [28–30]. This type IV secretion system is a prerequisite for translocation of pathogenetic virulence factors (e.g., CagA) of *H. pylori* into the epithelial cell [31, 32]. CagA is rapidly phosphorylated by host Srckinases and has subsequently the potential to change intracellular signal transduction and to disrupt epithelial cell junctions [33, 34]. The injected CagA leads to activation of the Ras-mitogen-activated protein kinase (MAPK) pathway, involving the Ras-dependent kinases ERK-1 and ERK-2, with further transactivation of hostrelated pathways [35-37]. CagA-dependent activation of the Ras-Erk cascade increases also IL-8 release and consequent NF-kB activation responsible for invasion of neutrophil granulocytes into the gastric mucosa [38]. NFκB-related carcinogenesis is enhanced by H. pylori-associated release of tumor necrosis factor- α -inducing protein (Tip- α) inducing high expression of TNF- α with further involvement of IL-8 and COX-2-dependent pathways [39, 40]. Interaction of CagA with the e-cadherin/ β -catenin system can lead to a direct transactivation of CDX1 and by this to metaplastic changes in the mucosa [41]. CagA is further thought to contribute to epithelial-mesenchymal transition, a hallmark of epithelial-derived carcinogenesis [42, 43].

Although the EPIC study has shown that the increased risk for gastric cancer in case of a positive CagA status is independent from environmental factors like diet, smoking habits, or body mass, recent data suggested that CagA expression can be increased under a high-salt diet leading also to increased IL8 secretion [44, 45]. Furthermore, it has been shown that genetic diversity of the Helicobacter plays an important role in H. pylori-driven carcinogenesis. Once injected in the host cell, CagA is phosphorylated at certain glutamate-isoleucine-tyrosine-alanine (EPIYA) motifs. Due to variations in the surrounding amino acid sequence, four distinct EPIYA-motifs are described (EPIYA-A, -B, -C, -D) [46, 47]. Prevalence of these motifs varies by region. They further influence the CagA-induced immune response as well as the related cancer risk. The odds ratio (OR) for gastric cancer development is 7.37 (95% CI 1.98-27.48) in case of one EPIYA-C segment and 32.5 (95% CI 8.41-125.58) in case of two or more segments [48].

A similar diversity has been identified for the vacuolating cytotoxin A (VacA) showing variations in its gene structure which can be divided into a signaling (s), a middle (m), and an intermediate (i) region [49]. After first identification of s1/m1 strains showing a higher attributable risk for gastric cancer development, more recently also, i1 strains have been demonstrated to be associated with not only dysplastic but also malignant invasive tissue formation [48, 50].

VacA has an inhibitory effect on GSK3 β - (glycogen synthase kinase 3- β)-regulated signaling pathways by phosphorylation through an Akt/PI3K- (phosphatidylinosi-tol-3-kinase) mediated pathway, which leads to a β -catenin release and furthermore modulation of apoptosis- and cell cycle-regulation [51, 52].

The outer-membrane protein BabA is expressed by 40– 95% of the *H. pylori* strains, in dependency of the geographic region [53]. Patients infected with a BabA positive strain show a higher density of bacterial colonization in the stomach and have an enhanced inflammation due to increased IL-8 levels [54]. *H. pylori* strains expressing all three genes (CagA, VacA, BabA) are associated with the highest risk for developing gastric cancer [55].

For a comprehensive review on further virulence factors of *H. pylori* and their interaction with intracellular signaling and host immune response mechanisms, see the article from Wroblewksi et al. [49].

Inflammatory response in the gastric mucosa

The inflammatory response to infection with *H. pylori* of the gastric mucosa is complex. A key role in the immune response to *H. pylori* and in the complex pathogenesis of *H. pylori*-related diseases is taken by regulatory T cells (Treg), mostly $CD4^+CD25^{+high}$ Treg [56].

Treg are associated with increasing bacterial colonization [57], chronic inflammatory changes [58, 59], and the expression of immuno-suppressive cytokines like IL-10, IL-17, and TGF- β [60, 61]. Eradication therapy of H. pylori infection leads to a significant reduction of Treg cells and corresponding cytokine levels in gastroduodenal mucosa [59]. In case of gastric cancer, numbers of Treg cells are increased both in the gastric mucosa and the peripheral blood [62-64]. Notably, the ratio of Th1/Th2derived cytokines is the highest in asymptomatic gastritis showing a steady decrease in gastric atrophy, intestinal metaplasia and intraepithelial neoplasia towards gastric adenocarcinoma. This is associated with a concomitant increase of the Treg cell compartment in the peripheral blood and the persistence of CagA positive strains favoring a Treg cell-mediated chronic inflammation [64].

A retrospective immunohistochemical study found increased numbers of FOXP3-expressing CD4⁺CD25⁺CD117^{low} Treg cells associated with vascular, lymphatic, and perineural invasion of gastric tumor cells. Higher numbers of Treg cells were correlated with advanced tumor stage and correlated negatively with overall survival of these patients [65].

Host factors

Gene polymorphisms of immune response genes

Genetic alterations of factors that modulate and mediate the inflammatory response to *H. pylori* infection have been suggested to play a major role in the development of gastric cancer. Among these are cytokine genes (e.g., *TNF-\alpha, IL-10, IL-8, IFN-\gamma*) involved in the adaptive immune system [66–68] and pattern recognition factors (*TLR-4, NOD-1, NOD-2*) initiating the innate immune system [69, 70]. Furthermore, variation of genes encoding for proteases [71], xenobiotic metabolism enzymes [72], cell cycle regulators [73, 74], mucins [75], HLA-molecules [76],

and DNA-repair enzymes [74, 77] have been reported to bear an increased risk for gastric cancer (Fig. 1).

IL-1 β is the most powerful proinflammatory cytokine produced in response to *H. pylori* infection, known to act also as a strong acid inhibitor on a molecular basis [78]. About a decade ago, it has been postulated that carriers of specific single nucleotide gene polymorphisms (SNPs) in the *IL-1\beta* gene or the gene of the IL-1-receptor antagonist (*IL-1RN*) have an up to fourfold increased risk for developing gastric cancer [79]. Numerous studies in various ethical groups have been published since and revealed conflicting results.

In a meta-analysis by Kamangar and colleagues, including 35 studies with about 5,500 patients and more than 7,800 controls, a decreasing association of these polymorphisms with gastric cancer risk was shown for the accumulative data until 2006 [80]. The OR for developing gastric cancer in case of present *IL-1* β gene-mutation varies from 0.82 to 1.99 depending on the geographic region of occurrence, the histological subtype of gastric cancer, and the genetic locus that is altered [80]. More recent metaanalyses report an increased risk for gastric cancer in case of SNPs of the locus *IL-1* β -511 *T* and of the *IL-1RN* for Caucasians if an adequate stratification of the included study populations is performed concerning ethnicity [81– 83]. We could not confirm this conclusion [84].

Furthermore, a functional relevance with respect to altered gene expression of defined cytokine haplotypes

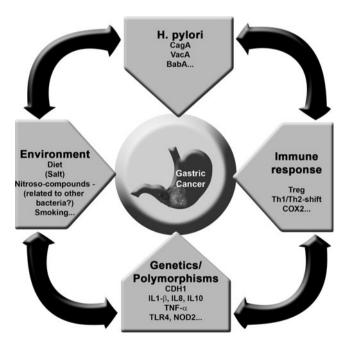


Fig. 1 Risk factors involved in gastric carcinogenesis. Gastric cancer is a multifactorial disease. Host genetic factors are mainly involved in modulation of the patient's immune response to *H. pylori* infection. The increased mucosal proliferation induced by exogen carcinogenic agents represents a further factor. There is a complex interplay between the main risk factors

was demonstrated for *IL-10* [85], *TNF-* α [86], and *IL-8* [87]. The majority of these studies identified significant associations between specific haplotypes and an increased risk for developing gastric cancer. In most cases (except *IL-10*), these cytokine polymorphisms result in a higher secretion of the corresponding cytokine leading to a stronger Th1-dominant immune response (Fig. 2).

A comprehensive review on host polymorphisms of immune-modulatory genes has recently been published by Wex et al. [88].

Interleukin-1 β and gastric physiology—interplay with H. pylori

The functional relevance of polymorphisms in the $IL-1\beta$ gene are based on several pleiotropic effects mediated by this cytokine. IL-1 β , mostly secreted by macrophages and to a lesser extent by epithelial and dentritic cells, induces the expression of other cytokines such as IL-12, TNF- α , IL-2, and interferons that subsequently shift the immune balance towards a mixed or Th1-predominant inflammation as it is seen in H. pylori-mediated gastritis [89-91]. The subsequent infiltration of granulocytes and lymphocytic cells leads to a chronic inflammatory condition that will last as long as the bacterium is colonizing the gastric mucosa. The degree of colonization and gastritis is dependent on various factors, like the presence and activity of regulatory T cells or the initial (naive) parietal cell mass reflecting the acid secretory capacity [60, 91-93]. An additional impact of IL-1ß is given by its cytoprotective, anti-ulcerative effects and capability to delay gastric emptying by modulating gastric motility [78]. Furthermore, IL-1ß

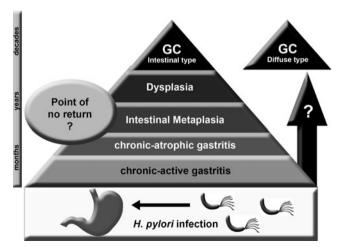


Fig. 2 Schematic image of the Correa-Model. Model of sequential mucosal alterations in the development of intestinal type gastric cancer on the basis of *H. pylori*-driven chronic active gastritis. In contrast to the adenoma-carcinoma sequence for colorectal cancer, order of appearance can be different and certain steps can be missed. A specific sequence leading to diffuse type gastric cancer is not described yet

regulates gastrin and histamine levels in the stomach and is capable to inhibit acid secretion from parietal cells [78, 94, 95]. Gastrin itself is important for an intact mucosal homeostasis in the stomach. A complex role in gastric carcinogenesis has been revealed, including the gastrininduced mediation of proliferation, angiogenesis, and tissue invasion (Fig. 3) [96].

Figueiredo et al. [97] were the first who explored the relationship between the *IL-1* polymorphisms and bacterial virulence factors. They analyzed different CagA and VacA genotypes of the *H. pylori* strains isolated from patients with gastric cancer or non-atrophic gastritis in context to the presence of the "IL-1 proinflammatory genotype" and identified combinations with either low or high risk, resulting in 6 to 87-fold differences in the OR for gastric cancer.

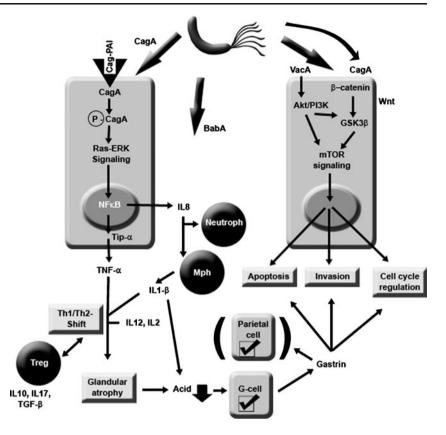
When *H. pylori* infection is predominantly located in the antrum, the proinflammatory effects of IL-1 β contribute to the development focal gastritis at this location. In this case, the acid secretion is mostly unchanged or can even be increased, resulting in "hyperchlorhydria", a condition that predisposes individuals to develop duodenal and gastric ulcer. If *H. pylori* colonization is predominantly located in the corpus of the stomach, the anti-secretory effect of IL-1 β leads to a vicious cycle by induction of hypochlorhydria which in turn facilitates the further spreading of *H. pylori* in the corpus and fundus mucosa. The resulting corpus-predominant inflammation goes hand in hand with mucosal atrophy and intestinal metaplasia.

Hwang et al. [98] showed the linkage between the "proinflammatory" genotype and elevated IL-1 β levels in human gastric mucosa. In a Japanese study, Furuta and colleagues identified higher atrophic and gastritis scores in *H. pylori*-infected patients with the proinflammatory $IL-1\beta$ -511 T/T genotype and demonstrated an elevation of the gastric juice pH as well as decreased pepsinogen I/II ratios [99]. Additionally, they showed that in the H. pylorinegative group, the *IL-1\beta* genotype did not have any effect on physiological or histological parameters of gastric mucosa [99]. Similar results were reported from Thailand, where the *IL-1\beta-511 TT* carriers had significantly higher IL- 1β levels in their antrum than corresponding controls, also influenced by bacterial factors (e.g., CagA type) [100]. In contrast, data from Korea showed higher mucosal IL-1ß levels in patients with "wildtype haplotype" compared to those with "proinflammatory haplotype" [101].

Environmental factors

Alimentary factors and the interaction with H. pylori

Animal studies have proven that ingestion of salt causes gastritis by destroying the mucosal barrier leading to inflammation and damage with diffuse erosions and Fig. 3 Overview on basic mechanisms of the mucosal response to *H. pylori*. Certain bacterial virulence factors of *H. pylori* can modulate intracellular signaling in the gastric epithelial cell. The resulting paracrine and endocrine mediators lead to a shift in the inflammatory response and may have a synergistic effect on the alteration of cell cycle regulation and the induction of proliferation and invasive tissue formation



epithelial degeneration [102-104]. The hypothesis tells that this leads to an increase of regeneration processes with the promotion of food-driven carcinogenesis. In 1997, the World Cancer Research Fund and the American Institute for Cancer Research analyzed 16 case-control studies, stating an association between salt or salted food and the risk of gastric cancer [105]. More recent case-control studies confirmed this association [106-109]. However, most studies including the presence of *H. pvlori* infection in the analysis demonstrated that a significant effect was only achieved in patients in which the gastric mucosa was already "pre-damaged" by H. pylori induced chronic-active inflammation [108, 110]. One prospective study examined the prevalence of gastric cancer in 2,476 men and women and evaluated the salt intake by a 70-item food frequency questionnaire. In this population, 93 cases of gastric cancer were identified, and the strongest effect of a high-salt diet was seen in patients with both H. pylori infection and atrophic changes of the mucosa [110].

H. pylori infection reduces the bioavailability of vitamin C, leading subsequently to decreased concentrations in the plasma and gastric juice [111, 112], whereas the luminal concentration of reactive oxygen species is increased [113]. In a randomised controlled trial carried out in Colombia, patients at high risk for gastric cancer were treated after *H. pylori* eradication with a combination of vitamin C and beta-carotene or placebo. In the group with vitamin C

supplementation, regression of premalignant lesions was observed [114]. The EPIC (European Prospective Investigation into Cancer and Nutrition) study, a large European prospective study including more than 500,000 participants in ten countries analyzed the association of plasma vitamin C with the risk for gastric cancer development, while taking into account factors like body mass index, total energy intake, smoking, and *H. pylori* status. This study did not confirm an association of vitamin C levels with gastric cancer development [115].

Chronic alcohol abuse is a major health care problem worldwide. More than 40 epidemiological, mostly retrospective studies have been carried out, and they did not confirm an association between chronic alcohol consumption and gastric cancer. Even intake of large amounts of alcohol (more than 200 g per day) was not significantly associated with increased risk for gastric adenocarcinomas, neither was the type of alcohol nor the concentration (percentage of ethanol) [116, 117]. In two prospective studies and four case–control studies, no significant correlation between alcohol consumption and cancer of the gastric cardia was reported [118].

A systematic review analyzed the relation between cigarette smoking and gastric cancer including 42 cohort, case–cohort, and case–control studies. The study provided solid evidence that smoking was significantly associated with an elevated relative risk (RR) for both gastric cardia (RR=1.87; 95% CI 1.31–2.67) and non-cardia cancers (RR=1.60; 95% CI 1.41–1.80). This conforms with a previous meta-analysis [119] and the results of the EPIC-study which estimated that 17.6% (95% CI 10.5–29.5%) of gastric cancer are related to smoking [120]. In conclusion, smoking seems to be the most important lifestyle risk factor for gastric cancer.

Gonzalez and colleagues recently published a review on the interaction of various environmental, especially alimentary carcinogenic agents with H. pylori infection [121]. Concerning smoking of tobacco, the OR for smoking and the development of gastric cancer varied between 1.04 and 9.2 in the studies analyzed, whereas the OR for H. pylori infection alone was higher than the OR for smoking in all but one study (OR 1.77-6.93). The OR for the combined presence of both risk factors was highest in all studies (2.3-19.0), although no trial revealed an additive effect, and the statistical analysis for risk factor interaction was negative [121]. The increased risk for gastric carcinogenesis in persons with a high intake of meat (total meat, processed meat, and red meat, OR 1.93-5.32) was demonstrated only in *H. pylori* positive subjects without the statistical term for interaction being positive [122].

Only for high-salt intake, a further direct influence on the risk for gastric cancer development by infection with H. *pylori* could be documented (p for interaction 0.047) [121].

The results for a negative association of high intake of fruits and vegetables with gastric carcinogenesis show especially for the plasma levels of vitamin C, vitamin E, and retinol, a protective effect only for the *H. pylori* positive group, although this interaction could again not be statistically confirmed [115, 123, 124].

Prevention of gastric cancer-clinical evidence

Eradication of H. pylori for gastric cancer prevention

It has been demonstrated by a recent meta-analysis that eradication of *H. pylori* has the potential to prevent gastric cancer [125]. In total, data of 6,695 patients were included showing that *H. pylori* eradication reduces gastric cancer risk (relative risk 0.65 [95% CI 0.43–0.98]). Overall, 56 of 3,307 (1.7%) untreated (control) participants developed gastric cancer compared with 37 of 3,388 (1.1%) treated patients.

Most data on the preventive effect of *H. pylori* eradication were generated in high incidence regions in Asia. A retrospective multicenter study from Japan analyzed the gastric cancer incidence in patients after *H. pylori* eradication for a 5-year follow-up in 23 centers including more than 3,000 patients [126]. Out of these, gastric cancer developed in 1% of patients who had been successfully eradicated and in 4% of patients with persistent infection

(OR 0.36; 95% CI 0.22–0.62). In an observational study by Uemura et al. [127], gastric cancer developed only in patients infected with *H*. pylori, but not in uninfected. Patients (1,526) with dyspeptic symptoms and an endoscopy-based diagnosis were followed up for 7.8 years. Thirty-six of the *H. pylori*-positive patients (2.9%) developed gastric cancer, but in no case (0%) gastric malignancy was detected among the *H. pylori*-negative patients [127].

In a prospective interventional study from Japan, patients with *H. pylori*-induced peptic ulcer disease have been followed up (n=1,342) for a median of 3.4 years [128]. Eradication therapy was not successful in 15% gastric cancer occurred in 0.8% of the successfully eradicated patients in contrast to 2.3% of patients with eradication failure [128].

The only prospective, randomized, placebo-controlled, population-based primary prevention study has been performed in China [129]. In total, 1,630 healthy individuals were recruited for randomization on either H. pylori eradication or placebo treatment. Within a follow-up period of 7.5 years, there have been 18 new cases of gastric cancer, 7 in the eradication group and 11 in the placebo group. Subgroup analysis revealed that all of the six patients with newly diagnosed gastric cancer presented with preneoplastic mucosal alterations (gastric atrophy, IM) at baseline, whereas no case of gastric cancer was diagnosed in without baseline mucosal changes (p=0.02) [129]. This gives the rational to search for the "point of no return" in the cascade of mucosal changes, when eradication of H. pylori will have no longer a protective effect to halt further progression of IM and atrophic gastritis towards gastric cancer (Fig. 2) [130].

The point of no return

A prospective observational study from Japan including 1,787 patients for a 9-year follow-up period described the clinical and histopathological characteristics of patients who still develop gastric cancer after eradication [131]. All of these patients presented with severe atrophic gastritis at baseline [131]. The risk for gastric carcinogenesis has been reported to be even increased with a significant correlation to the degree of baseline atrophy [132]. In a prospective study from Japan, 4,655 healthy, asymptomatic individuals have been followed up endoscopically for 7.7 years presenting a Hazard Ratio (HR) for gastric cancer of 7.13 in case of H. pylori infection without glandular atrophy, 14.85 if both conditions had been detected and 61.85 if H. pylori infection could not be detected any more due to the atrophic changes of the gastric mucosa [133]. It is noteworthy, that infection with H. pylori remains the major risk factor for the development of chronic atrophic gastritis [134].

Compared to chronic atrophic gastritis, there is less evidence for the influence of intestinal metaplasia on gastric cancer risk. However, in multiple logistic regression analysis, degree and distribution of IM have also been demonstrated to be independent risk factors for gastric cancer development [135]. A recent nationwide cohort study from the Netherlands, including more than 90,000 participants demonstrated a stepwise increase of gastric cancer incidence within 5 years follow-up and was 0.1% in patients with chronic atrophic gastritis, 0.25% in patients with IM, 0.6% in case of mild or moderate dysplasia, and 6% in case of severe dysplasia at baseline assessment, the latter resulting in a HR of 40.14 (95% CI 32.2–50.1) [136].

H. pylori eradication and reversibility of preneoplastic changes (atrophy, IM)

The degree of IM decreased after eradication therapy in patients endoscopically treated for early gastric cancer in a 3-year follow-up [137]. These results have been confirmed for healthy volunteers in China without any history of malignant disease [138]. Many studies have since been published with several limitations and flaws.

The data about the actual regression of IM or glandular atrophy is controversial. Several authors report an improvement only of inflammation within 1 year after eradication but no effect on the degree of metaplasia or atrophy [139, 140]. Rokkas and colleagues presented a meta-analysis on the long-term effect of eradication therapy on gastric histology [141]. The risk for atrophic gastritis was reduced by 45% in total (OR 0.554 [95% CI 0.372-0.825) and by almost 80% for alterations in the gastric corpus (OR 0.209 [95% CI 0.081-0.538]). An influence on IM could not be confirmed. This was also the case in a recent meta-analysis presented by Wang et al. [142]. The pooled data of the 12 studies analyzed showed that eradication of H. pylori led to significant improvement of atrophy in the gastric corpus, but not in the antrum, and to no improvement in IM. However, in this analysis, the degree of mucosal changes is not mentioned, which is crucial since only extensive atrophic alterations impact on gastric function.

It was suggested that the decisive factor whether or not there is an effect of *H. pylori* eradication is the time of follow-up. It has been shown that an improvement of mucosal inflammation can be documented within the first 6 to 12 months after eradication, whereas a follow-up period of more than 1 year is necessary to demonstrate an effect on IM and atrophic changes [143, 144]. Also, the side of biopsy sampling may be an important influencing factor [144]. Other risk factors for progression of IM/atrophy are age, male gender, abuse of alcohol, and drinking water from a well [145]. However, the issue of the necessary time of follow-up is not solved yet, since even after longer control periods, there might be no change or even progression in the degree of IM or atrophic gastritis, which refers one to the unsolved question of the "point of no return" [146].

The chance for prevention

The point of time at which *H. pylori* eradication still has an effect on gastric cancer prevention remains the burning question in this debate. Fukase and colleagues reported recently that even after endoscopic resection of early gastric cancer, recurrence of metachronous gastric cancer is significantly reduced by *H. pylori* eradication [147]. On the other hand, Wu et al. presented a cohort study with 80,255 patients included and came to the conclusion that the earlier *H. pylori* gets eradicated after peptic ulcer disease, the smaller is the risk for gastric cancer. Compared to the general population, patients that received early *H. pylori* eradication have no significant gastric cancer risk. Furthermore, independent protective factors for gastric cancer were identified in the frequent use of nonsteroidal anti-inflammatory drugs and aspirin [148, 149].

In the recently published updated Japanese guidelines for the management of *H. pylori* and related disease, it was finally stated that eradication is useful for the prevention of gastric cancer [150]. However, in case of present mucosal alterations at baseline assessment, a close and effective endoscopic follow-up and surveillance is mandatory, even after successful *H. pylori* eradication [151–153].

One important challenge of the near future is the development of efficient diagnostic tools for the identification of individuals at high risk for gastric cancer development to enable a cost-effective surveillance strategy even in low-incidence areas. The most promising tool is the noninvasive identification of subjects with chronic atrophic gastritis by blood analysis for pepsinogen 1 and 2 as well as the pepsinogen 1/2 ratio [154, 155].

Chemoprevention

The main focus in the issue of chemoprevention of gastric cancer has been on the effect of inhibition of cyclooxygenase-2 (COX-2) in the gastric mucosa. Expression of this enzyme is induced in case of gastric cancer development and shows an association to invasive growth, lymph vessel invasion, and nodal involvement, especially for intestinal type tumors [156–158]. Since changes in COX-2 expression can already be detected in early dyplastic lesions, induced by *H. pylori*-driven inflammation, they are considered to be an early event in gastric carcinogenesis and thus a target for prevention [159]. Several case–control and cohort studies have been carried out on this issue. The largest assessment was performed in the USA by Thun and colleagues who observed more than 650,000 individuals taking aspirin for different reasons for about 10 years [160]. A protective effect against gastric cancer, with an almost 50% reduction of gastric cancer incidence was observed in participants who took aspirin more often than 16 times a month (OR 0.53 [95% CI 0.34-0.81]). Non-aspirin NSAID use was analyzed from the population-based North Jutland prescription database and the Danish Cancer Registry, following more than 170,000 individuals for over 9 years showing a clear trend for gastric cancer risk reduction with reaching statistical significance (OR 0.70 [95% CI 0.4-1.1]) [161]. Several case-control studies confirm these data and an association between duration of aspirin or NSAID intake and gastric cancer risk reduction rose [162]. Even a positive effect on IM regression has been reported for patients on selective COX-2 inhibitors [163]. However, these data have not been transferred into clinical practice due to the reason of costeffectiveness and drug-induced morbidity, e.g., NSAIDinduced gastric ulcer bleeding, which has to be considered as a severe side-effect causing high morbidity and mortality.

Conclusion and a glimpse to the future

H. pylori eradication has proven to be effective in the prevention of gastric cancer and to be even more effective if provided before the development of atrophic changes and IM. A prophylactic vaccine as primary prevention is likely to represent the best strategy to protect from *H. pylori* and its consequences including gastric cancer. From the socio-economic point of view, the use of a prophylactic vaccine is cost-effective, and the vaccine development is more than desirable, especially considering decreasing eradication rates using antibiotic regimens [164, 165]. Animal studies as well as data from first trials on healthy human volunteers revealed promising results that deserve a committed follow-up [166, 167].

Conflicts of interest None.

References

- Parkin DM, Bray F, Ferlay J, Pisani P (2005) Global cancer statistics, 2002. CA Cancer J Clin 55:74–108
- Schmidt N, Peitz U, Lippert H, Malfertheiner P (2005) Missing gastric cancer in dyspepsia. Aliment Pharmacol Ther 21:813–820
- Coleman MP, Gatta G, Verdecchia A, Esteve J, Sant M, Storm H, Allemani C, Ciccolallo L, Santaquilani M, Berrino F (2003) EUROCARE-3 summary: cancer survival in Europe at the end of the 20th century. Ann Oncol 14(5):128–149

- Lee KJ, Inoue M, Otani T, Iwasaki M, Sasazuki S, Tsugane S (2006) Gastric cancer screening and subsequent risk of gastric cancer: a large-scale population-based cohort study, with a 13year follow-up in Japan. Int J Cancer 118:2315–2321
- Tashiro A, Sano M, Kinameri K, Fujita K, Takeuchi Y (2006) Comparing mass screening techniques for gastric cancer in Japan. World J Gastroenterol 12:4873–4874
- Dan YY, So JB, Yeoh KG (2006) Endoscopic screening for gastric cancer. Clin Gastroenterol Hepatol 4:709–716
- Barber M, Fitzgerald RC, Caldas C (2006) Familial gastric cancer—aetiology and pathogenesis. Best Pract Res Clin Gastroenterol 20:721–734
- Huntsman DG, Carneiro F, Lewis FR, MacLeod PM, Hayashi A, Monaghan KG, Maung R, Seruca R, Jackson CE, Caldas C (2001) Early gastric cancer in young, asymptomatic carriers of germ-line E-cadherin mutations. N Engl J Med 344:1904–1909
- 9. Suriano G, Yew S, Ferreira P, Senz J, Kaurah P, Ford JM, Longacre TA, Norton JA, Chun N, Young S, Oliveira MJ, Macgillivray B, Rao A, Sears D, Jackson CE, Boyd J, Yee C, Deters C, Pai GS, Hammond LS, McGivern BJ, Medgyesy D, Sartz D, Arun B, Oelschlager BK, Upton MP, Neufeld-Kaiser W, Silva OE, Donenberg TR, Kooby DA, Sharma S, Jonsson BA, Gronberg H, Gallinger S, Seruca R, Lynch H, Huntsman DG (2005) Characterization of a recurrent germ line mutation of the E-cadherin gene: implications for genetic testing and clinical management. Clin Cancer Res 11:5401–5409
- Lynch HT, Kaurah P, Wirtzfeld D, Rubinstein WS, Weissman S, Lynch JF, Grady W, Wiyrick S, Senz J, Huntsman DG (2008) Hereditary diffuse gastric cancer: diagnosis, genetic counseling, and prophylactic total gastrectomy. Cancer 112:2655–2663
- Park JG, Yang HK, Kim WH, Caldas C, Yokota J, Guilford PJ (2000) Report on the first meeting of the International Collaborative Group on Hereditary Gastric Cancer. J Natl Cancer Inst 92:1781– 1782
- Kolligs FT, Bommer G, Goke B (2002) Wnt/beta-catenin/tcf signaling: a critical pathway in gastrointestinal tumorigenesis. Digestion 66:131–144
- Graziano F, Humar B, Guilford P (2003) The role of the Ecadherin gene (CDH1) in diffuse gastric cancer susceptibility: from the laboratory to clinical practice. Ann Oncol 14:1705– 1713
- Malfertheiner P, Sipponen P, Naumann M, Moayyedi P, Megraud F, Xiao SD, Sugano K, Nyren O (2005) *Helicobacter pylori* eradication has the potential to prevent gastric cancer: a state-ofthe-art critique. Am J Gastroenterol 100:2100–2115
- (1994) Schistosomes, liver flukes and *Helicobacter pylori*. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Lyon, 7–14 June 1994. IARC Monogr Eval Carcinog Risks Hum 61:1–241
- Honda S, Fujioka T, Tokieda M, Satoh R, Nishizono A, Nasu M (1998) Development of *Helicobacter pylori*-induced gastric carcinoma in Mongolian gerbils. Cancer Res 58:4255–4259
- Watanabe T, Tada M, Nagai H, Sasaki S, Nakao M (1998) Helicobacter pylori infection induces gastric cancer in Mongolian gerbils. Gastroenterology 115:642–648
- Shimizu N, Inada K, Nakanishi H, Tsukamoto T, Ikehara Y, Kaminishi M, Kuramoto S, Sugiyama A, Katsuyama T, Tatematsu M (1999) *Helicobacter pylori* infection enhances glandular stomach carcinogenesis in Mongolian gerbils treated with chemical carcinogens. Carcinogenesis 20:669–676
- Maruta F, Sugiyama A, Ishida K, Ikeno T, Murakami M, Kawasaki S, Ota H, Tatematsu M, Katsuyama T (2000) Timing of *N*-methyl-*N*-nitrosourea administration affects gastric carcinogenesis in Mongolian gerbils infected with *Helicobacter pylori*. Cancer Lett 160:99–105

- Helicobacter and Cancer Collaborative Group (2001) Gastric cancer and *Helicobacter pylori*: a combined analysis of 12 case control studies nested within prospective cohorts. Gut 49:347– 353
- Huang JQ, Sridhar S, Chen Y, Hunt RH (1998) Meta-analysis of the relationship between *Helicobacter pylori* seropositivity and gastric cancer. Gastroenterology 114:1169–1179
- Eslick GD, Lim LL, Byles JE, Xia HH, Talley NJ (1999) Association of *Helicobacter pylori* infection with gastric carcinoma: a meta-analysis. Am J Gastroenterol 94:2373–2379
- Ekstrom AM, Held M, Hansson LE, Engstrand L, Nyren O (2001) *Helicobacter pylori* in gastric cancer established by CagA immunoblot as a marker of past infection. Gastroenterology 121:784–791
- 24. Huang JQ, Zheng GF, Sumanac K, Irvine EJ, Hunt RH (2003) Meta-analysis of the relationship between cagA seropositivity and gastric cancer. Gastroenterology 125:1636–1644
- Brenner H, Arndt V, Stegmaier C, Ziegler H, Rothenbacher D (2004) Is *Helicobacter pylori* infection a necessary condition for noncardia gastric cancer? Am J Epidemiol 159:252–258
- Bornschein J, Selgrad M, Warnecke M, Kuester D, Wex T, Malfertheiner P (2010) *H. pylori* infection is a key risk factor for proximal gastric cancer. Dig Dis Sci 55:3124–3131
- 27. Hansen S, Vollset SE, Derakhshan MH, Fyfe V, Melby KK, Aase S, Jellum E, Mccoll KE (2007) Two distinct aetiologies of cardia cancer; evidence from premorbid serological markers of gastric atrophy and *Helicobacter pylori* status. Gut 56:918–925
- Parsonnet J, Friedman GD, Orentreich N, Vogelman H (1997) Risk for gastric cancer in people with CagA positive or CagA negative *Helicobacter pylori* infection. Gut 40:297–301
- Enroth H, Kraaz W, Engstrand L, Nyren O, Rohan T (2000) Helicobacter pylori strain types and risk of gastric cancer: a case-control study. Cancer Epidemiol Biomark Prev 9:981–985
- 30. Blaser MJ, Perez-Perez GI, Kleanthous H, Cover TL, Peek RM, Chyou PH, Stemmermann GN, Nomura A (1995) Infection with *Helicobacter pylori* strains possessing cagA is associated with an increased risk of developing adenocarcinoma of the stomach. Cancer Res 55:2111–2115
- Backert S, Ziska E, Brinkmann V, Zimny-Arndt U, Fauconnier A, Jungblut PR, Naumann M, Meyer TF (2000) Translocation of the *Helicobacter pylori* CagA protein in gastric epithelial cells by a type IV secretion apparatus. Cell Microbiol 2:155–164
- Odenbreit S, Puls J, Sedlmaier B, Gerland E, Fischer W, Haas R (2000) Translocation of *Helicobacter pylori* CagA into gastric epithelial cells by type IV secretion. Science 287:1497–1500
- 33. Stein M, Bagnoli F, Halenbeck R, Rappuoli R, Fantl WJ, Covacci A (2002) c-Src/Lyn kinases activate *Helicobacter pylori* CagA through tyrosine phosphorylation of the EPIYA motifs. Mol Microbiol 43:971–980
- 34. Saadat I, Higashi H, Obuse C, Umeda M, Murata-Kamiya N, Saito Y, Lu H, Ohnishi N, Azuma T, Suzuki A, Ohno S, Hatakeyama M (2007) *Helicobacter pylori* CagA targets PAR1/ MARK kinase to disrupt epithelial cell polarity. Nature 447:330– 333
- 35. Higashi H, Tsutsumi R, Muto S, Sugiyama T, Azuma T, Asaka M, Hatakeyama M (2002) SHP-2 tyrosine phosphatase as an intracellular target of *Helicobacter pylori* CagA protein. Science 295:683–686
- 36. Tsutsumi R, Higashi H, Higuchi M, Okada M, Hatakeyama M (2003) Attenuation of *Helicobacter pylori* CagA x SHP-2 signaling by interaction between CagA and C-terminal Src kinase. J Biol Chem 278:3664–3670
- Hatakeyama M (2004) Oncogenic mechanisms of the *Helicobacter* pylori CagA protein. Nat Rev Cancer 4:688–694
- 38. Brandt S, Kwok T, Hartig R, Konig W, Backert S (2005) NFkappaB activation and potentiation of proinflammatory

Springer

responses by the Helicobacter pylori CagA protein. Proc Natl Acad Sci USA 102:9300-9305

- 39. Kuzuhara T, Suganuma M, Kurusu M, Fujiki H (2007) *Helicobacter pylori*-secreting protein Tipalpha is a potent inducer of chemokine gene expressions in stomach cancer cells. J Cancer Res Clin Oncol 133:287–296
- 40. Kim H, Lim JW, Kim KH (2001) *Helicobacter pylori*-induced expression of interleukin-8 and cyclooxygenase-2 in AGS gastric epithelial cells: mediation by nuclear factor-kappaB. Scand J Gastroenterol 36:706–716
- 41. Avidan B, Sonnenberg A, Schnell TG, Chejfec G, Metz A, Sontag SJ (2002) Hiatal hernia size, Barrett's length, and severity of acid reflux are all risk factors for esophageal adenocarcinoma. Am J Gastroenterol 97:1930–1936
- 42. Bagnoli F, Buti L, Tompkins L, Covacci A, Amieva MR (2005) *Helicobacter pylori* CagA induces a transition from polarized to invasive phenotypes in MDCK cells. Proc Natl Acad Sci USA 102:16339–16344
- Katoh M (2005) Epithelial-mesenchymal transition in gastric cancer (Review). Int J Oncol 27:1677–1683
- 44. Palli D, Masala G, Del Guidice G, Plebani M, Basso D, Berti D, Numans ME, Ceroti M, Peeters PH, Bueno de Mesquita HB, Buchner FL, Clavel-Chapelon F, Boutron-Ruault MC, Krogh V, Saieva C, Vineis P, Panico S, Tumino R, Nyren O, Siman H, Berglund G, Hallmans G, Sanchez MJ, Larranaga N, Barricarte A, Navarro C, Quiros J, Key T, Allen N, Bingham S, Khaw KT, Boeing H, Weikert C, Linseisen J, Nagel G, Overvad K, Thomsen RW, Tjonneland A, Olsen A, Trichoupoulou A, Trichopoulos D, Arvaniti A, Pera G, Kaaks R, Jenab M, Ferrari P, Nesi G, Carneiro F, Riboli E, Gonzalez CA (2007) CagA+ *Helicobacter pylori* infection and gastric cancer risk in the EPIC-EURGAST study. Int J Cancer 120:859–867
- Loh JT, Torres VJ, Cover TL (2007) Regulation of *Helicobacter* pylori cagA expression in response to salt. Cancer Res 67:4709– 4715
- 46. Higashi H, Yokoyama K, Fujii Y, Ren S, Yuasa H, Saadat I, Murata-Kamiya N, Azuma T, Hatakeyama M (2005) EPIYA motif is a membrane-targeting signal of *Helicobacter pylori* virulence factor CagA in mammalian cells. J Biol Chem 280:23130–23137
- 47. Naito M, Yamazaki T, Tsutsumi R, Higashi H, Onoe K, Yamazaki S, Azuma T, Hatakeyama M (2006) Influence of EPIYA-repeat polymorphism on the phosphorylation-dependent biological activity of *Helicobacter pylori* CagA. Gastroenterology 130:1181–1190
- Basso D, Zambon CF, Letley DP, Stranges A, Marchet A, Rhead JL, Schiavon S, Guariso G, Ceroti M, Nitti D, Rugge M, Plebani M, Atherton JC (2008) Clinical relevance of *Helicobacter pylori* cagA and vacA gene polymorphisms. Gastroenterology 135:91– 99
- Wroblewski LE, Peek RM Jr, Wilson KT (2010) *Helicobacter* pylori and gastric cancer: factors that modulate disease risk. Clin Microbiol Rev 23:713–739
- 50. Douraghi M, Talebkhan Y, Zeraati H, Ebrahimzadeh F, Nahvijoo A, Morakabati A, Ghafarpour M, Esmaili M, Bababeik M, Oghalaie A, Rakhshani N, Hosseini ME, Mohagheghi MA, Mohammadi M (2009) Multiple gene status in *Helicobacter pylori* strains and risk of gastric cancer development. Digestion 80:200–207
- 51. Nakayama M, Hisatsune J, Yamasaki E, Isomoto H, Kurazono H, Hatakeyama M, Azuma T, Yamaoka Y, Yahiro K, Moss J, Hirayama T (2009) *Helicobacter pylori* VacA-induced inhibition of GSK3 through the PI3K/Akt signaling pathway. J Biol Chem 284:1612–1619
- 52. Manente L, Perna A, Buommino E, Altucci L, Lucariello A, Citro G, Baldi A, Iaquinto G, Tufano MA, De LA (2008) The *Helicobacter pylori*'s protein VacA has direct effects on the

regulation of cell cycle and apoptosis in gastric epithelial cells. J Cell Physiol 214:582–587

- Prinz C, Hafsi N, Voland P (2003) *Helicobacter pylori* virulence factors and the host immune response: implications for therapeutic vaccination. Trends Microbiol 11:134–138
- 54. Rad R, Gerhard M, Lang R, Schoniger M, Rosch T, Schepp W, Becker I, Wagner H, Prinz C (2002) The *Helicobacter pylori* blood group antigen-binding adhesin facilitates bacterial colonization and augments a nonspecific immune response. J Immunol 168:3033–3041
- 55. Gerhard M, Lehn N, Neumayer N, Boren T, Rad R, Schepp W, Miehlke S, Classen M, Prinz C (1999) Clinical relevance of the *Helicobacter pylori* gene for blood-group antigen-binding adhesin. Proc Natl Acad Sci USA 96:12778–12783
- Vignali DA, Collison LW, Workman CJ (2008) How regulatory T cells work. Nat Rev Immunol 8:523–532
- Kandulski A, Malfertheiner P, Wex T (2010) Role of regulatory T-cells in *H. pylori*-induced gastritis and gastric cancer. Anticancer Res 30:1093–1103
- Jang TJ (2010) The number of Foxp3-positive regulatory T cells is increased in *Helicobacter pylori* gastritis and gastric cancer. Pathol Res Pract 15(206):34–38
- 59. Kindlund B, Sjoling A, Hansson M, Edebo A, Hansson LE, Sjovall H, Svennerholm AM, Lundin BS (2009) FOXP3expressing CD4(+) T-cell numbers increase in areas of duodenal gastric metaplasia and are associated to CD4(+) T-cell aggregates in the duodenum of *Helicobacter pylori*-infected duodenal ulcer patients. Helicobacter 14:192–201
- 60. Kandulski A, Wex T, Kuester D, Peitz U, Gebert I, Roessner A, Malfertheiner P (2008) Naturally occurring regulatory T cells (CD4+, CD25high, FOXP3+) in the antrum and cardia are associated with higher *H. pylori* colonization and increased gene expression of TGF-beta1. Helicobacter 13:295–303
- Robinson K, Kenefeck R, Pidgeon EL, Shakib S, Patel S, Polson RJ, Zaitoun AM, Atherton JC (2008) *Helicobacter pylori*induced peptic ulcer disease is associated with inadequate regulatory T cell responses. Gut 57:1375–1385
- 62. Enarsson K, Lundgren A, Kindlund B, Hermansson M, Roncador G, Banham AH, Lundin BS, Quiding-Jarbrink M (2006) Function and recruitment of mucosal regulatory T cells in human chronic *Helicobacter pylori* infection and gastric adenocarcinoma. Clin Immunol 121:358–368
- 63. Shen LS, Wang J, Shen DF, Yuan XL, Dong P, Li MX, Xue J, Zhang FM, Ge HL, Xu D (2009) CD4(+)CD25(+)CD127(low/-) regulatory T cells express Foxp3 and suppress effector T cell proliferation and contribute to gastric cancers progression. Clin Immunol 131:109–118
- 64. Wang SK, Zhu HF, He BS, Zhang ZY, Chen ZT, Wang ZZ, Wu GL (2007) CagA+ H pylori infection is associated with polarization of T helper cell immune responses in gastric carcinogenesis. World J Gastroenterol 13:2923–2931
- 65. Perrone G, Ruffini PA, Catalano V, Spino C, Santini D, Muretto P, Spoto C, Zingaretti C, Sisti V, Alessandroni P, Giordani P, Cicetti A, D'Emidio S, Morini S, Ruzzo A, Magnani M, Tonini G, Rabitti C, Graziano F (2008) Intratumoural FOXP3-positive regulatory T cells are associated with adverse prognosis in radically resected gastric cancer. Eur J Cancer 44:1875–1882
- 66. Hou L, El-Omar EM, Chen J, Grillo P, Rabkin CS, Baccarelli A, Yeager M, Chanock SJ, Zatonski W, Sobin LH, Lissowska J, Fraumeni JF Jr, Chow WH (2007) Polymorphisms in Th1-type cell-mediated response genes and risk of gastric cancer. Carcinogenesis 28:118–123
- 67. Crusius JB, Canzian F, Capella G, Pena AS, Pera G, Sala N, Agudo A, Rico F, Del GG, Palli D, Plebani M, Boeing H, Bueno-de-Mesquita HB, Carneiro F, Pala V, Save VE, Vineis P, Tumino R, Panico S, Berglund G, Manjer J, Stenling R,

Hallmans G, Martinez C, Dorronsoro M, Barricarte A, Navarro C, Quiros J, Allen N, Key TJ, Binghan S, Caldas C, Linseisen J, Kaaks R, Overvad K, Tjonneland A, Buchner FC, Peeters PH, Numans ME, Clavel-Chapelon F, Trichopoulou A, Lund E, Jenab M, Rinaldi S, Ferrari P, Riboli E, Gonzalez CA (2008) Cytokine gene polymorphisms and the risk of adenocarcinoma of the stomach in the European prospective investigation into cancer and nutrition (EPIC-EURGAST). Ann Oncol 19:1894–1902

- 68. Machado JC, Figueiredo C, Canedo P, Pharoah P, Carvalho R, Nabais S, Castro AC, Campos ML, Van Doorn LJ, Caldas C, Seruca R, Carneiro F, Sobrinho-Simoes M (2003) A proinflammatory genetic profile increases the risk for chronic atrophic gastritis and gastric carcinoma. Gastroenterology 125:364–371
- 69. Santini D, Angeletti S, Ruzzo A, Dicuonzo G, Galluzzo S, Vincenzi B, Calvieri A, Pizzagalli F, Graziano N, Ferraro E, Lorino G, Altomare A, Magnani M, Graziano F, Tonini G (2008) Toll-like receptor 4 Asp299Gly and Thr399Ile polymorphisms in gastric cancer of intestinal and diffuse histotypes. Clin Exp Immunol 154:360–364
- 70. Wex T, Ebert MP, Kropf S, Dierkes J, Schuttler K, Rocken C, Hocker M, Malfertheiner P (2008) Gene polymorphisms of the NOD-2/CARD-15 gene and the risk of gastric cancer in Germany. Anticancer Res 28:757–762
- Ye S (2000) Polymorphism in matrix metalloproteinase gene promoters: implication in regulation of gene expression and susceptibility of various diseases. Matrix Biol 19:623–629
- 72. Geddert H, Kiel S, Zotz RB, Zhang J, Willers R, Gabbert HE, Sarbia M (2003) Polymorphism of p16 INK4A and cyclin D1 in adenocarcinomas of the upper gastrointestinal tract. J Cancer Res Clin Oncol 131:803–808
- 73. Capella G, Pera G, Sala N, Agudo A, Rico F, Del GG, Plebani M, Palli D, Boeing H, Bueno-de-Mesquita HB, Carneiro F, Berrino F, Vineis P, Tumino R, Panico S, Berglund G, Siman H, Nyren O, Hallmans G, Martinez C, Dorronsoro M, Barricarte A, Navarro C, Quiros J, Allen N, Key T, Bingham S, Caldas C, Linseisen J, Nagel G, Overvad K, Tjonneland A, Boshuizen HC, Peeters PH, Numans ME, Clavel-Chapelon F, Trichopoulou A, Lund E, Jenab M, Kaaks R, Riboli E, Gonzalez CA (2008) DNA repair polymorphisms and the risk of stomach adenocarcinoma and severe chronic gastritis in the EPIC-EURGAST study. Int J Epidemiol 37:1316–1325
- 74. Agundez JA (2008) Polymorphisms of human *N*-acetyltransferases and cancer risk. Curr Drug Metab 9:520–531
- 75. Silva F, Carvalho F, Peixoto A, Seixas M, Almeida R, Carneiro F, Mesquita P, Figueiredo C, Nogueira C, Swallow DM, Amorim A, David L (2001) MUC1 gene polymorphism in the gastric carcinogenesis pathway. Eur J Hum Genet 9:548–552
- 76. Magnusson PKE, Enroth H, Eriksson I, Held M, Nyren O, Engstrand L, Hansson LE, Gyllensten UB (2001) Gastric cancer and human leukocyte antigen: distinct DQ and DR alleles are associated with development of gastric cancer and infection by *Helicobacter pylori*. Cancer Res 61:2684–2689
- 77. Ikeda S, Sasazuki S, Natsukawa S, Shaura K, Koizumi Y, Kasuga Y, Ohnami S, Sakamoto H, Yoshida T, Iwasaki M, Tsugane S (2008) Screening of 214 single nucleotide polymorphisms in 44 candidate cancer susceptibility genes: a case–control study on gastric and colorectal cancers in the Japanese population. Am J Gastroenterol 103:1476–1487
- El-Omar EM (2001) The importance of interleukin lbeta in Helicobacter pylori associated disease. Gut 48:743–747
- El-Omar EM, Carrington M, Chow WH, Mccoll KE, Bream JH, Young HA, Herrera J, Lissowska J, Yuan CC, Rothman N, Lanyon G, Martin M, Fraumeni JF Jr, Rabkin CS (2000) Interleukin-1 polymorphisms associated with increased risk of gastric cancer. Nature 404:398–402

- Kamangar F, Cheng C, Abnet CC, Rabkin CS (2006) Interleukin-1B polymorphisms and gastric cancer risk—a metaanalysis. Cancer Epidemiol Biomark Prev 15:1920–1928
- Wang P, Xia HH, Zhang JY, Dai LP, Xu XQ, Wang KJ (2007) Association of interleukin-1 gene polymorphisms with gastric cancer: a meta-analysis. Int J Cancer 120:552–562
- 82. Vincenzi B, Patti G, Galluzzo S, Pantano F, Venditti O, Santini D, Ruzzo A, Schiavon G, Caraglia M, Marra M, Graziano F, Tonini G (2008) Interleukin 1beta-511T gene (IL1beta) polymorphism is correlated with gastric cancer in the Caucasian population: results from a meta-analysis. Oncol Rep 20:1213–1220
- 83. Camargo MC, Mera R, Correa P, Peek RM Jr, Fontham ET, Goodman KJ, Piazuelo MB, Sicinschi L, Zabaleta J, Schneider BG (2006) Interleukin-1beta and interleukin-1 receptor antagonist gene polymorphisms and gastric cancer: a meta-analysis. Cancer Epidemiol Biomark Prev 15:1674–1687
- 84. Wex T, Leodolter A, Bornschein J, Kuester D, Kahne T, Kropf S, Albrecht C, Naumann M, Roessner A, Malfertheiner P (2010) Interleukin 1 beta (IL1B) gene polymorphisms are not associated with gastric carcinogenesis in Germany. Anticancer Res 30:505– 511
- 85. Suarez A, Castro P, Alonso R, Mozo L, Gutierrez C (2003) Interindividual variations in constitutive interleukin-10 messenger RNA and protein levels and their association with genetic polymorphisms. Transplantation 75:711–717
- Wilson AG, Symons JA, McDowell TL, McDevitt HO, Duff GW (1997) Effects of a polymorphism in the human tumor necrosis factor alpha promoter on transcriptional activation. Proc Natl Acad Sci USA 94:3195–3199
- 87. Ohyauchi M, Imatani A, Yonechi M, Asano N, Miura A, Iijima K, Koike T, Sekine H, Ohara S, Shimosegawa T (2005) The polymorphism interleukin 8–251 A/T influences the susceptibility of *Helicobacter pylori* related gastric diseases in the Japanese population. Gut 54:330–335
- Wex T, Bornschein J, Malfertheiner P (2009) Host polymorphisms of immune regulatory genes as risk factors for gastric cancer. Minerva Gastroenterol Dietol 55:395–408
- Dinarello CA (1998) Interleukin-1, interleukin-1 receptors and interleukin-1 receptor antagonist. Int Rev Immunol 16:457–499
- Lindholm C, Quiding-Jarbrink M, Lonroth H, Hamlet A, Svennerholm AM (1998) Local cytokine response in *Helicobacter pylori*-infected subjects. Infect Immun 66:5964–5971
- 91. Goll R, Gruber F, Olsen T, Cui G, Raschpichler G, Buset M, Asfeldt AM, Husebekk A, Florholmen J (2007) *Helicobacter pylori* stimulates a mixed adaptive immune response with a strong T-regulatory component in human gastric mucosa. Helicobacter 12:185–192
- Perasso A, Testino G, De AP, Augeri C, De GR (1991) Gastric chief cell mass in chronic gastritis. Count and relationships to parietal cell mass and functional indices. Hepatogastroenterology 38(1):63–66
- Testino G (1997) Parietal cell mass, hydrochloric acid secretion, and *Helicobacter pylori*. Am J Gastroenterol 92:1070–1071
- 94. Prinz C, Neumayer N, Mahr S, Classen M, Schepp W (1997) Functional impairment of rat enterochromaffin-like cells by interleukin 1 beta. Gastroenterology 112:364–375
- 95. Garcia-Zaragoza E, Hernandez C, Barrachina MD, Esplugues JV (2003) Interleukin 1 beta-induced inhibition of gastric acid secretion involves glutamate, NO and cGMP synthesis in the brain. Naunyn Schmiedebergs Arch Pharmacol 367:22–27
- 96. Watson SA, Grabowska AM, El-Zaatari M, Takhar A (2006) Gastrin—active participant or bystander in gastric carcinogenesis? Nat Rev Cancer 6:936–946
- 97. Figueiredo C, Machado JC, Pharoah P, Seruca R, Sousa S, Carvalho R, Capelinha AF, Quint W, Caldas C, Van Doorn LJ,

Carneiro F, Sobrinho-Simoes M (2002) *Helicobacter pylori* and interleukin 1 genotyping: an opportunity to identify high-risk individuals for gastric carcinoma. J Natl Cancer Inst 94:1680–1687

- 98. Hwang IR, Kodama T, Kikuchi S, Sakai K, Peterson LE, Graham DY, Yamaoka Y (2002) Effect of interleukin 1 polymorphisms on gastric mucosal interleukin 1beta production in *Helicobacter pylori* infection. Gastroenterology 123:1793–1803
- 99. Furuta T, El-Omar EM, Xiao F, Shirai N, Takashima M, Sugimura H (2002) Interleukin 1beta polymorphisms increase risk of hypochlorhydria and atrophic gastritis and reduce risk of duodenal ulcer recurrence in Japan. Gastroenterology 123:92– 105
- 100. Vilaichone RK, Mahachai V, Tumwasorn S, Wu JY, Graham DY, Yamaoka Y (2005) Gastric mucosal cytokine levels in relation to host interleukin-1 polymorphisms and *Helicobacter pylori* cagA genotype. Scand J Gastroenterol 40:530–539
- 101. Chang YW, Jang JY, Kim NH, Lee JW, Lee HJ, Jung WW, Dong SH, Kim HJ, Kim BH, Lee JI, Chang R (2005) Interleukin-1B (IL-1B) polymorphisms and gastric mucosal levels of IL-1beta cytokine in Korean patients with gastric cancer. Int J Cancer 114:465–471
- 102. Tatematsu M, Takahashi M, Hananouchi M, Shirai T, Hirose M (1976) Protective effect of mucin on experimental gastric cancer induced by *N*-methyl-*N*-nitro-*N*-nitrosoguanidine plus sodium chloride in rats. Gann 67:223–229
- 103. Takahashi M, Hasegawa R (1985) Enhancing effects of dietary salt on both initiation and promotion stages of rat gastric carcinogenesis. Princess Takamatsu Symp 16:169–182
- Liu C, Russell RM (2008) Nutrition and gastric cancer risk: an update. Nutr Rev 66:237–249
- 105. Glade MJ (1999) Food, nutrition, and the prevention of cancer: a global perspective. American Institute for Cancer Research/ World Cancer Research Fund, American Institute for Cancer Research, 1997. Nutrition 15:523–526
- 106. Ye WM, Yi YN, Luo RX, Zhou TS, Lin RT, Chen GD (1998) Diet and gastric cancer: a casecontrol study in Fujian Province, China. World J Gastroenterol 4:516–518
- 107. Ward MH, Lopez-Carrillo L (1999) Dietary factors and the risk of gastric cancer in Mexico City. Am J Epidemiol 149:925–932
- 108. Lee SA, Kang D, Shim KN, Choe JW, Hong WS, Choi H (2003) Effect of diet and Helicobacter pylori infection to the risk of early gastric cancer. J Epidemiol 13:162–168
- Kim HJ, Chang WK, Kim MK, Lee SS, Choi BY (2002) Dietary factors and gastric cancer in Korea: a case–control study. Int J Cancer 97:531–535
- 110. Shikata K, Kiyohara Y, Kubo M, Yonemoto K, Ninomiya T, Shirota T, Tanizaki Y, Doi Y, Tanaka K, Oishi Y, Matsumoto T, Iida M (2006) A prospective study of dietary salt intake and gastric cancer incidence in a defined Japanese population: the Hisayama study. Int J Cancer 119:196–201
- 111. Woodward M, Tunstall-Pedoe H, McColl K (2001) *Helicobacter pylori* infection reduces systemic availability of dietary vitamin C. Eur J Gastroenterol Hepatol 13:233–237
- 112. Banerjee S, Hawksby C, Miller S, Dahill S, Beattie AD, Mccoll KE (1994) Effect of *Helicobacter pylori* and its eradication on gastric juice ascorbic acid. Gut 35:317–322
- 113. Drake IM, Mapstone NP, Schorah CJ, White KL, Chalmers DM, Dixon MF, Axon AT (1998) Reactive oxygen species activity and lipid peroxidation in *Helicobacter pylori* associated gastritis: relation to gastric mucosal ascorbic acid concentrations and effect of *H. pylori* eradication. Gut 42:768–771
- 114. Correa P, Fontham ET, Bravo JC, Bravo LE, Ruiz B, Zarama G, Realpe JL, Malcom GT, Li D, Johnson WD, Mera R (2000) Chemoprevention of gastric dysplasia: randomized trial of

antioxidant supplements and anti-Helicobacter pylori therapy. J Natl Cancer Inst 92:1881–1888

- 115. Jenab M, Riboli E, Ferrari P, Sabate J, Slimani N, Norat T, Friesen M, Tjonneland A, Olsen A, Overvad K, Boutron-Ruault MC, Clavel-Chapelon F, Touvier M, Boeing H, Schulz M, Linseisen J, Nagel G, Trichopoulou A, Naska A, Oikonomou E, Krogh V, Panico S, Masala G, Sacerdote C, Tumino R, Peeters PH, Numans ME, Bueno-de-Mesquita HB, Buchner FL, Lund E, Pera G, Sanchez CN, Sanchez MJ, Arriola L, Barricarte A, Quiros J, Hallmans G, Stenling R, Berglund G, Bingham S, Khaw KT, Key T, Allen N, Carneiro F, Mahlke U, Del GG, Palli D, Kaaks R, Gonzalez CA (2006) Plasma and dietary vitamin C levels and risk of gastric cancer in the European Prospective Investigation into Cancer and Nutrition (EPIC-EURGAST). Carcinogenesis 27:2250–2257
- Franke A, Teyssen S, Singer MV (2005) Alcohol-related diseases of the esophagus and stomach. Dig Dis 23:204–213
- 117. Boeing H (1991) Epidemiological research in stomach cancer: progress over the last 10 years. J Cancer Res Clin Oncol 117:133–143
- Pollack ES, Nomura AM, Heilbrun LK, Stemmermann GN, Green SB (1984) Prospective study of alcohol consumption and cancer. N Engl J Med 310:617–621
- 119. Tredaniel J, Boffetta P, Buiatti E, Saracci R, Hirsch A (1997) Tobacco smoking and gastric cancer: review and meta-analysis. Int J Cancer 72:565–573
- 120. Ladeiras-Lopes R, Pereira AK, Nogueira A, Pinheiro-Torres T, Pinto I, Santos-Pereira R, Lunet N (2008) Smoking and gastric cancer: systematic review and meta-analysis of cohort studies. Cancer Causes Control 19:689–701
- 121. Gonzalez CA, Lopez-Carrillo L (2010) *Helicobacter pylori*, nutrition and smoking interactions: their impact in gastric carcinogenesis. Scand J Gastroenterol 45:6–14
- 122. Gonzalez CA, Jakszyn P, Pera G, Agudo A, Bingham S, Palli D, Ferrari P, Boeing H, Del GG, Plebani M, Carneiro F, Nesi G, Berrino F, Sacerdote C, Tumino R, Panico S, Berglund G, Siman H, Nyren O, Hallmans G, Martinez C, Dorronsoro M, Barricarte A, Navarro C, Quiros J, Allen N, Key TJ, Day NE, Linseisen J, Nagel G, Bergmann MM, Overvad K, Jensen MK, Tjonneland A, Olsen A, Bueno-de-Mesquita HB, Ocke M, Peeters PH, Numans ME, Clavel-Chapelon F, Boutron-Ruault MC, Trichopoulou A, Psaltopoulou T, Roukos D, Lund E, Hemon B, Kaaks R, Norat T, Riboli E (2006) Meat intake and risk of stomach and esophageal adenocarcinoma within the European Prospective Investigation Into Cancer and Nutrition (EPIC). J Natl Cancer Inst 98:345–354
- 123. Jenab M, Riboli E, Ferrari P, Friesen M, Sabate J, Norat T, Slimani N, Tjonneland A, Olsen A, Overvad K, Boutron-Ruault MC, Clavel-Chapelon F, Boeing H, Schulz M, Linseisen J, Nagel G, Trichopoulou A, Naska A, Oikonomou E, Berrino F, Panico S, Palli D, Sacerdote C, Tumino R, Peeters PH, Numans ME, Bueno-de-Mesquita HB, Buchner FL, Lund E, Pera G, Chirlaque MD, Sanchez MJ, Arriola L, Barricarte A, Quiros J, Johansson I, Johansson A, Berglund G, Bingham S, Khaw KT, Allen N, Key T, Carneiro F, Save V, Del GG, Plebani M, Kaaks R, Gonzalez CA (2006) Plasma and dietary carotenoid, retinol and tocopherol levels and the risk of gastric adenocarcinomas in the European prospective investigation into cancer and nutrition. Br J Cancer 95:406–415
- 124. Epplein M, Nomura AM, Hankin JH, Blaser MJ, Perez-Perez G, Stemmermann GN, Wilkens LR, Kolonel LN (2008) Association of *Helicobacter pylori* infection and diet on the risk of gastric cancer: a case–control study in Hawaii. Cancer Causes Control 19:869–877
- Fuccio L, Zagari RM, Eusebi LH, Laterza L, Cennamo V, Ceroni L, Grilli D, Bazzoli F (2009) Meta-analysis: can *Helicobacter*

pylori eradication treatment reduce the risk for gastric cancer? Ann Intern Med 151:121–128

- 126. Kato M, Asaka M, Nakamura T, Azuma T, Tomita E, Kamoshida T, Sato K, Inaba T, Shirasaka D, Okamoto S, Takahashi S, Terao S, Suwaki K, Isomoto H, Yamagata H, Nomura H, Yagi K, Sone Y, Urabe T, Akamatsu T, Ohara S, Takagi A, Miwa J, Inatsuchi S (2006) *Helicobacter pylori* eradication prevents the development of gastric cancer—results of a long-term retrospective study in Japan. Aliment Pharmacol Ther 24(suppl 4):203–206
- 127. Uemura N, Okamoto S, Yamamoto S, Matsumura N, Yamaguchi S, Yamakido M, Taniyama K, Sasaki N, Schlemper RJ (2001) *Helicobacter pylori* infection and the development of gastric cancer. N Engl J Med 345:784–789
- 128. Take S, Mizuno M, Ishiki K, Nagahara Y, Yoshida T, Yokota K, Oguma K, Okada H, Shiratori Y (2005) The effect of eradicating *Helicobacter pylori* on the development of gastric cancer in patients with peptic ulcer disease. Am J Gastroenterol 100:1037– 1042
- 129. Wong BC, Lam SK, Wong WM, Chen JS, Zheng TT, Feng RE, Lai KC, Hu WH, Yuen ST, Leung SY, Fong DY, Ho J, Ching CK, Chen JS (2004) *Helicobacter pylori* eradication to prevent gastric cancer in a high-risk region of China: a randomized controlled trial. JAMA 291:187–194
- Domellof L (1998) Reversal of gastric atrophy after *Helicobacter* pylori eradication: is it possible or not? Am J Gastroenterol 93:1407–1408
- 131. Kamada T, Hata J, Sugiu K, Kusunoki H, Ito M, Tanaka S, Inoue K, Kawamura Y, Chayama K, Haruma K (2005) Clinical features of gastric cancer discovered after successful eradication of *Helicobacter pylori*: results from a 9-year prospective follow-up study in Japan. Aliment Pharmacol Ther 21:1121–1126
- 132. Take S, Mizuno M, Ishiki K, Nagahara Y, Yoshida T, Yokota K, Oguma K (2007) Baseline gastric mucosal atrophy is a risk factor associated with the development of gastric cancer after *Helicobacter pylori* eradication therapy in patients with peptic ulcer diseases. J Gastroenterol 42(17):21–27
- 133. Ohata H, Kitauchi S, Yoshimura N, Mugitani K, Iwane M, Nakamura H, Yoshikawa A, Yanaoka K, Arii K, Tamai H, Shimizu Y, Takeshita T, Mohara O, Ichinose M (2004) Progression of chronic atrophic gastritis associated with *Heli-cobacter pylori* infection increases risk of gastric cancer. Int J Cancer 109:138–143
- Weck MN, Brenner H (2008) Association of *Helicobacter pylori* infection with chronic atrophic gastritis: meta-analyses according to type of disease definition. Int J Cancer 123:874–881
- 135. Cassaro M, Rugge M, Gutierrez O, Leandro G, Graham DY, Genta RM (2000) Topographic patterns of intestinal metaplasia and gastric cancer. Am J Gastroenterol 95:1431–1438
- 136. de Vries AC, van Grieken NC, Looman CW, Casparie MK, De VE, Meijer GA, Kuipers EJ (2008) Gastric cancer risk in patients with premalignant gastric lesions: a nationwide cohort study in the Netherlands. Gastroenterology 134:945–952
- 137. Uemura N, Mukai T, Okamoto S, Yamaguchi S, Mashiba H, Taniyama K, Sasaki N, Haruma K, Sumii K, Kajiyama G (1997) Effect of *Helicobacter pylori* eradication on subsequent development of cancer after endoscopic resection of early gastric cancer. Cancer Epidemiol Biomark Prev 6:639–642
- 138. Sung JJ, Lin SR, Ching JY, Zhou LY, To KF, Wang RT, Leung WK, Ng EK, Lau JY, Lee YT, Yeung CK, Chao W, Chung SC (2000) Atrophy and intestinal metaplasia 1 year after cure of *H. pylori* infection: a prospective, randomized study. Gastroenterology 119:7–14
- 139. Schenk BE, Kuipers EJ, Nelis GF, Bloemena E, Thijs JC, Snel P, Luckers AE, Klinkenberg-Knol EC, Festen HP, Viergever PP, Lindeman J, Meuwissen SG (2000) Effect of *Helicobacter pylori*

eradication on chronic gastritis during omeprazole therapy. Gut 46:615-621

- 140. Salih BA, Abasiyanik MF, Saribasak H, Huten O, Sander E (2005) A follow-up study on the effect of *Helicobacter pylori* eradication on the severity of gastric histology. Dig Dis Sci 50:1517–1522
- 141. Rokkas T, Pistiolas D, Sechopoulos P, Robotis I, Margantinis G (2007) The long-term impact of *Helicobacter pylori* eradication on gastric histology: a systematic review and meta-analysis. Helicobacter 12(2):32–38
- 142. Wang J, Xu L, Shi R, Huang X, Li SW, Huang Z, Zhang G (2011) Gastric atrophy and intestinal metaplasia before and after *Helicobacter pylori* eradication: a meta-analysis. Digestion 83:253–260
- 143. Ohkusa T, Fujiki K, Takashimizu I, Kumagai J, Tanizawa T, Eishi Y, Yokoyama T, Watanabe M (2001) Improvement in atrophic gastritis and intestinal metaplasia in patients in whom *Helicobacter pylori* was eradicated. Ann Intern Med 134:380–386
- 144. Sugiyama T, Sakaki N, Kozawa H, Sato R, Fujioka T, Satoh K, Sugano K, Sekine H, Takagi A, Ajioka Y, Takizawa T (2002) Sensitivity of biopsy site in evaluating regression of gastric atrophy after *Helicobacter pylori* eradication treatment. Aliment Pharmacol Ther 2:187–190
- 145. Leung WK, Lin SR, Ching JY, To KF, Ng EK, Chan FK, Lau JY, Sung JJ (2004) Factors predicting progression of gastric intestinal metaplasia: results of a randomised trial on Helicobacter pylori eradication. Gut 53:1244–1249
- 146. Zhou L, Sung JJ, Lin S, Jin Z, Ding S, Huang X, Xia Z, Guo H, Liu J, Chao W (2003) A five-year follow-up study on the pathological changes of gastric mucosa after *H. pylori* eradication. Chin Med J 116:11–14
- 147. Fukase K, Kato M, Kikuchi S, Inoue K, Uemura N, Okamoto S, Terao S, Amagai K, Hayashi S, Asaka M (2008) Effect of eradication of *Helicobacter pylori* on incidence of metachronous gastric carcinoma after endoscopic resection of early gastric cancer: an open-label, randomised controlled trial. Lancet 372:392–397
- 148. Wu CY, Kuo KN, Wu MS, Chen YJ, Wang CB, Lin JT (2009) Early *Helicobacter pylori* eradication decreases risk of gastric cancer in patients with peptic ulcer disease. Gastroenterology 137:1641–1648
- 149. Malfertheiner P, Megraud F, O'Morain C, Bazzoli F, El-Omar E, Graham D, Hunt R, Rokkas T, Vakil N, Kuipers EJ (2007) Current concepts in the management of *Helicobacter pylori* infection: the Maastricht III Consensus Report. Gut 56:772–781
- 150. Asaka M, Kato M, Takahashi S, Fukuda Y, Sugiyama T, Ota H, Uemura N, Murakami K, Satoh K, Sugano K (2010) Guidelines for the management of *Helicobacter pylori* infection in Japan: 2009 revised edition. Helicobacter 15:1–20
- 151. Fuccio L, Eusebi LH, Zagari RM, Bazzoli F (2009) *Helicobacter pylori* eradication treatment reduces but does not abolish the risk of gastric cancer. Am J Gastroenterol 104:3100–3102
- 152. de Vries AC, Kuipers EJ, Rauws EA (2009) Helicobacter pylori eradication and gastric cancer: when is the horse out of the barn? Am J Gastroenterol 104:1342–1345
- Cannizzaro R, De PP (2009) *Helicobacter pylori* eradication, endoscopic surveillance, and gastric cancer. Am J Gastroenterol 104:3100–3101

- 154. Miki K (2006) Gastric cancer screening using the serum pepsinogen test method. Gastric Cancer 9:245–253
- 155. Yanaoka K, Oka M, Ohata H, Yoshimura N, Deguchi H, Mukoubayashi C, Enomoto S, Inoue I, Iguchi M, Maekita T, Ueda K, Utsunomiya H, Tamai H, Fujishiro M, Iwane M, Takeshita T, Mohara O, Ichinose M (2009) Eradication of *Helicobacter pylori* prevents cancer development in subjects with mild gastric atrophy identified by serum pepsinogen levels. Int J Cancer 125:2697–2703
- 156. Murata H, Kawano S, Tsuji S, Tsuji M, Sawaoka H, Kimura Y, Shiozaki H, Hori M (1999) Cyclooxygenase-2 overexpression enhances lymphatic invasion and metastasis in human gastric carcinoma. Am J Gastroenterol 94:451–455
- 157. Joo YE, Oh WT, Rew JS, Park CS, Choi SK, Kim SJ (2002) Cyclooxygenase-2 expression is associated with welldifferentiated and intestinal-type pathways in gastric carcinogenesis. Digestion 66:222–229
- 158. Yamac D, Ayyildiz T, Coskun U, Akyurek N, Dursun A, Seckin S, Koybasioglu F (2008) Cyclooxygenase-2 expression and its association with angiogenesis, *Helicobacter pylori*, and clinicopathologic characteristics of gastric carcinoma. Pathol Res Pract 204:527–536
- 159. van Rees BP, Saukkonen K, Ristimaki A, Polkowski W, Tytgat GN, Drillenburg P, Offerhaus GJ (2002) Cyclooxygenase-2 expression during carcinogenesis in the human stomach. J Pathol 196:171–179
- 160. Thun MJ, Namboodiri MM, Calle EE, Flanders WD, Heath CW Jr (1993) Aspirin use and risk of fatal cancer. Cancer Res 53:1322–1327
- 161. Sorensen HT, Friis S, Norgard B, Mellemkjaer L, Blot WJ, McLaughlin JK, Ekbom A, Baron JA (2003) Risk of cancer in a large cohort of nonaspirin NSAID users: a population-based study. Br J Cancer 88:1687–1692
- 162. Nardone G, Rocco A, Malfertheiner P (2004) Review article: *Helicobacter pylori* and molecular events in precancerous gastric lesions. Aliment Pharmacol Ther 20:261–270
- 163. Yang HB, Cheng HC, Sheu BS, Hung KH, Liou MF, Wu JJ (2007) Chronic celecoxib users more often show regression of gastric intestinal metaplasia after *Helicobacter pylori* eradication. Aliment Pharmacol Ther 25:455–461
- 164. Rupnow MF, Chang AH, Shachter RD, Owens DK, Parsonnet J (2009) Cost-effectiveness of a potential prophylactic *Helicobacter pylori* vaccine in the United States. J Infect Dis 200:1311–1317
- 165. Del Guidice G, Malfertheiner P, Rappuoli R (2009) Development of vaccines against *Helicobacter pylori*. Expert Rev Vaccin 8:1037–1049
- 166. Malfertheiner P, Schultze V, Rosenkranz B, Kaufmann SH, Ulrichs T, Novicki D, Norelli F, Contorni M, Peppoloni S, Berti D, Tornese D, Ganju J, Palla E, Rappuoli R, Scharschmidt BF, Del Guidice G (2008) Safety and immunogenicity of an intramuscular *Helicobacter pylori* vaccine in noninfected volunteers: a phase I study. Gastroenterology 135:787–795
- 167. Aebischer T, Walduck A, Schroeder J, Wehrens A, Chijioke O, Schreiber S, Meyer TF (2008) A vaccine against *Helicobacter pylori*: towards understanding the mechanism of protection. Int J Med Microbiol 298:161–168