



Developmental and age-related changes to the elastic lamina of Bruch's membrane in mice

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Abstract

Background Fibrillin-1, tropoelastin, fibulin-5, and latent transforming growth factor beta-binding protein-2 and protein-4 (LTBP-2 and LTBP-4) are essential proteins for the elastic lamina (EL). In this study, we analyzed each of these molecules in the EL of Bruch's membrane (BM) through development and aging.

Methods C57BL/6 mice (embryonic (E) days E12.5, E15.5, and E18.5; postnatal (P) days P1, P4, and P7 and P3, P6, and P75 weeks of age) were used. To investigate localization, immunohistochemical staining (IH) was performed. Transmission electron microscopy (TEM) was used to evaluate the formation of microfibrils and tropoelastin. mRNA expression was determined by quantitative real-time PCR (qRT-PCR).

Results All five proteins were expressed in the EL of BM by IH except in embryonic mice. TEM results showed that tropoelastin co-stained with microfibrils. Between 3 and 6 weeks of age, microfibrils became longer and thicker. It was difficult to evaluate the EL of BM in senile mice at 75 weeks of age because of abundant deposits which correspond to drusen. mRNA levels of each protein increased dramatically from E15.5 to P1 days and plateaued by P3 weeks as shown by qRT-PCR.

Conclusions In conclusion, these five proteins are possibly involved in elastic fiber assembly in BM. We define the date of full assembly of the EL of BM as 3 weeks of age in mice.

Keywords Elastic lamina of Bruch's membrane · fibrillin1 · Fibulin-5 · Tropoelastin · LTBP-2 · LTBP-4

Introduction

Bruch's membrane (BM) has a thin and complex structure with five stratified layers that contain the basement membrane of the retinal pigment epithelium (RPE), the inner collagenous layer, the central elastic layer, the outer collagenous layer, and the basement membrane of the choriocapillaris endothelium (CCE) [1, 2]. Bruch's membrane plays a critical role in the eye. Transport of oxygen, glucose, growth factors, and fluids derived from choroidal vasculature to the retina/RPE and removal of waste

materials from the RPE through BM supports the function and health of RPE and photoreceptors [3–5]. Bruch's membrane also works as a tight barrier to prevent abnormal angiogenesis [6, 7] as the rupture of BM may induce choroidal neovascularization (CNV) [8–15]. Therefore, maintaining the five-layer structure of BM is essential for retinal health and normal visual function. Particularly, damage of elastic lamina and/or mutations in the genes encoding elastic fibers in BM is associated with severe vision loss by CNV formation according to previous studies [6, 15, 16]. Chong et al. reported that discontinuities within the elastic lamina of BM are significantly longer in patients with age-related macular degeneration (AMD) than in age-matched controls [6]. Sivaprasad et al. reported that serum levels of elastin-derived peptides in patients with AMD are significantly higher than that in non-AMD patients [16]. Jones et al. reported that mouse RPE overexpression of human high temperature requirement A 1 (HTRA1) induced polypoidal choroidal vasculopathy (PCV) which is a subtype of AMD [15]. The degeneration of the elastic lamina

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of BM induced by human HTRA1 as a multifunctional serine protease that works as elastase activity [17, 18] was the major factor for the occurrence of PCV.

Discontinuities of the elastic lamina of BM may be predictors for the occurrence of CNVs [6]. Among the five layers in BM, proper homeostasis and turnover of the elastic lamina of BM seems to be essential to prevent CNV. Therefore, it is important to improve understanding of the elastic lamina of BM in terms of histology and molecular biology characteristics.

Elastic fibers give connective tissue resilience, deformability, and recoil abilities without energy input [19, 20]. Elastic fibers play the principal role in the structural integrity and function of various organs [20]. Knowledge in the field of elastic fiber assembly-related molecules has spread in recent years owing to the identification of gene mapping of elastic fibers using in vivo studies (transgenic mouse models) and in vitro studies (biochemical examinations) [19–48]. Elastic fibers are composed of two major composite biomaterials including an outer lattice of microfibrillar mantle mainly consisting of polymers of fibrillin-1 and an inner amorphous core of cross-linking tropoelastin (elastin monomers) [20–22]. Several molecules associated with the coacervation and cross-linking between microfibrils and tropoelastin have been implicated in the process of elastic fiber assembly. Recent genetic studies have revealed that fibulin-5, latent transforming growth factor- β -binding protein-2 (LTBP-2), and LTBP-4 are involved in elastic fiber assembly in addition to microfibrils and tropoelastin [34, 38]. Schematic view of elastic fiber assembly involves recruitment and organized deposition of molecules including fibrillin-1, tropoelastin, fibulin-5, LTBP-2, and LTBP-4, as shown in Fig. 1. The complete molecular mechanism in elastic fiber assembly remains unclear. Structural components of these five molecules (fibrillin-1, fibulin-5, tropoelastin, LTBP-2, and LTBP-4) differ between organs. For example, ciliary zonules consist of rich microfibrils and LTBP-2 only [34]. There have been no reports about whether these five molecules are expressed in the elastic lamina of BM. Researching the elastic

lamina of BM may lead to elucidating the mechanism of CNV development.

In this study, we analyzed the relationships between these five molecules and the elastic lamina of BM through prenatal and postnatal development and in aging by both qualitative and quantitative assessments using wild-type mice.

Materials and methods

Animals

In the following descriptions, the day or week of embryonic or postnatal development were used (e.g., E15 to designate the 15th day after fertilization, P6 weeks to indicate the 6th week after birth). In this study, male and female C57BL/6 mice (each E12.5, E15.5, and E18.5 days; P1, P4, and P7 days; and P3, P6, and P75 weeks of age) were purchased from CLEA Japan (Tokyo, Japan). All animal experiments were carried out in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research and the Guideline for Animal Experimentation at Kansai Medical University. Major ocular structures appear and are able to be differentiated between E10 and E15 days [49]. Therefore, fetal mice were studied starting at E12.5 days in this study. Animals were housed with a normal laboratory diet in a temperature and humidity-controlled environment at 22 °C and 50% filtered air. All mice were kept in pathogen-free plastic cages with 12-h light-dark cycles and had continuous free access to water and food (CLEA Japan, Tokyo, Japan). All plastic cages, water and bedding feed were purified before use.

Sixteen eyes of eight mice per time point were used for each examination: immunohistochemistry, transmission electron microscopy (TEM), and quantitative real-time polymerase chain reaction (qRT-PCR).

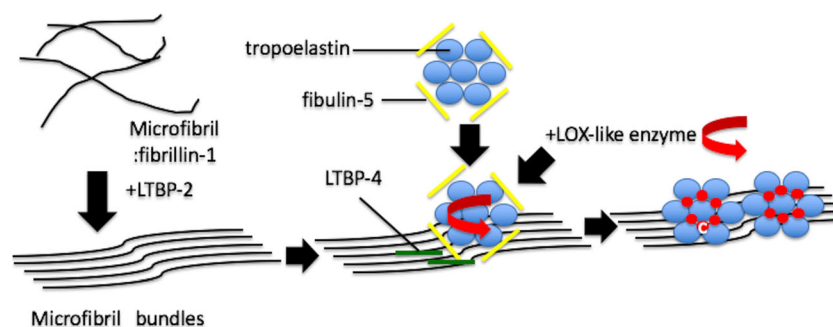


Fig. 1 Roles of elastogenic molecules in elastic fiber assembly. LTBP-2 leads each microfibril to the formation of a microfibril bundle. The coacervation of tropoelastin is carried out by fibulin-5. A cluster of tropoelastin and fibulin-5 is deposited onto microfibril bundles by

LTBP-4, which is localized on microfibrils. Then, LOX-like enzyme forms a cross-linked structure made up of the tropoelastin/fibulin-5 complex on microfibrils, which marks the completion of the elastic fiber assembly

We defined the embryonic period as three stages: early (E12.5 days), middle (E15.5 days), and late (E18.5 days).

Immunohistochemistry

All mice were killed by cervical dislocation after deep intraperitoneal injection of a mixture containing 7 μ l/mg of ketamine hydrochloride (Daiichisankyo, Tokyo, Japan) and 0.7 μ l/mg of xylazine (Bayer, Leverkusen, Germany). The eyes were enucleated immediately and frozen using dry ice, followed by embedding in optimal cutting temperature compound (Tissue-Tek OCT; Sakura Finetek, Torrance, CA, USA). The frozen blocks were preserved overnight at -80°C . Vertical and 8 μ m thin-sliced sections were cut with a cryostat (Leica CM3050S®; Leica Biosystems, Nußloch, Germany). To minimize tissue damage during dissection, Kawamoto's films (SECTION-LAB, Hiroshima, Japan) were used [50]. Each section with Kawamoto's film was air dried at room temperature for 20 min. Immunohistochemistry was performed by the indirect immunofluorescence technique. Briefly, cryostat sections were fixed with cold methanol at -20°C for 5 min, washed with phosphate-buffered saline (PBS) for 5 min, and blocked with 4% blocking agent prepared from lactoprotein (Sumitomo Dainippon Pharma, Osaka, Japan) for 20 min. The sections were then incubated with the primary antibodies anti-fibrillin-1 (1:100, original), mouse polyclonal anti-tropoelastin (1:100, Elastin Products Company), rabbit polyclonal anti-fibulin-5 (1:100, BSYN2473), anti-LTBP-2 (1:100, original) and goat polyclonal anti-LTBP-4 (1:100, R&D Systems) for 1 h at room temperature. After incubation, these sections were rinsed with PBS, and then incubated with a combination of Alexa Fluor®-488, Alexa Fluor®-546 (1:200, Thermo Fisher Scientific, Waltham, MA, USA), and DyLight®-650 (1:200, Leinco technologies, Fenton, MO, USA) secondary antibodies for 1 h at room temperature. The combinations of primary antibodies and secondary antibodies are shown in Table 1. After rinsing with PBS, the sections were mounted with ProLong®Gold mounting media

(Thermo Fisher Scientific, Waltham, MA, USA) for preventing fluorescence photobleaching. The sections were observed and images were taken with a fluorescence microscope (BZ-X700®; KEYENCE, Osaka, Japan). Negative control slides were stained by primary non-specific IgG (anti-goat, 144-09531; anti-rabbit, 148-09551) and/or IgY (anti-chicken, PSC-3707-1) antibodies one at a time to confirm positive staining of immunohistochemistry.

Triple staining with the immunofluorescent technique was performed; fibrillin-1 (Alexa Fluor®-488)-LTBP-4 (Alexa Fluor®-488)-LTBP-2 (DyLight®-650), tropoelastin (Alexa Fluor®-488)-LTBP-4 (Alexa Fluor®-488)-LTBP-2 (DyLight®-650), fibulin-5 (Alexa Fluor®-488)-LTBP-4 (Alexa Fluor®-488)-LTBP-2 (DyLight®-650).

TEM

After deep intraperitoneal injection as described above, all mice were killed and pre-fixed with perfusion fixation from the heart by 2% glutaraldehyde (Electron Microscopy Sciences, Hatfield, PA, USA), 1 mM CaCl_2 in 0.1 M sodium cacodylate (Electron Microscopy Sciences, Hatfield, PA, USA), and 1/15 M phosphate buffer without NaCl (pH 7.4). Immediately afterwards, each eye was enucleated. We dissected the eyes carefully under a microscope at the corneal limbus and removed the lens. Posterior eyecups were immersed in fixation liquid of 2% glutaraldehyde and 1 mM CaCl_2 in 0.1 M sodium cacodylate buffer (pH 7.4) and preserved at 4°C for 2 h. After pre-fixation, samples were washed twice by 0.2 M sucrose in 0.1 M sodium cacodylate buffer (pH 7.4) for 10 min. Posterior eyecups were postfixed for 2 h in 1% osmium tetroxide (Nissin EM, Tokyo, Japan) and 0.15 M sodium cacodylate buffer at 4°C , dehydrated through a series of subsequent high-grade ethanol concentrations, and embedded in epoxy resin (TAAB Laboratories Equipment Ltd., Aldermaston, Berks, England). Semi-thin sections (1 μ m thick) were cut with an ultramicrotome (REICHERT-NISSEI ULTRACUT S®; Leica AG, Wien, Austria). These semi-thin sections were stained with toluidine blue for identifying BM under a light microscope. Then, ultrathin sections were cut at 50 nm by an ultramicrotome with diamond knives (Diatome AG, Biel, Switzerland) and mounted on 100 mesh copper grids (Veco B.V., Karel van Gelreweg, Eerbeek, The Netherlands). These sections were stained with uranyl acetate (Cerac, Inc., Milwaukee, WI, USA) for 10 min and with lead citrate (TAAB Laboratories Equipment Ltd., Aldermaston, Berks, England) for 5 min. Then these sections were stained with tannic acid (Nacalai Tesque, Kyoto, Japan) and *p*-nitrophenol (Wako, Osaka, Japan) for 1 h to facilitate the visualization of tropoelastin as small black dot-like deposits. Electron micrographs of BM were viewed and taken with 80 kV under TEM (JEM-1400A®; JEOL, Tokyo, Japan).

Table 1 All primary and secondary antibodies used in this study for immunohistochemistry

Primary antibody	Secondary antibody
Fibrillin-1, tropoelastin, fibulin-5	Anti-rabbit Alexa Fluor® 488
LTBP-4	Anti-goat Alexa Fluor® 546
LTBP-2	Anti-chicken DyLight® 650

Anti-fibrillin-1, tropoelastin, and fibulin-5 primary antibodies were stained with Alexa Fluor® 488 as the secondary antibody. Anti-LTBP-4 primary antibody was stained with Alexa Fluor® 546 as the secondary antibody. Anti-LTBP-2 primary antibody was stained with Alexa Fluor® 650 as the secondary antibody

qRT-PCR

All mice were killed by cervical dislocation after deep intra-peritoneal injection as described above. Immediately, after each whole eye was enucleated, they were stored in RNAlater® (Thermo Fisher Scientific, Waltham, MA, USA) and homogenized by Biomasher (Nippi Inc., Tokyo, Japan). Total RNA was isolated using RNeasy Plus Micro® and Mini® Kit (Qiagen, Venlo, The Netherlands) according to the manufacturer's protocol. Total RNAs were reverse-transcribed to generate single-stranded complementary DNA (cDNA) using the SuperScript® VILO™ cDNA Synthesis Kit (Thermo Fisher Scientific, Waltham, MA, USA). The reverse-transcriptase cDNAs were then subjected to qRT-PCR (Thermal Cycler Dice® Real Time System II; TAKARA Bio, Otsu, Shiga, Japan). Each qRT-PCR was performed in a 20- μ l volume using the SYBR® Premix Ex TaqII™ PCR Kit (TAKARA Bio, Otsu, Shiga, Japan) protocol as follows: pre-denaturing at 95 °C for 30 s, 5-s denaturing at 95 °C, and 30-s annealing and extending each at 60 °C. Denaturing, annealing, and extending were repeated for 40 cycles.

Targeting sequences used in this study were amplified with the following primers.

Fibrillin-1:

- Forward, 5'-AATATCTCGGAGCCATTGTC-3'
- Reverse, 5'-CAGGTCTACGGCAGTTGTCA-3'

Tropoelastin:

- Forward, 5'-CTATGGAGGACCCCTTGGAG-3'
- Reverse, 5'-CACAGGATTTCCCAAAGCAG-3'

Fibulin-5:

- Forward, 5'-CCGATACCCTGGTGCCTATT-3'
- Reverse, 5'-GCACTGATAGGCCCTGTTTG-3'

LTBP-2:

- Forward, 5'-AAACCCCTCAGCGACCCGCGGCTGC-3'
- Reverse, 5'-TGCTTCTGTGAGGACCGGGTGCTCT-3'

LTBP-4:

- Forward, 5'-GCACAAATACTAAAGGCTCCTTCCAC-3'
- Reverse, 5'-GCACTCGTCAATGTCAAGGCAGGAG-3'

Cyclophilin:

- Forward, 5'-CAGACGCCACTGTCGCTTT-3'
- Reverse, 5'-TGTCTTTGCAACTTTGTCTGCAA-3'

Quantitative mRNA expression levels in each age group were normalized to the amount of cyclophilin mRNA as an internal control. Quantitative mRNA expression levels in each age group were calculated by relative quantification (standard curve method). We used the mRNA of the whole eye at P12 weeks as a standard sample. Quantitative mRNA expression levels in eye tissues was calculated with logarithmic transformation and analyzed by JMP software (SAS institute, Inc., Cary, NC, USA). All data were presented as mean \pm SD according to normal distribution by Bartlett's test. Statistical analyses were performed by Tukey-Kramer test. *P* values < 0.05 were considered statistically significant.

Results

Immunohistochemistry

Bruch's membrane was observed as the gap between RPE pigment and the choroid layer under bright-field light microscope (Fig. 2a). At embryonic stages, BM could not be detected clearly under the light microscope because BM was too faint and fragile (Fig. 2b). The BM gap was not obvious in the developing mouse at the embryonic early stage (Fig. 2b, left). However, this gap became visible from the embryonic middle stage and onwards (Fig. 2b, center and right).

At embryonic early and middle stages, staining of the five proteins (fibrillin-1, tropoelastin, fibulin-5, LTBP-2, LTBP-4) was not detected in BM (data not shown). However, fibrillin-1 showed a strong staining signal in the sclera at the embryonic early stage (data not shown). Tropoelastin, fibulin-5, LTBP-2, and LTBP-4 showed a strong staining signal in the sclera at the embryonic middle stage (data not shown). At the embryonic late stage, positive staining of fibrillin-1 and LTBP-4 in BM was isolated and weak (data not shown), but tropoelastin, fibulin-5, and LTBP-2 were positive in BM in some slides. Immunohistochemical staining confirmed the consistent expression of all five matrix proteins in BM except in embryonic stages, which indicated that production and/or deposition of the matrix proteins were dramatically enhanced at P1 days and onwards (Fig. 3). A summary of immunohistochemical staining in BM in each age group is shown in Fig. 4.

We further analyzed the localization pattern of each protein in BM. Figure 5 represents a summary of the characteristic staining pattern of each protein. Fibrillin-1 was detected as linear continuous long lines at postnatal stages (Fig. 5a). Tropoelastin was confirmed as a dot-like pattern at postnatal stages (Fig. 5b). LTBP-2 and LTBP-4 were co-localized with fibrillin-1 (Fig. 5c). Moreover, there was co-localization between tropoelastin and LTBP-2 by merged images (Fig. 5d). There was no clear difference in staining intensity among these five proteins with regard to exposing time, fluorescence intensity, or location (peripheral or center regions).

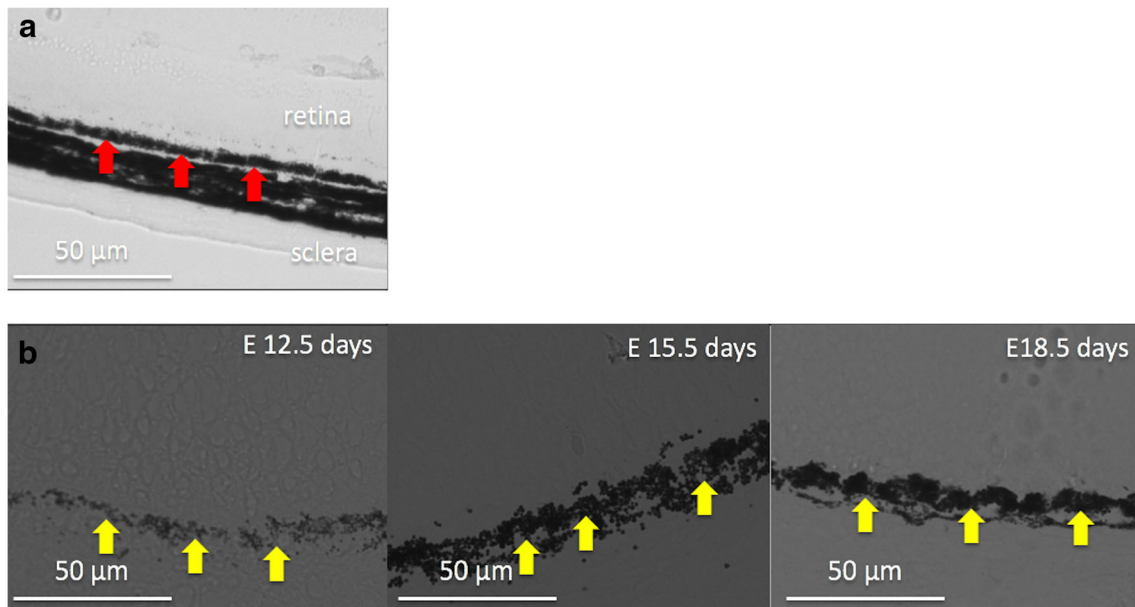


Fig. 2 Light microscopic appearance of Bruch's membrane during development. **a** Bruch's membrane of C57/BL6 mice is observed as the white layer between the retinal pigment epithelium and the choroid under a bright-field microscope (red arrow). **b** Three bright-field images show

embryonic early (E12.5 days), middle (E15.5 days), and late (E18.5 days) stages. A clear white layer confirms that Bruch's membrane is fully assembled by the embryonic late stage. All images were taken at $\times 400$ magnification

TEM

At the embryonic early stage (E12.5 days), BM was mostly observed as an empty space between the RPE and CCE. Single layer as the basement membrane of RPE was lined along with RPE. Short and small clusters of fibrous microfibrils were detected. Tropoelastin occasionally overlapped with clusters of microfibrils (Fig. 6a). At the embryonic middle stage (E15.5 days), another single layer inside of the basement membrane of CCE appeared parallel to CCE. Two layers of collagenous fibrils were observed in the space between the basement membrane of the RPE and elastic fibers. The basement membrane of the RPE was lined along with the CCE. Clusters of microfibrils connected by tropoelastin gradually began to accumulate in the middle layer of BM (Fig. 6b). At the embryonic late stage (E18.5 days), short and small clusters of immature elastic fibers began to increase and connect to each other. Two collagenous fibrils began to form a three-dimensional meshwork-like helix structure (Fig. 6c). At P1 day, extended microfibrils attached by tropoelastin grew and connected to form as cross-linked structure in BM (Fig. 6d). From P4 to P7 days, microfibrils with tropoelastin became thick central bundles, indicating that it became a layer of relatively uniform structure (Fig. 6e, f). From P3 to P6 weeks, a mostly consecutive single structure of microfibrils attached by tropoelastin became longer and thicker (Fig. 6g, h). Two collagenous layers became thick and formed a tight meshwork. Bruch's membrane was able to be identified as five discrete layers during P3 to P6 weeks. Their morphology indicated

that developmental formation of BM was completed from P3 to P6 weeks. The elastic lamina of BM could not be clearly identified at P75 weeks because abundant dense deposits in BM localized to the RPE and disturbed the ability to distinguish the structure of BM (Fig. 6i).

The elastic lamina of BM presented discontinuity from embryo to senile stages in this study. Furthermore, the thickness of BM varied in location (Fig. 7).

qRT-PCR

Fibrillin-1 mRNA

The mean mRNA level of fibrillin-1 (Fig. 8a) at E12.5 days was low (926.3 ± 249.1). Significant elevation of mRNA levels were observed at E15.5 days ($13,954.4 \pm 4371.5$) compared with those of other time points ($P < 0.01$). mRNA expression levels were maintained until P7 days even though mRNA expression level was reduced between E15.5 and P7 days. mRNA expression levels were reduced back to early embryonic levels by P3 weeks. There was no significant difference between E12.5 days and P6 weeks or between E12.5 days and P75 weeks.

Tropoelastin mRNA

The mean mRNA level of tropoelastin (Fig. 8b) at E12.5 days was relatively low (540.5 ± 258.6). Significant elevations of tropoelastin mRNA expression levels were observed at E15.5

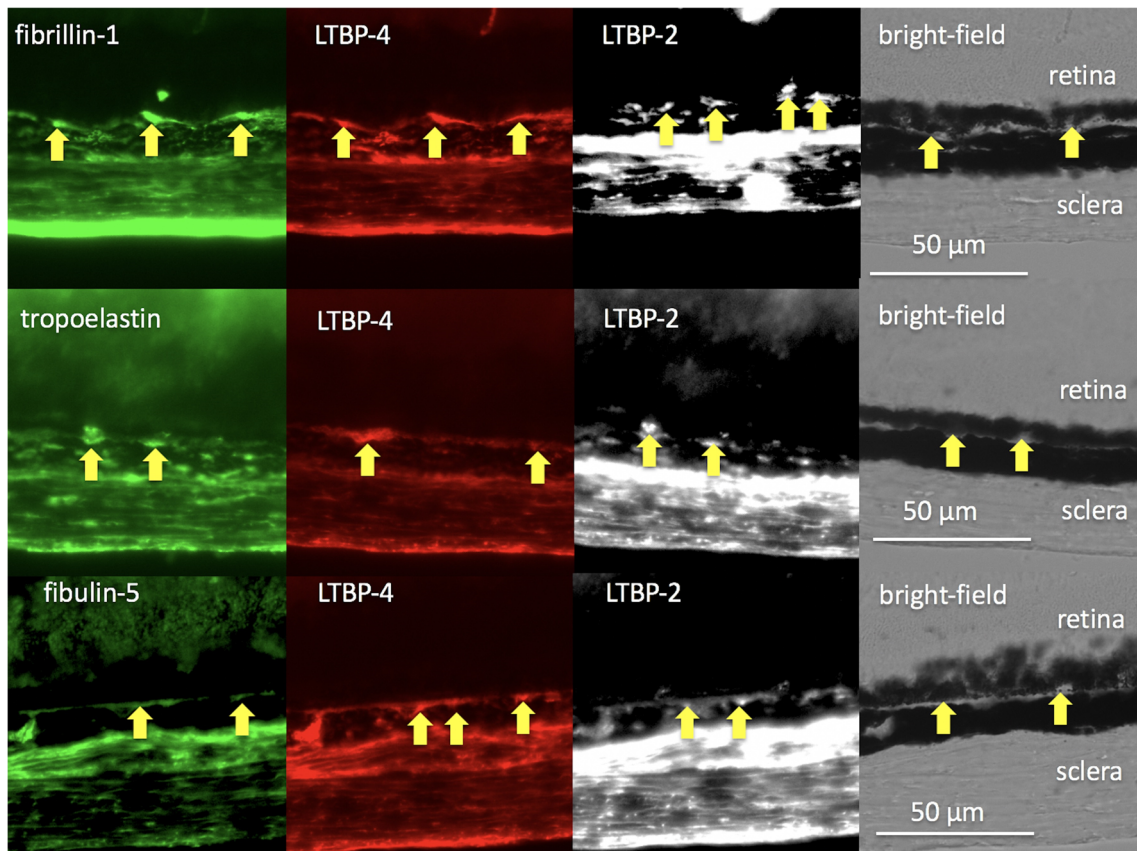


Fig. 3 Immunohistopathological findings in Bruch’s membrane. Figure 3 is an image from a 6-week-old mouse. From postnatal stages and onwards, all proteins in Bruch’s membrane clearly showed positive

expression by staining. In each image, Bruch’s membrane is shown as a yellow arrow. All images were taken at $\times 400$ magnification

days ($23,843.1 \pm 8030.1$) compared with E12.5 days ($P < 0.01$). The highest mRNA levels were observed at P1 day ($35,266.3 \pm 8700.0$). High mRNA levels were maintained between E15.5 and P7 days, and there were no significant differences in expression level between these time points. At P3 weeks, tropoelastin mRNA levels were reduced. No significant difference was detected between P75 weeks and E12.5 days.

Fibulin-5 mRNA

The mean mRNA level of fibulin-5 (Fig. 8c) were lowest at E12.5 days (150.8 ± 68.6). At E15.5 days, mRNA levels were elevated (2635.7 ± 689.1) and at their highest compared with those of other time points ($P < 0.01$). Fibulin-5 mRNA levels were significantly reduced at E18.5 days compared with E15.5 days ($P < 0.01$) and were maintained at high values

Fig. 4 Summary of immunohistochemistry results in Bruch’s membrane from each age group. The five proteins were found to be expressed in Bruch’s membrane by immunohistochemistry in postnatal mice; “+” positive staining, “-” negative staining, “±” between negative and positive

	fibrillin-1	tropoelastin	fibulin-5	LTBP-2	LTBP-4
E12.5 days	-	-	-	-	-
E15.5 days	-	-	-	-	-
E18.5 days	±	-	-	-	±
P1 day	+	+	+	+	+
P4 days	+	+	+	+	+
P7 days	+	+	+	+	+
P3 weeks	+	+	+	+	+
P6 weeks	+	+	+	+	+
P75 weeks	+	+	+	+	+

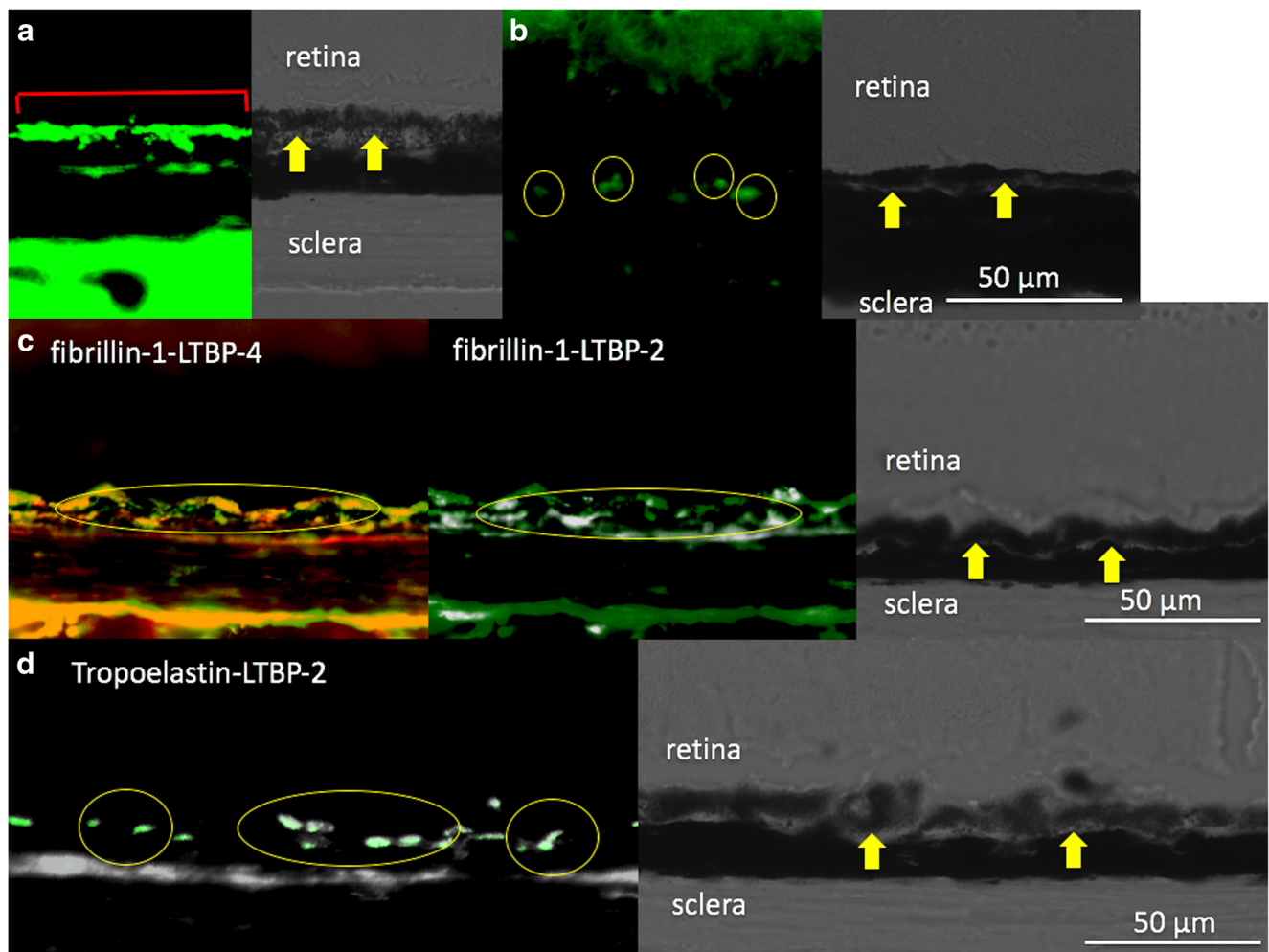


Fig. 5 Patterns of protein expression in Bruch’s membrane. **a** Fibrillin-1, **b** tropoelastin, **c** fibrillin-1, and LTBP-2 (left image) and fibrillin-1 and LTBP-2 (middle image); **d** tropoelastin and LTBP-2. Fibrillin-1 proteins show relatively long, linear staining (red bracket). Tropoelastin proteins show dot-like staining (yellow, round). Co-localization of fibrillin-1 and

LTBP-2, fibrillin-1 and LTBP-2, and tropoelastin and LTBP-2 is observed. LTBP-2 and LTBP-4 staining overlaps with fibrillin-1. Tropoelastin and LTBP-2 punctate staining overlapped. All images were taken at $\times 400$ magnification

between E18.5 days and P7 days (E18.5 days, 1406.7 ± 230.3 ; P1 day, 1751.8 ± 408.9 ; P4 days, 1341.2 ± 312.3 ; P7 days, 1653.9 ± 417.7) (no significant difference among time points). By P3 weeks, fibulin-5 mRNA levels showed significant reduction compared to that of P7 days ($P < 0.01$).

LTBP-2 mRNA

The mean mRNA level of LTBP-2 (Fig. 8d) was at its lowest at E12.5 days (40.0 ± 83.0) compared with those of other time points ($P < 0.01$). Significant elevations of mRNA levels were observed at E15.5 days (1859.9 ± 1206.9 , $P < 0.01$). The highest mRNA expression level was observed at E18.5 days (3639.6 ± 1442.7). High mRNA levels were maintained from E15.5 to P1 days. mRNA levels at P4 days were reduced significantly compared with P1 day ($P < 0.01$). By P4 days, mRNA expression levels plateaued.

LTBP-4 mRNA

The mean mRNA (Fig. 8e) level at E12.5 days was lowest (136.1 ± 41.7) compared with those of other time points ($P < 0.01$). Elevations of mRNA levels were observed at E15.5 days (1433.4 ± 707.8). By E15.5 days, mRNA levels plateaued.

Discussion

The number of AMD patients is increasing for many reasons such as change of lifestyle (e.g., a western diet), an increase in the senior population, and genetic factors [51–55]. Clinically, AMD is divided into three types: typical AMD, PCV, and retinal angiomatous proliferation (RAP) [51–55]. Moreover, the development of CNV in AMD is divided into two types [52]:

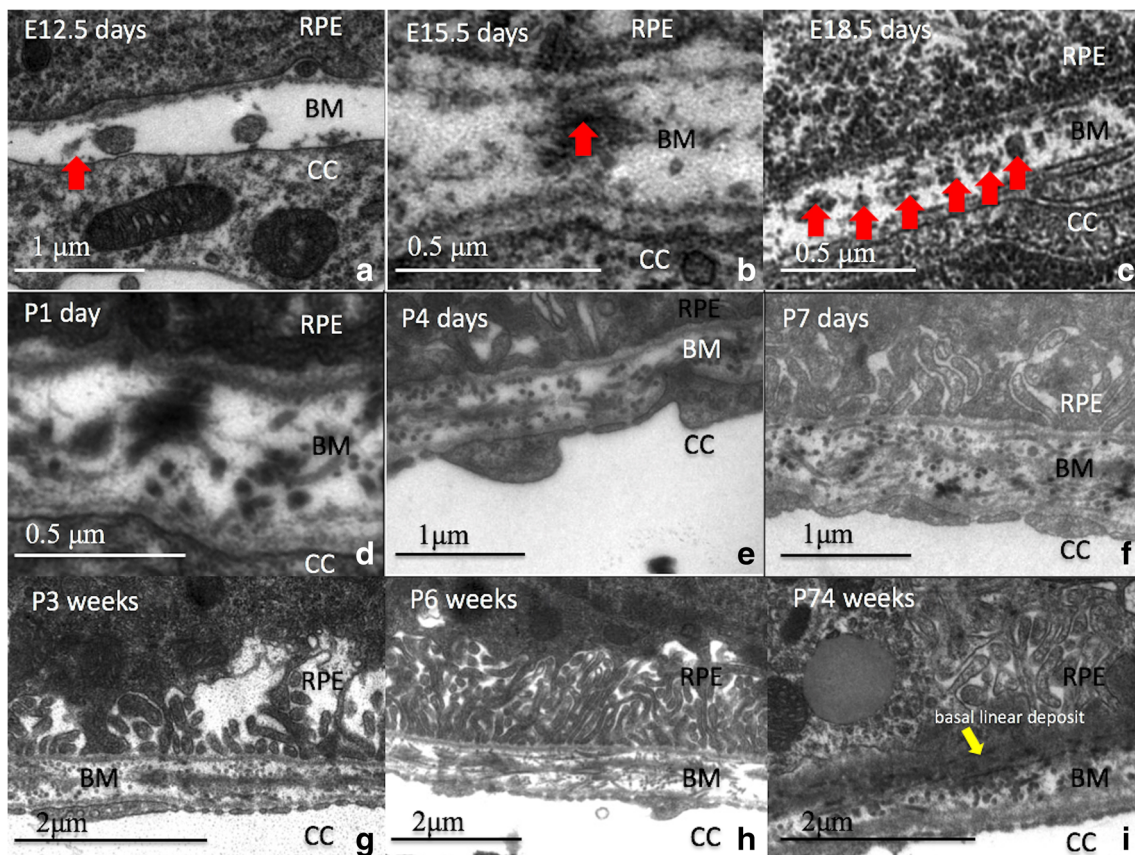


Fig. 6 Ultrastructural pre- and postnatal changes of Bruch's membrane in wild-type mice. Transmission electron micrographs of the elastic lamina of Bruch's membrane at embryonic 12.5 days (a), 15.5 days (b), 18.5 days (c), P1 day (d), P4 days (e), P7 days (f), P3 weeks (g), P6 weeks (h), and P74 weeks (i). Tropoelastin is stained as black dots by tannic acid and *p*-nitrophenol. A microfibril/tropoelastin (red arrow) complex appeared at embryonic early stage (a), increased in expression in the embryonic middle stage (b), and formed connections with one other by the embryonic

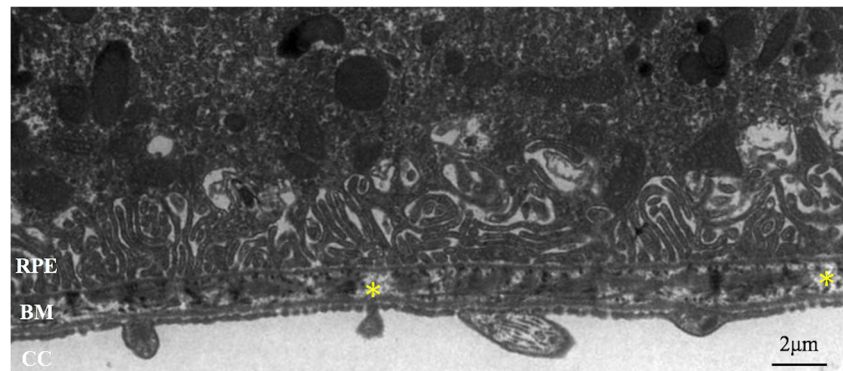
late stage (c, d). Extended microfibrils grew longer and thicker between P4 days and P3 weeks (e–g). Mature structure of the elastic lamina of Bruch's membrane revealed relatively continuous and thick fibers by P3 to 6 weeks (f, g). At P74 weeks (i), basal linear deposits prevented a clear elastic layer from being detected. Magnification—**a** $\times 28,700$, **b** $\times 90,400$, **c** $\times 59,200$, **d** $\times 74,800$, **e** $\times 28,700$, **f** $\times 28,700$, **g** $\times 21,400$, **h** $\times 14,200$, and **i** $\times 221,400$

type 1 CNV arises from choroid to reach the sub-RPE space, and type 2 CNV arises from choroid to reach the sub-retinal space. Typical AMD has type 1 and/or type 2 CNV separately [51, 52]. PCV has only type 1 CNV [53]. RAP has types 1 and 2 CNVs in addition to chorioretinal anastomosis [55]. Among the three subtypes of AMD, the number of PCV patients in Asia is larger than that in Western countries [51, 53]. PCV can cause severe visual loss due to massive hemorrhage in the eye by ruptured polypoid lesions [54]. There have been many reports about animal models of CNVs [8–15, 56]. Laser and surgically induced CNVs by penetrating BM have been widely used as experimental CNV models [8–11]. Recently, transgenic animal models have also been used for the investigation of CNVs [12–14]. Most of CNVs in these animal models belong to type 2 [8–14]. Conversely, only a few animal models of type 1 CNV have been produced [15, 56]. Moreover, there has been only 1 report about an animal model of PCV by Jones et al. This study found that the degeneration of the elastic lamina of BM is a fundamental factor for the occurrence of PCV [15]. There have

been many reports characterizing the elastic lamina of BM; however, most previous reports assessed BM by only histological evaluation. There have been no reports about the relationship between essential molecules (fibrillin-1, tropoelastin, fibulin-5, LTBP-2 and LTBP-4) for elastic fiber assembly in the elastic lamina of BM. In this study, we found that fibrillin-1, tropoelastin, fibulin-5, LTBP-2 and LTBP-4 proteins were expressed constitutively in BM at all postnatal stages by immunohistochemistry.

Fibrillins account for the majority of the microfibril complex [22–28]. Fibrillin monomers are long flexible molecules [24] encoded by three kinds of genes (fibrillin-1, fibrillin-2 and fibrillin-3) [27–29]. Each fibrillin-1 and fibrillin-2 plays a role in elastic fiber assembly. Fibrillin-1 and fibrillin-2 double knockout mice showed hypoplasia of elastic fibers, and mice deficient of either fibrillin gene resulted in elastogenesis [28, 31]. The molecular organization of fibrillin-1 and fibrillin-2 is unclear. They have been found in the cornea [32], crystalline lens [33] and ciliary zonule [34, 35].

Fig. 7 Maturation of the elastic lamina of Bruch's membrane at postnatal week 6. Transmission electron microscopy findings in Bruch's membrane of C57/BL mice at P6 weeks showed a relatively continuous elastic layer, with small gaps between long microfibrils (yellow asterisks). RPE, retinal pigment epithelium; BM, Bruch's membrane; CC, choriocapillaris. Magnification, $\times 7140$



Mutations in the fibrillin-1 gene cause Marfan syndrome, which is associated with ectopia lentis [35]. Whether fibrillin-3 is involved in elastic fiber assembly or not remains unknown [29]. Within the three isoforms of the fibrillin family, fibrillin-1 is mainly responsible for forming microfibrils [25]. Thus, we studied fibrillin-1. Fibrillin microfibrils were first described as small-diameter fibrils located close to the basement membranes and also at the periphery of elastic fibers under the electron microscope [23]. In this study, fibrillin-1-rich microfibrils were seen as long bundles in BM at all postnatal stages under TEM. This result corresponded with the appearance in immunohistochemistry, which indicated that fibrillin-1 protein presents as a long linear pattern in BM at all postnatal stages.

Tropoelastin is the major component of elastic fiber, and fibrillin-1 forms microfibrils that act as a scaffold for tropoelastin. Tropoelastin presents as the non-cross-linked soluble precursor of elastin at less than 20 °C. Tropoelastin represents two stages in elastic fiber assembly; coacervation and cross-linking. The aqueous solutions aggregate at 37 °C by interactions between hydrophobic domains. This process termed coacervation is reversible and thermodynamically controlled. The coacervate becomes steady and insoluble by cross-linking via lysyl oxidase (LOX)-like enzyme in an irreversible manner. LOX-like enzyme aligns tropoelastin molecules prior to cross-linking [19, 36, 37]. Thus, the microaggregation of tropoelastin deposited onto microfibrils and the cross-linked structure of tropoelastin by LOX-like enzyme are the major processes that form mature elastic fibers. In this study, the microaggregation of tropoelastin was observed as aggregation of black dots on the microfibril bundle using tannic acid and *p*-nitrophenol under TEM. This result corresponded with the appearance of tropoelastin in immunohistochemistry as well, in which tropoelastin protein stained as a dot-like pattern.

LTBPs are also integral components in elastic fiber assembly. There are four isoforms within the LTBP family (LTBP-1, LTBP-2, LTBP-3, and LTBP-4). The LTBP family regulates transforming growth factor- β (TGF- β) bioavailability by

binding and sequestering TGF- β [42]. Among the four isoforms of the LTBP family, LTBP-1 and LTBP-3 are not required for the elastic fiber assembly reported by studies on mice and humans that are deficient in these genes [43]. LTBP-2, however, does not bind to TGF- β [44]. Mutations in LTBP2 occur in ocular diseases including primary or secondary glaucoma, megalocornea, microspherophakia and ectopia lentis [45–47]. LTBP-2 facilitates the formation of thick microfibrils by binding fibrillin microfibrils into bundles in ciliary zonules [34].

In the absence of LTBP-4, the elastin/fibulin-5 complex is unable to linearly deposit on microfibrils, and misplaced aggregation of elastin/fibulin-5 forms globular structures [48]. In previous reports, interactions between fibrillin-1 and LTBP-2 or LTBP-4 indicated that LTBP-2 and LTBP-4 play a role in formation and maturation of microfibrils [34, 38]. In this study, we confirmed interaction between fibrillin-1 and LTBP-2 or LTBP-4 by localization in immunohistochemistry in BM. Moreover, we found a novel co-localization of LTBP-2 and tropoelastin. We could not confirm co-localization of fibulin-5 and tropoelastin or LTBP-4 by immunohistochemistry. Unfortunately, we did not have evidence for how LTBP-2 and tropoelastin interact with each other in BM.

LTBP-4 localized on microfibril bundles is essential for linear assembly of elastin through interaction with fibulin-5 [48]. LTBP-4 and fibulin-5 play a role in proper accumulation of tropoelastin to the microfibril scaffold [48]. Nakamura et al. reported that the elastic fiber assembly process requires fibulin-5 using fibulin-5-deficient mice. These knockout mice resulted in loose skin, tortuous vessels, and emphysematous lungs [38, 39]. Yanagisawa reported that fibulin-5 can strongly interact with tropoelastin [40]. Hirai reported that fibulin-5 not only binds with tropoelastin but also promotes coacervation of tropoelastin in cell culture [41]. In human donor eyes, the fibulin-5 protein was localized in BM using immunohistochemistry [57]. In this study, fibulin-5 protein was expressed in BM in postnatal mice similar to human BM.

We have shown that the elastic fibers in BM were divided into microfibrils and tropoelastin separately using TEM. Our

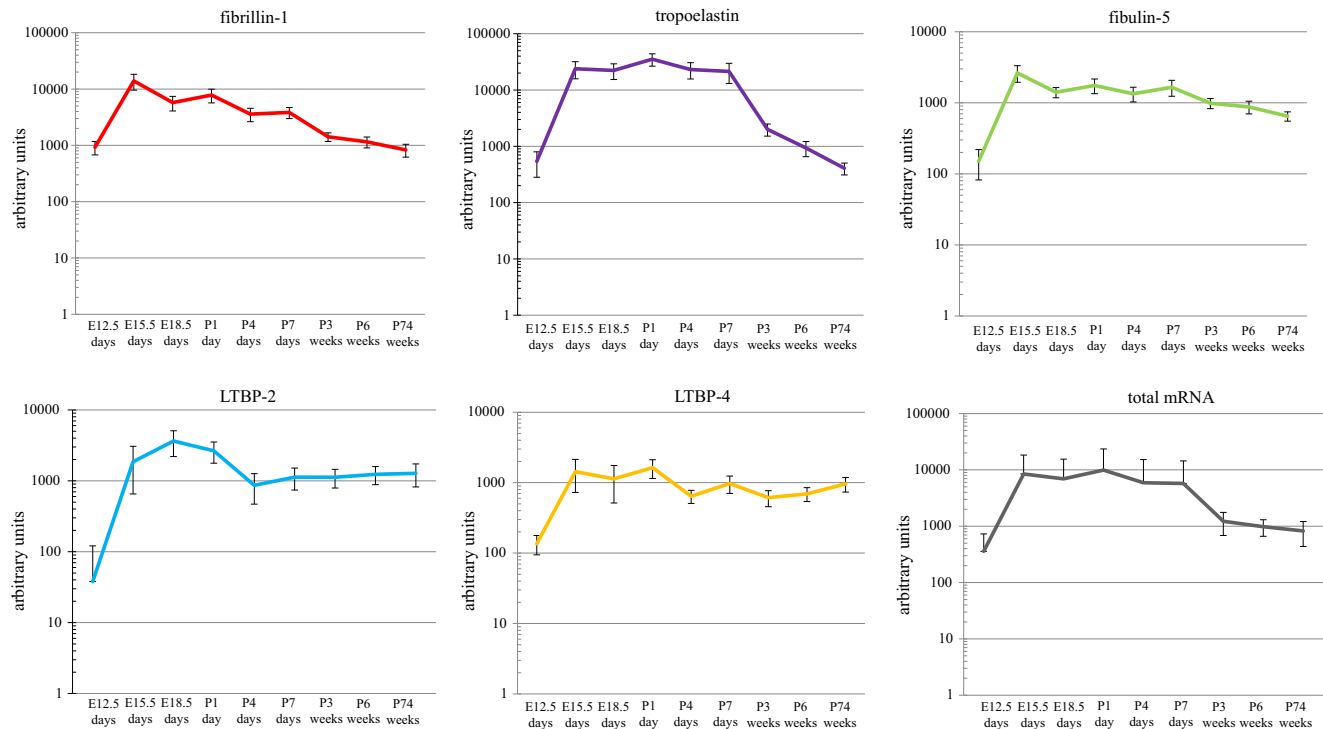


Fig. 8 mRNA abundance of each elastic fiber assembly-related protein in whole eye tissue. mRNA levels of fibrillin-1 (a), tropoelastin (b), fibulin-5 (c), LTBP-2 (d), LTBP-4 (e), and total mRNA (f) were evaluated at pre- and postnatal stages with quantitative real-time PCR. Total mRNA expression levels were upregulated before and after birth. From postnatal

weeks 3 to 6, mRNA expression plateaued at low levels, which indicates that production of each protein is finished in adult mice. The bars represent mean \pm SD. Statistics were analyzed by Tukey-Kramer ($n = 16$ mice/group, $P < 0.05$)

TEM showed that there were slightly negligible clusters of microfibrils in the hollow space of BM at the embryonic early stage, which was consistent with the negative staining in immunohistochemistry and low level of each mRNA in qRT-PCR at this same stage. TEM indicated that microfibril expression levels began to increase and tropoelastin began to deposit onto the cluster of microfibrils in the embryonic middle stage, which was consistent with the results of qRT-PCR. The expression levels of these five molecules (fibrillin-1, tropoelastin, fibulin-5, LTBP-2, and LTBP-4) were elevated in the embryonic middle stage.

Connections of each microfibril by tropoelastin between embryonic to postnatal stages allowed the elastic lamina of BM to gradually become mature fibers as shown under TEM. This TEM result coincides with immunohistochemistry, which shows positive staining of the five proteins by postnatal stages in immunohistochemistry.

Hirabayashi et al. defined the completion of the development of the elastic lamina of BM in normal mice as P6 weeks by the continuity of elastic fibers using TEM [2]. However, the elastic lamina of BM does not have full continuity from embryo to senile stages. In this study, we showed the maturation of the elastic lamina of BM using qualitative (TEM and immunohistochemistry) and quantitative (qRT-PCR) examinations. Relatively thick and continuous microfibrils with

tropoelastin were observed from P3 to P6 weeks under TEM. Immunohistochemistry of all five proteins showed positive staining starting at postnatal stages. The mRNA levels of fibrillin-1, tropoelastin, and fibulin-5 remained high until P7 days and decreased by P3 weeks as shown by qRT-PCR. The mRNA levels of LTBP-2 and LTBP-4 remained high until P4 days and decreased by P7 days as shown by qRT-PCR. We also found that mRNA levels of the five proteins involved in elastic fiber assembly decreased starting at either P7 days or P3 weeks. According to the results of TEM, immunohistochemistry, and qRT-PCR, we define the maturation of elastic lamina of BM at P3 to P6 weeks.

With age, BM loses normal function and leads to degenerative changes [6, 58–65]. Discontinuities of elastic fiber of BM were observed in senescence accelerated mice (SAM), aging mice, and elderly people according to numerous studies [6, 16–23]. Increased length of discontinuities of the elastic lamina of BM may be the predilection for the occurrence of CNVs [6]. Furthermore, lipid deposits and other debris which are likely to play a role in the pathogenesis of AMD accumulated in BM in aging eyes [66, 67]. In this study, we showed abundant dense deposits in BM of senile mice (P75 weeks). These deposits aligned sub-RPE and BM in a similar manner to the basal linear deposits of human AMD. It was difficult to detect the elastic lamina of BM clearly in senile mice because

of the abundant dense deposits located between the basement membrane of the RPE and the elastic lamina. Stone et al. hypothesized that fibulin-5 may be involved in basal deposit formation from the result that human patients with AMD have missense mutations in fibulin-5 [60]. The mechanism of how fibulin-5 mutations are related with AMD is not understood completely and must be addressed.

There are some limitations of this study. Further studies are needed to elucidate the elastic fiber assembly, age-related changes, and the formation of deposits in BM in the context of these five molecules. In this study, mRNA expression levels by qRT-PCR were measured from the whole eye. The eye itself contains various elastic fiber tissues such as the sclera, ciliary zonules, and BM. To strictly examine mRNA levels of the elastic lamina of BM, we would need to perform qRT-PCR using laser capture microdissection microscopy. Instead, we qualitatively assessed our proteins by using immunohistochemistry. To compare the quantitative protein expression levels, using Western blot or ELISA is recommended. In this study, we performed qRT-PCR on total ocular tissue because to collect proteins from such a small tissue as Bruch's membrane would yield a level of protein that is too small for analysis. In the future, we will have to study how each molecule interacts by using an immunoelectron microscope to fully understand how these proteins are assembled. Further studies including fibrillin-1, tropoelastin, fibulin-5, LTBP-2, and LTBP-4 transgenic mice may lead to understanding the elastic fiber formation in BM, which furthermore may lead to understanding the pathology of types 1 and 2 CNVs.

In summary, we studied immunohistochemical localization, ultrastructural changes, and mRNA levels of five proteins (fibrillin-1, tropoelastin, fibulin-5, LTBP-2, and LTBP-4) in the EL of BM. We conclude that the EL became mature from P3 to P6 weeks as formerly stated. The structure of EL remained discontinuous throughout the age groups. Elastic fibers are still poorly understood, particularly in the BM, because they are complex and difficult to illustrate. Degradation of elastic fibers in BM may cause age-related disease such as AMD. Future therapeutic interventions which regenerate elastic fibers may be used instead of, or alongside, intravitreal anti-vascular endothelial growth factor (VEGF), and/or placental growth factor (PIGF) for the treatment of CNV.

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Toyama. Data analysis was performed by Hidetsugu Mori. Hidetsugu Mori wrote the paper with support of Haruhiko Yamada, Kanji Takahashi, Tomoya Akama, and Tomoyuki Nakamura.

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Compliance with ethical standards

All animal experiments were carried out in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research and the Guideline for Animal Experimentation at Kansai Medical University.

Conflict of interest The authors declare that they have no conflict of interest.

Abbreviations LTBP, Latent transforming growth factor beta-binding protein; EL, Elastic lamina; BM, Bruch's membrane; IH, Immunohistochemical staining; TEM, Transmission electron microscopy; qRT-PCR, Quantitative real-time PCR; RPE, Retinal pigment epithelium; CCE, Choriocapillaris endothelium; CNV, Choroidal neovascularization; AMD, Age-related macular degeneration; HTRA1, Human high-temperature requirement A 1; PCV, Polypoidal choroidal vasculopathy; PBS, Phosphate-buffered saline; LOX, Lysyl oxidase; TGF- β , Transforming growth factor- β ; VEGF, Vascular endothelial growth factor; PIGF, Placental growth factor

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