GYNECOLOGIC ENDOCRINOLOGY AND REPRODUCTIVE MEDICINE



Bone turnover in lactating and nonlactating women

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Abstract

Purpose During lactation, bone turnover increases, reflecting the mobilization of Calcium from maternal skeletal stores and resulting in bone loss. However, mechanisms are not yet fully understood, and previous studies have been comparatively small. We aim to assess bone metabolism during lactation by comparing bone-metabolism-related-parameters between large cohorts of lactating and nonlactating women.

Methods In a retrospective cohort study, we recruited 779 postpartum women and 742 healthy, nonpregnant, nonlactating controls. Postpartum women were examined 3 and 6 months after delivery and retrospectively assigned to either the exclusively breastfeeding (exc-bf) group if they had exclusively breastfed or the nonexclusively breastfeeding (nonexc-bf) group if they had not exclusively breastfed up to the respective visit. Serum levels of PTH, Estradiol, total Calcium, Phosphate, and bone turnover markers (\u00e3CTX, P1NP, Osteocalcin) were compared between the groups.

Results Bone turnover markers were significantly increased in exc-bf and nonexc-bf women compared with the controls (all ps < .001). BCTX was approximately twice as high in exc-bf women than in the controls. PTH levels were marginally higher in exc-bf (p < .001) and nonexc-bf women (p = .003) compared with the controls (6 months). Estradiol was suppressed in exc-bf women compared with the controls (p < .001, 3 months).

Conclusion Exc-bf and even nonexc-bf states are characterized by an increase in bone formation and resorption markers. The PTH data distribution of exc-bf, nonexc-bf, and control groups in the underpart of the reference range suggest that lactational bone loss is relatively independent of PTH.

Keywords Lactation \cdot Bone turnover \cdot Breastfeeding \cdot Calciotropic hormones

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What does this study add to the clinical work

This retrospective study examines bone turnover during lactation and shows that both bone resorption and bone formation are increased in lactating women compared to healthy, nonlactating, nonpregnant controls. To our knowledge, this is the largest published data that describes bone turnover in the comparison of breastfeeding and nonbreastfeeding women.

Introduction

Lactation represents a time of major challenges for the maternal organism. In order to supply the newborn, the lactating mother secretes 300–400 mg of Calcium (Ca) into her breast milk every day [1]. A main source is the maternal

bone. Ca is resorbed transiently from the mother's skeleton, resulting in bone loss that is restored after weaning [2]. Studies found that bone mass decreased by up to 10% in women who breastfed exclusively for 6 months (m) [3, 4]. A main physiological mediator that is attributed to lactational bone loss is Estradiol (E_2) [5–7]. Lactation causes hypoestrogenemia by suppressing the hypothalamic-pituitary-ovarian axis, thereby inhibiting reproductive function. Different studies have reported significantly lower E₂ levels in breastfeeding (bf) compared with nonbf postpartum (pp) women and nonlactating, nonpregnant controls [8–14]. In this context, lactation-associated bone loss has been shown to be rather independent of calciotropic hormones, such as Parathyroid Hormone (PTH), which is either normal [15–17] or suppressed [11, 18, 19] in bf women. At the molecular level, bone turnover increases, as reflected by changes in bone turnover markers (BTM). Several studies have consistently found increased levels of bone resorption markers in bf women compared with nonbf women or controls [13, 15, 16, 19–23]. Whereas only a few authors have reported normal levels of bone formation markers [21], the majority of studies have shown significantly increased bone formation in bf women [13, 15, 16, 19, 20, 22, 23]. In a prospective cohort study examining bone turnover during lactation, bone resorption and bone formation, as measured with C-Telopeptide of Type 1 Collagen (BCTX), Bone-specific Alkaline Phosphatase, Osteocalcin (OC), and Procollagen Type 1 N Terminal Propeptide (P1NP), were significantly increased in women who lactated for 6-8 weeks (wk) compared with bottle-feeding women and controls [13]. However, this study used only a small sample, a trend that applies to most other studies as well, many of which are also outdated. Furthermore, the underlying hormonal mechanisms regulating Ca homeostasis during lactation are not yet fully understood. Findings on concentrations in bf and nonbf women are somewhat contradictory, as reviewed by Kovacs et al. [2]. Therefore, we aim to examine bone metabolism in a large cohort of exclusively bf (exc-bf) and nonexclusively bf (nonexc-bf) pp women as well as healthy, nonpregnant, nonlactating controls using the most current BTM BCTX, P1NP, and OC, as well as serum PTH, E₂, total Ca, and Phosphate (P) to provide a representative update on bone metabolism during lactation.

Methods

Study design and population

The data originated from the large "LIFE Child" study, which was initiated in 2011 in Leipzig, Germany. It is part of the "Leipzig Research Centre for Civilization Diseases (LIFE)" and is aimed at monitoring children's development,

growth, and healthiness while considering various lifestyle factors. To do so, children are recruited as early as the 24th week of gestation up to the age of 20 with annual follow-ups to allow for both cross-sectional and longitudinal approaches. The children's parents are recruited as well. Details have been described previously [24, 25]. For the current retrospective cohort study, data were collected from 04/2011 to 03/2020. A total of 812 women between the ages of 21 and 44 years old who had delivered recently were examined throughout the pp period (lactation cohort). Assessments of serum bone-metabolism-related-parameters (bmrp) (BCTX, P1NP, OC, total Ca, P, PTH, E₂) were performed at 3, 6, and 12 m pp. As controls, we included 947 nonpregnant, nonlactating control subjects between the ages of 20 and 45 years (control cohort) from the LIFE Adult study, which is also part of the "Leipzig Research Centre for Civilization Diseases (LIFE)." Due to the small number of bf women at the 12 m examination (n=7), data from this visit were not analyzed. Women who participated only at 12 m were excluded (Fig. 1). Participants with chronic diseases (e.g., diagnoses of chronic renal and hepatic diseases, hyperthyroidism, diabetes mellitus, osteoporosis, neoplastic diseases, or other endocrinological and metabolic disorders) or medication affecting bone metabolism (e.g., glucocorticoids, heparin, warfarin, immunosuppressants, except stable doses of thyroid hormone) were also excluded as shown in Fig. 1. After visual inspection, we decided to eliminate any participants who did not fit the distribution of Ca and PTH values (n=7). The cut-off values were defined as mean ± 3.5 standard deviations for the Ca and PTH values. The final sample included 779 women in the lactation cohort, with 116 participating with several pregnancies. A total of 742 women in the control cohort were included after the exclusion criteria were applied (Fig. 1).

Anthropometric measurements

Anthropometric measurements were performed by professional health care staff using standardized procedures. For the lactation cohort, we used the prepregnancy weight from their maternity log. Body mass index (BMI) was determined by weight (kilograms) divided by height (meters) squared. Participants also completed questionnaires about their medical history, lifestyle habits, and demographic characteristics.

Breastfeeding status

At each visit (3 m and 6 m), pp women were interviewed about their infant feeding practice. They were assigned either to the exc-bf group if they had exclusively breastfed at the respective time of examination or to the nonexc-bf group if they had not exclusively breastfed anymore or had stopped breastfeeding completely at that time. For both visits, exc-bf Fig. 1 Selection of participants from the lactation and control cohorts. Flowchart showing the exclusion of individuals



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women and nonexc-bf women were compared with each other and with the nonpregnant, nonlactating control group. The numbers of women who participated from the lactation cohort were 618 at 3 m and 590 at 6 m (Table 1). Controls were set as the reference for the comparisons exc-bf versus controls and nonexc-bf versus controls. Nonexc-bf women were used as the reference group for the comparison exc-bf versus nonexc-bf.

Laboratory measurements

Venous blood samples were collected in the morning (7:30) a.m. to 10:00 a.m.) under fasting conditions. Serum parameters were measured at the Institute of Laboratory Medicine, Clinical Chemistry, and Molecular Diagnostics of the University of Leipzig, using the following methods for routine patient diagnostics: Serum samples P1NP, BCTX, OC, and intact PTH were measured with electrochemiluminescence assays (ECLIA; Elecsys, Cobas 801, Roche Diagnostics, Mannheim, Germany). The mean interassay coefficient of variation for these four parameters was between 1.95 and

13.10% from 4 QC cycles across the representative 4 months (82–164 runs). Details can be found in Geserick et al. [26]. Total serum Ca and P were assessed with a Roche Cobas C701 analyzer (Roche Diagnostics GmbH, Mannheim, Germany). E2 was quantified by liquid chromatography-tandem mass spectrometry (LC-MS/MS). A detailed description of the method can be found in Gaudl et al. [27]. Reference levels were chosen according to the manufacturer and refer to nonpregnant, nonlactating women. We used reference levels for E2 from Bae et al. [28].

Statistical analysis

Descriptive statistics were given as the mean and standard deviation for continuous variables and counts for categorical variables, stratified by bf status. Bmrp levels were compared between groups at each visit using simple linear regression (univariate analyses). Subsequently, we evaluated the association between bmrp levels and bf status using linear mixed effect model. These associations were corrected for age and BMI. The subject was added as random intercept to account for

Table 1	Anthropometric and	breastfeeding	characteristics	of the	lactation	and control	cohorts
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Lactation cohort					Control cohort			
N (data)		Age in years $(mean \pm SD)$	BMI in kg/m ² (mean \pm SD)	N (data)	Age in years $(mean \pm SD)$	BMI in kg/m ² (mean \pm SD)		
779 (N total)			31.9 (4.27)	22.7 (3.44)	742	39.2 (6.52) ***	24.4 (4.56) ***	
Visit	Subgroup							
	Exc-bf	Nonexc-bf						
3 m	570	48						
6 m	360	230						

Descriptives are presented as counts for categorical variables and as means (SD) for continuous variables (age, BMI). The lactation cohort was set as the statistical reference. The means of the controls were significantly different from those of the lactation cohort: ***p <.001. exc-bf exclusive breastfeeding, nonexc-bf nonexclusive breastfeeding, BMI body mass index, SD standard deviation, m months

multiple measurements per woman. All effect sizes are given as group differences (β). The number of cases we included varied slightly between models because of missing values in the parameters (Suppl. Table 1) [29]. Means and standard deviations for the respective parameter, cohort and visit are presented in Supplemental Table 2 [29]. Statistical analyses were conducted with R, version 4.0.5. Ggplot2 was used to create figures. The significance level was set to α =0.05.

Results

Study population

Anthropometric and breastfeeding characteristics are presented in Table 1. The final sample consisted of 1,521 participants divided into a lactation cohort (n = 779) and controls (n = 742). The majority of the lactation cohort were still breastfeeding exclusively at 3 m pp (92.2%, n = 570). The proportion of exclusively breastfeeding participants decreased to 61.0% (n = 360) at 6 m. Mean age and BMI were significantly lower in the lactation cohort compared with controls (p < .001, Table 1). At 6 m pp, controls had a mean BMI (\pm SD) of 24.4 (4.56) kg/m². BMI values were above 25 kg/m² in 34.0% and above 30 kg/m² in 10.3% of the control group. For the lactation cohort, exc-bf women had a mean BMI (\pm SD) of 22.4 (3.15) kg/m² whereas nonexc-bf women had a mean BMI (\pm SD) of 22.8 kg/m² (3.47) at 6 m. BMI values were above 25 kg/m² in 13.7% (exc-bf) or 19.4% (nonexc-bf) and above 30 kg/m² in 3.7% (exc-bf) or 5.7%(nonexc-bf) of the lactation cohort. Regarding BMI, there was no significant difference between exc-bf and nonexcbf women at 6 m ($\beta = 0.3$, p = .652), while both groups had significantly lower BMI levels compared with the controls $(\beta = -1.7 \text{ [exc-bf vs. controls]}, \beta = -1.5 \text{ [nonexc-bf vs. con$ trols], both *ps* < 0.001).

Bone-metabolism-related parameters and breastfeeding status

Results from univariate and multivariate analyses are presented in Table 2. In general, BTM levels were significantly higher in exc-bf and nonexc-bf women compared with the controls. Results were similar at 3 m and 6 m. β CTX reached distinctly higher levels in exc-bf women (β =419, p < .001 at 6 m) than in the controls in the univariate analyses. Levels in exc-bf mothers were about twice as high as those of the controls (Fig. 2a). There were also distinctly higher levels in the nonexc-bf mothers (β =350, p < .001 at 6 m) than in the controls. Furthermore, the difference between nonexcbf and exc-bf mothers was statistically significant. β CTX levels were significantly higher in the exc-bf group by about 70 pg/ml (β =69, p < .001 at 6 m) compared with nonexc-bf mothers. Effects persisted after we corrected for age and BMI (Table 2), except for the difference between nonexc-bf and exc-bf mothers at 3 m, where statistical significance was no longer achieved (β =36, p=.308). For P1NP, levels in bf subjects were significantly higher by about 60 ng/ml when compared with the controls at 6 m in the univariate analyses (β =60 [exc-bf vs. controls], β =61 [nonexc-bf vs. controls] at 6 m, both *ps*<0.001).

Results were comparable at the 3 m examination. We found similar patterns for OC. Effects persisted after we corrected for age and BMI (Fig. 2b, c). We did not detect a difference between exc-bf and nonexc-bf mothers in the univariate or multivariate analyses for either P1NP or OC (see Table 2).

For PTH, we did not find significant differences between the groups at 3 m in the univariate analyses, whereas the levels were marginally lower in exc-bf mothers compared with nonexc-bf mothers after we adjusted for age and BMI ($\beta = -0.5$, p = .049). For the multivariate analyses, nonexc-bf mothers also showed significantly higher levels than the controls ($\beta = 0.6$, p = .030) At 6 m, PTH reached approximately 0.3 pmol/L higher levels in exc-bf mothers $(\beta = 0.3, p = .005)$ compared with nonexc-bf mothers and approximately 0.5 pmol/L higher levels when compared with the controls ($\beta = 0.5$, p < .001) in the univariate analyses. Nonexc-bf mothers showed marginally but not significantly higher levels than the controls ($\beta = 0.1, p = .211$). However, this effect became statistically significant after we corrected for the other predictors in the multivariate models ($\beta = 0.4$, p=.003). Moreover, the difference between exc-bf mothers and controls was also slightly larger after we corrected for age and BMI ($\beta = 0.6, p < .001$), whereas the difference between nonexc-bf and exc-bf women was no longer statistically significant ($\beta = 0.2, p = .175$; Fig. 3a).

For E₂, exc-bf women had significantly lower levels compared with the controls in the univariate analyses ($\beta = -124$, p < .001 at 3 m, $\beta = -82$, p = .012 at 6 m). After adjusting for age and BMI, this effect remained significant only at 3 m ($\beta = -150$, p < .001; Fig. 3b). The E₂ levels of nonexc-bf women were also lower when compared with the controls; however, this effect was only significant at 6 m in the univariate analyses ($\beta = -95$, p = .010). We did not find a significant difference between nonexc-bf and exc-bf mothers.

Total serum Ca and P were significantly higher in exc-bf and nonexc-bf mothers than in the controls at 3 m and 6 m in the univariate analyses. Effects persisted after we corrected for age and BMI except for the difference between nonexcbf women and the controls at 3 m for total serum Ca levels (Table 2). There were no differences between exc-bf and nonexc-bf mothers in the univariate or multivariate models (Suppl. Figures 1 a and b) [29].

Model	Visit (m)	Comparisons	Comparisons Univariate		Multivariate		
			estimate	<i>p</i> -value	estimate	<i>p</i> -value	
PTH (pmol/l)	3	Exc-bf-nonexc-bf	$\beta = -0.4; 95\% \text{ CI} [-0.8, 0]$	p=.053	$\beta = -0.5; 95\%$ CI $[-1, 0]$	<i>p</i> = .049	
		Exc-bf-controls	β=-0.1; 95% CI [-0.3, 0]	p = .074	β=0; 95% CI [-0.2, 0.3]	p = .647	
		Nonexc-bf-controls	β=0.3; 95% CI [-0.1, 0.7]	p = .202	$\beta = 0.6; 95\%$ CI [0.1, 1.1]	p = .030	
	6	Exc-bf-nonexc-bf	β=0.3; 95% CI [0.1, 0.6]	p = .005	$\beta = 0.2; 95\%$ CI $[-0.1, 0.5]$	p = .175	
		Exc-bf-controls	β=0.5; 95% CI [0.3, 0.7]	p < .001	β=0.6; 95% CI [0.4, 0.9]	p < .001	
		Nonexc-bf-controls	β=0.1; 95% CI [-0.1, 0.3]	p = .211	$\beta = 0.4; 95\%$ CI [0.1, 0.7]	p = .003	
E ₂ (pmol/l)	3	Exc-bf-nonexc-bf	β=-46; 95% CI [-122, 30]	p=.233	β=-49; 95% CI [-197, 100]	p = .522	
		Exc-bf-controls	β=-124; 95% CI [-159, -88]	p < .001	β=-150; 95% CI [-207, -94]	p < .001	
		Nonexc-bf-controls	β=-78; 95% CI [-157, 1]	p = .054	β=-102; 95% CI [-248, 45]	p = .174	
	6	Exc-bf-nonexc-bf	β=12; 95% CI [-52, 76]	p = .704	β=81; 95% CI [-76, 237]	p = .313	
		Exc-bf-controls	β=-82; 95% CI [-146, -18]	p = .012	β=-55; 95% CI [-172, 62]	p = .358	
		Nonexc-bf-controls	β=-95; 95% CI [-166, -23]	p = .010	β=-135; 95% CI [-278, 7]	p = .063	
ßCTX (pg/ml)	3	Exc-bf-nonexc-bf	β=77; 95% CI [13, 142]	<i>p</i> =.019	β=36; 95% CI [33, 105]	p = .308	
		Exc-bf-controls	β=410; 95% CI [385, 434]	p < .001	β=323; 95% CI [295, 351]	p < .001	
		Nonexc-bf-controls	β=332; 95% CI [268, 397]	p < .001	β=287; 95% CI [219, 356]	p < .001	
	6	Exc-bf-nonexc-bf	β=69; 95% CI [33, 104]	p < .001	β=63; 95% CI [23, 103]	p = .002	
		Exc-bf-controls	β=419; 95% CI [391, 446]	p < .001	β=334; 95% CI [301, 367]	p < .001	
		Nonexc-bf-controls	β=350; 95% CI [318, 382]	p < .001	β=271; 95% CI [234, 308]	p < .001	
P1NP (ng/ml)	3	Exc-bf-nonexc-bf	$\beta = 1;95\%$ CI [7, 9]	<i>p</i> =.835	β=2; 95% CI [-7, 10]	<i>p</i> =.711	
		Exc-bf-controls	β=53; 95% CI [50, 56]	<i>p</i> < .001	β=43; 95% CI [39, 46]	p < .001	
		Nonexc-bf-controls	β=52; 95% CI [44, 60]	p < .001	β=41; 95% CI [32, 50]	p < .001	
	6	Exc-bf-nonexc-bf	β=-1; 95% CI [-5, 4]	<i>p</i> =.716	$\beta = -4;95\%$ CI $[-10, 1]$	p = .094	
		Exc-bf-controls	β=60; 95% CI [56, 63]	<i>p</i> < .001	β=50; 95% CI [46, 55]	p < .001	
		Nonexc-bf-controls	β=61; 95% CI [57, 65]	p < .001	β=55; 95% CI [50, 60]	p < .001	
OC (ng/ml)	3	Exc-bf-nonexc-bf	$\beta = 1;95\%$ CI $[-1, 4]$	p=.317	β=2; 95% CI [-1, 6]	p = .122	
		Exc-bf-controls	$\beta = 19;95\%$ CI [18, 20]	p < .001	$\beta = 15; 95\%$ CI [14, 17]	p < .001	
		Nonexc-bf-controls	$\beta = 17;95\%$ CI [14, 20]	p < .001	$\beta = 13;95\%$ CI [10, 16]	p < .001	
	6	Exc-bf-nonexc-bf	$\beta = 1;95\%$ CI [0, 3]	p = .140	β=0; 95% CI [-2, 2]	p = .972	
		Exc-bf-controls	β=21; 95% CI [19, 22]	p < .001	$\beta = 17;95\%$ CI [15, 18]	p < .001	
		Nonexc-bf-controls	β=19; 95% CI [18, 21]	p < .001	$\beta = 16;95\%$ CI [15, 18]	p < .001	
Ca (mmol/l)	3	Exc-bf – nonexc-bf	$\beta = 0.02; 95\%$ CI $[-0.01, 0.04]$	<i>p</i> =.212	$\beta = 0.02; 95\%$ CI [-0.01, 0.05]	<i>p</i> =.171	
		Exc-bf - controls	$\beta = 0.06; 95\%$ CI [0.05, 0.07]	p < .001	$\beta = 0.05; 95\%$ CI [0.04, 0.06]	p < .001	
		Nonexc-bf-controls	β=0.04; 95% CI [0.02, 0.07]	p < .001	β=0.03; 95% CI [0, 0.06]	p = .088	
	6	Exc-bf-nonexc-bf	β=0; 95% CI [-0.02, 0.01]	p = .741	$\beta = 0;95\%$ CI $[-0.02, 0.01]$	p = .699	
		Exc-bf-controls	β=0.03; 95% CI [0.02, 0.04]	p < .001	β=0.02; 95% CI [0.01, 0.04]	p < .001	
		Nonexc-bf-controls	β=0.04; 95% CI [0.02, 0.05]	p < .001	β=0.03; 95% CI [0.01, 0.04]	p < .001	
P (mmol/l)	3	Exc-bf-nonexc-bf	β=0.04; 95% CI [-0.01, 0.08]	p=.085	β=0.04; 95% CI [-0.02, 0.1]	p=.189	
		Exc-bf-controls	β=0.22; 95% CI [0.2,0.23]	p < .001	β=0.2; 95% CI [0.18,0.22]	p < .001	
		Nonexc-bf-controls	β=0.18; 95% CI [0.14,0.22]	p < .001	β=0.16; 95% CI [0.1,0.22]	p < .001	
	6	Exc-bf-nonexc-bf	β=0.01; 95% CI [-0.02,0.03]	p=.482	β=0; 95% CI [-0.04,0.03]	p=.820	
		Exc-bf-controls	β=0.16; 95% CI [0.14,0.18]	p < .001	β=0.13; 95% CI [0.11,0.16]	p < .001	
		Nonexc-bf-controls	β=0.15; 95% CI [0.13,0.17]	p < .001	β=0.14; 95% CI [0.11,0.17]	p < .001	

BMI and age were controlled for in the multivariate analyses. Comparisons were between exclusively breastfeeding (exc-bf) women, nonexclusively bf (nonexc-bf) women, and nonpregnant, nonlactating controls. *BMI* body mass index, *m* months



Fig. 2 Serum β CTX (**a**), P1NP (**b**), and OC (**c**) levels at 6 months postpartum. Exclusively breastfeeding (exc-bf) and non-exclusively breastfeeding (nonexc-bf) women were compared with controls at each visit. Results of multivariate analyses. Grey lines indicate reference levels. Bone turnover marker levels in exc-bf and

nonexc-bf women were significantly higher than those of controls (all ps < .001) and either close to (BCTX, OC) or above the upper reference limit (P1NP). BCTX was significantly higher in exc-bf women than in nonexc-bf women (p = .002)

Discussion

The aim of this study was to examine serum PTH, E_2 , BTM, Ca, and P in bf women and controls to extend the understanding of how molecular mechanisms regulate Ca metabolism during lactation. To do so, we compared bmrp levels between exc-bf and nonexc-bf women as well as healthy, nonpregnant, nonlactating controls.

In previous studies, lactational bone loss has frequently been attributed to low levels of estrogen rather than to classic calciotropic hormones including PTH and Calcitriol [6, 7, 18, 20, 30–34]. For PTH, in earlier studies, either no difference was found between bf and nonbf women or controls (a finding that is consistent with our finding at 3 m when we compared exc-bf women with controls) [15–17], or else the hormone was suppressed in lactating women [11, 18–20,





Fig.3 Serum PTH (**a**) and serum Estradiol (E_2) (**b**) at the respective visit. Exclusively breastfeeding (exc-bf) and nonexclusively breastfeeding (nonexc-bf) women were compared with controls at each visit. Results of multivariate analyses. Grey lines indicate refer-

35]. Although our data showed significantly higher PTH levels in exc-bf and nonexc-bf women when compared with the controls at 6 m, we believe that this effect has only minor clinical and biological relevance, as the distributions of data in all three groups existed largely in the lower half of the reference range of the assay (Fig. 3a). Therefore, we surmise that lactational bone loss is relatively independent of PTH [36]. With respect to E_2 , lactation causes hypoestrogenemia by suppressing the hypothalamic-pituitary-ovarian axis. In agreement with other authors [6, 11, 13], our results supported this effect, as we found suppressed levels in bf women compared with controls. The difference was significant only for exc-bf women at 3 m, most likely due to the smaller case number with available E₂ values in the exc-bf group at 6 m (Suppl. Table 1) [29]. Although the findings were not significant, E_2 levels were also suppressed in the nonexc-bf women, a finding that could be attributed to the effect of partial bf on the hypothalamic-pituitary-ovarian axis.

During lactation, bone turnover is increased as reflected by high rates of osteoclast-driven bone resorption and osteoblast-driven bone formation [5–7, 12, 13, 16, 21, 23, 37]. Similarly, our study demonstrates elevated rates of bone resorption markers. In exc-bf women, we found increased ßCTX levels that were twice as high as those of nonpregnant, nonlactating controls, confirming prior findings by Carneiro et al. [13]. In our data, ßCTX was also higher in exc-bf women when compared with nonexc-bf women, but this difference was significant only at 6 m which might

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ence levels. PTH was significantly increased in exc-bf and nonexc-bf women compared with controls (p < .001 and p = .003, respectively), whereas E_2 levels were suppressed in exc-bf and nonexc-bf women,

once again reside in the smaller sample size of the nonexc-bf women at 3 m (Suppl. Table 1) [29].

below levels of controls (p < .001 and p = .174, respectively)

Similar to findings from previous papers [9, 13, 15, 20–22], our bone formation markers, assessed as OC and P1NP, were significantly elevated in exc-bf women when compared with the controls. Both OC and P1NP levels were either at the upper limit (OC) of the reference range or even considerably above it (P1NP). The substantially elevated rates of bone formation and bone resorption markers in exc-bf women support the concept of high bone turnover during lactation.

We also observed increased bone formation markers (P1NP, OC) in nonexc-bf women with levels significantly above those of the controls and almost equal to those of exc-bf women. We surmise that this finding is due to the effect of partial bf. However, the significantly lower levels of BCTX in nonexc-bf compared with exc-bf women indicate a decrease in bone resorption that might be attributable to reduced bf as Kent et al. and others reported a normalization of bone resorption in weaning mothers [2, 15]. Nevertheless, we cannot completely rule out the possibility that the elevation in BTM in the lactation cohort was due to the effect of a previous pregnancy, as we did not have a pregnant nonbf control group, and bone turnover has been shown to also be increased in late pregnancy [9, 21, 38–40]. Further research is needed to compare bmrp in exc-bf and nonexc-bf women as well as in nonbf pregnant women.

Attributable to skeletal Ca mobilization, serum total Ca was significantly increased in exc-bf and nonexc-bf women

compared with the controls but remained within normal limits. Previous studies have also reported increased or normal levels [13, 17, 19–21, 31]. Results were similar for serum P. Increased bone resorption, along with decreased renal P excretion, is thought to be the cause of increased serum P levels in lactating women, as reported by others [2].

This study has some advantages and limitations. The greatest strength of the present study was that we had largesized lactation and control cohorts. To our knowledge, this is the largest published data that describes bone turnover in the comparison of breastfeeding and non-breastfeeding women. We were thereby able to provide an up-to-date overview on bone metabolism during lactation and re-evaluate the findings of other studies, some of which have been contradictory and have only been conducted in small samples [15, 17, 19, 21, 32, 41–43]. However, we lack measurements of Prolactin, PTHrP, Calcitonin and Calcitriol, which are also involved in bone turnover in animal or human models [2]. Missing data on other factors potentially affecting bone metabolism, such as menstrual status, breast milk volume, or Ca and P intake, must be considered when interpreting our results. In addition, we did not measure bone mineral density, so changes in BTM could not be directly correlated with bone loss. Furthermore, the number of cases we had in our lactation cohort varied by visit and lactation subgroup. In particular, the size of the group of nonexc-bf women at 3 m was comparatively small (Table 1), as the World Health Organization (WHO) recommends exclusive bf for 6 m [44]. Moreover, for the nonexc-bf group, we did not have any information about the frequency of bf. Both must be considered when interpreting our results. Lastly, the lactation and control cohorts differed significantly in BMI and age. The control cohort was significantly older and heavier. Consequently, we cannot completely rule out that the differences in bmrp levels between the lactation and control cohorts are related to the higher age and BMI of the controls. To summarize, this study shows that bone formation and bone resorption are substantially higher in exc-bf and nonexc-bf women compared with nonpregnant, nonlactating controls, indicating high bone turnover during lactation. Our data suggest that lactational bone loss is relatively independent of PTH as the data distribution of exc-bf, nonexc-bf, and control groups were in the underpart of the reference range.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00404-023-07189-0.

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Data availability Some or all datasets generated during and/or analyzed during the current study are not publicly available but are available from the corresponding author on reasonable request.

Declarations

Conflict of interest The authors have no relevant financial or non-financial interests to disclose.

Ethical approval The LIFE Child study protocol was designed in accordance with the Declaration of Helsinki and was approved by the Ethics Committee of the Medical Faculty of Leipzig University (Reg. No. 264/10-ek).

Consent to participate Informed consent was obtained from all individual participants included in the study.

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References

- Wysolmerski JJ (2010) Interactions between breast, bone, and brain regulate mineral and skeletal metabolism during lactation. Ann N Y Acad Sci 1192:161–169. https://doi.org/10.1111/j.1749-6632.2009.05249.x
- Kovacs CS (2016) Maternal mineral and bone metabolism during pregnancy, lactation, and post-weaning recovery. Physiol Rev 96:449–547. https://doi.org/10.1152/physrev.00027.2015
- Sowers M (1996) Pregnancy and lactation as risk factors for subsequent bone loss and osteoporosis. J Bone Miner Res 11:1052– 1060. https://doi.org/10.1002/jbmr.5650110803
- Kalkwarf HJ, Specker BL, Bianchi DC et al (1997) The effect of calcium supplementation on bone density during lactation and after weaning. N Engl J Med 337:523–528. https://doi.org/10. 1056/NEJM199708213370803

- Kovacs CS, Kronenberg HM (1997) Maternal-fetal calcium and bone metabolism during pregnancy, puerperium, and lactation. Endocr Rev 18:832–872. https://doi.org/10.1210/edrv.18.6.0319
- Sowers MF, Hollis BW, Shapiro B et al (1996) Elevated parathyroid hormone-related peptide associated with lactation and bone density loss. JAMA 276:549–554
- VanHouten JN, Wysolmerski JJ (2003) Low estrogen and high parathyroid hormone-related peptide levels contribute to accelerated bone resorption and bone loss in lactating mice. Endocrinology 144:5521–5529. https://doi.org/10.1210/en.2003-0892
- Kovacs CS (2012) The role of vitamin D in pregnancy and lactation: insights from animal models and clinical studies. Annu Rev Nutr 32:97–123. https://doi.org/10.1146/annur ev-nutr-071811-150742
- Møller UK, Streym S, Mosekilde L et al (2013) Changes in calcitropic hormones, bone markers and insulin-like growth factor I (IGF-I) during pregnancy and postpartum: a controlled cohort study. Osteoporos Int 24:1307–1320. https://doi.org/10.1007/ s00198-012-2062-2
- Kalkwarf HJ, Specker BL (1995) Bone mineral loss during lactation and recovery after weaning. Obstet Gynecol 86:26–32
- Krebs NF, Reidinger CJ, Robertson AD, Brenner M (1997) Bone mineral density changes during lactation: maternal, dietary, and biochemical correlates. Am J Clin Nutr 65:1738–1746. https://doi. org/10.1093/ajcn/65.6.1738
- Ritchie LD, Fung EB, Halloran BP et al (1998) A longitudinal study of calcium homeostasis during human pregnancy and lactation and after resumption of menses. Am J Clin Nutr 67:693–701. https://doi.org/10.1093/ajcn/67.4.693
- Carneiro RM, Prebehalla L, Tedesco MB et al (2010) Lactation and bone turnover: a conundrum of marked bone loss in the setting of coupled bone turnover. J Clin Endocrinol Metab 95:1767– 1776. https://doi.org/10.1210/jc.2009-1518
- McNeilly AS, Tay CC, Glasier A (1994) Physiological mechanisms underlying lactational amenorrhea. Ann N Y Acad Sci 709:145–155. https://doi.org/10.1111/j.1749-6632.1994.tb303 94.x
- Kent GN, Price RI, Gutteridge DH et al (1990) Human lactation: Forearm trabecular bone loss, increased bone turnover, and renal conservation of calcium and inorganic phosphate with recovery of bone mass following weaning. J Bone Miner Res 5:361–369. https://doi.org/10.1002/jbmr.5650050409
- Kalkwarf HJ, Specker BL, Ho M (1999) Effects of calcium supplementation on calcium homeostasis and bone turnover in lactating women. J Clin Endocrinol Metab 84:464–470. https://doi.org/ 10.1210/jcem.84.2.5451
- Greer FR, Tsang RC, Searcy JE et al (1982) Mineral homeostasis during lactation- relationship to serum 1,25-dihydroxyvitamin D, 25-hydroxyvitamin D, parathyroid hormone and calcitonin. Am J Clin Nutr 36:431–437. https://doi.org/10.1093/ajcn/36.3.431
- Kovacs CS, Chik CL (1995) Hyperprolactinemia caused by lactation and pituitary adenomas is associated with altered serum calcium, phosphate, parathyroid hormone (PTH), and PTH-related peptide levels. J Clin Endocrinol Metab 80:3036–3042. https:// doi.org/10.1210/jcem.80.10.7559893
- Affinito P, Tommaselli GA, di Carlo C et al (1996) Changes in bone mineral density and calcium metabolism in breastfeeding women: a one year follow-up study. J Clin Endocrinol Metab 81:2314–2318. https://doi.org/10.1210/jcem.81.6.8964870
- Dobnig H, Kainer F, Stepan V et al (1995) Elevated parathyroid hormone-related peptide levels after human gestation: relationship to changes in bone and mineral metabolism. J Clin Endocrinol Metab 80:3699–3707. https://doi.org/10.1210/jcem.80.12.85306 22

- Cross NA, Hillman LS, Allen SH et al (1995) Calcium homeostasis and bone metabolism during pregnancy, lactation, and postweaning: a longitudinal study. Am J Clin Nutr 61:514–523. https://doi.org/10.1093/ajcn/61.3.514
- Paoletti AM, Orrù M, Floris L et al (2003) Pattern of bone markers during pregnancy and their changes after delivery. Horm Res 59:21–29. https://doi.org/10.1159/000067935
- Sowers M, Eyre D, Hollis BW et al (1995) Biochemical markers of bone turnover in lactating and nonlactating postpartum women. J Clin Endocrinol Metab 80:2210–2216. https://doi.org/10.1210/ jcem.80.7.7608281
- 24. Quante M, Hesse M, Döhnert M et al (2012) The life child study: a life course approach to disease and health. BMC Public Health 12:1021
- the LIFE Child study team, Poulain T, Baber R, et al (2017) The LIFE Child study: a population-based perinatal and pediatric cohort in Germany. Eur J Epidemiol 32:145–158. https://doi.org/ 10.1007/s10654-016-0216-9
- 26. Geserick M, Vogel M, Eckelt F et al (2020) Car. Bone 132:115124. https://doi.org/10.1016/j.bone.2019.115124
- Gaudl A, Kratzsch J, Bae YJ et al (2016) Liquid chromatography quadrupole linear ion trap mass spectrometry for quantitative steroid hormone analysis in plasma, urine, saliva and hair. J Chromatogr A 1464:64–71. https://doi.org/10.1016/j.chroma.2016.07.087
- Bae YJ, Zeidler R, Baber R et al (2019) Reference intervals of nine steroid hormones over the life-span analyzed by LC-MS/MS: effect of age, gender, puberty, and oral contraceptives. J Steroid Biochem Mol Biol 193:105409. https://doi.org/10.1016/j.jsbmb. 2019.105409
- Nerius L, Vogel M, Ceglarek U et al (2023) Supplemental Material 2023: bone turnover in lactating and non lactating women. Zenodo. https://doi.org/10.5281/zenodo.7865186
- VanHouten JN, Dann P, Stewart AF et al (2003) Mammary-specific deletion of parathyroid hormone-related protein preserves bone mass during lactation. J Clin Invest 112:1429–1436. https:// doi.org/10.1172/JCI19504
- Grill V, Hillary J, Ho PMW et al (1992) Parathyroid hormonerelated protein: a possible endocrine function in lactation. Clin Endocrinol 37:405–410. https://doi.org/10.1111/j.1365-2265. 1992.tb02350.x
- Sowers M, Zhang D, Hollis BW et al (1998) Role of calciotrophic hormones in calcium mobilization of lactation. Am J Clin Nutr 67:284–291. https://doi.org/10.1093/ajcn/67.2.284
- Bucht E, Rong H, Bremme K et al (1995) Midmolecular parathyroid hormone-related peptide in serum during pregnancy, lactation and in umbilical cord blood. Eur J Endocrinol 132:438–443. https://doi.org/10.1530/eje.0.1320438
- Lippuner K, Zehnder HJ, Casez JP et al (1996) PTH-related protein is released into the mother's bloodstream during lactation: evidence for beneficial effects on maternal calcium-phosphate metabolism. J Bone Miner Res 11:1394–1399. https://doi.org/ 10.1002/jbmr.5650111004
- Hillman L, Sateesha S, Haussler M et al (1981) Control of mineral homeostasis during lactation: interrelationships of 25-hydroxyvitamin D, 24,25-dihydroxyvitamin D, 1,25-dihydroxyvitamin D, parathyroid hormone, calcitonin, prolactin, and estradiol. Am J Obstet Gynecol 139:471–476. https://doi.org/10.1016/0002-9378(81)90327-6
- Retallack RW, Jeffries M, Kent GN et al (1977) Physiological hyperparathyroidism in human lactation. Calcif Tissue Res 22(Suppl):142–146. https://doi.org/10.1007/BF02064055

- Kent GN, Price RI, Gutteridge DH et al (1993) Effect of pregnancy and lactation on maternal bone mass and calcium metabolism. Osteoporosis Int 3:44–47. https://doi.org/10.1007/BF016 21861
- Cross NA, Hillman LS, Allen SH, Krause GF (1995) Changes in bone mineral density and markers of bone remodeling during lactation and postweaning in women consuming high amounts of calcium. J Bone Miner Res 10:1312–1320. https://doi.org/10. 1002/jbmr.5650100907
- 39. Kaur M, Godber IM, Lawson N et al (2003) Changes in serum markers of bone turnover during normal pregnancy. Ann Clin Biochem 40:508–513. https://doi.org/10.1258/000456303322326 416
- Black AJ, Topping J, Durham B et al (2000) A detailed assessment of alterations in bone turnover, calcium homeostasis, and bone density in normal pregnancy. J Bone Miner Res 15:557–563. https://doi.org/10.1359/jbmr.2000.15.3.557

- Dobnig H, Turner RT (1997) The effects of programmed administration of human parathyroid hormone fragment (1–34) on bone histomorphometry and serum chemistry in rats. Endocrinology 138:4607–4612. https://doi.org/10.1210/endo.138.11.5505
- 42. More C, Bhattoa HP, Bettembuk P, Balogh A (2003) The effects of pregnancy and lactation on hormonal status and biochemical markers of bone turnover. Eur J Obstetr Gynecol Reprod Biol 106:209–213. https://doi.org/10.1016/S0301-2115(02)00237-3
- Grill V, Hillary J, Ho PM et al (1992) Parathyroid hormone-related protein: a possible endocrine function in lactation. Clin Endocrinol (Oxf) 37:405–410. https://doi.org/10.1111/j.1365-2265. 1992.tb02350.x
- Hoddinott P, Tappin D, Wright C (2008) Breast feeding. BMJ 336:881–887. https://doi.org/10.1136/bmj.39521.566296.BE

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