CORRESPONDENCE



## Gene transcript fusions are associated with clinical outcomes and molecular groups of meningiomas

Naomi Zakimi<sup>1,2,3</sup> · Minh P. Nguyen<sup>1,2,3</sup> · David R. Raleigh<sup>1,2,3</sup>

Received: 21 January 2024 / Revised: 15 February 2024 / Accepted: 15 February 2024 © The Author(s) 2024

The discovery of molecular groups of meningiomas that are associated with distinct biological drivers, therapeutic vulnerabilities, and clinical outcomes provides a framework for redefining the classification of the most common primary intracranial tumor [2–4, 6, 14, 16, 18–20, 22, 25]. Meningiomas from the Merlin-intact molecular group with favorable clinical outcomes encode recurrent short somatic variants targeting *TRAF7*, *KLF4*, *PI3K*, *POLR2A*, or the Hedgehog pathway [2, 4, 18, 21, 22, 33, 34]. Recurrent short somatic variants in meningiomas from molecular groups with poor clinical outcomes, such as *TERT* promoter mutations or *CDKN2A/B* deletions [10, 24, 29], are rare. These data suggest that alternative genomic mechanism may contribute to the formation or progression of aggressive meningiomas.

Gene fusions form when independent DNA or RNA sequences are juxtaposed through (1) chromosome structural rearrangements such as translocations, deletions, duplications, or inversions, (2) transcription read-through of neighboring genes, or (3) pre-mRNA slicing [12]. Many gene fusions are oncogenic, and gene fusions have been reported in meningiomas [11, 32], particularly *NF2* structural rearrangements in radiation-induced meningiomas [1, 26], and *YAP1* fusions in rare pediatric meningiomas [28, 30]. Associations between gene transcript fusions and clinical outcomes across molecular groups of meningiomas are unknown.

To define the landscape of meningioma gene transcript fusions, paired-end RNA sequencing data from 302 consecutive frozen meningiomas with matched DNA methylation profiling and targeted gene expression data from The University of Hong Kong [2–4] were analyzed using Arriba (Supplementary Table 1, online resource), a highly accurate bioinformatic pipeline for fusion detection in cancer transcriptomes [8]. Single-end RNA sequencing reduces the accuracy of gene transcript fusion detection [8]. Nevertheless, 81.9% of gene transcript fusions that were identified in 2 or more meningiomas using Arriba to analyze single-end RNA sequencing data from 200 frozen meningiomas from the University of California San Francisco [2–4] were also identified in paired-end RNA sequencing data from meningiomas from The University of Hong Kong (Supplementary Table 2, online resource).

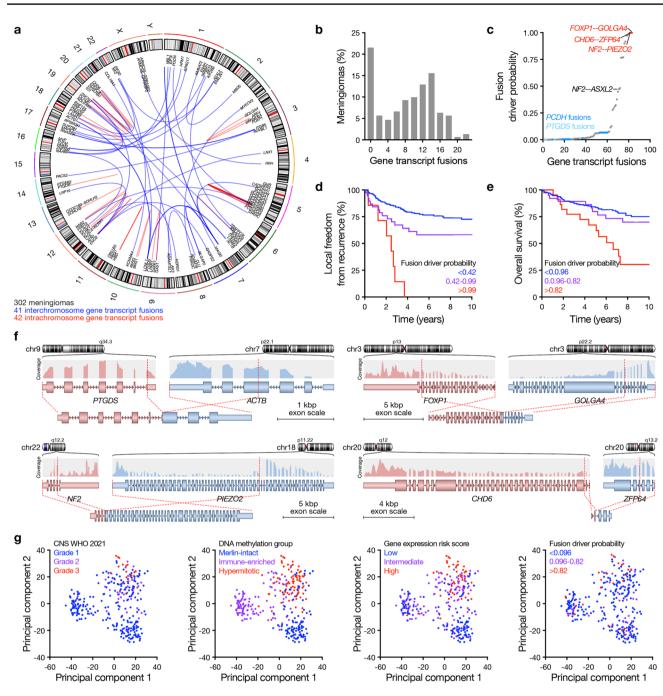
To distinguish gene transcript fusions with oncogenic potential from passenger fusion sequences, the Oncofuse bioinformatic pipeline was used to calculate the probability that meningioma gene transcript fusions were biological drivers based on features in known oncogenic fusions [27]. These analyses identified 83 gene transcript fusions that were present in an average of 20 meningiomas each (range 2-186 meningiomas/fusion) across the 302 samples with high quality paired-end RNA sequencing data from The University of Hong Kong (Fig. 1a and Supplementary Table 3, online resource). There were no gene transcript fusions in 21.5% of meningiomas (n=65) (Fig. 1b). Radiation-induced meningiomas (n = 15, average 11 fusions/ meningioma) had a higher burden of gene transcript fusions than sporadic meningiomas (n = 287, average 8 fusions/ meningioma, p = 0.0322, Student's t test) (Supplementary Table 3, online resource). The most common gene transcript fusions were at the protocadherin (PCDH) locus on chromosome 5, a degenerative genomic region with multiple transcription start sites and alternative splicing patterns that may be particularly susceptible to genomic instability (n = 1299 of 1701 gene transcript fusion events across 302meningiomas, 76.3%) (Fig. 1a and Supplementary Table 3, online resource). There were no associations between PCDH fusions and meningioma DNA molecular group [4], CNS

David R. Raleigh david.raleigh@ucsf.edu

<sup>&</sup>lt;sup>1</sup> Department of Radiation Oncology, University of California San Francisco, San Francisco, CA, USA

<sup>&</sup>lt;sup>2</sup> Department of Neurological Surgery, University of California San Francisco, San Francisco, CA, USA

<sup>&</sup>lt;sup>3</sup> Department of Pathology, University of California San Francisco, San Francisco, CA, USA



**Fig. 1** The landscape of meningioma gene transcript fusions. **a** Circos plot showing interchromosome (blue, n=41) and intrachromosome (red, n=42) Oncofuse results from paired-end RNA sequencing of 302 consecutive frozen meningiomas that underwent surgery. **b** Number of Oncofuse gene transcript fusions per meningioma. **c** Oncofuse driver probability of 83 gene transcript fusions across 302 consecutive meningiomas. **d** Local freedom from recurrence according to recursive partitioning analysis (RPA) of gene fusions with the highest Oncofuse driver probability per meningioma (p < 0.0001, Logrank test) (see also Supplementary Fig. 1a, online resource). **e** Overall

survival according to RPA of gene fusions with the highest Oncofuse driver probability per meningioma (see also Supplementary Fig. 1b, online resource) (p=0.0006, Log-rank test). **f** The structure of common gene fusions (*PTGDS*–*ACTB*) and gene fusions with the highest Oncofuse driver probabilities (*FOXP1*–*GOLGA4*, *NF2*–*PIEZO2*, *CHD6*–*CFP64*) across 302 consecutive meningiomas. **g** Principal component analysis from paired-end RNA sequencing of 302 consecutive frozen meningiomas shaded by CNS WHO 2021 grade, DNA methylation group, gene expression risk score, or Oncofuse driver probability

World Health Organization (WHO) 2021 grade [14], or gene expression risk score [2]. The second most common gene transcript fusions were translocations between prostaglandin D2 synthase (PTGDS) on chromosome 9 and actin genes on chromosomes 7 or 17 (n = 160 of 1701 gene transcript fusions events across 302 meningiomas, 9.4%) (Fig. 1a and Supplementary Table 3, online resource). PTGDS expression has been associated with meningioma development [9], and PTGDS fusions were more common in Merlin-intact meningiomas (n = 29 of 104 meningiomas, 27.9%) compared to Immune-enriched meningiomas (n = 13 of 105 meningiomas, 11.3%) or Hypermitotic meningiomas (n = 11 of 83 meningiomas, 13.3%, p=0.0037, Fischer's exact test). There were no associations between PTGDS fusions and meningioma CNS WHO 2021 grade (p=0.1712) [14] or meningioma gene expression risk score (p = 0.3644, Fisher's exact tests) [2]. These data suggest that PCDH gene transcript fusions may be a common finding in meningiomas that is not relevant to molecular characteristics, while PTGDS fusions may contribute to the molecular group of meningiomas with the best clinical outcomes.

To determine if meningioma gene transcript fusions were associated with clinical outcomes, fusion sequences were ranked by Oncofuse driver potential, which suggested that PCDH and PTGDS were low on the list of biologically-relevant fusions (Fig. 1c). Recursive partitioning analysis (RPA) was used to predict local freedom from recurrence or overall survival from the highest Oncofuse driver probability per meningioma (Supplementary Fig. 1 and Supplementary Table 4, online resource). Kaplan-Meier analysis showed that Oncofuse driver probability was significantly associated with meningioma local freedom from recurrence (Fig. 1d) and overall survival (Fig. 1e), with higher fusion driver probability associated with worse clinical outcome. The gene transcript fusions with the highest driver probabilities were novel inversions inactivating the transcription factor FOXP1 through juxtaposition next to GOLGA4 on chromosome 3 (n=2) or the chromatin remodeler CHD6 through juxtaposition next to ZFP64 on chromosome 20 (n=2), and novel translocations likely inactivating NF2 on chromosome 22 through juxtaposition next to *PIEZO2* on chromosome 18 (n=3)(Fig. 1a, c, f and Supplementary Table 3 and 4, online resource). A novel translocation inactivating NF2 through juxtaposition next to ASXL2 on chromosome 2 was also identified (n=2) (Fig. 1a, c, and Supplementary Table 3, online resource). Gene transcript fusions with high driver potential were validated using annoFuse, a complementary bioinformatic pipeline for annotation and prioritization of biologically relevant fusions [7] (Supplementary Table 5, online resource). In sum, these results suggest that oncogenic gene transcript fusions are minimally conserved across meningiomas, but that bioinformatic approaches can be used to group gene transcript fusions according to their predicted biological relevance and shed light on meningioma clinical outcomes.

Principal component analysis of differentially expressed genes and annotation of CNS WHO 2021 grade [14], DNA methylation group [4], gene expression risk score [2], and gene fusion driver probability across the 302 meningiomas from The University of Hong Kong showed that meningiomas with oncogenic fusions clustered with high-grade meningiomas and molecular groups of meningiomas that are associated with poor clinical outcomes (Fig. 1g). In support of these findings, univariate Cox regression analysis showed that the highest Oncofuse driver probability per meningioma was significantly associated with local freedom from recurrence (hazard ratio 2.73, 95% confidence interval 1.48–5.04, p = 0.001) and overall survival (hazard ratio 1.99, 95% confidence interval 1.02-3.87, p = 0.042). These findings were not conserved on multivariate Cox regression analysis, where unifying polygenic molecular features such as gene expression risk score [2] remained significant but Oncofuse driver probability did not (Supplementary Table 6, online resource). There were no significant differences in meningioma gene transcript fusion burden or driver probability according to extent of resection, prior treatment, or tumor size (Supplementary Table 4, online resource).

In conclusion, these data demonstrate that gene transcript fusions are associated with clinical outcomes and molecular groups of meningiomas. Our results shed new light on genomic mechanisms that may contribute to the formation or progression of meningiomas, but it remains to be determined if gene transcript fusions can be targeted to improve treatments or clinical outcomes for patients with meningiomas. None of the gene fusions reported here or in previous publications [1, 11, 26, 28, 32], with rare exceptions [23], juxtapose kinases or other oncogenes that may be susceptible to small molecule inhibition. Thus, mechanistic interrogation of biochemical pathways that may be dysregulated in response to gene fusions, and whether such pathways are conserved in meningiomas with divergent oncogenic gene fusions, may be necessary to translate these findings into a therapeutic framework.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00401-024-02708-y.

Acknowledgements D.R.R. is supported by the Trenchard Family Charitable Fund and NIH grants R01 CA262311 and P50 CA097257.

**Data availability** RNA sequencing, DNA methylation profiling, and targeted gene expression profiling data for all meningiomas analyzed in this study have been deposited to the NCBI Gene Expression Omnibus under accession numbers GSE183656, GSE101638, GSE212666, and GSE222054.

## Acta Neuropathologica (2024) 147:57

## Declarations

**Conflict of interest** The authors declare they have no competing interests related to this study.

**Ethical standards** This study was approved by the Committee on Human Research at the University of California San Francisco.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

## References

- 1. Agnihotri S, Suppiah S, Tonge PD, Jalali S, Danesh A, Bruce JP et al (2017) Therapeutic radiation for childhood cancer drives structural aberrations of NF2 in meningiomas. Nat Comms 8:186. https://doi.org/10.1038/s41467-017-00174-7
- Chen WC, Choudhury A, Youngblood MW, Polley M-YC, Lucas C-HG, Mirchia K et al (2023) Targeted gene expression profiling predicts meningioma outcomes and radiotherapy responses. Nat Med. https://doi.org/10.1038/s41591-023-02586-z
- Choudhury A, Chen WC, Lucas C-HG, Bayley JC, Harmanci AS, Maas SLN et al (2022) Hypermitotic meningiomas harbor DNA methylation subgroups with distinct biological and clinical features. Neuro Oncol. https://doi.org/10.1093/neuonc/noac224
- Choudhury A, Magill ST, Eaton CD, Prager BC, Chen WC, Cady MA et al (2022) Meningioma DNA methylation groups identify biological drivers and therapeutic vulnerabilities. Nat Genet 54:649–659. https://doi.org/10.1038/s41588-022-01061-8
- Dobin A, Davis CA, Schlesinger F, Drenkow J, Zaleski C, Jha S et al (2013) STAR: ultrafast universal RNA-seq aligner. Bioinformatics 29:15–21. https://doi.org/10.1093/bioinformatics/bts635
- Driver J, Hoffman SE, Tavakol S, Woodward E, Maury EA, Bhave V et al (2021) A Molecularly Integrated Grade for Meningioma. Neuro Oncol. https://doi.org/10.1093/neuonc/noab213
- Gaonkar KS, Marini F, Rathi KS, Jain P, Zhu Y, Chimicles NA et al (2020) annoFuse: an R Package to annotate, prioritize, and interactively explore putative oncogenic RNA fusions. BMC Bioinform 21:577. https://doi.org/10.1186/s12859-020-03922-7
- Haas BJ, Dobin A, Li B, Stransky N, Pochet N, Regev A (2019) Accuracy assessment of fusion transcript detection via readmapping and de novo fusion transcript assembly-based methods. Genome Biol 20:213. https://doi.org/10.1186/s13059-019-1842-9
- Kalamarides M, Stemmer-Rachamimov AO, Niwa-Kawakita M, Chareyre F, Taranchon E, Han Z-Y et al (2011) Identification of a progenitor cell of origin capable of generating diverse meningioma histological subtypes. Oncogene 30:2333–2344. https://doi. org/10.1038/onc.2010.609
- Khan AB, English CW, Chen WC, Athukuri P, Bayley JC, Brandt VL et al (2023) Even heterozygous loss of CDKN2A/B greatly accelerates recurrence in aggressive meningioma. Acta Neuropathol 145:501–503. https://doi.org/10.1007/s00401-023-02543-7

- Khan AB, Gadot R, Shetty A, Bayley JC, Hadley CC, Cardenas MF et al (2020) Identification of novel fusion transcripts in meningioma. J Neuro-Oncol 149:219–230. https://doi.org/10.1007/ s11060-020-03599-1
- Latysheva NS, Babu MM (2016) Discovering and understanding oncogenic gene fusions through data intensive computational approaches. Nucleic Acids Res 44:4487–4503. https://doi.org/10. 1093/nar/gkw282
- Liao Y, Smyth GK, Shi W (2014) featureCounts: an efficient general purpose program for assigning sequence reads to genomic features. Bioinformatics 30:923–930. https://doi.org/10.1093/ bioinformatics/btt656
- Louis DN, Perry A, Wesseling P, Brat DJ, Cree IA, Figarella-Branger D et al (2021) The 2021 WHO Classification of Tumors of the Central Nervous System: a summary. Neuro Oncol 23:1231–1251. https://doi.org/10.1093/neuonc/noab106
- Love MI, Huber W, Anders S (2014) Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. Genome Biol 15:550. https://doi.org/10.1186/s13059-014-0550-8
- Maas SLN, Stichel D, Hielscher T, Sievers P, Berghoff AS, Schrimpf D et al (2021) Integrated Molecular-Morphologic Meningioma Classification: A Multicenter Retrospective Analysis. J Clin Oncol, Retrospectively and Prospectively Validated. https:// doi.org/10.1200/jco.21.00784
- Martin M (2011) Cutadapt removes adapter sequences from highthroughput sequencing reads. EMBnetJ 17:10–12. https://doi.org/ 10.14806/ej.17.1.200
- Nassiri F, Liu J, Patil V, Mamatjan Y, Wang JZ, Hugh-White R et al (2021) A clinically applicable integrative molecular classification of meningiomas. Nature 597:119–125. https://doi.org/ 10.1038/s41586-021-03850-3
- Olar A, Wani KM, Wilson CD, Zadeh G, DeMonte F, Jones DTW et al (2017) Global epigenetic profiling identifies methylation subgroups associated with recurrence-free survival in meningioma. Acta Neuropathol 133:431–444. https://doi.org/10.1007/ s00401-017-1678-x
- Ostrom QT, Price M, Neff C, Cioffi G, Waite KA, Kruchko C et al (2022) CBTRUS Statistical Report: Primary Brain and Other Central Nervous System Tumors Diagnosed in the United States in 2015–2019. Neuro Oncol 24:v1–v95. https://doi.org/10.1093/ neuonc/noac202
- Paramasivam N, Hübschmann D, Toprak UH, Ishaque N, Neidert M, Schrimpf D et al (2019) Mutational patterns and regulatory networks in epigenetic subgroups of meningioma. Acta Neuropathol 208:345–414. https://doi.org/10.1007/s00401-019-02008-w
- Patel AJ, Wan Y-W, Al-Ouran R, Revelli J-P, Cardenas MF, Oneissi M et al (2019) Molecular profiling predicts meningioma recurrence and reveals loss of DREAM complex repression in aggressive tumors. Proc Natl Acad Sci USA 116:21715–21726. https://doi.org/10.1073/pnas.1912858116
- Sadagopan NS, Nandoliya KR, Youngblood MW, Horbinski CM, Ahrendsen JT, Magill ST (2023) A novel BRAF::PTPRN2 fusion in meningioma: a case report. Acta Neuropathol Commun 11:194. https://doi.org/10.1186/s40478-023-01668-w
- Sahm F, Schrimpf D, Olar A, Koelsche C, Reuss D, Bissel J et al (2016) TERT Promoter Mutations and Risk of Recurrence in Meningioma. J Natl Cancer Inst. https://doi.org/10.1093/jnci/ djv377
- Sahm F, Schrimpf D, Stichel D, Jones DTW, Hielscher T, Schefzyk S et al (2017) DNA methylation-based classification and grading system for meningioma: a multicentre, retrospective analysis. Lancet Oncol. https://doi.org/10.1016/s1470-2045(17) 30155-9
- Sahm F, Toprak UH, Hübschmann D, Kleinheinz K, Buchhalter I, Sill M et al (2017) Meningiomas induced by low-dose radiation carry structural variants of NF2 and a distinct mutational

signature. Acta Neuropathol 134:155–158. https://doi.org/10. 1007/s00401-017-1715-9

- Shugay M, de Mendíbil IO, Vizmanos JL, Novo FJ (2013) Oncofuse: a computational framework for the prediction of the oncogenic potential of gene fusions. Bioinformatics 29:2539–2546. https://doi.org/10.1093/bioinformatics/btt445
- Sievers P, Chiang J, Schrimpf D, Stichel D, Paramasivam N, Sill M et al (2019) YAP1-fusions in pediatric NF2-wildtype meningioma. Acta Neuropathol 91:520–524. https://doi.org/10.1007/ s00401-019-02095-9
- Sievers P, Hielscher T, Schrimpf D, Stichel D, Reuss DE, Berghoff AS et al (2020) CDKN2A/B homozygous deletion is associated with early recurrence in meningiomas. Acta Neuropathol 140:409–413. https://doi.org/10.1007/s00401-020-02188-w
- Szulzewsky F, Arora S, Arakaki AKS, Sievers P, Bonnin DAA, Paddison PJ et al (2022) Both YAP1-MAML2 and constitutively active YAP1 drive the formation of tumors that resemble NF2 mutant meningiomas in mice. Genes Dev 36:857–870. https:// doi.org/10.1101/gad.349876.122
- Uhrig S, Ellermann J, Walther T, Burkhardt P, Fröhlich M, Hutter B et al (2021) Accurate and efficient detection of gene fusions from RNA sequencing data. Genome Res. https://doi.org/10.1101/ gr.257246.119

- 32. Viaene AN, Zhang B, Martinez-Lage M, Xiang C, Tosi U, Thawani JP et al (2019) Transcriptome signatures associated with meningioma progression. Acta Neuropathol Commun 7:67–13. https://doi.org/10.1186/s40478-019-0690-x
- Youngblood MW, Duran D, Montejo JD, Li C, Omay SB, Ozduman K et al (2019) Correlations between genomic subgroup and clinical features in a cohort of more than 3000 meningiomas. J Neurosurg 1:1–10. https://doi.org/10.3171/2019.8.jns191266
- Youngblood MW, Erson-Omay Z, Li C, Najem H, Cokun S, Tyrtova E et al (2023) Super-enhancer hijacking drives ectopic expression of hedgehog pathway ligands in meningiomas. Nat Commun 14:6279. https://doi.org/10.1038/s41467-023-41926-y
- Zhang H, Meltzer P, Davis S (2013) RCircos: an R package for Circos 2D track plots. BMC Bioinform 14:244. https://doi.org/10. 1186/1471-2105-14-244

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.