



## Correction to: Synthesis of amphiphilic block copolymers containing ferrocene–boronic acid and their micellization, redox-responsive properties and glucose sensing

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The authors wish to correct the following mistakes or unclear description in their publication:

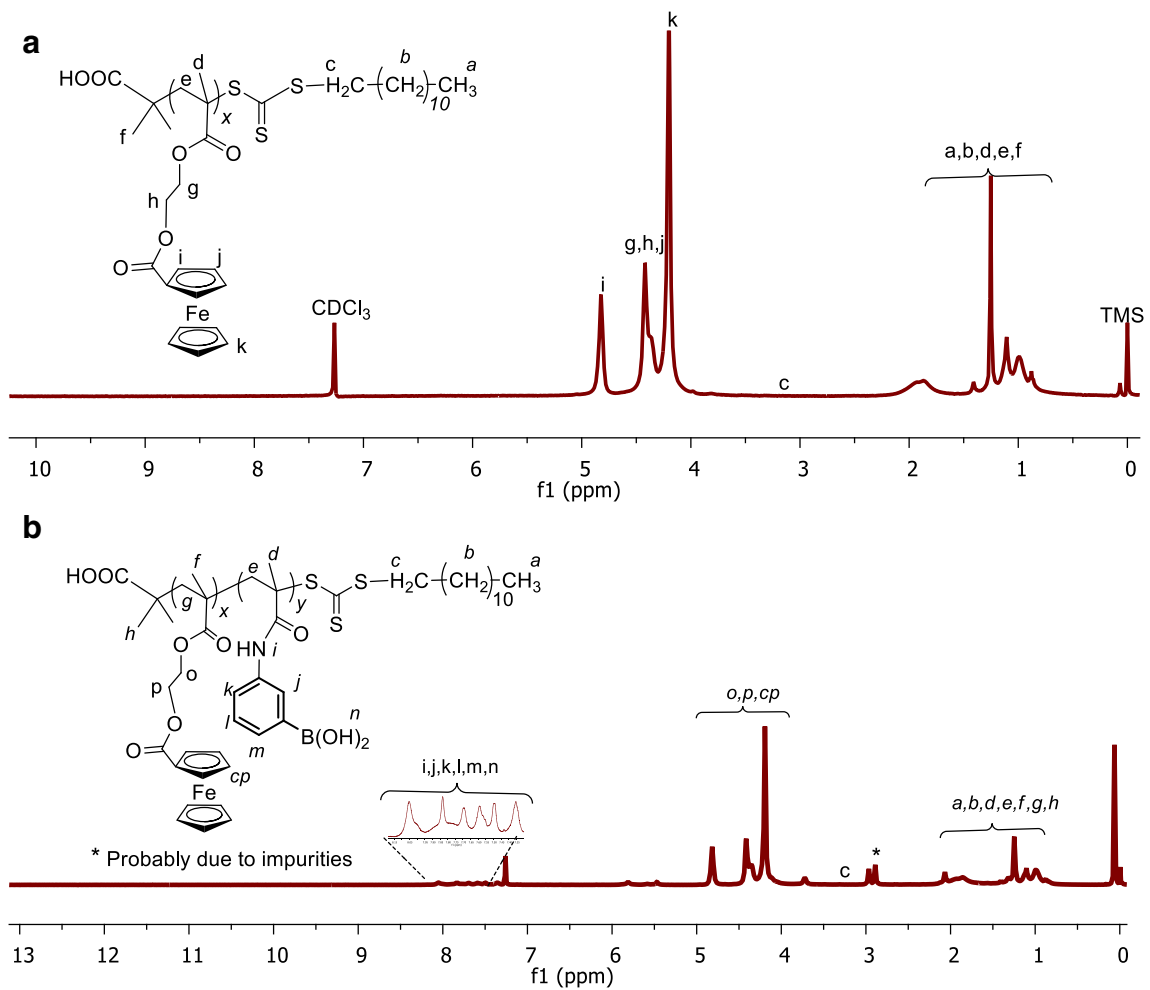
1. The boronic acid containing monomer is not a methylvinylamine (e.g. in the first paragraph of the section materials). Instead, its name is 3-methacrylamido phenylboronic acid.
2. Polymer PMAEFc is not a macro initiator, as stated on page 1000, right column, line 4, but a macro chain transfer agent.
3. On page 997, in the last paragraph of the right column, the 15 mg of PMAEFc correspond to  $1.901 \times 10^{-6}$  mol and not to 0.0438 mmol. As shown in Table 2, the Mn of PMAEFc is  $7.89 \times 10^3$  g/mol.
4. In Fig. 2B, the signal at about 3 ppm is not due to the protons labeled “d”. Rather, the signal of the protons of type “d” is part of the complex signal group between 0.9 and 2.1 ppm. The signal at 2.9 ppm is most probably due to impurities. The revised Fig. 2 is given below.
5. The signals of the dodecyl chain of the RAFT agent comprise 3 types of chemical shifts of the protons: -S-CH<sub>2</sub>-(CH<sub>2</sub>)<sub>10</sub>-CH<sub>3</sub>. The protons from the -S-CH<sub>2</sub>- group in low molar mass RAFT agent starting compound give rise to a separate signal at about 3.2 ppm in CDCl<sub>3</sub>. The molecular weights of PMAEFc and PMAEFc-*b*-PMVAPBA are  $7.89 \times 10^3$  g/mol and  $1.92 \times 10^5$  g/mol. Probably, due to their relatively high molecular weights, the peaks of this group are not obvious in Fig. 2.
6. On page 1000, in the 3rd paragraph of the right column, the relationship between the conversion rate and the reaction time in Fig. 3 is not linear. Therefore, the process is not well-controlled, which is possibly due to the structure of the monomers.
7. The size of the PMAEFc-*b*-PMVAPBA micelles, which was found in the range of 530–721 nm by DLS (page 1001, 2nd paragraph of the left column) may be due to aggregates.
8. Additional discussion about the change of the micellar sizes in Table 4 concerning the increase of the aggregate sizes by a factor of 3 in diameter (and thus by a factor of ca. 27 in volume): The data were obtained by DLS. It is possible that, when the ferrocene group is oxidized, it is more hydrophilic compared to the neutral state. The micellar core would become looser and the size would increase. Another possible reason is that the loose structure of the micelles in the oxidized state would facilitate the aggregation and would result in a much larger size in DLS measurement.
9. The cartoons should be removed from Fig. 5 and Fig. 7 since they may be unclear.

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**Fig. 2**  $^1\text{H}$  NMR spectra. (A) PMAEFc and (B) PMAEFc-*b*-PMVAPBA