#### **ORIGINAL CONTRIBUTION**



# Iterative assessment of a sports rehydration beverage containing a novel amino acid formula on water uptake kinetics

Mark P. Funnell<sup>1</sup> · Loris A. Juett<sup>1</sup> · Kirsty M. Reynolds<sup>1</sup> · Drusus A. Johnson<sup>1</sup> · Ruth M. James<sup>2</sup> · Stephen A. Mears<sup>1</sup> · Samuel N. Cheuvront<sup>3,4</sup> · Robert W. Kenefick<sup>3</sup> · Lewis J. James<sup>1</sup>

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#### Abstract

**Purpose** Rapid gastric emptying and intestinal absorption of beverages is essential for rapid rehydration, and certain amino acids (AA) may augment fluid delivery. Three sugar-free beverages, containing differing AA concentrations (AA + PZ), were assessed for fluid absorption kinetics against commercial sugar-free (PZ, GZ) and carbohydrate-containing (GTQ) beverages. **Methods** Healthy individuals (n = 15-17 per study) completed three randomised trials. Three beverages (550–600 mL) were ingested in each study (Study 1: AA + PZ [17.51 g/L AA], PZ, GZ; Study 2: AA + PZ [6.96 g/L AA], PZ, GZ; Study 3: AA + PZ [3.48 g/L AA], PZ, GTQ), containing 3.000 g deuterium oxide (D<sub>2</sub>O). Blood samples were collected pre-, 2-min, 5-min, and every 5-min until 60-min post-ingestion to quantify maximal D<sub>2</sub>O enrichment (Cmax), time Cmax occurred (Tmax) and area under the curve (AUC).

**Results** Study 1: AUC (AA + PZ: 15,184 ± 3532 8% vs. VSMOW; PZ: 17,328 ± 3153 8% vs. VSMOW; GZ: 17,749 ± 4204 8% vs. VSMOW;  $P \le 0.006$ ) and Tmax ( $P \le 0.005$ ) were lower for AA + PZ vs. PZ/GZ. Study 2: D<sub>2</sub>O enrichment characteristics were not different amongst beverages ( $P \ge 0.338$ ). Study 3: Cmax (AA + PZ: 440 ± 94 8% vs. VSMOW; PZ: 429 ± 83 8% vs. VSMOW; GTQ: 398 ± 81 8% vs. VSMOW) was greater (P = 0.046) for AA + PZ than GTQ, with no other differences ( $P \ge 0.106$ ).

**Conclusion** The addition of small amounts of AA (3.48 g/L) to a sugar-free beverage increased fluid delivery to the circulation compared to a carbohydrate-based beverage, but greater amounts (17.51 g/L) delayed delivery.

Keywords Rate of absorption · Rehydration · Deuterium · Recovery · Fluid balance

## Introduction

Rapid gastric emptying and intestinal absorption of ingested fluid is essential for quickly replacing fluid losses incurred during exercise, heat stress and illness [1]. The rate at which ingested fluids are available to replace fluid losses is dependent on the speed the fluid empties from the stomach (i.e.

Lewis J. James L.James@lboro.ac.uk

- <sup>1</sup> National Centre for Sport and Exercise Medicine, School of Sport, Exercise and Health Sciences, Loughborough University, Leicestershire LE11 3TU, UK
- <sup>2</sup> Sport, Health and Performance Enhancement Research Centre, School of Science and Technology, Nottingham Trent University, Nottingham NG11 8NS, UK
- <sup>3</sup> Entrinsic Bioscience, LLC, Norwood, MA 02062, USA
- <sup>4</sup> Sports Science Synergy, LLC, Franklin, MA 02038, USA

gastric emptying rate) and the rate of absorption at the site of the intestine [2]. The constituent solutes of a beverage significantly affect the gastric emptying rate and intestinal absorption, and thus, how quickly the fluid enters the circulation [1–3]. Rehydration beverages typically contain a mixture of electrolytes and carbohydrate [3] and are formulated to promote rapid gastric emptying and intestinal absorption, consequently facilitating delivery of fluids to the circulation as quickly as possible [4–6].

Beverages spiked with heavy water (deuterium oxide,  $D_2O$ ) provide an integrated measure of both gastric emptying and intestinal absorption of fluids [7]. Although  $D_2O$  does not provide a quantitative value for the amount of water delivered into the vasculature at any given time point, the temporal accumulation and kinetics of  $D_2O$  can be mathematically described allowing for a quantitative comparison of differences in fluid absorption amongst beverages [8–11]. Studies have demonstrated superior  $D_2O$  absorption kinetics when ingesting dilute carbohydrate beverages (i.e. sports beverages; < 6% carbohydrate) compared to water [11, 12]. Although higher glucose-containing beverages (> 6% carbohydrate) slow gastric emptying when compared to water [8, 9, 12], the active co-transport of two sodium molecules for every glucose molecule accelerates water absorption along the small intestine when compared to passive water absorption [13]. Nevertheless, growing health concerns over sugar sweetened beverages [14] have led to the wide-spread sale and consumption of sugar-free alternatives.

Artificially sweetened sugar-free beverages maintain palatability, improve voluntary fluid consumption, and minimise dehydration as well as sugar-containing beverages [15]. However, in the absence of carbohydrate carriers, both artificially sweetened sugar-free beverages and water are absorbed more slowly across the intestine than dilute sugar-containing beverages [11]. The inclusion of artificial non-nutritive sweeteners has no impact on fluid absorption [16], and therefore, artificially sweetened sugar-free beverages may be sub-optimal in scenarios where rapid delivery of ingested fluid into the circulation is required. At least when compared to low-concentration glucose beverages [11, 12].

One alternative to the inclusion of carbohydrate within rehydration beverages is the use of amino acids. The small intestine has the capacity for large-scale absorption of amino acids, dipeptides, and tripeptides, which can enhance the absorption of sodium and water across the small intestine [3, 17]. However, the selection and concentration of amino acids are important for a few reasons; (1) sodium stoichiometry varies amongst amino acids; (2) certain amino acids compete for the same intestinal transporters; (3) the density of amino acid transporters varies along the length of the small intestine; and (4) transporter saturation kinetics vary by amino acid [18–21]. Consequently, the inclusion of strategically selected amino acids into artificially sweetened sugar-free beverages may augment water absorption across the intestine [18], facilitating delivery to the circulation.

Therefore, the aim of the present study was to investigate the effect of the addition of differing amounts of a novel sixamino acid formula to a commercially available sugar-free rehydration beverage on water absorption, assessed via gastrointestinal  $D_2O$  kinetics, and subsequent fluid balance markers. It was hypothesised that the addition of select amino acids would increase the rate of absorption of the sugar-free beverage, and the greater the amount of novel amino acid formula, the quicker the fluid absorption.

#### Methods

#### **Design of studies**

Three sequential studies assessing sugar-free rehydration beverages containing differing amounts of a novel amino acid formula were conducted. All studies received ethical approval from the Loughborough University Ethics Approvals (Human Participants) Sub-Committee (Study 1 ID: LEON3151; Study 2 ID: LEON1416; Study 3 ID: LEON3151-2859) and were registered with Clinical Trials (clinicaltrials.gov; Study 1 ID: NCT04819334; Study 2 ID: NCT04509388; Study 3 ID: NCT05698849). For Study 1 and 2, the amino acid beverages were compared to two commercially available sugar-free beverages, Powerade Zero<sup>TM</sup>(PZ) and Gatorade Zero<sup>TM</sup> (GZ). For Study 3, the novel amino acid beverage was compared to a commercially available sugar-free rehydration beverage (PZ) and a commercially available 6% carbohydrate-electrolyte beverage (Gatorade Thirst Quencher<sup>TM</sup>; GTQ). In all three studies, subjects completed a screening visit, and three experimental trials commencing at the same time of day (standardised within subjects between 08:00 and 09:00) in a randomised order, separated by  $\geq 6$  days.

#### **Screening visit**

Before commencement of each study, subjects provided written informed consent, consent to publish, and completed a medical screening questionnaire. Subjects were healthy (according to a medical screening questionnaire), non-smokers and had no known history of cardiovascular, metabolic, digestive, or renal disease. During the screening visit, body mass (AFW-120 K, Adam Equipment Co., Milton Keynes, UK) and height (Seca 216, Hamburg, Germany) were measured, whilst body fat was estimated using skinfold measurements (Harpenden Skinfold Caliper, HaB International Ltd., Southam, UK) at the biceps, triceps, sub-scapula and supra-iliac [22], and subjects self-reported their activity levels. All skinfold measurements were taken by the same accredited The International Society for the Advancement of Kinanthropometry (ISAK) anthropometrist. The subject characteristics for the three studies are displayed in Table 1. There was no control for menstrual cycle phase as ovarian hormones/menstrual cycle phase do not appear to affect gastric emptying [23] or hydration outcomes [24].

#### **Pre-trial standardisation**

In each study, subjects completed a diet and physical activity record for the 24 h preceding their first experimental trial

#### Table 1 Subject characteristics for Study 1, 2 and 3 (mean $\pm$ SD)

	Study 1			Study 2			Study 3		
	Males	Females	Group mean	Males	Females	Group mean	Males	Females	Group mean
Subject number	10	5	15	10	7	17	12	3	15
Age (y)	27 <u>+</u> 4	$25 \pm 1$	27±3	27±3	$25 \pm 1$	$26\pm3$	$29\pm5$	27±3	$28 \pm 4$
Height (m)	$1.81 \pm 0.08$	$1.63 \pm 0.06$	$1.75 \pm 0.11$	$1.83 \pm 0.08$	$1.64 \pm 0.06$	$1.75 \pm 0.12$	$1.79 \pm 0.07$	$1.64 \pm 0.10$	$1.76 \pm 0.09$
Body mass (kg)	$77.0 \pm 12.2$	$54.8 \pm 4.7$	69.6±14.8	$80.2 \pm 12.0$	$59.8 \pm 8.0$	71.8±14.5	$81.2 \pm 11.1$	$56.3 \pm 5.2$	$76.2 \pm 14.4$
BMI (kg/m <sup>2</sup> )	$23.5 \pm 3.0$	$20.7 \pm 1.7$	$22.6 \pm 3.0$	$23.8 \pm 2.8$	$22.3 \pm 3.0$	$23.2 \pm 2.9$	$25.2 \pm 2.5$	$20.9 \pm 0.7$	24.4±2.9
Sum of 4-site skinfolds (mm)	$29.7 \pm 9.2$	$36.8 \pm 10.1$	$32.1 \pm 9.8$	36.1±15.1	$48.6 \pm 17.8$	41.3±16.9	38.6±18.4	$36.5 \pm 10.8$	38.2±16.9
Estimated body fat (%)	$12.2 \pm 3.5$	$14.8 \pm 3.4$	$13.0 \pm 3.6$	$14.0 \pm 5.0$	17.8±4.8	15.6±5.1	$14.7 \pm 5.0$	14.7 ± 3.8	14.7 <u>+</u> 4.7
Activity level									
Training sessions per week	4±1	4±3	4±2	5±1	4±3	$5\pm 2$	$4\pm1$	5±3	4±2
Training volume (h/ week)	4±2	$5\pm 6$	$4\pm3$	$6\pm 2$	$5\pm3$	$6\pm 2$	$5\pm 2$	$5\pm4$	$5\pm 2$

Sum of 4-site skinfolds (mm) = biceps, triceps, sub-scapular and supra-iliac

and replicated these patterns before the second and third experimental trials. Adherence was verbally checked on arrival for trials. Strenuous exercise or alcohol intake were not permitted during this period. The day before trials, subjects were instructed to consume a minimum of 40 mL/kg body mass of fluid [25, 26]. This volume included any fluid, i.e. water, juice, coffee, tea, carbonated drinks, etc. Subjects stopped eating and drinking at least 10 h before arrival at the laboratory.

#### **Experimental trials**

Upon arrival at the laboratory, subjects voided their bladder into a plastic container, before nude body mass was recorded, and a flexible 20-gauge cannula was inserted into an antecubital/forearm vein for subsequent blood sampling. Subjects sat on a treatment bed with their legs flat on the bed and the backrest raised at ~  $55^{\circ}$  (i.e. a semi-upright Fowler's position). After 30 min, a baseline blood sample was taken. All blood samples were ~7.5 mL, and immediately, following each sample, the cannula was flushed with ~7.5 mL isotonic sterile saline (BD Biosciences, New Jersey, USA). A 550 mL (Study 1) or 500 mL bolus (Study 2 and 3) of the experimental beverage was then given, containing 3.000 g of deuterated water (Deuterium Oxide 99.9 atom % D, Sigma-Aldrich, St. Louis, USA), followed by a further 50 mL of the experimental beverage, which was used to swill around the drink vessel to ensure all deuterium oxide was ingested. Subjects were instructed to consume the beverage as quickly as possible, but to prioritise not spilling any. Subjects remained on the treatment bed in the semi-upright Fowler's position for a further 60 min; the timer began at the commencement of drinking. Additional ~7.5 mL blood samples were taken at 2, 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55 and 60 min. A second urine sample was collected after the final blood sample. Ambient temperature and relative humidity (Kestrel 4400, Nielsen-Kellerman Co., Philadelphia, USA) were recorded at 0, 30 and 60 min.

#### **Experimental beverages and blinding**

Experimental beverages were administered in a double-blind manner, prepared by an investigator not involved in the data collection or analysis, and served in an opaque bottle. The composition of beverages for the three studies is detailed in Table 2. Protein was calculated from the sum of elemental amino acid gram weights (molecular weight x mM), which included in descending order by concentration, aspartic acid, serine, valine, isoleucine, threonine, and tyrosine. The proprietary amino acid ratios were held constant when adjusting mM and beverage gram weights up or down. The energy density was also estimated from the energy equivalent for whole proteins, allowing for small errors [27].

#### Sample analysis

From each ~7.5 mL blood sample, ~1 mL was dispensed into a tube containing  $K_2$  EDTA (1.75 mg/mL; Teklab, Durham, UK). This was used to determine haemoglobin concentration and haematocrit via the cyanmethemoglobin method and microcentrifugation, respectively. These values were used to estimate changes in plasma volume relative to baseline [28]. These data were collected, and plasma volume estimated, at 10 timepoints in Study 1 (0, 2, 5, 10, 15, 20, 25, 30, 45, 60 min), 14 timepoints in Study 2 (0, 2, 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60 min), and 5 timepoints in Study 3 (0, Table 2Composition of sugar-<br/>free rehydration beverages<br/>containing differing amounts<br/>of a novel amino acid formula<br/>(AA + PZ) and commercially<br/>available control beverages (PZ,<br/>GZ and GTQ) for Study 1, 2<br/>and 3

	AA+PZ			Control Beverages		
	1	2	3	PZ	GZ	GTQ
Study inclusion	1	2	3	1, 2, 3	1, 2	3
Energy (kJ/L)	292	116	58	0	0	1002
Protein (g/L)	17.51	6.96	3.48	0	0	0
Carbohydrate (g/L)	0	0	0	0	0	60
Fat (g/L)	0	0	0	0	0	0
Sodium (mmol/L)	$18 \pm 1$	$19\pm0$	$18 \pm 1$	$18 \pm 1$	$19 \pm 1$	$19 \pm 1$
Potassium (mmol/L)	$2\pm0$	$2\pm0$	$2\pm 0$	$2\pm 0$	$3\pm0$	$3\pm0$
Chloride (mmol/L)	$20 \pm 1$	19 <u>±</u> 1	$18 \pm 1$	19 <u>±</u> 1	$9\pm0$	12 <u>+</u> 1
Osmolality (mOsm/kg H <sub>2</sub> O)	$209 \pm 2$	$109 \pm 2$	$87 \pm 3$	$61 \pm 3$	$49 \pm 1$	$343 \pm 33$

AA + PZ = a sugar-free rehydration beverage (PZ) containing differing amounts of a novel amino acid formula. PZ=Powerade Zero<sup>TM</sup>. GZ=Gatorade Zero<sup>TM</sup>. GTQ=Gatorade Thirst Quencher<sup>TM</sup>. Energy and macronutrient composition obtained from manufacturer information. Protein and energy density equivalents calculated from the sum of elemental amino acid gram weights (i.e. not whole protein). Sodium and potassium concentrations were analysed via flame photometry (M410C Flame Photometer, Sherwood Ltd., Cambridge, UK). Chloride concentrations were analysed via a chloride meter (M926S Chloride Analyser, Sherwood Ltd., Cambridge, UK). Osmolality was analysed via freezing-point depression (Gonotec Osmomat 030 Cryoscopic Osmometer, Gonotec, Berlin, Germany)

5, 15, 30, 60 min). For consistency, plasma volume is displayed at 5 timepoints for each of the three studies (Fig. 3). The reduction of plasma volume data from 10 and 14 timepoints to 5 timepoints in Study 1 and 2, respectively, did not alter the statistical outcomes/findings. From the remaining ~ 6.5 mL of whole blood, ~ 5 mL was dispensed into a second tube containing K<sub>2</sub> EDTA (1.6 mg/mL; Sarstedt AG & Co., Nümbrecht, Germany), and ~ 1.3 mL was dispensed into a tube containing lithium heparin (0.25 mg/mL; Sarstedt AG & Co., Nümbrecht, Germany). Plasma was separated from both tubes by centrifugation (2500 g, 20 min, 4 °C) and frozen (-80 °C) for subsequent analysis. Plasma samples used for D<sub>2</sub>O enrichment analysis were stored in glass vials.

Freezing-point depression (Gonotec Osmomat 030 Cryoscopic Osmometer; Gonotec, Berlin, Germany) was used to determine the osmolality of plasma from lithium heparin tubes. Urine specific gravity of both baseline and 60 min urine samples was measured on the day of trials (PAL-10S, Digital Urine Specific Gravity Refractometer, Atago Co. Ltd., Tokyo, Japan).

Plasma  $D_2O$  enrichment was determined in duplicate using the Europa Scientific ANCA-GSL sample preparation unit and 20–20 isotope ratio mass spectrometry (Sercon Ltd., Cheshire, UK). In brief, an appropriate sample volume was pipetted into Exetainer tubes and an insert vial containing 5% platinum on alumina was added. The tubes were sealed and subsequently filled with pure hydrogen. Samples were left for an equilibration period, during which the isotopes in the solution exchanged with the hydrogen gas in the headspace. A sample of the headspace gas was then analysed by continuous-flow isotope ratio mass spectrometry. The isotopic enrichment data are expressed as  $\delta\%_0$  against the international water standard Vienna Standard Mean Ocean Water (VSMOW). The CV of this measurement was 0.23%. Plasma  $D_2O$  enrichment area under the curve (AUC<sub>60</sub>) was calculated, and the maximal plasma  $D_2O$  enrichment concentration observed at any measured time point (Cmax) and the time Cmax occurred (Tmax) were derived [29].

Additional results are provided in the supplementary material for Study 1 and 2 (plasma amino acids [Supplementary Figs. 1 and 2], glucose [Supplementary Fig. 3], lactate [Supplementary Fig. 4], creatinine [Supplementary Fig. 5]), and Study 2 only (plasma sodium [Supplementary Fig. 6A] and potassium [Supplementary Fig. 6B], and urine D<sub>2</sub>O concentration). Plasma amino acid concentrations were determined at 0, 15, 30, 45 and 60 min using a Biochrom 30 + high-performance liquid chromatography ion exchange system (Biochrom, Cambourne, UK). Plasma glucose, lactate, and creatinine at 0, 15, 30, 45 and 60 min were determined via enzymatic colorimetric method (ABX Pentra C400, Horiba Medical, Northampton, UK). Plasma sodium and potassium were determined at 0, 2, 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55 and 60 min via flame photometry (M410C Flame Photometer, Sherwood Ltd., Cambridge, UK). Urine D<sub>2</sub>O concentration was determined via the method described above for plasma D<sub>2</sub>O enrichment; pre-trial urine D<sub>2</sub>O enrichment was subtracted from posttrial urine D<sub>2</sub>O enrichment to nullify any remaining D<sub>2</sub>O in the body water pool from previous experimental visits.

#### **Statistical analysis**

Data were initially checked for normality of distribution using a Shapiro–Wilk test. Data containing two factors (Trial\*Time) were initially analysed using two-way repeated measures analysis of variance (ANOVA) (SPSS version 27, SPSS Inc., Illinois, USA). Data containing one factor were initially analysed using one-way repeated measures ANOVA (normally distributed data) or Friedman's ANOVA (nonnormally distributed data). Where the assumption of sphericity was violated, the degrees of freedom were corrected using the Greenhouse-Geisser estimate. Significant ANOVA interaction (two-way ANOVA) and main (one-way ANOVA) effects were followed-up by post hoc paired t tests for normally distributed data, and Wilcoxon signed-rank tests for non-normally distributed data. The Holm-Bonferroni correction was applied to post hoc tests to control the family-wise error rate. A a-priori sample size estimation was performed using the data of Hill et al. [30] and Jeukendrup et al. [11], an  $\alpha$  of 0.05, and a statistical power of 0.80. It was estimated that 15 subjects would be required per study to reject the null hypothesis for D<sub>2</sub>O kinetic parameters (e.g. AUC). Statistical significance was accepted when P < 0.05. All data are displayed as mean  $\pm$  SD.

## Results

#### **Trial conditions**

No differences were present for ambient temperature or relative humidity between trials in each of the three studies ( $P \ge 0.130$ ; Table 3). There were no differences between

trials for pre-trial body mass, urine specific gravity (Table 3) or plasma osmolality ( $P \ge 0.168$ ; Fig. 2A–C), indicating subjects were in a similar hydration state at the beginning of trials in each of the three studies.

#### Plasma D<sub>2</sub>O enrichment

Study 1: There were main effects of time (P < 0.001), trial (P=0.001), and a trial by time interaction effect (P<0.001)for plasma D<sub>2</sub>O enrichment. Plasma D<sub>2</sub>O enrichment was lower at 15 and 20 min with AA + PZ compared to PZ  $(P \le 0.023)$ , and lower at 15, 20 and 25 min with AA+PZ compared to GZ ( $P \le 0.006$ ). There were no differences between PZ and GZ ( $P \ge 0.240$ ) for plasma D<sub>2</sub>O enrichment (Fig. 1A). There were main effects of trial for plasma  $D_2O$ enrichment total AUC<sub>60</sub> (P < 0.001) and Tmax (P = 0.005). Plasma D<sub>2</sub>O enrichment total AUC<sub>60</sub> was lower with AA + PZ compared to PZ (P = 0.002) and GZ (P = 0.006), but there was no difference between PZ and GZ (P = 0.553). Tmax was greater with AA + PZ compared to PZ (P = 0.027) and GZ (P = 0.044), but there was no difference between PZ and GZ (P = 0.849). There was no difference in Cmax amongst trials (P = 0.612; Table 4).

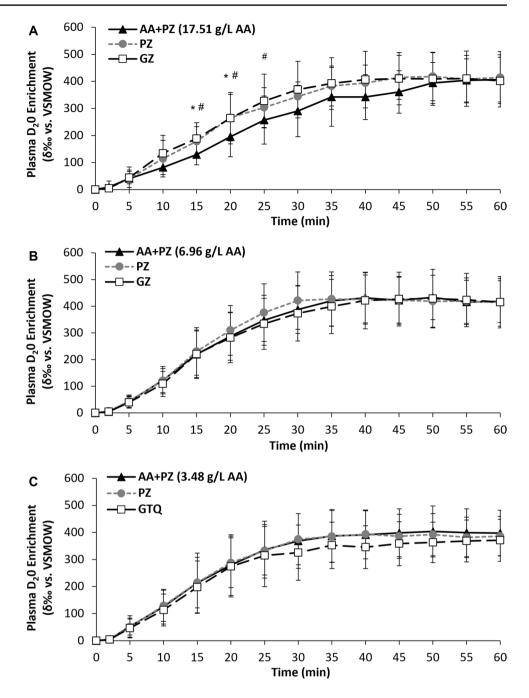
Study 2: There was a main effect of time (P < 0.001), with an initial increase in plasma D<sub>2</sub>O enrichment until ~ 35 min before reaching a plateau. There were no trial or trial by time interaction effects for plasma D<sub>2</sub>O enrichment ( $P \ge 0.252$ ; Fig. 1B). There were no differences in plasma D<sub>2</sub>O characteristics amongst trials ( $P \ge 0.338$ ; Table 4).

Table 3Mean ambienttemperature, mean relativehumidity, pre-trial bodymass and pre-trial urinespecific gravity for the threeexperimental trials for Study 1,2 and 3

	AA+PZ	PZ	GZ	GTQ	p value (ANOVA)
Study 1	17.51 g/L AA				
Ambient temperature (°C)	$23.2 \pm 0.9$	$23.2 \pm 0.9$	$23.3 \pm 0.8$	_	0.967
Relative humidity (%)	$32.2 \pm 6.2$	$34.4 \pm 6.9$	$32.9 \pm 6.3$	_	0.938
Pre-trial body mass (kg)	$68.3 \pm 14.7$	$68.3 \pm 14.8$	$68.5 \pm 14.8$	_	0.191
Pre-trial USG	$1.022 \pm 0.005$	$1.021 \pm 0.006$	$1.022 \pm 0.004$	_	0.863
Study 2	6.96 g/L AA				
Ambient temperature (°C)	$24.6 \pm 0.6$	$24.6\pm0.5$	$24.3 \pm 0.7$	_	0.751
Relative humidity (%)	$49.2 \pm 9.1$	$47.3 \pm 8.7$	$49.5 \pm 9.3$	_	0.130
Pre-trial body mass (kg)	$70.3 \pm 14.2$	$70.5 \pm 14.1$	$70.4 \pm 14.0$	_	0.509
Pre-trial USG	$1.021 \pm 0.006$	$1.021\pm0.005$	$1.022 \pm 0.005$	_	0.985
Study 3	3.48 g/L AA				
Ambient temperature (°C)	$23.0 \pm 1.2$	$22.8\pm0.9$	_	$22.8 \pm 1.3$	0.186
Relative humidity (%)	$40.1 \pm 14.1$	$44.8 \pm 10.9$	-	$42.7 \pm 13.9$	0.442
Pre-trial body mass (kg)	$75.6 \pm 14.5$	$75.6 \pm 14.5$	_	$75.5 \pm 14.3$	0.675
Pre-trial USG	$1.019\pm0.005$	$1.019\pm0.005$	-	$1.020\pm0.004$	0.746

Bold italic indicates amino acid concentration of AA-PZ of that study

Data are mean  $\pm$  SD. USG=urine specific gravity. AA=amino acids. AA+PZ=a sugar-free rehydration beverage (PZ) containing differing amounts of a novel amino acid formula. PZ=Powerade Zero<sup>TM</sup>. GZ=Gatorade Zero<sup>TM</sup>. GTQ=Gatorade Thirst Quencher<sup>TM</sup>. Ambient temperature and relative humidity data are mean of 0, 30 and 60 min time-points collapsed together Fig. 1 Plasma D<sub>2</sub>O enrichment ( $\delta\%$  vs. VSMOW) over time after ingesting the three expeirmental beverages for Study 1 (A), Study 2 (B) and Study 3 (C). \*=AA+PZ significantly different to PZ. #=AA+PZ significnatly different to GZ. AA+PZ=a sugar-free rehydration beverage (PZ) containing differing amounts of a novel amino acid formula. PZ=Powerade Zero<sup>TM</sup>. GZ=Gatorade Zero<sup>TM</sup>. GTQ=Gatorade Thirst Quencher<sup>TM</sup>



Study 3: There was a main effect of time (P < 0.001), with an initial increase in plasma D<sub>2</sub>O enrichment until ~ 35 min before reaching a plateau. There were no trial or trial by time interaction effects for plasma D<sub>2</sub>O enrichment ( $P \ge 0.108$ ; Fig. 1C). There were no differences amongst trials for total AUC<sub>60</sub> or Tmax ( $P \ge 0.114$ ). There was a main effect of trial for Cmax (P = 0.030), which was greater with AA + PZ compared to GTQ (P = 0.046), but not PZ (P = 0.498). There was no difference in Cmax between GTQ and PZ (P = 0.106; Table 4).

## Plasma osmolality

Study 1: There was a main effect of time (P < 0.001) for plasma osmolalility, with an initial decrease in plasma osmolality until ~ 35 min before reaching a plateau. There was a trial by time interaction effect (P < 0.001), but no effect of trial (P = 0.142) for plasma osmolality (Fig. 2A). Post hoc tests revealed no significant differences between trials after correction for multiple comparisons ( $P \ge 0.137$ ).

Table 4Plasma  $D_2O$ enrichment characteristicsfor the three experimentalbeverages for Study 1, 2 and 3

	AA+PZ	PZ	GZ	GTQ	<i>p</i> value (ANOVA)
Study 1	17.51 g/L AA				
AUC <sub>60</sub> (δ% vs. VSMOW)	$15,184 \pm 3532*$	$17,328 \pm 3153$	$17,\!749 \pm 4204$	-	< 0.001
Cmax (δ% vs. VSMOW)	$434 \pm 91$	$447 \pm 86$	$453 \pm 110$	-	0.612
Tmax (min)	51±11*	$45 \pm 12$	$45 \pm 12$	-	0.005
Study 2	6.96 g/L AA				
AUC <sub>60</sub> (δ% vs. VSMOW)	$18,655 \pm 3541$	$19,063 \pm 4308$	$18,\!290\pm\!4355$	-	0.584
Cmax (δ% vs. VSMOW)	$469 \pm 80$	$476 \pm 110$	$458 \pm 104$	-	0.573
Tmax (min)	$41 \pm 13$	$41 \pm 11$	$45 \pm 11$	-	0.338
Study 3	3.48 g/L AA				
AUC <sub>60</sub> (δ% vs. VSMOW)	$17,731 \pm 4101$	$17,564 \pm 3732$	-	16,195±4209	0.114
Cmax (δ% vs. VSMOW)	$440 \pm 94^{\#}$	$429 \pm 83$	_	398±81	0.030
Tmax (min)	$45 \pm 15$	$40 \pm 14$	-	$45 \pm 12$	0.206

Bold italic indicates amino acid concentration of AA-PZ of that study

Data are mean  $\pm$  SD. AUC<sub>60</sub>=total area under the curve for 60 min. Cmax=maximal concentration of plasma D<sub>2</sub>O enrichment. Tmax=the time which Cmax occurred. AA=amino acids. AA+PZ=a sugarfree rehydration beverage (PZ) containing differing amounts of a novel amino acid formula. PZ=Powerade Zero<sup>TM</sup>. GZ=Gatorade Zero<sup>TM</sup>. GTQ=Gatorade Thirst Quencher<sup>TM</sup>. \*=AA+PZ significantly different to PZ and GZ. <sup>#</sup>=AA+PZ significantly different to GTQ

Study 2: There was a main effect of time (P < 0.001) for plasma osmolalility, with an initial decrease in plasma osmolality until ~ 35 min before reaching a plateau. There were no trial or trial by time interaction effects for plasma osmolality ( $P \ge 0.228$ ; Fig. 2B).

Study 3: There were main effects for time (P < 0.001), trial (P = 0.022), and a trial by time interaction effect (P = 0.007) for plasma osmolality (Fig. 2C). Plasma osmolality was greater ( $P \le 0.024$ ) at 30 min and 50 min after consumption of GTQ compared to PZ, post hoc tests revealed no further differences between trials ( $P \ge 0.076$ ). After consumption of AA + PZ, plasma osmolality was not different from baseline for the first 15 min ( $P \ge 0.153$ ), but was significantly lower than baseline from 20 min onwards ( $P \le 0.032$ ). After consumption of PZ, plasma osmolality was not different from baseline for the first 25 min ( $P \ge 0.096$ ), but was significantly lower than baseline from 30 min onwards ( $P \le 0.039$ ). After consumption of GTQ, plasma osmolality was not different to baseline at any time point ( $P \ge 0.192$ ).

#### **Plasma volume**

Study 1: There was a main effect of time (P < 0.001) for change in plasma volume; with plasma volume significantly lower than baseline at 5 and 15 min ( $P \le 0.001$ ), but not different to baseline at 30 and 60 min ( $P \ge 0.163$ ). There were no trial (P = 0.102) or trial by time interaction effects (P = 0.124) for change in plasma volume (Fig. 3A).

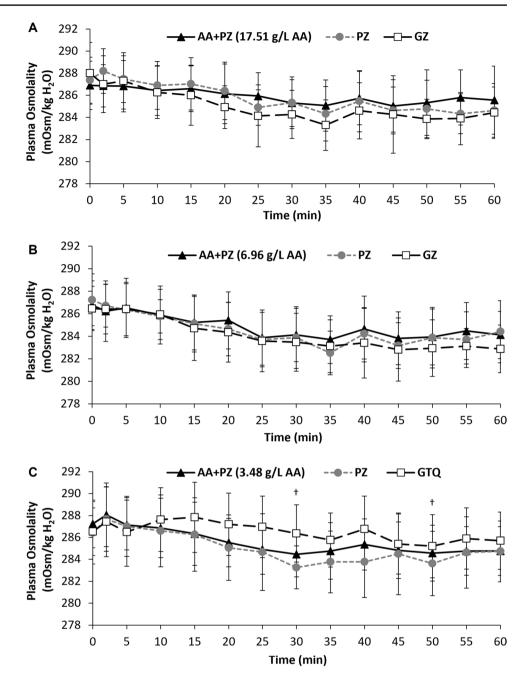
Study 2: There was a main effect of time (P < 0.001) for change in plasma volume; with plasma volume initially decreasing below baseline at 5 and 15 min ( $P \le 0.001$ ),

before increasing above baseline at 30 and 60 min  $(P \le 0.029)$ . There were no trial (P = 0.468) or time by trial interaction effects (P = 0.474) for change in plasma volume (Fig. 3B).

Study 3: There were main effects for time (P < 0.001), trial (P = 0.028) and a trial by time interaction effect (P = 0.002) for change in plasma volume (Fig. 3C). Plasma volume was greater at 30 min after consumption of GTQ compared to PZ and AA + PZ ( $P \le 0.045$ ), with no further differences between trials ( $P \ge 0.123$ ). Plasma volume was not different from baseline at any time point after consumption of AA + PZ ( $P \ge 0.074$ ). After consumption of PZ, plasma volume was significantly lower than baseline at 5 and 15 min ( $P \le 0.019$ ), but was not different to baseline at 30 and 60 min ( $P \ge 0.176$ ). After consumption of GTQ, plasma volume was not different to baseline at 5 and 15 min ( $P \ge 0.470$ ), but was greater than baseline at 30 and 60 min ( $P \ge 0.036$ ).

#### Urine specific gravity and urine volume

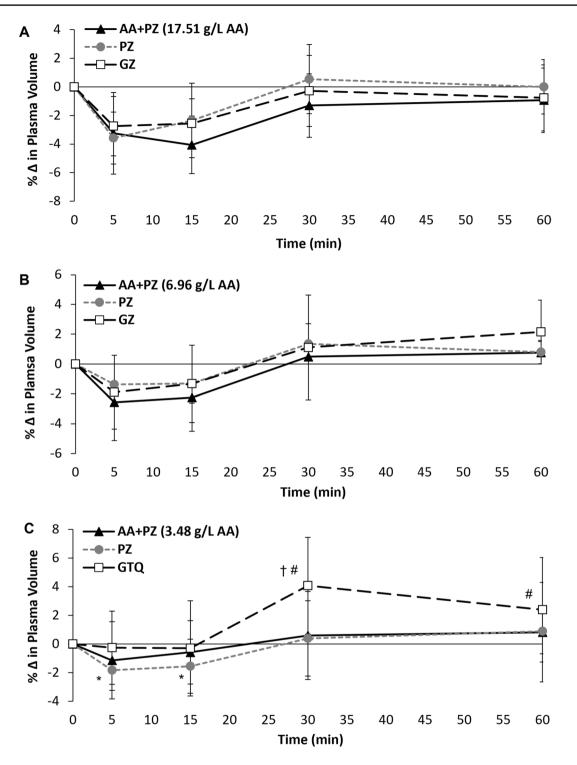
No differences were present between trials for post-trial urine specific gravity in any of the three studies ( $P \ge 0.406$ ; Table 5). No differences were present between trials in Study 1 and 2 for post-trial urine volume ( $P \ge 0.841$ ). Posttrial urine volume was significantly different between trials in Study 3 (P = 0.033; Table 5), with lower volume after consumption of GTQ compared to PZ (P = 0.046), but not AA + PZ (P = 0.149). There was no difference in post-trial urine volume between AA + PZ and PZ (P > 0.999). Fig. 2 Plasma osmolality (mOsm/kg H<sub>2</sub>O) over time after ingesting the three experimental beverages for Study 1 (A), Study 2 (B) and Study 3 (C).  $\dagger$ =GTQ significantly different from PZ. AA + PZ = a sugar-free rehydration beverage (PZ) containing differing amounts of a novel amino acid formula. PZ=Powerade Zero<sup>TM</sup>. GZ=Gatorade Zero<sup>TM</sup>. GTQ=Gatorade Thirst Quencher<sup>TM</sup>



## Discussion

Three studies were conducted to assess the addition of differing amounts of a novel amino acid formula to a sugar-free rehydration beverage on fluid absorption. Fluid absorption was assessed via gastrointestinal  $D_2O$  kinetics, and the effect on subsequent fluid markers was measured. It was hypothesised that the rate of absorption of a sugar-free rehydration beverage would increase as the amount of amino acids added to the beverage increased. However, opposed to the hypothesis, the addition of greater amounts of the amino acid mixture (17.51 g/L) to a sugar-free rehydration beverage delayed water delivery to the circulation and neither lower concentration amino acid beverage demonstrated differences in water uptake kinetics compared to the sugar-free beverages. In contrast, when compared to a 6% carbohydrate beverage (GTQ), the addition of a smaller amount of amino acids (3.48 g/L) to a sugar-free rehydration beverage increased fluid delivery, evidenced by a greater maximal plasma  $D_2O$  enrichment concentration.

Water-soluble organic molecules, which include certain amino acids, dipeptides and tripeptides, when absorbed from the small intestine enhance the absorption of electrolytes and water [20, 31-33]. The aim of Study 1 was to provide



**Fig. 3** Change in plasma volume (%) relative to 0 min after ingesting the three experimental beverages for Study 1 (**A**), Study 2 (**B**) and Study 3 (**C**).  $\dagger$ =GTQ significantly different from AA+PZ and PZ. \*=time point significantly different from 0 min within PZ trial.

#=time point significantly different from 0 min within GTQ trial. AA+PZ=a sugar-free rehydration beverage (PZ) containing differing amounts of a novel amino acid formula. PZ=Powerade Zero<sup>TM</sup>. GZ=Gatorade Zero<sup>TM</sup>. GTQ=Gatorade Thirst Quencher<sup>TM</sup>

as many grams of the amino acid formula as possible within the sugar-free rehydration beverage; based upon solubility and flavour, 17.51 g/L of amino acid formula was delivered in a 600 mL bolus (i.e. a total of 10.5 g of amino acids). The delayed time of maximal plasma  $D_2O$  enrichment concentration, and lower plasma  $D_2O$  enrichment AUC, demonstrate

	AA+PZ	PZ	GZ	GTQ	p value (ANOVA)
Study 1	17.51 g/L AA				
Urine volume (mL)	$342 \pm 109$	$355 \pm 128$	$332 \pm 98$	_	0.841
USG	$1.006 \pm 0.003$	$1.005 \pm 0.002$	$1.005 \pm 0.002$	_	0.676
Study 2	6.96 g/L AA				
Urine volume (mL)	$251 \pm 116$	$256 \pm 125$	$260 \pm 113$	_	0.954
USG	$1.009 \pm 0.006$	$1.008 \pm 0.004$	$1.009 \pm 0.005$	_	0.901
Study 3	3.48 g/L AA				
Urine volume (mL)	$335 \pm 134$	$336 \pm 110$	_	$268 \pm 79*$	0.033
USG	$1.005 \pm 0.003$	$1.005 \pm 0.002$	-	$1.006 \pm 0.003$	0.406

Bold italic indicates amino acid concentration of AA-PZ of that study

Data are mean $\pm$ SD. USG=urine specific gravity. AA=amino acids. AA+PZ=a sugar-free rehydration beverage (PZ) containing differing amounts of a novel amino acid formula. PZ=Powerade Zero<sup>TM</sup>. GZ=Gatorade Zero<sup>TM</sup>. GTQ=Gatorade Thirst Quencher<sup>TM</sup>. \*=GTQ significantly lower than PZ. \*=GTQ significantly lower than PZ

a delay in water delivery into the circulation after consumption of the amino acid beverage compared to two commercially available sugar-free rehydration beverages.

The use of a D<sub>2</sub>O tracer is an integrated measure of gastric emptying and intestinal absorption. Therefore, the delay in D<sub>2</sub>O appearance in the circulation after consumption of the novel amino acid beverage was a result of delayed gastric emptying, and/or intestinal amino acid transporter saturation, or a combination of both [1, 20]. The greater energy density, and potentially osmolality, of the novel amino acid beverage may have delayed gastric emptying [1, 34], impeding fluid delivery to the proximal intestine and consequently circulation. Once emptied from the stomach, fluids delivered to the duodenum are quickly brought into osmotic equilibrium with the circulating plasma, and the jejunum is relatively permeable to electrolytes and water [1]. Therefore, although the initial beverage was hypotonic  $(209 \pm 2 \text{ mOsm}/$ kg  $H_2O$ ), if a portion of the water and electrolytes from the beverage were rapidly absorbed in the proximal intestine, a high concentration of amino acids would remain in the small intestine. This would raise the osmolality of the small intestine above that of the plasma [35], resulting in 'osmotic backflow' (fluid osmotically moving from the extracellular fluid to the intestinal lumen [20], potentially negating any beneficial effects of increased absorption induced by amino acid transport along the intestine [1, 2, 35]. Although there is limited data on beverage amino acid content and gastrointestinal D<sub>2</sub>O kinetics, similar inferior D<sub>2</sub>O absorption kinetics have been observed with higher glucose-containing beverages (>6% carbohydrate) compared to more dilute carbohydrate beverages and water [8, 9, 11, 12, 35].

For Study 2, a smaller amount of amino acid formula (6.96 g/L) within the sugar-free rehydration beverage was delivered in a marginally smaller 550 mL bolus (i.e. a total of 3.83 g of amino acids). The addition of 6.96 g/L of

amino acids into a rehydration beverage resulted in comparable water delivery into the circulation compared to two commercially available sugar-free rehydration beverages. Receptors in the duodenum and ilium are sensitive to macronutrient content, pH, and osmotic pressure [1, 36], and the activity of these receptors can delay gastric emptying by initiating hormonal and neural responses that alter gastric and duodenal muscular contraction [1]. Therefore, increasing macronutrient content and energy density of a beverage can delay gastric emptying [36, 37]. One hypothesis is that the inclusion of amino acids in the rehydration beverage may have delayed gastric emptying whilst concurrently increasing net sodium and water transport across the intestine [1]. Therefore, the beneficial effect of increased intestinal absorption may have been negated by a decreased gastric emptying rate. On the contrary, a second hypothesis is that, due to the relatively low amino acid content of the beverage, gastric emptying may have been similar between beverages, resulting in a comparable amount of fluid rapidly absorbed in the duodenum, and subsequently similar fluid delivery between beverages. However, with limited research on amino acid containing beverages and fluid delivery, it is difficult to fully ascertain.

Nevertheless, due to the delayed or equivalent water delivery of the novel amino acid beverages in Study 1 and 2, respectively, the amount of amino acid formula within the sugar-free rehydration beverage was further reduced to 3.48 g/L in a 550 mL bolus (i.e. a total of 1.91 g of amino acids). Additionally, the results from Study 1 and 2 demonstrate that both sugar-free rehydration beverages (PZ and GZ) have similar gastrointestinal D<sub>2</sub>O kinetics, a commercially available 6% carbohydrate–electrolyte beverage (GTQ) and a sugar-free rehydration beverage (PZ) were used for comparison in Study 3. The addition of a smaller amount of novel amino acid formula to a sugar-free rehydration beverage increased fluid delivery compared to the commercially available carbohydrate–electrolyte beverage, evidenced by greater maximal plasma  $D_2O$  enrichment. However, there were no differences in  $D_2O$  delivery into the circulation between the sugar-free rehydration beverage and amino acid beverage.

The small intestine has the capacity for fast and largescale absorption of amino acids, dipeptides, and tripeptides, which can enhance the absorption of sodium and water across the small intestine [3, 17]. The low concentration of amino acids within the beverage likely emptied rapidly from the stomach and increased water and sodium transport across the intestine, resulting in greater water delivery compared to the carbohydrate–electrolyte beverage [18–20]. However, there may also have been inhibitory feedback from duodenal osmoreceptors and the glucose-sodium cotransporter (SGLT1) in the jejunal epithelium (to prevent the absorptive capacity of the proximal intestine becoming overwhelmed) that could have reduced gastric emptying after consumption of the carbohydrate-electrolyte beverage [12]. The greater maximal plasma D2O enrichment with the amino acid beverage, but not the sugar-free rehydration beverage, compared to the carbohydrate-electrolyte beverage suggests the amino acids may have marginally accelerated water delivery and warrants further investigation.

Plasma  $D_2O$  enrichment does not reflect absorption per se, as it requires extra- and intra-cellular volumes to remain constant during the sampling period [11]. This was likely not the case, for example in Study 3, an expansion in plasma volume occurred 30–60 min after consumption of the 6% carbohydrate–electrolyte beverage. If plasma volume expanded, an increase in fluid absorption from the intestine could have occurred without a parallel increase in plasma  $D_2O$  enrichment [11]. Due to potential alterations in extraand intra-cellular volumes, the  $D_2O$  uptake results should be treated with caution.

The lower post-trial urine output following consumption of the 6% carbohydrate-electrolyte beverage in Study 3 indicates greater beverage retention. Plasma osmolality influences circulating arginine vasopressin concentrations, and arginine vasopressin concentrations are responsible for the re-absorption of water in the kidney and thus urine production [35, 38]. Lessening urine production is pivotal in maximising rehydration beverage retention [39], and this occurs by minimising the reduction in plasma osmolality and associated circulating arginine vasopressin concentrations following beverage consumption [4, 40, 41]. Plasma osmolality did not differ from baseline after consumption of the carbohydrate-electrolyte beverage, whereas plasma osmolality decreased below baseline following consumption of the sugar-free rehydration beverage (with or without amino acids). Therefore, the greater plasma osmolality and expected greater associated circulating arginine vasopressin

concentrations, after consumption of the 6% carbohydrate beverage was likely responsible for the lower urine output [41, 42]. Given the similarity in electrolyte composition of the beverages used across the studies, these effects are likely directly attributable to the differences in carbohydrate content [42]. Interestingly, previous studies [43, 44] suggest larger carbohydrate concentrations (> 10%) are required to decrease post-ingestion urine output, in contrast to the present findings. It could be that the lack of a difference in these previous studies is explained by the larger drink volume (1000 mL vs. 550 mL in Study 3) causing a volumeinduced diuresis that masked the more subtle effects of lower carbohydrate contents.

The greater plasma osmolality and potentially decreased urine output, following consumption of the 6% carbohydrate–electrolyte beverage was likely responsible for the observed increase in plasma volume at 30 and 60 min. However, the greater plasma volume at 30 min post-consumption of the 6% carbohydrate–electrolyte beverage was unlikely the result of increased gastrointestinal absorption of the beverage as the D<sub>2</sub>O enrichment characteristics were not superior to the other beverages. Therefore, the increase in plasma volume likely derived from movement of interstitial or intracellular fluid augmented by the higher plasma osmolality, and not from the beverage itself.

## Conclusion

In conclusion, the addition of a small amount of a novel amino acid formula (3.48 g/L) to a sugar-free rehydration beverage increased water delivery into the circulation compared to a 6% carbohydrate-containing rehydration beverage. However, the addition of greater amounts of amino acids (17.51 g/L) to a sugar-free rehydration beverage delayed fluid delivery, potentially due to delayed gastric emptying and/or intestinal transporter saturation.

Future research should assess a sugar-free rehydration beverage containing a small amount of a novel amino acid formula in scenarios where rapid delivery of water into the circulation is required (i.e. illness, heat stress or exercise). The gastrointestinal water uptake kinetics of differing amalgams of amino acids, dipeptides, and tripeptides, within rehydration solutions, with the aim to further increase the rate of fluid delivery, should also be investigated. Additionally, age-related differences in fluid retention [42], potentially due to changes in renal function, and/or fluid absorption kinetics [45], mean it would be prudent to confirm the current findings in differing populations (i.e. older adults).

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s00394-024-03325-x. Author contributions Authors significantly contributed to this study in the following areas. Study conceptualisation and design were contributed by MPF, LJJ, RWK, SNC. Data collection and analysis were contributed by MPF, LAJ, KMR, DAJ, RMJ, LJJ, SAM. Drafting the manuscript was contributed by MPF, LJJ. All authors read and approved the final manuscript.

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**Data availability** Data is available upon reasonable request to the corresponding author.

## Declarations

**Conflict of interest** LJJ has current funding from Entrinsic Beverage Company LLC, Entrinsic Bioscience, LLC, Herbalife Europe Ltd, Bridge Farm Nurseries and Decathlon SA, and has previously received funding from PepsiCo Inc., Volac International and British Summer Fruits; has performed consultancy for PepsiCo Inc. and Lucozade, Ribena Suntory, and has received conference fees from PepsiCo Inc. and Danone Nutricia. In all cases, monies have been paid to LJJs institution and not to LJJ. The funders were involved in the design of the study, but were not involved in the collection, analyses, or interpretation of data, in the writing of the manuscript; or in the decision to publish the results. RWK and SNC are employees of Entrinsic Bioscience, LLC, who funded the study. They were involved in the study design and have read and agreed the final manuscript.

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