ORIGINAL CONTRIBUTION



Associations of long chain polyunsaturated fatty acids with bone mineral density and bone turnover in postmenopausal women

Orlagh Feehan¹ • Pamela Jane Magee¹ • Laura Kirsty Pourshahidi¹ • David John Armstrong^{1,2} • Mary Martina Slevin¹ • Philip James Allsopp¹ • Marie Catherine Conway¹ • J J Strain¹ • Emeir Mary McSorley¹

Received: 30 September 2021 / Accepted: 7 June 2022 / Published online: 30 July 2022 © The Author(s) 2022

Abstract

Purpose The immunomodulatory properties of n-3 long chain polyunsaturated fatty acids (LCPUFA) are reported to reduce bone loss through alteration of bone remodelling and n-3 LCPUFA, therefore, may benefit bone health in post-menopausal women, a vulnerable group at high risk of osteoporosis.

Methods Measures of bone mineral density (BMD) were determined using dual energy X-ray absorptiometry (DEXA) in 300 post-menopausal women. The bone turnover markers osteocalcin (OC), C-terminal telopeptides of type 1 collagen (CTX) and total alkaline phosphatase were quantified in serum along with urinary creatinine corrected deoxypyridinoline (DPD/Cr) and CTX/Cr and the CTX:OC ratio calculated. Total serum n-6 PUFA (LA + AA) and n – 3 LCPUFA (ALA + EPA + DPA + DHA) were measured and the n-6:n-3 ratio was calculated.

Results Mean (SD) age and body mass index (BMI) were 61 (6.4) years and 27.4 (4.8) kg/m², respectively with participants being 12.6 (7.6) years post-menopause. Multiple regression analysis identified no association between n-3 LCPUFA and any of the measures of T-score or BMD albeit a significant positive association between total n – 3 LCPUFA and femur BMD (β =0.287; p=0.043) was observed within those women with a low n – 6:n – 3 ratio. There was a significant inverse association between ALA and urinary DPD/Cr (β = –0.141; p=0.016).

Conclusion A favourable low n-6: n-3 ratio was associated with higher femur BMD and a higher n-3 LCPUFA (ALA) was associated with lower bone resorption. These results support a beneficial role for n-3 LCPUFA in reducing postmenopausal bone resorption and favourably influencing BMD.

Trial number & date of registration ISRCTN63118444, 2nd October 2009, "Retrospectively registered".

Keywords LCPUFA · Bone mineral density · Postmenopausal women · Bone resorption

Introduction

Osteoporosis is a chronic debilitating condition characterised by a decrease in bone mineral density (BMD) and altered bone microarchitecture, which can lead to structural fragility and a subsequent increased risk of fracture [1, 2]. Osteoporosis affects 3 million people in the United Kingdom (UK) and annually over 8.9 million osteoporotic fractures occur

 worldwide [3, 4]. Postmenopausal women are at increased risk of developing osteoporosis and associated fractures as a result of the natural decline of oestrogen after the menopause [5]. Declining oestrogen concentrations increase bone resorption by enhancing osteoclastogenesis and also reduce new bone formation activity by osteoblasts, causing a net loss of bone tissue [6]. Greater expression of pro-inflammatory cytokines as seen after menopause and in later age is one possible contributing factor in the development of osteoporosis. Hormones (such as oestrogen), pro- and anti-inflammatory cytokines play an important role in regulating osteoblast and osteoclast differentiation and activity. A balance of these systems is needed to prevent the development of osteoporosis [7, 8]. It has been postulated that n – 3 LCPUFA can up-regulate the absorption of calcium from



Nutrition Innovation Centre for Food and Health (NICHE), Ulster University, Coleraine BT52 1SA, UK

Department of Rheumatology, Altnagelvin Hospital, Western Health and Social Care Trust, Londonderry BT47 6SB, UK

the intestine and reduce the production of prostaglandin E2 (PGE2) which is a key eicosanoid involved in the modulation of inflammation [9]. Elevated concentrations of PGE_2 can lead to a reduction in the production of osteoprotegerin (OPG), an important osteoclastogenesis inhibitory factor expressed by osteoblasts. This production in turn can increase the expression of receptor activator of nuclear factor kappa-B ligand (RANKL) as a result of decreased OPG [10] leading to overall negative effects on bone health.

Nutrition is a modifiable factor important in optimising bone health [11] with well-defined roles for protein, calcium and vitamin D in reducing the risk of osteoporosis and fragility fractures post menopause [12]. Recent evidence suggests that n-3 long chain polyunsaturated fatty acids (LCPUFA) may benefit bone health and prevent bone loss [13–16]; however, there is limited evidence in postmenopausal women. Furthermore, the majority of research to date has focused on dietary intakes of n-3 LCPUFA rather than the biological status of n-3 LCPUFA in postmenopausal women. Dietary n – 3 LCPUFA have been proposed to benefit overall bone health [15] and have a role in reducing the risk of hip fracture in all age groups [17]. A higher intake of total n-3 LCPUFA and fish have been positively associated with BMD [17–19] and a reduced risk of hip fracture [20] as well as total fracture risk in postmenopausal women [11, 21]. Positive associations between n-3 LCPUFA status, dietary n-3 LCPUFA and skeletal outcome measures such as lumbar spine and femur BMD have been demonstrated in older people and in postmenopausal women and are postulated to be owing to higher fish consumption [13, 22]. Total dietary n-6 PUFA and the n-6:n-3 ratio have been shown to be negatively associated with spine and femur BMD in cohorts of older adults [13, 23] and postmenopausal women [24] albeit studies were limited by use of a nutrient database or dietary records to quantify dietary n-3 LCPUFA intake rather than status [23, 24]. Relative amounts of n-6 and n-3 LCPUFA should be taken into consideration for overall health, with research suggesting a lower n-6:n-3 ratio could be of greater benefit to bone health [23].

Recent analysis of the UK biobank including 492,713 adults showed a significant association between habitual fish oil supplement use and lower risk of both incident and recurrent fractures [25]. A limited number of supplementation studies with n-3 LCPUFA have been conducted in postmenopausal women with the majority reporting an inverse association between n-3 LCPUFA and markers of bone resorption including C-terminal telopeptides of type 1 collagen (CTX) and urinary pyridinoline (Pyd), and others reporting beneficial effects of n-3 supplementation on the bone formation markers bone-specific alkaline phosphatase (BALP) and osteocalcin (OC) in postmenopausal women [2, 26]. Although the reported changes were small in magnitude, they warrant further investigation particularly given

that inclusion of rich sources of n-3 LCPUFA, such as oily fish, in the diet could be a simple effective dietary alteration in these women [26]. To date, the evidence is limited by small sample sizes and short study durations and research is needed to further investigate this association between n-3 LCPUFA and BMD in post-menopausal women [27]. Therefore, the aim of this study was to investigate the relationship between serum n-3 LCPUFA status with BMD and BTMs in a large cohort of postmenopausal women.

Materials and methods

Study design

A convenient sample of a total of 300 non-osteoporotic postmenopausal women (45-75 years) were recruited to take part in a study conducted in Ulster University to investigate the effect of a combination of AquaminTM and Nutraflora® on bone health [28]. The current study utilises baseline data from this trial. Women were excluded if they were pre- or peri-menopausal, went through menopause earlier than 40 years old, used medication or supplements known to affect bone health, had osteoporosis or a medical condition known to affect bone health. Dietary intake of total daily fish (g/day) and lifestyle factors were determined by a 4-day semi-quantitative food diary and food frequency questionnaires. Food composition database Weighed Intake Software Package (WISP) (WISP for WINDOWS, version 3; Tinuviel Software, Anglesey, United Kingdom) was used to analyse the dietary intakes. Information on total physical activity was assessed by means of a shortened version of the International Physical Activity Questionnaire [29] and was represented as MET (metabolic equivalent tasks) hours per week. Participants also completed a questionnaire about past/current medical history. Ethical approval was received from Ulster University Research Ethics Committee (REC/08/0083) and the original study was registered as a clinical trial (www. controlled-trials.com as ISRCTN63118444). The study was conducted in accordance with the declaration of Helsinki and written informed consent was obtained from all participants.

Measurements

A non-fasted blood sample was collected from each participant. Duplicate weight (kg) and height (cm) were taken to calculate body mass index (BMI) (weight (kg)/height (m)²). Lumbar spine and femur BMD (g/cm²) and T-scores were determined using dual energy X-ray absorptiometry (DEXA) (Lunar corporation, Madison, WI). BMD was recorded from the total hip or the femoral neck depending on which was the lowest value. According to the cut-offs established by



the World Health organisation [30], osteoporosis is defined as BMD at least 2.5 SD below that of a healthy young adult (T-score is < -2.5), osteopenia when the T-score is between -1.0 and -2.5 and a normal bone density value above -1.0. BMD was expressed to 3 decimal places and T-scores to 1 decimal place according to the International Society for Clinical Densitometry [31]. Diagnosis was made according to the site with the lowest T-score.

Sample analysis

Blood and urinary bone turnover markers

Blood samples were collected from all participants for analysis. Serum bone formation marker osteocalcin (OC) (normal reference value: 26.5 (12.8-55.0 95% CI) ng/ml) and bone resorption marker C-terminal telopeptides of type 1 collagen (CTX) (normal reference value: 0.439 (0.142–1.351 95% CI) ng/ml) were measured using enzyme-linked immunosorbent assay (ELISA) (Immunodiagnostics Systems Ltd (IDS); intra-assay CV was 1.8% & 4.7%; respectively). Serum total alkaline phosphatase was quantified by Roche Modular Analyser and expressed as a bone formation marker (u/L). Urinary bone resorption markers CTX (normal reference range: 324 (121–874 95% CI) µg/mmol/Cr) and deoxypyridinoline (DPD) (7.56 (2.27)/7.94 (3.25) nmol DPD/mmol Cr) were measured using enzyme immunosorbent assay (EIA) (Urine CrossLaps EIA, IDS, intra-assay CV was 4.7% and Urine DPD EIA, Metra DPD EIA kit; Quidel, intra-assay CV was 4.3%). Urinary CTX and DPD results were both corrected for urinary concentrations of creatinine (Cr). CTX:OC ratio was calculated. A high CTX:OC ratio means higher bone resorption relative to bone formation.

Polyunsaturated fatty acid (LCPUFA) analysis

Serum LCPUFA were quantified using an adaption of the method by Folch et al. [32]. In brief, fatty acid methyl esters were detected and quantified by gas chromatography-mass spectrometry (7890A-5975C; Agilent) using heptadecanoic acid (C17:0) as the internal standard [33]. All analytic standards were of ≥99% purity and purchased from Sigma-Aldrich. Serum LCPUFA status was chosen as a biomarker to encompass recent LCPUFA concentrations of the triacylglycerol fraction. The individual n – 6 PUFAs; linoleic acid; 18:2 n - 6 (LA), arachidonic acid; 20:4 n - 6 (AA) and the individual n-3 LCPUFA, α -linolenic acid; 18:3 n-3(ALA), eicosapentaenoic acid; 20.5 n - 3 (EPA), docosapentaenoic acid; 23.5 n - 3 (DPA) and docosahexaenoic acid; 22:6 n - 3 (DHA) were quantified. Results were presented as milligrams per millilitre (mg/mL). Total n - 6:n - 3 ratio was calculated. A high n-6:n-3 ratio is considered less favourable as this means higher total n-6 PUFA relative to n-3 LCPUFA.

Statistical methods

All data analysis was performed using Statistical Package for the Social Sciences (SPSS, IBM SPSS Statistics version 25). Nominal data were presented as mean (SD) and categorical variables presented as frequencies (*n*) and percentages (%). Data were tested for normality using Kolmogorov–Smirnov statistic test and variables were transformed using the natural logarithm where appropriate. Pearson correlation analysis was conducted to test for correlation sample size.

Primary analysis assessing the associations between the natural logarithm of LCPUFA (total n-3, n-6, n-6:n-3ratio) with femur and lumbar spine BMD and T-score was conducted using Spearman correlation analysis. Separate multiple regression analysis was also performed between femur and lumbar spine BMD and T-score as a dependent variable and LCPUFA additionally adjusting for age, BMI, years' post-menopause, total physical activity and smoking status as independent variables. Secondary analysis was conducted to test for associations between the natural logarithm of LCPUFA (total n-3, n-6, n-6:n-3 ratio) and BTMs (OC, CTX, DPD/Cr, CTX/Cr, CTX:OC ratio) using the same statistical approach. Tertiles were created and participants were classified as having a high (≥ 6.5), medium (between 4.9 and 6.5) or low (\leq 4.9) n – 6:n – 3 ratio. Participants who were classified as having a low, medium or high n-6:n-3 ratio were selected as individual groups and regression analysis was repeated to take into consideration relative amounts of n-6 and n-3 LCPUFA and to determine if n-6 or n-3 LCPUFA was associated with bone parameters and BTMs, after adjusting for independent variables. To better satisfy regression assumptions, we used the natural logarithmic transformation of all markers in the models. As some of the T-scores were negative values, we added a constant of +3 to each individual T-score data prior to natural logarithmic transformation. All values presented in tables are log transformed. Significance was set at p < 0.05.

Results

Baseline characteristics

Table 1 summarises the characteristics for the 300 post-menopausal women. Data for BTMs and PUFA concentrations were available for 299 postmenopausal women. Mean (SD) age and BMI of subjects were 61 (6.4) years and 27.4 (4.8) kg/m², respectively. Mean values for years post-menopause and physical activity were 12.6 (7.6) years and 25.4 (24.3) MET hours/week. Age of menarche and menopause were



Table 1 Descriptive characteristics of postmenopausal women (n=300)

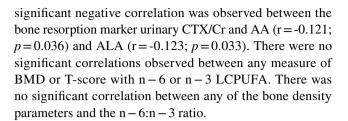
Descriptive	Mean (SD)
Age (years)	61.0 (6.4)
Height (cm)	160.6 (6.2)
Weight (kg)	70.7 (12.8)
Body mass index (kg/m ²)	27.4 (4.8)
Years post-menopause	12.6 (7.6)
Total physical activity (MET hours/week)	25.4 (24.3)
Femur BMD (g/cm ²)	0.895 (0.1)
Lumbar spine BMD (g/cm ²)	1.099 (0.1)
Femur T-score	-0.7(0.9)
Lumbar spine T-score	-0.6(1.2)
n Normal based on T-score (%)	127 (42.3)
n Osteopenia based on T-score (%)	173 (57.7)
BTMs	n = 299
Serum CTX (ng/mL)	0.63 (0.20)
Serum total alkaline Phosphatase (u/L)	77.3 (19.5)
Serum Osteocalcin (ng/mL)	19.68 (0.22)
Urinary CTX/Cr (ug/mmoL Cr)	309.2 (146.6)
Urinary DPD/Cr (nmol DPD/mmol Cr)	8.1 (2.3)
CTX:Osteocalcin ratio	3.4 (1.1)
LCPUFA	
Total $n-3 \text{ (mg/mL)}$	0.241 (0.110)
Total $n-6$ (mg/mL)	1.328 (0.375)
LA (C18) (mg/mL)	1.014 (0.304)
AA (C20) (mg/mL)	0.313 (0.099)
ALA (C18) (mg/mL)	0.030 (0.017)
EPA (C20) (mg/mL)	0.071 (0.064)
DPA (C23) (mg/mL)	0.037 (0.015)
DHA (C22) (mg/mL)	0.103 (0.045)
n-6:n-3 ratio	6.4 (3.3)

SD standard deviation, MET metabolic equivalent of task, BMD bone mineral density, BTMs bone turnover markers, serum CTX serum C-terminal telopeptides of type 1 collagen, urinary CTX/Cr urinary C-terminal telopeptides of type 1 collagen/creatinine, urinary DPD/Cr urinary deoxypyridinoline/creatinine, LCPUFA long chain polyunsaturated fatty acids, total n3 total omega 3, total n6 total omega 6, LA linoleic acid (18:2 n – 6); AA, arachidonic acid (20:4 n – 6); ALA, α -linolenic acid (18:3 n – 3); EPA, eicosapentaenoic acid (20:5 n – 3); DPA, docosapentaenoic acid (23:5 n – 3); DHA, docosahexaenoic acid (22:6 n – 3); n – 6:n – 3 ratio, total n – 3:total n – 6 ratio

13.1 (1.5) and 48.5 (5.2) years. Mean (SD) total dietary intake of fish was 30.2 (31.8) g/day. Mean (SD) dietary intake of polyunsaturated fat was 11.43 (4.57) g/day.

Correlation analysis

A significant negative correlation was observed between the bone resorption marker urinary DPD/Cr and total n-3LCPUFA (r=-0.137; p=0.018) as well as with EPA (r=-0.150; p=0.009) and DPA (r=-0.118; p=0.042). A



Covariates

Age was a significant negative predictor of femur ($\beta = -0.221$; p = 0.012) and lumbar spine BMD ($\beta = -0.208$; p = 0.019) but not a significant predictor of BTMs. BMI was a significant positive predictor of femur ($\beta = 0.261$; P < 0.001) and lumbar spine BMD ($\beta = 0.327$; P < 0.001). BMI was also a significant negative predictor of serum CTX ($\beta = -0.324$; P < 0.001), serum osteocalcin ($\beta = -0.249$; P < 0.001) and urinary CTX/Cr (β =-0.396; P<0.001). The number of years' post-menopause was a significant negative predictor of serum total alkaline phosphatase ($\beta = -0.194$; p = 0.033) and total physical activity was a significant negative predictor of urinary DPD/Cr ($\beta = -0.124$; p = 0.044) but neither were significant predictors of bone parameters. Smoking status was a significant negative predictor of femur T-score $(\beta = -0.494; p = 0.002)$ but was not a significant predictor of BTMs.

Relationships between bone parameters and LCPUFA

Associations between LCPUFA and bone parameters are outlined in Table 2. After adjusting for age, BMI, physical activity, smoking status and years' post-menopause, the n-3 LCPUFA, ALA, EPA, DPA, DHA and n-6 PUFA, AA, LA were not significantly associated with the bone parameters femur and lumbar spine BMD or with T-score (P>0.05). There was also no significant association observed between the n-6:n-3 ratio and bone parameters after adjusting for covariates. When split into tertiles according to the n-6:n-3 ratio, a significant positive association between total n-3 LCPUFA and femur BMD (β =0.287; p=0.043) within those with a low n-6:n-3 ratio was observed (Fig. 1).

Relationships between BTMs and LCPUFA

Associations between BTMs and serum LCPUFA concentrations are summarised in Table 3. After adjusting for covariates, there were no significant associations observed between total n-3 LCPUFA, EPA, DPA or DHA with any of the markers of bone turnover or the CTX:OC ratio. There was a significant inverse association between the n-3 LCPUFA ALA and urinary DPD/Cr (β =-0.141; p=0.016). Each



Table 2 Associations between LCPUFA and femur and lumbar spine bone mineral density and T-score in postmenopausal women (n = 299)

	Femur	BMD (g/c	cm ²)	Lumba cm ²)	ar spine BN	MD (g/	Femur	T-score		Lumba	Lumbar spine T-score		
Multivariate model	R^2	Std β	p value	R^2	Std β	p value	R^2	Std β	p value	R^2	Std β	p value	
Total n – 3 LCPUFA (mg/mL)	0.140	0.018	0.751	0.126	0.030	0.599	0.149	0.023	0.686	0.127	0.022	0.700	
ALA (mg/mL) (C18)	0.142	-0.047	0.399	0.125	-0.013	0.815	0.151	-0.049	0.379	0.127	-0.028	0.618	
EPA (mg/mL) (C20)	0.140	0.015	0.784	0.126	0.033	0.561	0.149	0.017	0.754	0.126	0.017	0.762	
DPA (mg/mL) (C23)	0.141	-0.033	0.549	0.125	-0.007	0.899	0.149	-0.020	0.724	0.126	-0.015	0.792	
DHA (mg/mL) (C22)	0.142	0.042	0.462	0.127	0.050	0.386	0.150	0.039	0.493	0.127	0.035	0.542	
Total n – 6 PUFA (mg/mL)	0.146	0.075	0.175	0.126	0.033	0.556	0.153	0.065	0.240	0.129	0.051	0.366	
LA (mg/mL) (C18)	0.145	0.068	0.221	0.126	0.019	0.733	0.151	0.049	0.375	0.127	0.033	0.550	
AA (mg/mL) (C20)	0.146	0.076	0.177	0.129	0.066	0.245	0.157	0.094	0.093	0.134	0.088	0.119	
n-6: n-3 ratio	0.141	0.031	0.577	0.125	-0.008	0.886	0.149	0.020	0.723	0.126	0.011	0.843	

Standardised Coefficients Beta, BMD bone mineral density, LCPUFA long chain polyunsaturated fatty acids, total n3 total omega 3, total n6 total omega 6, LA linoleic acid (18:2 n – 6), AA arachidonic acid (20:4 n – 6), ALA α -linolenic acid (18:3 n – 3), EPA, eicosapentaenoic acid (20:5 n – 3), DPA docosapentaenoic acid (23:5 n – 3), DHA docosapentaenoic acid (22:6 n – 3), n – 6: n – 3 ratio, total n – 3:total n – 6 ratio

^{*}Denotes significance (p < 0.05). Multivariate model adjusted for age, BMI, physical activity, smoking status and years' post-menopause

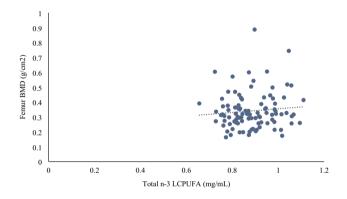


Fig. 1 Significant positive association between total n-3 LCPUFA and femur BMD within those with a low n-6:n-3 ratio (n=100). Significance determined using Multiple regression adjusted for age, BMI, physical activity, smoking status, years' post-menopause and n-6 PUFA ($\beta=0.287$, R^2 with covariates only =0.122, total model R^2 with LCPUFA =0.162, p=0.043)

unit increase in ALA was associated with a -0.141 mg/mL decrease in urinary DPD/Cr. The n-6: n-3 ratio was not significantly associated with any BTMs. Furthermore, there was no significant association between total n-6 PUFA, AA and LA and any of the markers of bone turnover. When split into tertiles for the n-6: n-3 ratio, a significant negative association between total n-3 LCPUFA and osteocalcin (β =-0.767; p=0.036) within those with a medium n-6: n-3 ratio was observed. There was a significant positive association between total n-3 LCPUFA and urinary CTX/Cr (β =0.224; p=0.024) within those with a high n-6: n-3 ratio.

Discussion

This observational study showed no significant association between n-3 LCPUFA and any of the measured bone parameters in this cohort of postmenopausal women; however total n-3 LCPUFA was shown to be positively associated with femur BMD in those postmenopausal women with a low n-6: n-3 ratio. Consideration should also be given to reducing n-6: n-3 ratio through improving dietary intake of n-3 LCPUFA along with reducing n-6 PUFA. In terms of BTMs, a higher ALA concentration (n-3 LCPUFA) was significantly associated with lower urinary DPD/Cr, suggesting lower bone resorption. Overall, this study highlights that increasing n-3 LCPUFA may have benefits to bone health in postmenopausal women, a vulnerable group at high risk of developing osteoporosis and associated fractures.

A number of studies have investigated associations between total PUFA status or n-3 LCPUFA dietary intake and BMD in women [18, 19, 23, 34, 35] as well as in postmenopausal women [21, 22, 24, 36]. A significant positive association between dietary n-3 LCPUFA and lumbar spine and femoral neck BMD has previously been reported [19, 22], but only after adjusting for covariates [22] including age, BMI, duration of menopausal state, grip strength, and intakes of calcium, vitamin D, vitamin K, n-6 PUFA, polyunsaturated fatty acid, serum N-terminal propeptide of type I collagen, and urinary type-I collagen cross-linked N-telopeptide. These studies assessed dietary intake of n-3LCPUFA rather than serum n-3 LCPUFA concentrations and therefore were not able to verify the correlation between n-3 LCPUFA and BMD using a validated biomarker of status [19, 22]. In a recent study in 301 Spanish postmenopausal women, a significant positive association between



Table 3 Associations between LCPUFA and bone turnover markers in postmenopausal women (n=299)

	Serum	Serum CTX (ng/mL)	/mL)	Serum	Serum OC (ng/mL)	L)	Serum alkalir phatase (u/L)	Serum alkaline phosphatase (u/L)	-soq	Urinary mmol/L)	Jrinary CTX/CR nmol/L)	(ng/	Urinary DPD/m	Urinary DPD/CR DPD/mmol Cr)	lomu)	CTX:0	CTX:0C ratio	
Multivariate model	\mathbb{R}^2	Std ß	Std β P value R^2	\mathbb{R}^2	Std ß	P value	\mathbb{R}^2	Std β	P value	\mathbb{R}^2	Std β	P value	\mathbb{R}^2	Std β	P value	\mathbb{R}^2	Std β	P value
Total n-3 PUFA (mg/mL) 0.098 -0.010 0.865 0.061	0.098	-0.010	0.865	0.061	0.022	0.705	0.004	-0.034	0.565	0.169	- 0.031	0.57	0.061	-0.104	0.078	0.014	-0.039	0.511
ALA (mg/mL) (C18)	0.100	0.100 - 0.056 0.332	0.332	0.061	-0.015	0.792	0.036	-0.021	0.721	0.169	-0.040	0.469	0.070	-0.141	0.016*	0.015	-0.047	0.429
EPA (mg/mL) (C20)	0.098	0.098 - 0.008 0.893	0.893	0.062	0.036	0.535	0.035	-0.003	996.0	0.172	0.069	0.209	0.062	-0.105	0.072	0.015	-0.055	0.362
DPA (mg/mL) (C23)	0.098	0.098 -0.022 0.699	0.699	0.061	0.020	0.732	0.036	-0.025	0.670	0.168	-0.007	968.0	0.059	-0.088	0.131	0.015	-0.052	0.389
DHA (mg/mL) (C22)	0.098	0.098 -0.012 0.831	0.831	0.062	0.031	0.603	0.042	-0.081	0.174	0.168	-0.03	0.591	0.055	-0.062	0.297	0.015	-0.054	0.378
Total n-6 PUFA (mg/mL)	0.098	0.098 -0.016 0.775	0.775	0.065	-0.063	0.275	0.037	-0.039	0.503	0.174	-0.080	0.141	0.051	-0.004	0.942	0.016	90.0	0.315
LA (mg/mL) (C18)	0.098	0.098 0.003	0.710	990.0	-0.071	0.222	0.037	-0.037	0.528	0.174	-0.082	0.131	0.051	-0.006	0.916	0.016	0.063	0.287
AA (mg/mL) (C20)	0.098	0.098 0.003	0.963	0.061	-0.024	0.688	0.037	-0.037	0.537	0.171	-0.059	0.285	0.051	0.001	0.983	0.013	0.033	0.586
n-6: $n-3$ ratio	0.098	0.098 -0.001 0.988	0.988	0.065	-0.063	0.280	0.036	0.008	0.888	0.168	-0.021	0.702	0.061	0.101	0.088	0.018	0.078	0.193

creatinine, urinary DPD/Cr, urinary deoxypyridinoline/creatinine, CTX: OC ratio, serum C-terminal telopeptides of type 1 collagen: serum osteocalcin, LCPUFA long chain polyunsaturated fatty Bandardised Coefficients Beta, serum CTX C-terminal telopeptides of type 1 collagen, serum OC serum osteocalcin, urinary CTX/Cr urinary C-terminal telopeptides of type 1 collagen acids, total n3 total omega 3, total omega 6, LA linoleic acid (18:2 n – 6); AA arachidonic acid (20:4 n – 6), ALA \alpha-linolenic acid (18:3 n – 3), EPA eicosapentaenoic acid (20:5 n – 3) total n-3:total n-6 ratio

Denotes significance (p < 0.05). Multivariate model adjusted for age, BMI, physical activity, smoking status and years' post-menopause

plasma n-3 LCPUFA and spine and femur neck BMD was observed after adjusting for covariates [21]. It is important to note that the Spanish cohort included osteoporotic women whereas anyone with osteoporosis was excluded from this study making it more challenging to determine associations with BMD. Nevertheless, taken together, this research supports a beneficial role n-3 LCPUFA for bone health in postmenopausal women.

It has been postulated that higher intakes of n-6 PUFA results in raised pro-inflammatory cytokine production that stimulates osteoclastic activity and thereby negatively impacting on bone health [37, 38]. Additionally, PGE2 produced from arachidonic acid (n-6 LCPUFA), is the main prostaglandin involved with the bone turnover cycle with lower PGE2 promoting bone formation whereas higher concentrations of PGE2 have been shown to inhibit bone formation [18, 39]. The optimal n-6: n-3 PUFA ratio for favourable effects on bone health is currently unknown; however, a contemporary western diet high in n-6 PUFA and therefore a higher n-6: n-3 ratio, may not contain the optimum amount of n-3 LCPUFA needed to benefit bone health [40]. It has been hypothesised that a balance between n-6 and n-3 LCPUFA is of greater importance for resultant health benefits than either class of PUFA by itself [41]. This hypothesis has been supported in our study where in participants with a low n-6: n-3 ratio, a significant positive association between n-3 LCPUFA and higher femur BMD was observed. Overall, circulating n-6: n-3 ratio was approximately 6:1 in the present study and this might suggest women in our cohort had a favourable dietary intake of n-3 LCPUFA as the circulating ratio is relatively low when compared to a western diet ratio of around 15-20:1 [42, 43]. Previous studies in similar groups of older women have reported a range of dietary n-6: n-3 ratios, between 4:1 to 8:3 [21, 22, 24, 34]. In the current study, dietary intake of fish was 30.2 g/day which is equivalent to 211.4 g/week. Dietary intake of rich n-3 LCPUFA sources such as oily fish can be low, with the current average adult intake in the United Kingdom ~ 56 g/week which is 2.5 times lower than the current guidance of at least one portion of oily fish/week (140 g) [44]. Although fish intakes in our study were shown to be sufficiently high to meet the UK recommendation, data were not available on the types of fish consumed and therefore it was not possible to quantify oily fish consumption. Fish intakes may have been primarily from non-oily fish sources. Furthermore, the inclusion of fish in the diet provides a significant source of protein and other nutrients with known benefits for musculoskeletal health and although this research was focused on PUFA it is plausible that other factors may be adding to the benefits seen. It has been reported that greater intakes of dietary n – 3 LCPUFA or use of supplementation may be essential for resultant benefits to bone health [24]. Dietary recommendations for fish consumption



are based on the nutritional benefits of consuming fish especially as a rich source of n-3 LCPUFA but also take into consideration the risks from potential pollutants [45]. In addition, there are no recommendations for n-3 supplements [46] and therefore it is difficult to recommend an optimal n-3 LCPUFA supplementation dose for benefit to bone heath.

BMD and BTMs have been assessed as outcome variables in several studies to investigate relationships between n-3LCPUFA and bone health. BTMs change more rapidly than BMD and therefore BTMs can be used to observe response to treatment before changes in BMD occur [47]. Although n-3 LCPUFA was not significantly associated with bone formation markers, ALA (n-3 LCPUFA) was shown to be significantly associated with less bone resorption in the present study. Similar findings have previously been reported in a group of elderly patients in which no significant association between n-3 LCPUFA on the bone formation marker BALP but an inverse association with the bone resorption marker TRAP-5b [13]. Although intervention with a low dose (1.2 g) of n-3 LCPUFA or fish oil showed no significant effect on bone formation markers [26, 48], intervention with 900 mg n-3 LCPUFA for 6 months led to a significant decrease in the bone resorption marker urinary Pyd when compared to a control group in postmenopausal women [26]. In addition, intervention with much higher doses of EPA + DHA (4 g) has been reported to significantly reduce deoxypyridinoline (DPD) from baseline to 6 months when compared to the control in postmenopausal women [27] suggesting a potential beneficial role for n-3 LCPUFA in reducing bone resorption in postmenopausal women. In those with a medium n-6: n-3 ratio, we observed a significant negative association between total n-3 LCPUFA and bone formation marker osteocalcin. In addition, in those with a high n-6: n-3 ratio, we observed a significant positive association between total n-3 LCPUFA and bone resorption marker urinary CTX/Cr which is contrary to what others have reported [49, 50].

We observed a significant negative association between the n-3 LCPUFA ALA concentrations and urinary DPD/Cr. It has been postulated that n-3 LCPUFA may lessen bone resorption by decreasing urinary excretion of calcium and reducing inflammatory cytokine production which are significant stimuli for osteoclastic activity [51, 52]. ALA may affect bone resorption through PGE2 which in turn can prevent activation of RANKL, a key growth factor that stimulates and promotes osteoclast (bone-resorbing cells) formation [53]. The cause and effect to explain the reported relationship between n-3 LCPUFA and bone health has not been conclusively determined but it has been proposed that the immunomodulatory effects of the PUFAs may contribute to the inhibition of osteoclast formation and in turn have a beneficial effect on preventing bone loss. Advancing age and

menopause are associated with greater pro-inflammatory cytokine expression and chronic inflammation, in addition to other mechanisms which can accelerate bone loss and contribute to the development of osteoporosis [54, 55]. Pro-inflammatory cytokines play a significant role in oestrogen depletion-associated bone loss in postmenopausal women [50, 56]. The LCPUFA have an immunomodulatory effect through the regulation of pro-inflammatory cytokines IL-1 β , IL-6, IL-8 and TNF- α through the production of eicosanoids including prostaglandins (PGs), thromboxanes (TXs), leukotrienes (LTs) and resolvins [26, 57]. Therefore, enhancing LCPUFA in the diet, particularly n – 3 LCPUFAs such as EPA and DHA, could have resultant benefits to the inflammatory milieu [57] and favourable effects on bone remodelling in postmenopausal women [50].

Research assessing the beneficial effect of LCPUFA on bone has frequently used dietary recall methods such as a food frequency questionnaires and food diaries to determine the dietary intake of LCPUFA. These dietary instruments can lead to variations in results owing to the methodological confinements of dietary recall [58]. In this study, PUFA were quantified in serum using the gold standard method of gas chromatography-mass spectrometry; however other measures such as plasma, phospholipids and red blood cells could be alternative, cost effective and readily available methods for PUFA analysis [13]. The cross-sectional nature of this study negates the ability to determine causality. Based on Pearson's correlation coefficient analysis, this study had over 90% power to detect correlation coefficients larger in magnitude than 0.20. Due to the challenging nature of interpreting n-6: n-3 ratio, secondary analysis in this study should be interpretated with caution. Measuring EPA and DHA within the red cell membrane for determination of the n-3index may be useful in fully elucidating the benefits of n-3PUFA on bone health without the confounding effect of the more pro-inflammatory n-6 PUFA. In addition, it is possible we have residual confounding owing to lack of inclusion of additional confounding predictors of bone health e.g. vitamin D, calcium & protein. Future studies with larger sample sizes should take this into consideration. Our analysis could also have benefited from including a sub-group of postmenopausal women with osteoporosis to further explore associations between serum PUFA and osteoporotic postmenopausal women, albeit this study still adds to the body of biological research exploring the beneficial role of dietary PUFA on bone in postmenopausal women.

In conclusion, the positive association between a low n-6: n-3 ratio and femur BMD alongside the negative association between ALA with urinary DPD/Cr highlights a potentially important role for n-3 LCPUFA status on maintenance of bone health in postmenopausal women. Our findings warrant further investigation using long-term intervention studies to confirm this possible beneficial role



for dietary intake of n-3 LCPUFA in reducing postmenopausal bone loss which could provide an evidence base to promote the intake of n-3 LCPUFA rich dietary sources to support bone health.

Authors' contributions Conceptualisation, Methodology: EM.McS., PJ.A., JJ.S. PJ.M., LK.P. Writing- Original draft preparation, Project administration: EM.McS., MM.S., O.F. Supervision: EM.McS., DJ.A., PJ.M., LK.P. Writing- Reviewing and Editing: EM.McS., PJ.A., PJ.M., LK.P., JJ.S., MC.C., DJ.A. Investigation: O.F., MM.S., MC.C., Formal analysis, Data curation: O.F. MM.S.

Funding Support by the Department for the Economy, Marigot (Cork, Ireland) and Corn Products International (Westchester, IL).

Availability of data and materials The authors confirm that the data supporting the findings of this study are available in this article.

Code availability Not applicable.

Declarations

Conflict of interest Dr McSorley, Dr Magee & Dr Pourshahidi reported a grant from BetterYou Ltd, outside the work submitted. Dr Pourshahidi reported a grant from Devenish Nutrition Ltd, outside the work submitted. Professor Armstrong reported Royal Osteoporosis Society (advisory board), National Osteoporosis Guideline Group (advisory board), personal fees from Internis Pharma (speaker), UCB Pharma (advisory board) and stock ownership (GlaxoSmithKline), outside of the work submitted.

Ethics approval Ethical approval was received from Ulster University Research Ethics Committee (REC/08/0083) and was performed in accordance with the ethical standards laid down in the 1964 Declaration of Helsinki and its later amendments. The original study was registered as a clinical trial (www.controlled-trials.com as ISRCTN63118444). Written informed consent was obtained from all participants prior to their inclusion in the study.

Consent to participate The study was conducted in accordance with the declaration of Helsinki and written informed consent was obtained from all participants.

Consent for publication All authors have read and approved the final manuscript.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Kruger MC, Wolber FM (2016) Osteoporosis: modern paradigms for last century's bones. Nutrients 8(6):376–376. https://doi.org/ 10.3390/nu8060376
- Shen D, Zhang X, Li Z, Bai H, Chen L (2017) Effects of omega-3 fatty acids on bone turnover markers in postmenopausal women: systematic review and meta-analysis. Climacteric 20(6):522–527. https://doi.org/10.1080/13697137.2017
- Johnell O, Kanis JA (2006) An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. Osteoporos Int 17(12):1726–1733. https://doi.org/10.1007/s00198-006-0172-4
- Hernlund E, Svedbom A, Ivergård M, Compston J, Cooper C, Stenmark J, McCloskey EV, Jönsson KJA, B, (2013) Osteoporosis in the European Union: Medical management, epidemiology and economic burden: A report prepared in collaboration with the International Osteoporosis Foundation (IOF) and the European Federation of Pharmaceutical Industry Associations (EFPIA). Arch Osteoporos 8(1):136. https://doi.org/10.1007/s11657-013-0136-1
- Ji M, Yu Q (2015) Primary osteoporosis in postmenopausal women. Chronic Dis Transl Med 1(1):9–13. https://doi.org/10. 1016/j.cdtm.2015.02.006
- Cauley JA (2015) Estrogen and bone health in men and women. Steroids 99:11–15. https://doi.org/10.1016/j.steroids.2014.12. 010
- Hofbauer LC, Schoppet M (2004) Clinical implications of the osteoprotegerin/RANKL/RANK system for bone and vascular diseases. J Am Med Assoc 292(4):490–495. https://doi.org/10. 1001/jama.292.4.490
- Boyce BF, Xing L (2007) Biology of RANK, RANKL, and osteoprotegerin. Arthr Res Ther 9:S1. https://doi.org/10.1186/ar2165
- Baggio B, Budakovic A, Nassuato MA, Vezzoli G, Manzato E, Luisetto G, Zaninotto M (2000) Plasma phospholipid arachidonic acid content and calcium metabolism in idiopathic calcium nephrolithiasis. Kidney Int 58(3):1278–1284. https://doi.org/10.1046/j. 1523-1755.2000.00283.x
- Li X, Pilbeam CC, Pan L, Breyer RM, Raisz LG (2002) Effects of prostaglandin E2 on gene expression in primary osteoblastic cells from prostaglandin receptor knockout mice. Bone 30(4):567–573. https://doi.org/10.1016/s8756-3282(02)00683-x
- Orchard TS, Cauley JA, Frank GC, Neuhouser ML, Robinson JG, Snetselaar L, Tylavsky F, Wactawski-Wende J, Young AM, Lu B, Jackson RD (2010) Fatty acid consumption and risk of fracture in the Women's Health Initiative. Am J Clin Nutr 92(6):1452–1460. https://doi.org/10.3945/ajcn.2010.29955
- Rizzoli R, Bischoff-Ferrari H, Dawson-Hughes B, Weaver C (2014) Nutrition and bone health in women after the menopause.
 Womens Health 10(6):599–608. https://doi.org/10.2217/whe.14.
- Kim B, Kwak MK, Lee SH, Koh J, Yoo HJ, Kim SJ, Park SJ, Hamrick MW, Isales CM, Ahn SH (2019) Association of blood n-3 fatty acid with bone mass and bone marrow TRAP-5b in the elderly with and without hip fracture. Osteoporos Int 30(5):1071– 1078. https://doi.org/10.1007/s00198-019-04881-3
- Sharma T, Mandal CC (2020) Omega-3 fatty acids in pathological calcification and bone health. J Food Biochem 44(8):e13333. https://doi.org/10.1111/jfbc.13333
- Martyniak K, Wei F, Ballesteros A, Meckmongkol T, Calder A, Gilbertson T, Orlovskaya N, Coathup MJ (2021) Do polyunsaturated fatty acids protect against bone loss in our aging and osteo-porotic population? Bone 143:115736. https://doi.org/10.1016/j.bone.2020.115736



- Romero-Márquez JM, Varela-López A, Navarro-Hortal MD, Badillo-Carrasco A, Forbes-Hernández TY, Giampieri F, Domínguez I, Madrigal L, Battino M, Quiles JL (2021) Molecular interactions between dietary lipids and bone tissue during aging. Int J Mol Sci 22(6473):6473–6473. https://doi.org/10.3390/ijms2 2126473
- Sadeghi O, Ghorabi S, Shab-Bidar S, Djafarian K, Khodadost M, Nasiri M (2019) Dietary intake of fish, n-3 polyunsaturated fatty acids and risk of hip fracture: a systematic review and meta-analysis on observational studies. Crit Rev Food Sci Nutr 59(8):1320–1333. https://doi.org/10.1080/10408398.2017.14059 08
- Järvinen R, Erkkilä AT, Penttinen P, Tuppurainen M, Kärkkäinen M, Salovaara K, Kröger H, Jurvelin JS (2012) Associations of dietary polyunsaturated fatty acids with bone mineral density in elderly women. Eur J Clin Nutr 66(4):496–503. https://doi.org/10. 1038/ejcn.2011.188
- Lavado-García J, Roncero-Martin R, Moran JM, Pedrera-Canal M, Aliaga I, Leal-Hernandez O, Rico-Martin S, Canal-Macias M (2018) Long-chain omega-3 polyunsaturated fatty acid dietary intake is positively associated with bone mineral density in normal and osteopenic Spanish women. PLoS ONE 13(1):e0190539. https://doi.org/10.1371/journal.pone.019053
- Feskanich D, Willet WC, Colditz GA (2003) Calcium, vitamin D, milk consumption, and hip fractures: a prospective study among postmenopausal women. Am J Clin Nutr 77(2):504–511. https://doi.org/10.1093/ajcn/77.2.504
- Roncero-Martín R, Aliaga I, Moran JM, Puerto-Parejo LM, Rey-Sánchez P, Luz Canal-Macías M, Sánchez-Fernández A, Pedrera-Zamorano JD, López-Espuela F, Vera V, Cerrato-Carretero P, Lavado-García JM (2021) Plasma fatty acids and quantitative ultrasound, DXA and pQCT derived parameters in postmenopausal spanish women. Nutrients 13(1454):1454– 1454. https://doi.org/10.3390/nu13051454
- Nawata K, Yamauchi M, Takaoka S, Yamaguchi T, Sugimoto T (2013) Association of n-3 polyunsaturated fatty acid intake with bone mineral density in postmenopausal women. Calcif Tissue Int 93(2):147–154. https://doi.org/10.1155/2015/737521
- Weiss LA, Barrett-Connor E, Von Mühlen D (2005) Ratio of n-6 to n-3 fatty acids and bone mineral density in older adults: the Rancho Bernardo study. Am J Clin Nutr 81(4):934–938. https:// doi.org/10.1093/ajcn/81.4.934
- Harris M, Farrell V, Going S, Houtkooper L, Lohman T (2015)
 Associations of polyunsaturated fatty acid intake with bone mineral density in postmenopausal women. J Osteoporos. https://doi.org/10.1155/2015/737521
- Mei Z, Chen G, Hu J, Lin C, Sun Z, Liu C, Geng X, Yuan C, Qi Q, Zheng Y (2021) Habitual use of fish oil supplements, genetic predisposition, and risk of fractures: a large population-based study. Am J Clin Nutr. https://doi.org/10.1093/ajcn/nqab157
- Sharif PS, Asalforoush M, Ameri F, Larijani B, Abdollahi M (2010) The effect of n-3 fatty acids on bone biomarkers in Iranian postmenopausal osteoporotic women: a randomised clinical trial. Age 32(2):179–186. https://doi.org/10.1007/s11357-009-9122-3
- Hutchins-Wiese H, Picho K, Watkins BA, Kenny AM, Li Y, Tannenbaum S, Claffey K (2014) High-Dose eicosapentaenoic acid and docosahexaenoic acid supplementation reduces bone resorption in postmenopausal breast cancer survivors on aromatase inhibitors: a pilot study. Nutr Cancer 66(1):68–76. https://doi.org/10.1080/01635581.2014.847964
- 28. Slevin MM, Allsopp PJ, Magee PJ, Bonham MP, Naughton VR, Strain JJ, Duffy ME, Wallace JM, McSorley EM (2014) Supplementation with calcium and short-chain fructo-oligosaccharides affects markers of bone turnover but not bone mineral density in

- postmenopausal women. J Nutr 144:297–304. https://doi.org/10.3945/jn.113.188144
- Hagstromer M, Oja P, Sjostrom M (2006) The International Physical Activity Questionnaire (IPAQ): a study of concurrent and construct validity. Public Health Nutr 9(6):755–762. https://doi.org/10.1079/phn2005898
- World Health Organisation (1994) Assessment of fracture risk and its application to screening for postmenopausal osteoporosis: report of a WHO study group [meeting held in Rome from 22 to 25 June 1992]. World Health Organisation Study Group. https:// apps.who.int/iris/handle/10665/39142/
- Writing Group for the ISCD Position Development Conference (2004) Nomenclature and decimal places in bone densitometry. J Clin Densitom 7(1):45–50. https://doi.org/10.1385/jcd:7:1:45
- Folch J, Lees M, Slone Stanley GH (1957) A simple method for the isolation and purification of total lipides from animal tissues. J Biol Chem 226(1):497–509. https://doi.org/10.1016/S0021-9258(18)64849-5
- 33. Strain JJ, Davidson PW, Bonham MP, Duffy EM, Stokes-Riner A, Thurston SW, Wallace JMW, Robson PJ, Shamlaye CF, Georger LA, Sloane-Reeves J, Cernichiari E, Canfield RL, Cox C, Huang LS, Janciuras J, Myers GJ, Clarkson TW (2008) Associations of maternal long-chain polyunsaturated fatty acids, methyl mercury, and infant development in the seychelles child development nutrition study. Neurotoxicology 29(5):776–782. https://doi.org/10.1016/j.neuro.2008.06.002
- 34. Farina EK, Kiel DP, Roubenoff R, Schaefer EJ, Cupples LA, Tucker KL (2011) Protective effects of fish intake and interactive effects of long-chain polyunsaturated fatty acid intakes on hip bone mineral density in older adults: the Framingham osteoporosis study. Am J Clin Nutr 93(5):1142–1151. https://doi.org/10.3945/ajcn.110.005926
- Macdonald HM, Golden MHN, Reid DM, Campbell MK, New SA (2004) Nutritional associations with bone loss during the menopausal transition: Evidence of a beneficial effect of calcium, alcohol, and fruit and vegetable nutrients and of a detrimental effect of fatty acids. Am J Clin Nutr 79(1):155–165. https://doi.org/10.1093/ajcn/79.1.155
- Bassey EJ, Littlewood JJ, Rothwell MC, Pye DW (2000) Lack of effect of supplementation with essential fatty acids on bone mineral density in healthy pre- and postmenopausal women: two randomised controlled trials of Efacal® v. calcium alone. Br J Nutr 83(6):629–635. https://doi.org/10.1017/S000711450 0000805
- Sun D, Krishnan A, Zaman K, Lawrence R, Bhattacharya A, Fernandes G (2003) Dietary n-3 fatty acids decrease osteoclastogenesis and loss of bone mass in ovariectomised mice. J Bone Miner Res 18(7):1206–1216. https://doi.org/10.1359/jbmr.2003. 18 7 1206
- Genuis SJ, Schwalfenberg GK (2007) Picking a bone with contemporary osteoporosis management: nutrient strategies to enhance skeletal integrity. Clin Nutr 26(2):193–207. https://doi. org/10.1016/j.clnu.2006.08.004
- 39. Watkins BA, Lippman HE, Le Bouteiller L, Li Y, Seifert MF (2001) Bioactive fatty acids: role in bone biology and bone cell function. Prog Lipid Res 40(1–2):125–148. https://doi.org/10.1016/s0163-7827(00)00016-3
- Blasbalg TL, Hibbeln JR, Ramsden CE, Majchrzak SF, Rawlings RR (2011) Changes in consumption of omega-3 and omega-6 fatty acids in the United States during the 20th century.
 Am J Clin Nutr 93(5):950–962. https://doi.org/10.3945/ajcn. 110.006643
- Simopoulos AP (2008) The importance of the omega-6/omega-3 fatty acid ratio in cardiovascular disease and other chronic diseases. Exp Biol Med 233(6):674–688. https://doi.org/10.3181/ 0711-MR-311



- Stark AH, Reifen R, Crawford MA (2008) Update on alpha-lino-lenic acid. Nutr Rev 66(6):326–332. https://doi.org/10.1111/j. 1753-4887.2008.00040.x
- Molendi-Coste O, Legry V, Leclercq IA (2011) Why and how meet n-3 PUFA dietary recommendations? Gastroenterol Res Pract. https://doi.org/10.1155/2011/364040
- 44. National Diet and Nutrition Survey (2019) Results from Years 5 to 9 (combined) of the Rolling Programme for Northern Ireland (2012/2013-2016/2017) and time trend and income analysis (Years 1 to 9; 2008/2009-2016/2017) https://www.food.gov.uk/sites/default/files/media/document/national-diet-and-nutrition-survey-northern-ireland-y5-9-full-report_2.pdf. Accessed 15 Sept 2020.
- Scientific Advisory Committee for Nutrition (2004) Advice on fish consumption: benefits & risks. Report of committee on Toxicity. London: TSO. https://cot.food.gov.uk/sites/default/files/ cot/fishreport200401.pdf. Accessed 26 May 2021.
- British Dietetic Association (2020) Omega-3: food fact sheet. https://www.bda.uk.com/resource/omega-3.html. Accessed 15 July 2021.
- 47. Konstantinidis I, Papageorgiou SN, Kyrgidis A, Tzellos T, Kouvelas D (2013) Effect of non-steroidal anti-inflammatory drugs on bone turnover: an evidence-based review. Rev Recent Clin Trials 8(1):48–60. https://doi.org/10.2174/1574887111308010008
- 48. Dong H, Kleppinger A, Annis K, Kenny AM, Liva E, Lammi-Keefe C, Durham H, Feinn R, Hutchins-Wiese H (2015) Effects of omega-3 polyunsaturated fatty acid supplementation on bone turnover in older women. Int J Vitam Nutr Res 84(3–4):124–132. https://doi.org/10.1024/0300-9831/a000199
- Griel AE, Kris-Etherton PM, Hilpert KF, Zhao G, West SG, Corwin RL (2007) An increase in dietary n-3 fatty acids decreases a marker of bone resorption in humans. Nutr J 6:2. https://doi.org/10.1186/1475-2891-6-2
- Kajarabille N, Díaz-Castro J, Hijano S, López-Frías M, López-Aliaga I, Ochoa JJ (2013) A new insight to bone turnover: role

- of -3 polyunsaturated fatty acids. Sci World J. https://doi.org/10.1155/2013/589641
- Salari P, Abdollahi M, Rezaie A, Larijani B (2008) A systematic review of the impact of n-3 fatty acids in bone health and osteoporosis. Med Sci Monitor 14(3):RA37–RA44
- Appleton KM, Fraser WD, Rogers PJ, Ness AR, Tobias JH (2011) Supplementation with a low-moderate dose of n-3 long-chain PUFA has no short-term effect on bone resorption in human adults. Br J Nutr 105(8):1145–1149. https://doi.org/10.1017/ S0007114510004861
- Coetzee M, Haag M, Kruger MC (2007) Effects of arachidonic acid, docosahexaenoic acid, prostaglandin E2 and parathyroid hormone on osteoprotegerin and RANKL secretion by MC3T3-E1 osteoblast-like cells. J Nutr Biochem 18(1):54–63. https://doi.org/ 10.1016/j.jnutbio.2006.03.002
- Ginaldi L, Di Benedetto MC, De Martinis M (2005) Osteoporosis, inflammation and ageing. Immunity and Ageing 2:14. https://doi. org/10.1186/1742-4933-2-14
- Orchard TS, Pan X, Cheek F, Ing SW, Jackson RD (2012) A systematic review of omega-3 fatty acids and osteoporosis. Br J Nutr 107:S253–S260. https://doi.org/10.1017/S0007114512001638
- Al-Daghri N, Yakout S, Al-Shehri E, Al-Fawaz H, Aljohani N, Al-Saleh Y (2014) Inflammatory and bone turnover markers in relation to PTH and vitamin D status among Saudi postmenopausal women with and without osteoporosis. Int J Clin Exp Med 7(10):3528–3535
- Stephenson JA, Al-Taan O, Arshad A, Morgan B, Metcalfe MS, Dennison AR (2013) The multifaceted effects of omega-3 polyunsaturated fatty acids on the hallmarks of cancer. J Lipids. https:// doi.org/10.1155/2013/261247
- Dao MC, Subar AF, Warthon-Medina M, Cade JE, Burrows T, Golley RK, Forouhi NG, Pearce M, Holmes BA (2019) Dietary assessment toolkits: an overview. Public Health Nutr 22(3):404– 418. https://doi.org/10.1017/S1368980018002951

