

Interactions between arbuscular mycorrhizal fungi and phosphate-solubilizing fungus (*Mortierella* sp.) and their effects on *Kosteletzkya virginica* growth and enzyme activities of rhizosphere and bulk soils at different salinities

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Abstract This study investigated the interactions between two arbuscular mycorrhizal fungi (AMF) (*Glomus aggregatum* and *Glomus mosseae*) and a P-solubilizing fungus (*Mortierella* sp.), with respect to their effects on growth of *Kosteletzkya virginica* and urease, invertase, neutral phosphatase, alkaline phosphatase, and catalase activities of rhizosphere and bulk soils at different salinity levels (i.e., 0, 100, 200, and 300 mM NaCl). Percentage of AMF colonization, *Mortierella* sp. populations, pH, electrical conductivity, and available P concentration in soil were also determined. Combined inoculation of AMF and *Mortierella* sp. increased the percentage of AMF colonization and *Mortierella* sp. populations under salt stress (i.e., 100, 200, and 300 mM NaCl). The dual inoculation of *Mortierella* sp. with AMF (*G. aggregatum* or *G. mosseae*) had significant effects on shoot and root dry weights and available P concentrations, pH values, and electrical conductivities of rhizosphere and bulk soils under salt stress. The inoculation of *Mortierella* sp. significantly enhanced the positive effects

of AMF on some enzyme activities (i.e., neutral phosphatase, alkaline phosphatase, and catalase in bulk soil; neutral phosphatase and urease in rhizosphere soil); on the contrary, it produced negative effects on urease activities in bulk soil and invertase activities in bulk and rhizosphere soils. The results indicated that the most effective co-inoculation was the dual inoculation with *Mortierella* sp. and *G. mosseae*, which may help in alleviating the deleterious effects of salt on plants growth and soil enzyme activities.

Keywords Arbuscular mycorrhizal fungus · Enzyme activity · Phosphate-solubilizing fungus · Plant growth · Salt stress

Introduction

Phosphorus (P) is one of the most essential macronutrients limiting plant growth owing to its low bioavailability in soils (Feng et al. 2004; Rodriguez and Fraga 1999). Nevertheless, the repeated and not proper applications of chemical P fertilizers is expensive and can lead to the loss of soil fertility by disturbing microbial diversity and reducing crops yield (Gyaneshwar et al. 2002a). Consequently, the use of environment-friendly alternative to applications of P fertilizers is gaining increasing attention (Barea et al. 1997). Phosphorous-solubilizing microorganisms (PSM) are able to replenish soil solution P by solubilizing insoluble native and applied rock phosphate (Rao 1992; Gyaneshwar et al. 2002b; Khan et al. 2007). Although P-solubilizing bacteria (PSB) have received greater attention than P-solubilizing fungi (PSF), the latter are more effective in solubilizing P compounds (Kucey 1983; Whitelaw et al. 1997). Moreover,

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PSF do not lose the P dissolving activities upon repeated subculturing under laboratory conditions as the PSB do (Kucey 1983; Rashid et al. 2004).

Approximately 7% of the global land surface is covered with saline soil (Ruiz-Lozano et al. 1996); the soil salinity is becoming one of the major limiting factors for crop production and an appropriate solution to this situation is to use PSF as bioinoculants. Rietz and Haynes (2003) and Tripathi et al. (2006) had reported that soil microbial communities and their activities are greatly influenced by salinity. Therefore, poor growth and survival of PSF may be a conceivable result. Nevertheless, no information is available on PSF isolated from saline soil.

Arbuscular mycorrhizal fungi (AMF) can form a symbiotic association with most plant species and improve the efficiency of associated plants to assimilate P from the soil solution (Manjunath et al. 1989; Habte and Osorio 2001). Studies have showed that AMF widely occur in salt environments (Rozema et al. 1986; Wang and Liu 2001) and can improve growth and productivity in both glycophytes and halophytes under salt stress (Asghari et al. 2005; Sannazzaro et al. 2006; Giri et al. 2007). Rodriguez and Fraga (1999) suggested that the inoculated rhizobacteria could release P ions from insoluble P sources and the released P was taken up by the external arbuscular mycorrhizal mycelium because the rhizobacteria cannot transfer P to roots. Previous studies showed that co-inoculation of AMF and PSF could enhance plant biomass, P concentration of plant tissues (e.g., Piccini and Azcon 1987; Singh and Kapoor 1999; Osorio and Habte 2001; Kohler et al. 2007), soil microbial activities (Kohler et al. 2007), and other soil properties (Matiasa et al. 2009). Thus, it is necessary to study the effects of AMF and PSF on microbial activities of saline soil and plant growth for the development of sustainable management of saline soils. To the authors' knowledge, however, there is no information of AMF and PSF combined inoculation in saline soil.

Soil enzyme activities are sufficiently sensitive to perturbations caused by microbial inoculation (Naseby and Lynch 1997), and these measurements can give insights on mineralization of important nutrient elements, such as carbon, nitrogen, and phosphorus (Ceccanti et al. 1994). The measurement of enzyme activities and the integration of the measured activities in a single index can provide information on microbial activity in soil (Nannipieri et al. 2002, 2003). Microorganisms are activated in the rhizosphere soil (Cordier et al. 2000). Therefore, it is important to determine enzyme activities of both rhizosphere and bulk soils, which can affect the concentrations of available P and other nutrients in soil (Naseby and Lynch 1997; Ceccanti et al. 1994). However, there is no information regarding the use of such parameters for monitoring the ecological impact of AMF and PSF inoculation in saline soil.

Kosteletzkya virginica (L.) Presl. is a perennial herbaceous halophyte native to brackish portions of coastal tidal marshes of the mid-Atlantic and southeastern United States and belongs to the family Malvaceae (Gallagher 1985). In 1992 it was transplanted on the saline land along coastal region of Jiangsu Province by Halophyte Research Lab of Nanjing University, China, to improve coastal tidal flats and develop biomass energy industry (Ruan et al. 2005). A few studies of salinity tolerance, delayed selfing, and plant regeneration of *K. virginica* have been conducted (Blits and Gallagher 1990; Ruan et al. 2009a, b). No information is available on the effects of AMF and PSF on the growth responses of *K. virginica* in saline soil. We have hypothesized that (1) inoculation of AMF and PSF can promote growth of the typical halophyte species (e.g., *K. virginica*) under salt stress; and (2) inoculated microorganisms can synergistically influence enzyme activities (i.e., invertase, catalase, urease, neutral phosphatase, and alkaline phosphatase) and available P concentrations of rhizosphere saline soil. To verify these hypotheses, two AMF [*Glomus mosseae* (Gm) and *Glomus aggregatum* (Ga)] and one PSF [*Mortierella* sp. (Ms)], which was isolated from salt-affected coastal soil samples collected from seashore of Jiangsu province, were inoculated in soil–sand culture watered with four levels of salinity.

Materials and methods

Plant culture

The experiment was conducted from the beginning of April to the end of December 2009 in a greenhouse at Halophyte Research Lab of Nanjing University (32°30' N and 118°42' E), Nanjing, China. The used substrate is a mixture of washed sand and soil (1:1, v:v), which was sterilized by autoclaving for 1 h at 121°C twice, on two consecutive days and then sieved (2 mm). Soil was collected from the botanical garden of Nanjing University with the following characteristics: pH 7.3, electrical conductivity (EC) 0.02 dS m⁻¹, organic matter 0.4%, total N 13 mg kg⁻¹, available P 3 mg kg⁻¹, and extractable potassium (K) 25 mg kg⁻¹. *K. virginica* was used as the host plant. Plants were grown in a greenhouse under controlled conditions: 16 h at day light intensity of 220 μEm⁻² s⁻¹ at 28°C and 8 h night at 18°C. The relative humidity in the greenhouse during the experiment was 65–85%.

Fungal inocula

Both *G. mosseae* (Nicol. and Gerd., originated from saline soil of Hebei province, China) and *G. aggregatum* (originated from saline soil of Hongkong, China) were

obtained from Bank of Glomales in China. The inocula consisted of spores (1,256 spores per 100 g⁻¹ soil), hyphae, and colonized root fragments, and were collected from a 6-month-old pot culture of *G. mosseae* and *G. aggregatum* grown on sorghum in sterile sandy soil.

To prepare liquid inoculum of *Mortierella* sp., the first step was to activate strains on slants. The fungus was inoculated on solid Martin culture medium (K₂HPO₄ 1 g, MgSO₄·7H₂O 0.5 g, NaCl 11.5 g, peptone 5 g, glucose 10 g, gelose 10 g, 1/30,000 Bengal red water solution 100 ml, and demineralized water 900 ml), which had been autoclaved for 30 min at 121°C and then cultured in the dark at 28°C for 4 days. After activation, 3 ml sterile water was added to test tube, and then the mixture was poured into 50 ml Martin broth (MB) which was added to 1.15% NaCl; *Mortierella* sp. was grown on a rotating shaker at 180 rpm for 48 h, and this was the starter culture. It was added (5% of volume) to MB and then we added 1.15% NaCl, and the MB was cultured on a shaker for 96 h at 180 rpm. At the end it contained 2.3 × 10⁵ colony forming units mL⁻¹ and the solution was stored at 4°C until use.

Experimental design and biological treatments

To study the effects of AMF and PSF on *K. virginica* under salt stress, the seedlings of *K. virginica* were watered with NaCl solutions. The experimental design was full factorial, with four salinity levels (0, 100, 200, and 300 mM) × eight microbial treatments: inoculated with 10 g of Gm, inoculated with 10 g of Ga, inoculated with 10 ml of Ms, inoculated with 5 g of *G. mosseae* and 5 g of *G. aggregatum* (Gm+Ga), inoculated with 10 g of *G. mosseae* and 10 ml of *Mortierella* sp. (Gm+Ms), inoculated with 10 g of *G. aggregatum* and 10 ml of *Mortierella* sp. (Ga+Ms), inoculated with 5 g of *G. mosseae* and 5 g of *G. aggregatum* and 10 ml of *Mortierella* sp. (Gm+Ga+Ms), and no inoculated control (CK). Each treatment was replicated three times in a randomized block design and each replicate had three plants.

On April 3, 2009, seeds of *K. virginica* were surface-sterilized by soaking in 5% NaOCl solution for 10 min and rinsed with sterile distilled water. They were then transferred aseptically to petri dishes filled with water and incubated for 4 days at 25°C. One week after germination, seedlings of uniform size were transferred to pots (diameter 15 cm, height 15 cm) filled with 900 g sterilized sandy soil; each pot had three seedlings. According to the design, 10 g mycorrhizal inoculum or/and 10 ml *Mortierella* sp. were placed in soil below the *K. virginica* seedlings prior to planting. Meanwhile, 10 g mycorrhizal inoculum and 10 ml Ms inoculum, both autoclaved at 121°C for 90 min three times, were added to control pots. Each pot was put on a 2-cm-deep plate and incubated in greenhouse on April 10, 2009.

Plants were irrigated with water for 4 weeks. Then, pots were watered by a modified (all nutrient solutions without P) solution of Hoagland and Arnon (1950). For the salinity treatments, NaCl (Fisher ACS) was added to the Hoagland nutrient solution to give final NaCl concentrations of 0, 100, 200, and 300 mM, respectively. The salt concentration is within the range of the habitat of *K. virginica*, which extends from Louisiana to Florida and along the Atlantic Coast to Delaware (Blits and Gallagher 1990). The pH of the nutrient solution for all treatments was 7.2. All nutrient solutions were prepared using demineralized water. Plants were watered with 25 ml NaCl solution at 3-day intervals for 24 days. The soil was salinized stepwise to avoid osmotic shock. The leachate was collected and added back to soil to maintain the salinity near the target level; distilled water was added as necessary to maintain soil moisture.

Soil samples collection

After 90 days, according to Riley and Barber (1969, 1970), whole plants were extracted from pots. The soil obtained by gently shaking roots and collected in sterilized culture dish was considered “bulk soil.” The rest soil that adhered to roots was then collected in another sterile culture dish and termed “rhizosphere soil.” Soil samples were divided into two parts; one part was stored at 4°C for biological and biochemical analyses and the other was air-dried at room temperature for chemical analysis.

Plant analyses

All shoots tissues were dried in a forced-air oven at 80°C for 72 h for biomass determination. Shoot biomass was the sum of leaves and stems. The fresh root mass of six plants was used for determining root dry weight. To assess AMF colonization, roots from three plants were cleared with 10% KOH and stained with 0.05% trypan blue (Phillips and Hayman 1970). The percentage of root length colonized by AMF was calculated as reported by McGonigle et al. (1990). The roots were cut into 1-cm segments, and 30 fragments were examined for AMF colonization by a compound microscope (100 magnifications). Positive counts for AMF colonization included the presence of vesicles, arbuscules or typical mycelium.

Soil chemical, biological, and biochemical analysis

The available P concentration of soil was determined using the sodium bicarbonate-extractable P colorimetric method (Olsen et al. 1954). Soil pH value was measured using a glass electrode and a soil to water ratio of 1:2.5 (Dick et al. 2000). EC of soil was measured with a conductivity meter (Model DDS-11A; Leizi, Shanghai, China). The pour-plate

technique was applied for estimation of Ms populations in soil (Pikovskaya 1948). The activities of soil invertase (E.C. 3.2.1.26), catalase (E.C. 1.11.1.6), urease (E.C. 3.5.1.5), and phosphatase (E.C. 3.1.3.2) were determined according to Ohshima et al. (2007), Trasar-Cepeda et al. (1999), Nannipieri et al. (1980), and Kandeler et al. (1999), respectively. The activities of soil enzymes were assayed within 1 week after sampling.

Statistical analysis

All data were statistically analyzed by analysis of variance using the SPSS software package (SPSS 10.0 for Windows 2001) and means separated by Duncan test at p values <0.05.

Results

AM colonization

Structures characteristic of AMF were not observed in roots of controls and plants inoculated with Ms alone (data not shown). Plants inoculated with Ga alone showed significantly higher root colonization (41.6%) than plants only inoculated with Gm (30%) at 100 mM NaCl, while there was an opposite result at 300 mM NaCl ($p < 0.05$, Fig. 1a). Except for 0 mM NaCl, the root colonization of plants co-inoculated with AMF (Gm and/or Ga) and Ms was significantly higher than plants inoculated with AMF alone ($p < 0.05$). Plants inoculated with Gm and Ms showed significantly higher root colonization (59.6% and 46%) than those inoculated with Ga and Ms (45.7% and 36.7%) and those inoculated with the three fungi (50.2% and 38%) at 200 and 300 mM NaCl, respectively. The percentages of root colonization declined by increasing salinity level ($p < 0.05$, Fig. 1a).

Ms populations

Both rhizosphere and bulk soils of plants inoculated with Gm and Ms showed significantly higher Ms populations than soils of other plants ($p < 0.05$) at each salinity level (Fig. 1b, c). Rhizosphere soil of plants inoculated with three fungi showed significantly lower Ms populations than rhizosphere soil of plants inoculated with AMF and Ms ($p < 0.05$, Fig. 1c). The highest Ms populations were presented in rhizosphere and bulk soils of plants inoculated with Gm and Ms at 100 mM NaCl.

Plant growth

At all levels of salinity, shoot and root dry weights of plants inoculated with AMF and/or Ms were higher than those of control plants. Root dry weights of plants inoculated with

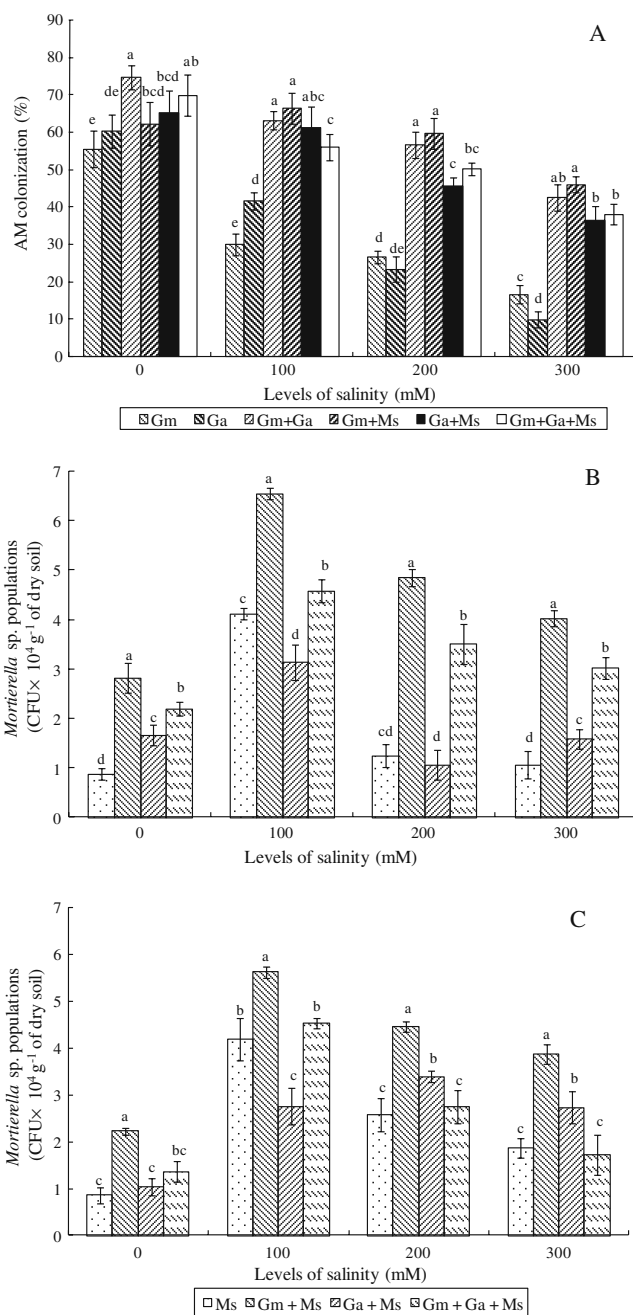


Fig. 1 Effects of arbuscular mycorrhizal fungi and P-solubilizing fungus (*Mortierella* sp.) on AM colonization (a), *Mortierella* sp. populations of bulk soil (b), and rhizosphere soil (c) at different salinities. Data are means \pm SE of three replicates. Means followed by the same letter are not significantly different at the same salinity level ($p < 0.05$) as determined by Duncan test

Ga alone were significantly higher than those of plants inoculated with Gm at 0 and 100 mM NaCl, while an opposite result was observed at 200 and 300 mM NaCl ($p < 0.05$, Fig. 2b). The inoculation of Ms significantly improved positive effects of AMF (Gm or Ga) on shoot and root dry weights under salt stress (i.e., 100, 200, and 300 mM NaCl, as well as below; $p < 0.05$, Fig. 2a, b).

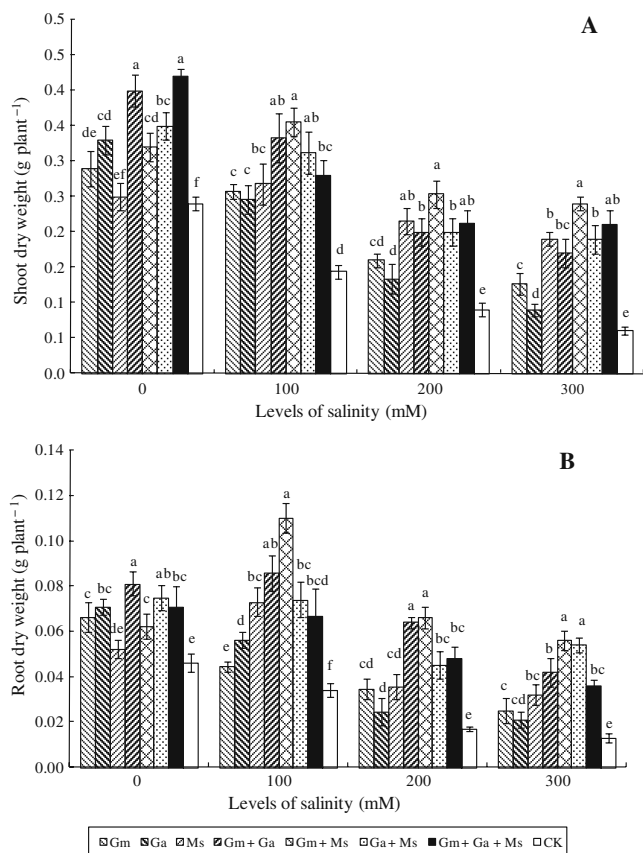


Fig. 2 Effects of arbuscular mycorrhizal fungi and P-solubilizing fungus (*Mortierella* sp.) on shoot dry weight (a) and root dry weight (b) at different salinities. Data are means ± SE of three replicates. Means followed by the same letter are not significantly different at the same salinity level ($p < 0.05$) as determined by Duncan test

Inoculation with Gm and Ms showed significant higher plant biomass than inoculation of Ga and Ms (e.g., shoot dry weights at 200 and 300 mM NaCl and root dry weights at 100 and 200 mM NaCl, $p < 0.05$).

Soil enzyme activities

All inoculation treatments enhanced urease activities of soils with respect to noninoculated controls at all salinity levels ($p < 0.05$, Fig. 3a, b). *Mortierella* sp. significantly increased urease activities of rhizosphere soil when inoculated alone or with dual AMF under salt stress ($p < 0.05$, Fig. 3b). Urease activities increased from 0 to 100 mM NaCl, and then decreased from 100 to 200 and 300 mM NaCl.

Except for inoculation with Gm and Ms, the combined inoculations enhanced invertase activities of all soils with respect to those of controls. In all soils, Ms decreased the positive effects of AMF on invertase activities, and invertase activity showed the lowest values in each salinity level in the Gm and Ms treatment ($p < 0.05$; Fig. 3c, d).

Invertase activities showed a trend similar to that of urease activities under salt stress.

All inoculation treatments at all salinity levels ($p < 0.05$) significantly increased neutral and alkaline phosphatase activities of both soils with respect to controls (Fig. 3e–h). *Mortierella* sp. significantly increased the positive effects of AMF on neutral phosphatase activity at all salinity levels ($p < 0.05$).

Catalase activities were reduced by increasing salinity; and those of both bulk and rhizosphere soils of inoculated with AMF and/or Ms were significantly higher than those of controls at all salinity levels ($p < 0.05$; Fig. 3i, j). *Mortierella* sp. significantly enhanced the positive effects of Gm at 300 mM NaCl, Ga at 0 and 300 mM NaCl, and Gm plus Ga at 200 and 300 mM NaCl on catalase activities in bulk soil whereas it had no effect in rhizosphere soil ($p < 0.05$, Fig. 3i).

Available P concentrations

Except for 0 mM NaCl, single inoculation with Ms significantly increased available P concentrations of all soils with respect to single inoculation with AMF and controls ($p < 0.05$; Fig. 4a, b). In all soils at each salinity level, inoculation with Gm and Ms significantly increased available P concentrations with respect to single inoculation with Gm or Ms and dual inoculation with Ga and Ms ($p < 0.05$). The highest available P concentrations were recorded when plants were inoculated with Gm and Ms at 100 mM NaCl. Bulk soils from treatments involving triple inoculation had significantly higher available P concentrations than those of dual inoculation with Ga and Ms. Similar results were observed in rhizosphere soil only at 100 mM NaCl ($p < 0.05$).

Soil pH and electricity conductivity

Except for the 300 mM NaCl treatment, in bulk soil, plants inoculated with Gm and Ms showed significantly lower pH values than those of other treated soils ($p < 0.05$, Fig. 5a). The pH values of all inoculated soils were lower than those of control soils at each salinity level (Fig. 5a, b). The pH values of rhizosphere soil declined by increasing salinity.

Electrical conductivities of all rhizosphere or bulk soil were significantly higher than those of control soils at all salinity level ($p < 0.05$; Fig. 5c, d). The highest EC values of soils were recorded when plants were inoculated with Gm and Ms at all salinity level. For all salinity treatments, bulk soil of plants inoculated with Ms showed significantly lower EC values than those of other inoculated soils ($p < 0.05$, Fig. 5c). A similar effect was observed in rhizosphere soil.

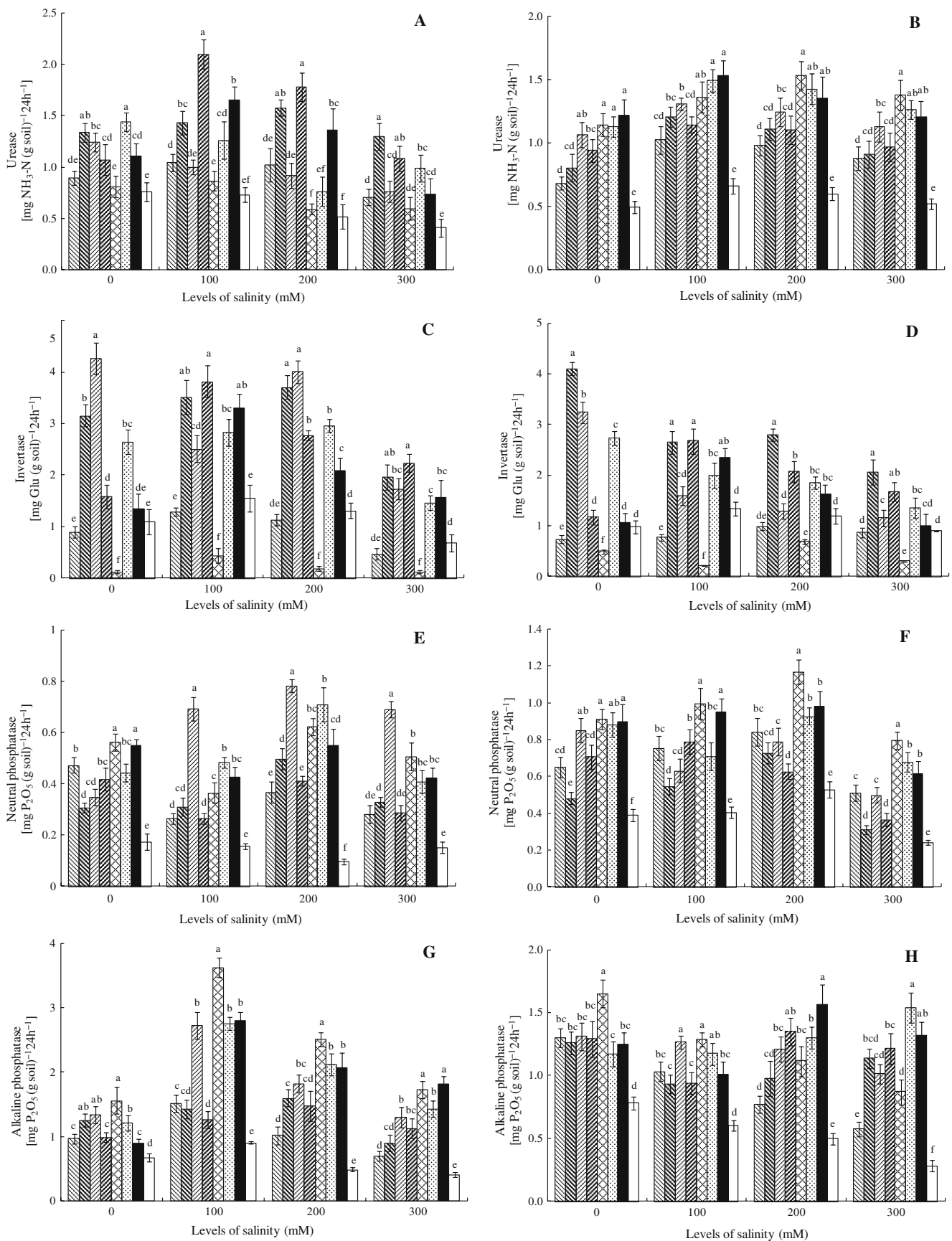


Fig. 3 Effects of arbuscular mycorrhizal fungi and P-solubilizing fungus (*Mortierella* sp.) on soil enzyme activities at different salinities. Urease (a), invertase (c), neutral phosphatase (e), alkaline phosphatase (g), and catalase (i) activities of bulk soil; urease (b), invertase (d), neutral phosphatase (f), alkaline phosphatase (h), and catalase (j) activities of rhizosphere soil. Data are means ± SE of three replicates. Means followed by the same letter are not significantly different at the same salinity level ($p < 0.05$) as determined by Duncan test

Discussion

Interaction between AMF and Ms under salt stress

Mycorrhizal symbiosis acts as a key component in helping plants to cope with adverse environmental condition (Giri et al. 2003), but the addition of various salts to soils inhibits hyphal growth with a subsequent decrease in the mycorrhizal colonization of plants (Ruiz-Lozano and Azcón 2000). We have shown that salt stress significantly inhibited AMF colonization of *K. virginica* roots, probably because salt inhibited germination of AMF spore (Figs. 1 and 6), colonization of the plant roots by the fungi and formation of functional mycorrhizal structures within the roots (Juniper and Abbott 2006). The mycorrhizal root colonization under salt stress promoted by combined inoculation of AMF and Ms, confirmed the finding of Garbaye and Bowen (1987) in *Pinus radiate*. The enhanced mycorrhizal root colonization by inoculation with Ms may be attributed to phytohormone production to the increase of soil soluble P promoted by Ms in saline soil (Figs. 2 and 6, Jacobsen et al. 1992; Toro et al. 1996). Inoculation with AMF and Ms also significantly increased Ms populations of soil, probably because mycorrhized plants released carbonaceous substances used as a C source by PSM in the rhizosphere soil (Linderman 1988; Azcon and Barea 1996; Singh and Kapoor 1999). However, plants inoculated with three fungi showed significantly lower mycorrhizal colonization and Ms pop-

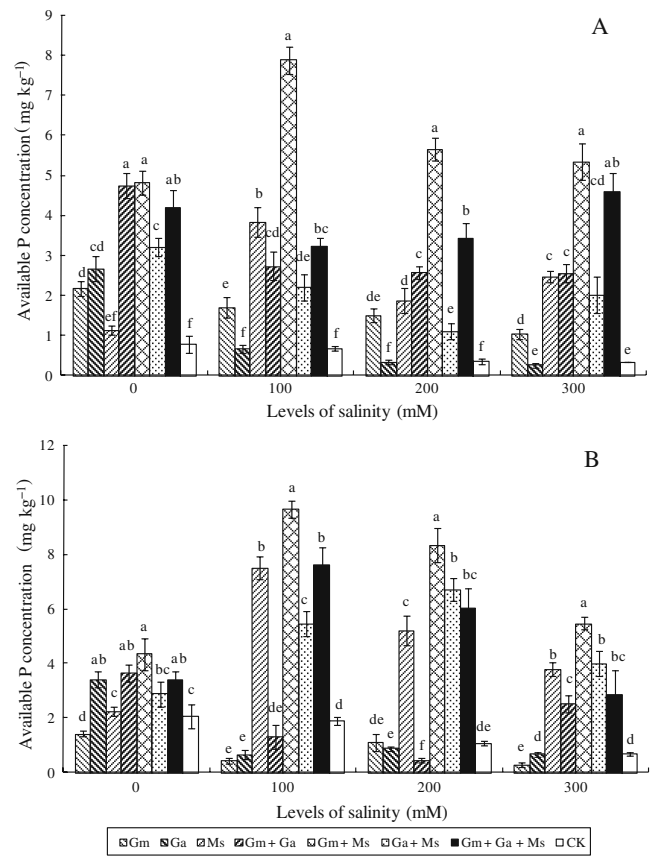


Fig. 4 Effects of arbuscular mycorrhizal fungi and P-solubilizing fungus (*Mortierella* sp.) on available P concentrations in bulk soil (a) and rhizosphere soil (b) at different salinities. Data are means ± SE of three replicates. Means followed by the same letter are not significantly different at the same salinity level ($p < 0.05$) as determined by Duncan test

ulations in rhizosphere soil than plants inoculated with Gm and Ms. This may be due to the competition among microorganisms for root exudates or nutrients under salt stress. It is important to underline that the used method for

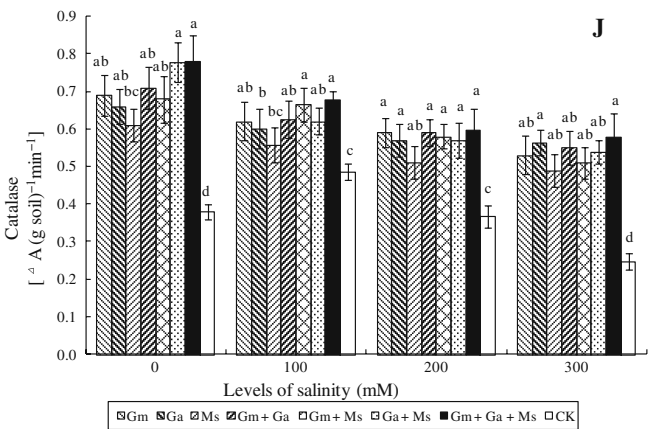
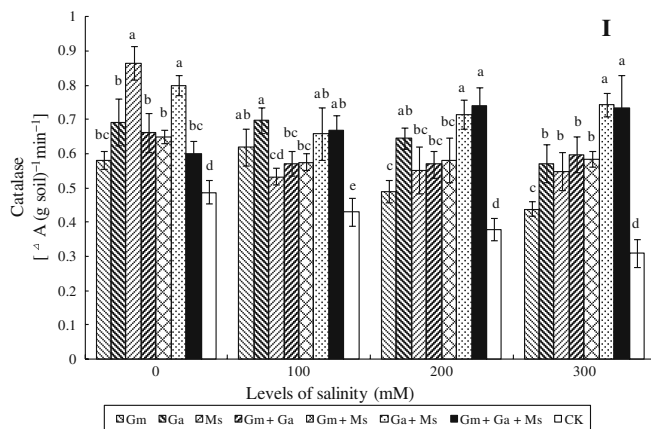


Fig. 3 (continued)

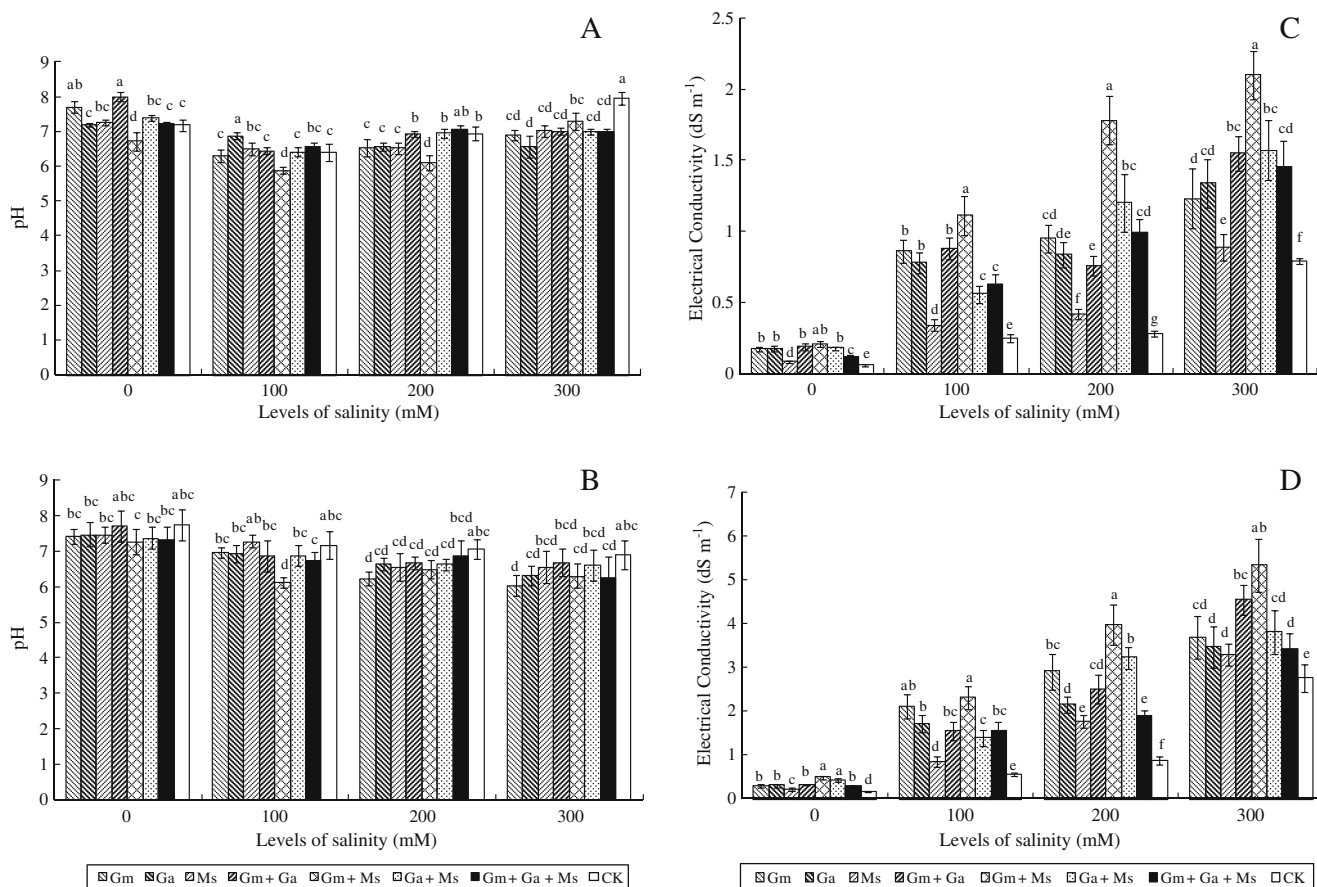


Fig. 5 Effects of arbuscular mycorrhizal fungi and P-solubilizing fungus (*Mortierella* sp.) on pH and electrical conductivity of soil at different salinities. pH (a) and electrical conductivity (c) of bulk soil; pH (b) and electrical conductivity (d) of rhizosphere soil. Data are

means \pm SE of three replicates. Means followed by the same letter are not significantly different at the same salinity level ($p < 0.05$) as determined by Duncan test

isolating PSM and determining Ms populations is based on the use of tricalcium phosphate (Pikovskaya 1948), and this method has been criticized by Collavino et al. (2010).

Effect of inoculation on plant growth under salt stress

Inoculation with AMF improves growth of plants under a variety of salt stress conditions (Al-Karaki et al. 2001; Giri et al. 2003; Sannazzaro et al. 2006). We have observed the same for shoot and root dry weights of *K. virginica* inoculated with AMF and/or Ms at all salinity levels. The dual inoculation with AMF and PSF had positive effects on alfalfa (Piccini and Azcon 1987), wheat (Singh and Kapoor 1999), and *Lactuca sativa* (Kohler et al. 2007). Osorio and Habte (2001) suggested a synergistic interaction between PSM and AMF. Although Ga had higher colonization rates than Gm at 100 mM NaCl, plants inoculated with Gm showed a better resistance to salt stresses than those colonized by Ga. This may partly explain why the highest shoot and root biomasses were obtained when plants were inoculated with Ms and Gm. Plants inoculated with three

fungi had significantly lower root dry matter than plants inoculated with Gm and Ms, probably due to the lower mycorrhizal root colonization of the former plants. However, inoculation of AMF and Ms may increase root development and nutrient uptake, and this may decrease root/shoot ratios (Kothari et al. 1990; Berta et al. 1995).

Changes of soil enzyme activities with inoculation under salt stress

Eizavi and Tabatabai (1977) had reported that acid phosphatase is predominant in acid soils and alkaline phosphatase is predominant in alkaline soils. Therefore, we have only determined neutral and alkaline phosphatase activities since soil pH was 7.3. Catalase (Frankenberger and Bingham 1982), alkaline phosphatase (Garcia and Hernandez 1996), invertase, and urease (Omar et al. 1994) activities were inhibited by increasing soil salinity. Similarly, we have observed that soil enzyme activities of control soil decreased proportionately by increasing salinity. The reduced enzyme activities may depend on the

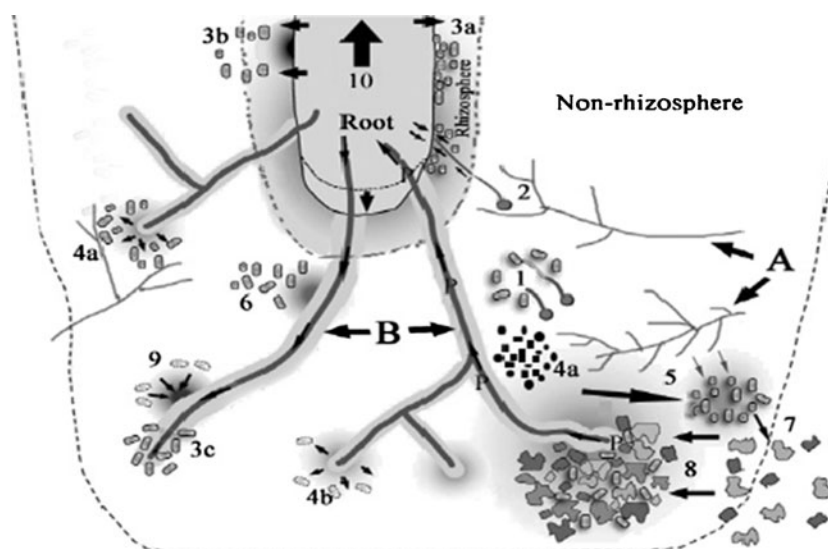


Fig. 6 Schematic view of possible mechanism of co-inoculation of AMF and P-solubilizing fungus in saline soil. *A* hyphae of *Mortierella* sp.; *B* hyphae of AMF; 1 inhibition of salt on AMF growth and colonization; 2 promotion of *Mortierella* sp. on AMF growth and colonization; 3 changes in root structure: 3a stimulatory or 3b inhibitory compounds present in root exudation, 3c supply of

energy-rich carbohydrates via AM fungi hyphae; 4 promotion (4a) or inhibition (4b) of microbial activity; 5 stimulation of soil enzyme activities; 6 pH changes; 7 effects on insoluble P; 8 uptake of soluble P; 9 uptake of other nutrients; 10 stimulation of plant (Fig. 5 is modified from Jonas et al. 2004)

decrease in enzyme synthesis due to the decrease in abundance and activity of soil microorganisms and/or to the “salting-out” effects on enzyme proteins (Omar et al. 1994). Microbial inoculation promoted neutral phosphatase, alkaline phosphatase, urease, and catalase activities in both bulk and rhizosphere soils (Figs. 5 and 6). Dodd et al. (1987) and Ezawa and Yoshida (1994) also showed that phosphatase activities of rhizosphere soil of AMF colonized plants were higher than those of rhizosphere soil of noncolonized plants. Microbial inoculation may alleviate the negative effects of increasing concentration of salt solutions on enzyme activities, probably because both AMF (Allen et al. 1981) and Ms inoculations increased soil microbial biomass. However, further research is required to understand the relative mechanisms.

In bulk soil, inoculation with Ms significantly increased the positive effects of AMF on neutral phosphatase, alkaline phosphatase, and catalase activities under salt stress, while invertase and urease activities were reduced. The effects of AMF on neutral phosphatase and urease activities were enhanced by Ms inoculations in rhizosphere soil, but reduced on invertase activities. It is difficult to explain the different responses of enzyme activities of bulk and rhizosphere soils inoculated with PSF and AMF, because the measured enzyme activities depend on the contribution of both extracellular and intracellular enzyme activities (Nannipieri et al. 2003), and stable extracellular enzyme activities are associated with soil colloids and persist even in harsh environments that would limit intracellular microbiological activity (Nannipieri et al. 2002).

Osorio and Habte (2001) reported that acid production was the major mechanism in the solubilization of rock phosphate by the Ms, and the pH value of the growth medium decreased as a result of acid production by PSM [Fig. 6 (6)]. We have shown that pH values of all inoculation treatments were lower than those of controls at each salinity level. Soil enzyme activities can be increased, decreased, or not affected by soil acidity (Kang and Freeman 1999; Acosta-Martinez and Tabatabai 2000). In addition, AMF can modify quantity and quality of root exudation (Mar et al. 2000).

Changes of available P concentrations and electricity conductivity with inoculation

Arbuscular mycorrhizal fungi can absorb P from the soil solution, as plant roots do, and there is no evidence of their ability to solubilize insoluble soil P minerals (Bolan 1991; Cardoso et al. 2006). However, we have shown that inoculation with Gm significantly increased available P concentrations in bulk soil and we are not able to explain the relative mechanisms. As discussed before, the use of tricalcium phosphate for determining PSF or PSB has been criticized (Collavino et al. 2010). The type of phosphate, the analytical procedure for determining the solubilized P, the incubation conditions, and medium are all important factors affecting P solubilization (Ogut et al. 2010). Soils inoculated with Gm and Ms had significantly higher available P concentrations than soils only inoculated with Gm or Ms. Probably, the effectiveness of inoculated PSF

can be stimulated by the mycorrhiza (Tarafdar and Marschner 1995; Osorio and Habte 2001). The P solubilization occurs in the rhizosphere soil where organic compounds are released by roots or mycorrhizae (Nguyen 2003; Amos and Walters 2006). Many rhizosphere microorganisms are heterotrophs and might use these organic substrates to produce organic acids, which would increase P concentrations in solution (Hameeda et al. 2006; Reyes et al. 2006). Probably Gm was more effective than Ga in stimulating P solubilization by Ms, but we are unable to explain the underlying mechanisms. When plants were inoculated with three fungi, however, there were no significant effects on available P concentrations in rhizosphere soil. Probably, changes in quantities and compositions of rhizodepositions and microbial secretions, which would affect the activities of PSM in solubilizing insoluble inorganic or organic phosphate, were responsible for the different response with three inoculants. Single inoculation with Ms did not significantly increase available P concentrations of the bulk and rhizosphere soils compared to single inoculation with AMF at 0 mM NaCl. Probably, Ms was not activated by the nonsaline conditions, being isolated from saline soil of seashore. The result that the highest Ms populations were observed at 100 mM NaCl indirectly confirms the above assumption. It is conceivable that 100 mM NaCl was the most effective saline treatment in stimulating Ms activities.

All inoculation treatments significantly enhanced soil EC. On the contrary, Giri et al. (2003) and Buwalda et al. (1983) reported that inoculation of mycorrhiza significantly reduced soil EC. Our results can depend on the fact that mycorrhized plants decreased the uptake of Na (Al-Karaki 2006; Colla et al. 2008).

Conclusion

The dual inoculation of arbuscular mycorrhizal fungi and *Mortierella* sp. significantly affected plant growth, but the triple inoculation was less effective. The increased enzyme activities and available P of soil after inoculation with arbuscular mycorrhizal fungi and *Mortierella* sp. may alleviate the negative effects of increasing salinity on plant growth. To obtain the maximum benefit for plant growth, it is needed to select the right combination of arbuscular mycorrhizal fungi species and phosphorous-solubilizing microorganisms to be inoculated in saline soils. However, further research is needed to understand the underlying mechanisms.

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