

Comparative study of hydroxyapatite and octacalcium phosphate coatings deposited on metallic implants by PLD method

W. Mróz · A. Bombalska · B. Budner · S. Burdyńska · M. Jedyński · A. Prokopiuk ·
E. Menaszek · A. Ścisłowska-Czarnecka · A. Niedzielska · K. Niedzielski

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Abstract The aim of the presented study was an analysis of two apatite coatings: hydroxyapatite (HA) and octacalcium phosphate (OCP) as coatings materials for metallic implants. Both layers were deposited by means of the PLD method. As a target material, synthetic, powdered and pressed hydroxyapatite was used. HA was deposited on 316L steel substrate in two temperature ranges for obtaining different coatings: $150 \pm 30^\circ\text{C}$ and $430 \pm 30^\circ\text{C}$ for OCP and HA, respectively. As an intermediate layer, the nanocrystalline diamond layer (NCD) was deposited. Examined calcium phosphate layers were tested for adhesion of osteoblast cell culture (MG-63). Analytical methods (AFM, FTIR) showed the usefulness of the PLD method for deposition of the apatite layers on metallic implants. Both examined layers showed biocompatibility with human osteoblast

cells and presented favorable conditions for their proliferation.

1 Introduction

Hydroxyapatite (HA) and other calcium phosphate ceramics (Ca–P) are widely used in medicine as coating materials for metallic implants because of their osteoconductive properties and biocompatibility with surrounding tissues.

The widest application, in coating of the metallic implants, was found for two phases of calcium phosphates: hydroxyapatite (HA) and octacalcium phosphate (OCP). OCP, in biological environment, is a precursor of HA and takes part in early stages of tissue calcification process [1–4]. In the past few years, OCP has drawn special attention as an alternative material to HA. In biological environment, it is more soluble than HA and as a coating material it would present faster resorption process and stimulation of the growth of the new bone [5–7]. In vitro tests showed correct adhesion, morphology and proliferation of the osteoblasts and fibroblasts on OCP coating [8]. Comparison of the materials composed of OCP, HA and Mn-HA showed positive in vitro results. In addition, the experiments also showed potential application of OCP as a biomimetic material for implant coating [9].

Between a metallic implant and an apatite layer, a transition layer is being introduced. Its role is to combine the materials with different structural and mechanical properties. One of these materials is nanocrystalline diamond (NCD). NCD layers turned to be highly resistive to bacteria colonization as compared to medical steel or titanium, which has an important advantage considering biomedical application of this material [10], especially in implantology where implant infection might lead to its rejection. NCD is applied

W. Mróz · A. Bombalska (✉) · B. Budner · S. Burdyńska ·
M. Jedyński · A. Prokopiuk
Institute of Optoelectronics, Military University of Technology,
Gen. S. Kaliskiego str. 2, 00-908 Warsaw, Poland
e-mail: abombalska@wat.edu.pl
Fax: +48-666-89-50

E. Menaszek
Department of Cytobiology and Histochemistry,
Jagiellonian University, Medyczna 9 str., 30-688 Cracow, Poland

A. Ścisłowska-Czarnecka
Department of Anatomy, Academy of Physical Education,
Al. Jana Pawła II 78 str., 31-571 Cracow, Poland

A. Niedzielska
Faculty of Mechanical Engineering, Technical University
of Łódź, Żeromskiego 116 str., 90-924 Lodz, Poland

K. Niedzielski
Department of Orthopaedics, Polish Mother Memorial Hospital,
Medical University, Rzgowska 281/289 str., 93-338 Lodz, Poland

in covering surgical tools in stomatology, cardiology, orthopaedics, vascular disease and also in biosensors in tissue engineering and implantology [11–18].

Comparative analysis of two apatite materials was the aim of the present study. Both materials, deposited by the PLD method, were characterized before and after cell culturing with use of AFM imaging. Also the adhesive properties were tested with the CV test, and the influence on growth and proliferation of human osteoblastic cells MG-63 was estimated.

2 Material and methods

2.1 Coatings deposition

The calcium phosphate layers were deposited by means of an excimer ArF laser operating at 193 nm and pulse repetition of 5 Hz. The laser pulse duration was 20 ns (FWHM), the laser fluence was about 7 J/cm² with pulse energy of 300 mJ. The substrate surface was parallel to target surface, and the distance between them was 60 mm in the case of HA and 50 mm in the case of OCP. In the case of HA, the substrate temperature was 430 ± 30°C and in the case of OCP the substrate temperature was 150 ± 30°C. The experiment was carried out with water vapor as a reactive atmosphere under a pressure of 30 ± 5 Pa. All layers were deposited on 316L stainless steel substrates with a 300 nm nanocrystalline diamond (NCD) buffer layer. The NCD layers were deposited by RF PACVD method described by Mitura et al. [19].

2.2 Cell culture

For the in vitro study, osteoblast-like cells MG-63 were seeded at the initial density of 7 × 10³ cells per one milliliter of the medium, DMEM culture medium (PAA, Austria) supplemented with 10% fetal bovine serum, 1% L-glutamine and antibiotics: penicillin (100 IU/mL) and streptomycin (100 µg/mL). As a control tissue, culture polystyrene plates (TCPS) were used. Cultures were grown for 3 or 7 days at 37°C in a 5% CO₂ and 95% air atmosphere. After 3 and 7 days of incubation, cell adherence test was performed. Estimation of adherent cell mass was tested by crystalline violet absorption test, the CV test [20, 21]. For the CV test, performance cells cultured on studied materials were washed twice with PBS, fixed in 2% paraformaldehyde for 1 h, stained with 0.5% crystal violet in 20% methanol for 5 min, washed with water and dried. The dye was extracted from the cells with 100% methanol (POCh, Poland) and optical density (OD) was measured at 570 nm. The absorbance is proportional to the cell mass, which adhered on the surface; the stronger the signal observed the greater the degree of the adherence.

3 Results

3.1 Surface characterization after deposition process

The phase composition of the obtained coatings was analyzed by means of an FTIR spectrometer (Perkin Elmer Spectrum GX) with a resolution of 4 cm⁻¹. The spectra were summed 10 times in order to improve the signal-to-noise ratio. The topography of deposited layers was imaged by using AFM (Veeco Multimode IV SPM).

After deposition the OCP layer was characterized by well developed structure with round shaped grains having size of about 200 nm. The height of the grains is diverse (Fig. 1), visible grains are homogeneously distributed over the surface. Phase composition pictured by AFM (Fig. 1(b)) shows one material phase. HA layer is composed of smaller grains as compared to OCP layer (Fig. 2).

Size of the grains is about 100 nm and their distribution is also morphologically homogeneous. In this case, the shape of grains is irregular. Phase composition indicates that more than one material phase is present in examined samples with overwhelming majority of HA phase.

Figure 3 depicts the absorption spectra (FTIR) of HA and OCP. The bands corresponding to PO₄ group consist of two

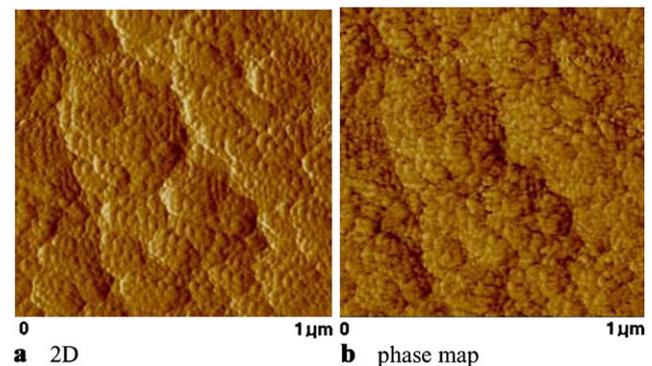


Fig. 1 OCP deposited on 316L stainless steel with NCD buffer layer: (a) 2D AFM image, (b) phase map

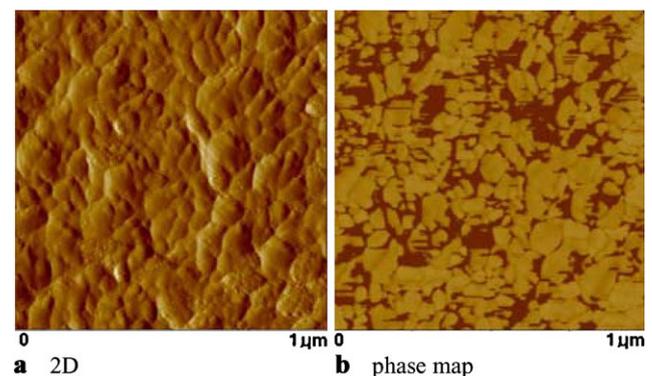
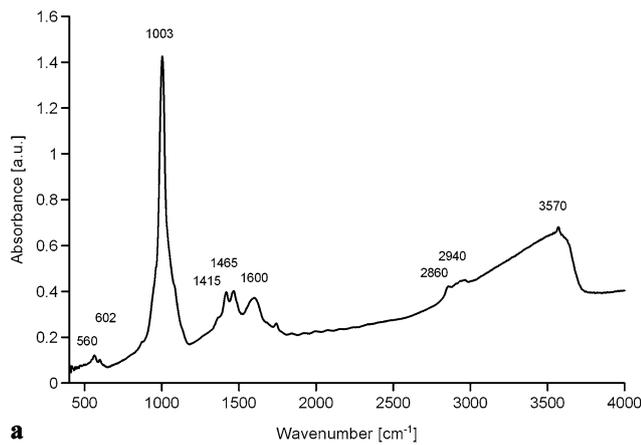
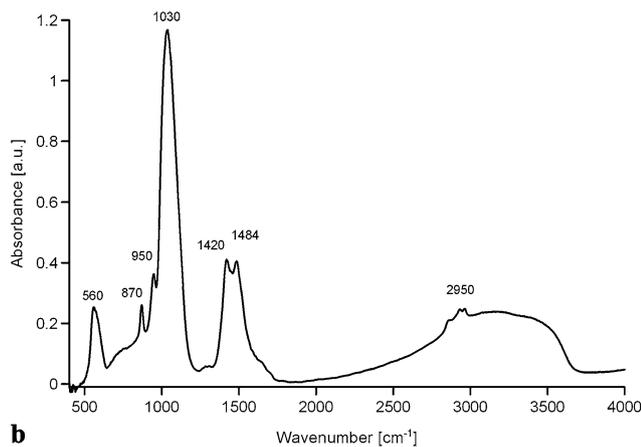


Fig. 2 HA deposited on 316L stainless steel with NCD buffer layer: (a) 2D AFM image, (b) phase map



a



b

Fig. 3 Fourier spectra of deposited calcium phosphate layers: (a) HA, (b) OCP

main regions. The first one is characterized by two peaks at 560 and 602 cm^{-1} , which reflect a bending mode. The second region is between 900 and 1200 cm^{-1} . The peaks at 560 and 602 cm^{-1} are visible in the case of the HA layer; nevertheless, their intensity is low (Fig. 3(a)). In the case of OCP, the peaks at 560 and 602 cm^{-1} are higher and broadened, forming one band with a maximum at 560 cm^{-1} (Fig. 3(b)). This peak is higher than in the case of HA. The second region of PO_4 mode in the HA spectrum has only one narrow peak at about 1003 cm^{-1} . In the case of OCP, this region consists of two peaks attributed to PO_4 vibrations, namely at 950 and 1030 cm^{-1} . The peak at 1030 cm^{-1} is visibly broadened and shifted towards higher wavenumbers (Fig. 3(b)). Moreover, the thin nature of PO_4 peak at 1003 cm^{-1} indicates that HA layer has polycrystalline structure and OCP is characterized by a less textured and ordered structure.

3.2 Surface characterization after cell culture

Surface of examined layers is visibly reconstructed both after 3 and 7 days of culturing. The HA surface is macrogranu-

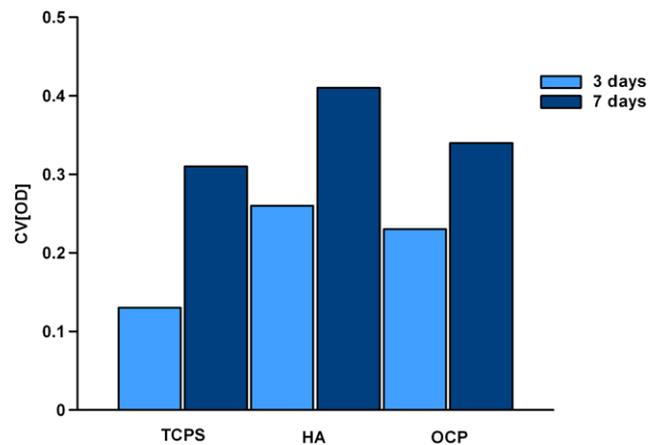


Fig. 4 Adhesion of the osteoblastic cells to the HA and OCP surface

lar, built out of triangular formations with material phase different than the initial one. These observations suggest biomimetic growth. In case of OCP, the surface is microgranular with uniform structure. Fine grains, uniformly distributed on the implant surface, are built out of one material phase.

Based on the CV test one can conclude that osteoblasts cultured on HA and OCP after 3 days of culturing possess similar adherence, substantially better compared to TCPS (Fig. 4). After 7 days of culturing, osteoblastic cells present better adhesion to HA than to OCP. Cells on both materials present correct morphology.

4 Summary

The PLD method was used for apatite layer deposition. Apatite layers were deposited from the same target material, namely hydroxyapatite. The FTIR and AFM diagnostics of the deposited layers characterized them as continuous, uniform structures with fine grains. For OCP, only one material phase was observed, while for HA, AFM phase image showed two, with HA as a major one. Both layers were appropriate for osteoblast cell culture. Both layers were visibly reconstructed. From macroscopic observations, based on topography analysis and the CV test, one can conclude that cells showed good adhesion to the surface. Cell adhesion to both apatites is similar after three days of culturing; however, after 7 days, effects are more diverse and optical density is higher for the HA material, pointing that this material is characterized as having better adhesion than OCP and TCPS.

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