

## Treatment of rheumatic diseases and hepatitis B virus coinfection

Anna Felis-Giemza · Marzena Olesińska ·  
Katarzyna Świerkocka · Ewa Więsik-Szewczyk ·  
Ewa Haładyj

Received: 26 September 2014 / Accepted: 17 December 2014 / Published online: 31 December 2014  
© The Author(s) 2014. This article is published with open access at Springerlink.com

**Abstract** We often encounter rheumatological patients coinfecting with hepatitis B in daily practice. In this paper, we will discuss the basic characteristics of the virus of hepatitis B, course of infection, the safety of rituximab, tocilizumab, abatacept treatment and therapeutic recommendations in management of patients with rheumatic diseases.

**Keywords** Rituximab · Tocilizumab · Abatacept · Biological therapy · Hepatitis B · Rheumatic diseases

### Introduction

The introduction of biological agents significantly increased the efficiency of the treatment of rheumatic diseases and limited permanent damage to organs in the course of the inflammatory process. Particular efficacy is achieved by depletion of B cells the monoclonal anti-CD20 (rituximab), antagonism of interleukin-6 (IL-6R) (tocilizumab) and inhibition of T cell activation molecule, CTLA-4-Ig (abatacept). These methods allow for the effective treatment of a growing number of patients with active forms of rheumatoid arthritis (RA), juvenile idiopathic arthritis (JIA), psoriatic arthritis (PsA) and ankylosing spondylitis (AS).

Coexistence of severe rheumatoid disease with chronic viral hepatitis (hepatitis) type B makes the treatment of abatacept, tocilizumab or rituximab risky because it can lead to reactivation of viral infection.

Knowledge of areas endemic for hepatitis infections, increased risk groups, the natural history of infection and the impact of the treatment of rheumatic diseases on the course of hepatitis increases the effectiveness and safety of treatment in patients eligible for the treatment of biological agents.

### Hepatitis B

Hepatitis B virus (HBV)

HBV belongs to the family Hepadnaviridae and comprises a circular double-stranded DNA and DNA polymerase surrounded by the core antigen (HBcAg) [1]. The HBcAg consists e antigen (HBeAg). HBcAg is surrounded by a shell containing the lipoprotein surface antigen (HBsAg) [1, 2].

Important features of HBV infection affecting the course of HBV DNA are the ability to integrate into the DNA of infected hepatocytes and the replicative capacity. It is estimated that the chronically infected blood stream is expelled section approximately 1011 viral particles. Of particular note is the fact that the HBV reverse transcriptase has a low error correction capability of reading, resulting in a high rate of mutation [3].

Areas endemic for HBV infection are a southeast Asia, Africa and other regions of the world outside North America and Western Europe, and Australia [7]. In Poland, 1.5 % of the population is chronically infected with HBV, which is about 700 thousand [8].

### Course of HBV infection

HBV infection can occur in a variety of clinical forms—from asymptomatic, by explicitly acute viral hepatitis,

A. Felis-Giemza · M. Olesińska · K. Świerkocka ·  
E. Więsik-Szewczyk · E. Haładyj (✉)  
Institute of Rheumatology, Connective Tissue Department,  
1 Spartanska Street, Warsaw, Poland  
e-mail: ehaladyj@o2.pl

chronic inflammation, which can result in cirrhosis and liver cancer [1, 2, 4]. The majority of adult patients (>95 %) of host defense mechanisms allow for total eradication HBV [12].

There are the following phases of HBV infection [5, 6]:

1. Immune tolerance phase—due to the low immune response, aminotransferase levels are low, and inflammatory changes in liver histopathology not significantly increased despite a high viral load of HBV DNA (106–109 copies/ml). If this occurs during the integration of HBV DNA with the host DNA in hepatocytes, the likelihood of HBV eradication decreases—both spontaneous and induced by treatment [12];
2. Immune clearance phase—active immune response against HBV-infected hepatocytes, leading to an elevated aminotransferase levels and severe necro-inflammatory changes in liver histopathology. HBs antigen is always present, and HBe antigen—positive or negative—antibodies to HBe may be present. HBV DNA viral load is moderately high (105–107 copies/ml). The majority of patients in this phase (about 90 %) present a spontaneous loss of HBeAg antigen and the appearance of antibodies to HBeAg, called seroconversion [12];
3. Immune control phase—after HBeAg seroconversion antibodies to HBe are present, HBV DNA replication is low (<105 copies/ml), transaminase activity is normal or slightly increased, and the histopathological changes in the liver are absent or minimal;
4. Phase reactivation of infection—dominate the mutant HBV DNA, which do or emit a small amount of HBe antigen. The observed increase in serum HBV DNA viral load, increased transaminases and liver biopsies are present necro-inflammatory changes.

In the initial period of hepatitis B infection, which lasts several weeks, HBV remains hidden from the immune system of the host. Then, a strong innate (NK cytotoxic lymphocytes and NKT—natural killer—natural killer T) and acquired (mainly CD8 + T cells) immune response is directed against infected hepatocytes secreting antigens HBV [9]. The virus removal process involves many cytokines, such as interferon- $\alpha$  (IFN- $\alpha$ ), interferon- $\beta$  (IFN- $\beta$ ), interferon- $\gamma$  (IFN- $\gamma$ ), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) or interleukin-6 (IL-6) [9–11].

#### Treatment of hepatitis B

Currently, the treatment of chronic hepatitis B, the classical recombinant IFN- $\alpha$  and its pegylated form, and nucleoside and nucleotide analogs, such as lamivudine, adefovir, tenofovir and entecavir, result in the suppression of

HBV replication. In case of the selection of strains resistant to lamivudine, adefovir, tenofovir or entecavir may be used [8].

#### HBsAg carrier state

After HBeAg seroconversion, patients can go in HBV carrier state. At this stage, the disease is characterized by the presence of HBsAg in the serum, low or undetectable levels of HBV DNA, normal aminotransferase activity and lack HBeAg [13]. Fewer than 1 % of inactive HBV carriers per year will undergo spontaneous seroconversion HBsAg [14]. In the carriers of HBsAg, the use of immunosuppressive therapies can lead to reactivation of infection.

#### Hidden HBV infection

Hidden HBV infection can be described as the presence of HBV DNA in patients without HBsAg, and with or without anti-HBc and/or anti-HBs. In patients with the presence HBV and HBsAg is undetected, HBV DNA viral load is low, even of the order of several virus copies per milliliter of plasma or liver tissue, impossible to detect with a serological test and detectable by molecular tests. Another reason for latent HBV infection may be a mutant HBV infection, which is not recognized by the serological test.

#### HBV reactivation

This reactivation of HBV hepatitis occurs during immunosuppressive therapy or shortly after its cessation, with concomitant increase in the levels of HBV DNA ( $\geq$ tenfold increase) or the absolute height of more than 9 log 10 copies/ml or sudden twofold increase in the activity of alanine transaminase (ALT) baseline and fivefold with respect to the upper limit of normal value or greater than 300 IU/L. It needs exclusion of delta virus superinfection, acute infection with HAV, HCV, HEV, toxic hepatitis and other liver diseases of other than viral aetiology [15, 16]. To monitor HBV reactivation, the level of HBV DNA and ALT activity should be monitored. The increase in HBV viral load always precedes an average of 2–3 weeks growth of ALT activity [17]. Reactivation of infection may occur without any clinical signs (only increase of HBV DNA, the presence of HBsAg), reveals as increase in liver transaminases, or even jaundice until the fatal fulminant hepatitis patients [18].

Immunosuppressive agents by inhibiting host immune responses lead to enhanced replication of HBV in the liver and to enhanced expression of the viral antigenic epitope [19]. Hepatitis may occur both during immunosuppressive therapy and thereafter, because of the renewal of the immune response against viral infected hepatocytes

[20]. Also the use of glucocorticoids is associated with an increased risk of HBV reactivation [17].

Glucocorticoids (GKS) are well-known risk factor for infection by inducing immunosuppression. The degree of suppression of the immune system increases with dose and duration of treatment. For the treatment of longer than 2 weeks, the dose above 20 mg/day of prednisolone or its equivalent is generally considered clinically significant to induce immunosuppression [37], but has been shown that the cumulative dose of 500 mg or below the average daily dose of less than 10 mg of prednisolone does not increase the risk of infectious complications, and can be stated that are potent immunosuppressive [38]. GKS are usually served with disease-modifying drugs (DMARDs) or TNF- $\alpha$  inhibitors in the treatment of rheumatic diseases, so that data on monotherapy are limited [39]. GKS influence on HBV reactivation cannot be denied. In 30 of 144 patients (one with a history of infection and the other with chronic HBV), HBV reactivation occurred after a median time of 9.8 months. HBV markers in serum were determined before steroid therapy in 67 patients, of whom one was a history of HBV infection and the remaining patients had chronic infection. In addition, Bae et al. [39, 40] reported a case of fatal hepatitis B, and it was a reactivation of infection during long-term, low-dose treatment with GKS of inactive carrier of HBV. Most of the patients treated in combination with glucocorticosteroids DMARDs (sulfasalazyna and chloroquine), but HBV reactivation was mainly associated with glucocorticoid therapy. There are two described cases [38, 39] of patients with RA-treated GKS alone. None of the patients was described prophylactic antiviral therapy. In both cases, HBV reactivation occurred in 5 and 9 months.

#### Non-biological disease-modifying drugs (nb-DMARD)

Based on previous data, it can be concluded that the non-biological DMARDs treatment of patients with serological profile HBsAg (+)/HBsAg (–) and anti-HBc (+) may increase the risk of reactivation of HBV. Non-biological DMARD therapy is relatively safe in patients with a low risk of reactivation—a low level of HBV DNA, untreated GKS, anti-HBs (+) and the use of prophylaxis in these patients are not recommended. In the group of 224 patients treated with nb-DMARDs, 192 underwent HBV infection and 32 had diagnosed with chronic hepatitis B [38]. Only four patients with chronic hepatitis B received prophylactic antiviral therapy. HBV reactivation was observed in 10 patients (4.46 %) with a median follow-up of 13.2 months [42–46]. Methotrexate (MTX) was administered to eight (out of ten) in patients with HBV reactivation [42–46]. Antiviral therapy was started after HBV reactivation. Efavirenz was the preferred drug in most of the patients. In one case, death of the patient with fulminant hepatitis

after a 28-month follow-up was reported, despite treatment with IFN- $\beta$ . There are also isolated reports of HBV reactivation during treatment MTX [41, 49–52], leflunomide [44], azathioprine [52], chloroquine [53] and sulfasalazine [44]. At the same time, there are early reports on the role of sulfasalazine in the intensification of apoptosis of cells secreting antigen HBV [48]. There is no evidence of HBV reactivation in patients treated with gold agents and d-penicillamine.

#### Biological DMARDs

##### Anti-TNF $\alpha$ agents

TNF- $\alpha$  molecule is a homotrimer, consisting of three units of a weight of 17 kDa each [57]. TNF- $\alpha$  in RA patients is synthesized in the joints synovium, blood vessels and articulate pannus [58]. High levels of TNF- $\alpha$  in joint space stimulate osteoclasts, which lead to bone resorption [58]. The use of biologics in RA patients coinfecting with HBV or HCV is not fully secure. Although TNF- $\alpha$  inhibitors did not show hepatotoxicity, their use carries the risk of activation or exacerbation of hepatitis [59].

TNF- $\alpha$  inhibits the replication of HBV DNA. Low concentrations of TNF- $\alpha$  abolish its desired biological effect. The lack of balance between the concentration of TNF- $\alpha$  and interferon- $\gamma$  may reduce the effectiveness of the elimination of the HBV [60–62].

It is believed that inhibition of TNF- $\alpha$  decreases the inflammatory cell response against a viral infection, and viral replication is extended and prevents the destruction of infected cell [63].

The risk of reactivation of HBV during therapy with anti-TNF- $\alpha$  increases. Therapy with anti-TNF- $\alpha$  may cause HBV reactivation in patients with HBsAg (+) or HBsAg (–) with detectable HBV DNA and additionally with the status of HBcAg (+) and HBsAg (–). At the same time, treatment of anti-TNF- $\alpha$  can be used safely in patients with HBsAg (+) with undetectable DNA HBV [64]. The current long-term observation of patients with a serological profile HBsAg (–) and anti-HBc (+) treated with anti-TNF- $\alpha$  suggests that the percentage of reactivation is low. This confirms the cohort study 72 patients observed from 43 to 21 weeks, in which no reactivation was observed [65].

In a study by Perez-Alvarez et al. [66], 257 patients were analyzed—89 carriers of HBsAg and 168 hidden carriers (anti-HBc+) treated with anti-TNF- $\alpha$  due to rheumatoid arthritis or inflammatory bowel diseases. Among carriers of HBsAg, HBV reactivation was observed in 35 (39 %), including acute liver failure in 5 (5.6 %). The incidence of HBV reactivation was higher in patients under immunosuppressive treatment (96 vs. 70 %,  $p = 0.033$ ) and lower in

patients who received antiviral prophylaxis (23 vs. 62 %,  $p = 0.003$ ). In the hidden HBV carriers, reactivation was observed in 9 (5 %) patients, including one patient developed acute liver failure.

Data in the literature concerning the reactivation of infection during treatment with anti-TNF- $\alpha$  show that infliximab often causes HBV reactivation as compared with etanercept. More often, the reactivation of these infections occurs during the third dose administration [67, 68]. Greater probability of HBV reactivation during treatment of infliximab than that of etanercept can be explained by pharmacological and biochemical differences between these molecules. HBV reactivation associated with infliximab is more frequent in HBV carriers compared with those infected with concealed HBV [66]. Etanercept has a shorter half-life than infliximab, which allows for faster elimination of the drug after cessation of treatment. In case of infliximab discontinuation, the rebirth of an increased pool of macrophages and T lymphocytes occurs, resulting in an acute response in relation to replicating virus, which can potentially cause damage to the fulminant hepatitis [69]. Thimm et al. [70] studying hepatitis B in animals have found that several months after the depletion of CD8 + T cells, there was an unexpected increase in the number of these cells to baseline. He was accompanied by an increase in liver enzymes and increased the elimination of HBV DNA without the formation of neutralizing antibodies anti-HBs. These studies suggest that elimination of the HBV DNA is associated with restoration previously eradicated CD8 + cells [70]. The combination of infliximab with methotrexate therapy can reduce the elimination of CD8 + T lymphocytes specific for HBV present in the hepatocytes [20]. HBV reactivation may appear during infliximab monotherapy [71]. As infliximab concentration decreases, the risk of exacerbation of HBV infection increases, because the remaining CD8 + T cells respond to the viral replication, previously inhibited by the action of TNF- $\alpha$  [72].

#### Rituximab

Rituximab is a chimeric human–murine monoclonal anti-CD20 antibody used in the treatment of rheumatoid arthritis and other rheumatic diseases such as vasculitis. Data on the safety of rituximab in patients with rheumatic diseases and HBV are scarce, but study on patients treated for hematological malignancies indicates a high risk of HBV reactivation in patients undergoing no prophylaxis (27–80 %) [21–23]. Four cases of HBV reactivation were described in patients with RA treated with rituximab [24, 33–35]. In two patients with chronic hepatitis B treated with rituximab with prior prophylaxis, no HBV reactivation was described [25]. The risk of HBV reactivation increases

with concomitant use of corticosteroid or chemotherapy [26]. To date observations of patients with hematologic malignancies showed that the HBV reactivation can occur with rituximab alone [26, 27]. There are also reports that show the opposite—rituximab alone HBV reactivation risk is greater in comparison with the same chemotherapy [26, 28].

#### Abatacept

Abatacept is a fusion protein obtained by genetic engineering. It consists of the extracellular domain of human CTLA4 and a modified Fc fragment of human immunoglobulin G1 for preventing the complement fixation. Abatacept, in contrast to TNF- $\alpha$  inhibitors, directly neutralizes pro-inflammatory cytokines, but specifically inhibits T cell activation in recent retrospective study described in eight patients with chronic hepatitis B treated with abatacept, respectively, four of them received prior anti-viral prophylaxis. In all patients who did not receive antiviral prophylaxis, HBV reactivation occurred [32]. One case of a patient treated with abatacept was described, with a serological profile HBsAg (–), HBeAg (–), anti-HBc (+), anti-HBs (–) and anti-HBe (+) during qualifying for the therapy. After 10 months of treatment, HBsAg (+) and HBV DNA were detected, hepatitis B was diagnosed, abatacept was stopped, and successful treatment with lamivudine was entered [28]. In second case, serological profile was HBsAg (–), HBeAg (–), anti-HBc (+), and anti-HBs (+); serum HBV DNA became undetectable 4 months after initiation of tenofovir treatment [34].

#### Tocilizumab

Tocilizumab (TCZ) is a humanized monoclonal antibody directed against the receptor for interleukin 6. RA patients coinfecting with HBV were not included to clinical studies with tocilizumab. For this reason, data on the safety of tocilizumab therapy in patients infected with HBV are scarce. Signaling pathway inhibition effect of IL-6 on the course of HBV infection has not been investigated; however, IL-6 is associated with both liver damage in the course of HBV infection and virus removal [37]. Two cases of patients who received tocilizumab with hepatitis B treated with antiviral prophylaxis were published in Japan, and no HBV reactivation was reported [29–31]. Therefore, in patients with chronic hepatitis B and normal liver function, antiviral therapy should be considered before treatment with tocilizumab. Prevention is also indicated in HBsAg carriers and in patients with hidden hepatitis B. Nakamura [47] study of 18 patients treated TCZ with a serological profile HBsAg (–)/HBsAg (+) anti-HBc (+), in two patients HBV reactivation was observed.

## Antiviral prophylaxis

Currently, there are no formal guidelines on eligibility for treatment of HBV patients with rituximab, abatacept and tocilizumab [35]. In these patients, it is recommended the initial determination of the presence of HBsAg and anti-HBc and anti-HBs. Patients with current Hbs antigen should receive antiviral treatment. Available data on this subject for rituximab in rheumatic diseases are limited, and for tocilizumab and abatacept inaccessible.

In each patient eligible for the treatment of anti-TNF- $\alpha$ , HBsAg and anti-HBc should be determined [73–75]. Patients with present isolated anti-HBc must also make a determination of HBV DNA by PCR. The carriers of HBsAg should monitor the activity of transaminases and viremia up to 3 months after completion of therapy medicines called anti-TNF [75].

Antiviral therapy for HBV infections led to a reduction in viral load, in spite of chronic drug therapy from the group of anti-TNF- $\alpha$ . Antiviral therapy may be implemented either preventively, at the same time as the anti-TNF- $\alpha$  or to be reserved for cases in which there is an viremia increase [76]. It should be borne in mind that the development of HBV mutations during the course of therapy leads to resistance to antiviral therapy and the development of severe liver inflammation [69]. It is recommended that patients with potential risk of belonging to the group of “hidden carriers,” before treatment with anti-TNF- $\alpha$ , should carry out detailed analysis of the clinical and laboratory signs of HBV reactivation (including the determination of HBs antigen) [77].

The HBsAg carriers and hidden HBV carriers before inclusion of anti-TNF- $\alpha$  should use prophylactic antiviral treatment. This is confirmed by a randomized study of patients undergoing chemotherapy [69, 78, 79], while in patients with rheumatic diseases, several clinical cases and reviews were performed [66, 80–85].

Based on the review of the published cases of patients with rheumatic diseases treated with anti-TNF- $\alpha$  ( $n = 255$ ), made by Ramos-Casals in 2010, it can be concluded that the risk of HBV reactivation is approximately 38 % ( $n = 87$ ) in patients HBsAg (+), without preventive antiviral treatment [80]. There are lack of reliable data on the safety and efficacy of antiviral treatment as protection against HBV reactivation [86]. Zingarelli et al.'s [87] literature review estimated that antiviral prophylaxis with lamivudine in HBsAg (+) patients reduces the risk of reactivation (lamivudine 14 % vs. without prophylaxis 69 %). Prophylaxis may be considered in the later treatment with antiretroviral agents with higher genetic barrier, with a low risk of viral resistance and a higher efficacy (tenofovir or entecavir). During therapy, patients should be monitored for HBV DNA (every 3–6 months) and ALT (every 3 months) [19]. These

drugs should be discontinued after 6–12 months after treatment of anti-TNF- $\alpha$  [20, 21]. In conclusion, tenofovir or entecavir is a good choice among active carriers with high HBV DNA in the case of reactivation or development of hepatitis, while lamivudine is recommended as prophylaxis in inactive and hidden carriers [21, 22].

The 2008 guidelines of the American College of Rheumatology (ACR), updated in 2012, do not recommend the use of anti-TNF- $\alpha$  in pharmacologically untreated chronic hepatitis B and treated with considerable damage to the liver (Child class-Pugh B and C). There are no clear recommendations on their use in adjusted cirrhosis (Child–Pugh class A). The novelty in relation to the recommendations of the ACR 2008 is to allow the possibility of the use of etanercept in the treatment of RA patients with hepatitis C [74].

Anti-TNF- $\alpha$  should not be used in patients with chronic HBV infection, according to the guidelines European League Against Rheumatism (EULAR). If during treatment with anti-TNF- $\alpha$  patient is diagnosed with hepatitis B, you can implement an antiviral drug.

The prevention can be considered for treatment with newer antiretroviral agents with higher genetic barrier with a low risk of viral resistance and higher efficiency (tenofovir or entecavir). During therapy, patient should be monitored for the presence of viral DNA (every 3–6 months) and ALT (every 3 months) [28]. These drugs should be discontinued after 6–12 months after the completion of biological treatment [36–38].

Observational studies in oncology suggest extending the prophylaxis to the 12th month after the discontinuation of immunosuppressive treatment [55]. In case reports of patients treated with rituximab, HBV reactivation occurred later [53, 54]. According to the onco-hematological recommendations, prior to starting RTX treatment, all patients should be screened for HBV infection. While HBsAg-positive active carriers should receive long-term antiviral treatment with entecavir or tenofovir, inactive carriers are candidates for universal prophylaxis with lamivudine, or entecavir or tenofovir in selected cases, to prevent hepatitis reactivation. Conversely, for HBsAg-negative anti-HBc-positive carriers, that is, those with resolved HBV infection, universal prophylaxis with lamivudine is recommended for those with onco-hematological diseases, whereas watchful monitoring of HBsAg/HBV DNA levels is advisable for all the other indications [56].

In conclusion, tenofovir or entecavir is a good choice among active carriers with high HBV DNA, in the case of reactivation or development of hepatitis, while lamivudine is recommended as prophylaxis in inactive and hidden carriers [38, 39]. It should be emphasized that the final diagnosis of hepatitis and implementation of antiviral treatment should be carried out in close collaboration with a specialist in the field of infectious diseases.

## Summary

Available data based on a small number of case reports in the literature are not always consistent. Therefore, before starting treatment HBsAg, anti-Hbc, and anti-Hbs should be determined, especially in people at increased risk. In patients with positive tests for the presence of virus, it is recommended to consult with physicians experienced in the treatment of viral hepatitis. Throughout the period of biological treatment and a few months after treatment, patients should be closely monitored for signs and symptoms of active HBV infection and, if necessary, take appropriate treatment. The risk of reactivation of hepatitis B appears to be large, and it can reduce the prophylactic use of lamivudine.

**Open Access** This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

## References

- Juszczyk J (2000) Clinical course and consequences of hepatitis B infection. *Vaccine* 18:S23–S25
- Tiollais P, Charnay P, Vyas GN (1981) Biology of hepatitis B virus. *Science* 213:406–411
- Stuyver LJ, Locarnini SA, Lok A, Richman DD, Carman WF, Dienstag JL, Schinazi RF (2001) Nomenclature for antiviral resistant of human hepatitis B virus mutations in the polymerase region. *Hepatology* 33:751–757
- Ferir G, Kaptein S, Neyts J, De Clercq E (2008) Antiviral treatment of chronic hepatitis B virus infections: the past, the present and the future. *Rev Med Virol* 18:19–34
- Morgan M, Keeffe EB (2009) Diagnosis and treatment of chronic hepatitis B: 2009 update. *Minerva Gastroenterol Dietol* 55:5–22
- Pungpapong S, Kim WR, Poterucha JJ (2007) Natural history of hepatitis B virus infection: an update for clinicians. *Mayo Clin Proc* 82:967–975
- Winthrop KL, Calabrese LH (2011) Let the fog be lifted: screening for hepatitis B virus before biological therapy. *Ann Rheum Dis* 70:1701–1703
- Szczeklik A (2010) Choroby wewnętrzne. *Medycyna Prakt*, pp 967–980
- Chisari FV, Isogawa M, Wieland S (2010) Pathogenesis of hepatitis B virus infection. *Pathol Biol (Paris)* 58(4):258–266
- Hösel M, Quasdorff M, Wiegmann K, Webb D, Zedler U, Broxtermann M, Tedjokusumo R, Esser K, Arzberger S, Kirschning CJ, Langenkamp A, Falk C, Büning H, Rose-John S, Protzer U (2009) Not interferon, but interleukin-6 controls early gene expression in hepatitis B virus infection. *Hepatology* 50(6):1773–1782
- Puro R, Schneider RJ (2007) Tumor necrosis factor activates a conserved innate antiviral response to hepatitis B virus that destabilizes nucleocapsids and reduces nuclear viral DNA. *J Virol* 81(14):7351–7362
- Yapali S, Talaat N, Lok AS (2014) Management of hepatitis B: our practice and how it relates to the guidelines. *Clin Gastroenterol Hepatol* 12(1):16–26
- de Franchis R, Hadengue A, Lau G et al (2003) EASL Jury. EASL International consensus conference on hepatitis B. Consensus statement (long version). *J Hepatol* 39(Suppl. 1):S3–S25
- Tang JR, Hsu HY, Lin HH et al (1998) Hepatitis B surface antigenemia at birth: a long-term followup study. *J Pediatr* 133:374–377. [www.who.int](http://www.who.int)
- Yeo W, Chan PKS, Zhong S, Ho WM, Steinberg JL, Tam JS, Hui P, Leung NWY, Zee B, Johnson PJ (2000) Frequency of hepatitis B virus reactivation in cancer patients undergoing cytotoxic chemotherapy: a prospective study of 626 patients with identification of risk factors. *J Med Virol* 62:299–307
- Yeo W, Johnson PJ (2006) Diagnosis, prevention and management of hepatitis B virus reactivation during anticancer therapy. *Hepatology* 43(2):209–220
- Brojer E (2009) Ukryte zakażenie wirusem HBV w hematologii i transfuzjologii. *Acta Haematol Pol* 40(2):435–449
- Vassilopoulos D (2011) Should we routinely treat patients with autoimmune/rheumatic diseases and chronic hepatitis B virus infection starting biologic therapies with antiviral agents? *Yes*. *Eur J Int Med* 22:572–575
- Perrillo RP (2001) Acute flares in chronic hepatitis B: the natural and unnatural history of an immunologically mediated liver disease. *Gastroenterology* 120:1009–1022
- Evens AM, Jovanovic BD, Su YC et al (2011) Rituximab-associated hepatitis B virus (HBV) reactivation in lymphoproliferative diseases: meta-analysis and examination of FDA safety reports. *Ann Oncol* 22:1170–1180
- Mendez-Navarro J, Corey KE, Zheng H et al (2011) Hepatitis B screening, prophylaxis and reactivation in the era of rituximab-based chemotherapy. *Liver Int* 31:330–339
- Chen XQ, Peng JW, Lin GN et al (2012) The effect of prophylactic lamivudine on hepatitis B virus reactivation in HBsAg—positive patients with diffuse large B-cell lymphoma undergoing prolonged rituximab therapy. *Med Oncol* 29:1237–1241
- Pyrpasopoulou A, Douma S, Vassiliadis T et al (2011) Reactivation of chronic hepatitis B virus infection following rituximab administration for rheumatoid arthritis. *Rheumatol Int* 31:403–404
- Mitroulis I, Hatzara C, Kandili A et al (2013) Long-term safety of rituximab in patients with rheumatic diseases and chronic or resolved hepatitis B virus infection. *Ann Rheum Dis* 72:308–310
- Tsutsumi Y, Shigematsu A, Hashino S et al (2009) Analysis of reactivation of hepatitis B virus in the treatment of B cell non-Hodgkin's lymphoma in Hokkaido. *Ann Hematol* 88(4):375–377
- Yang SH, Kuo SH (2008) Reactivation of hepatitis B virus during rituximab treatment of a patient with follicular lymphoma. *Ann Hematol* 87(4):325–327
- Yeo W, Chan TC, Leung NWY et al (2009) Hepatitis B virus reactivation in lymphoma patients with prior resolved hepatitis B undergoing anticancer therapy with or without rituximab. *J Clin Oncol* 27(4):605–611
- Nagashima T, Minota S (2008) Long-term tocilizumab therapy in a patient with rheumatoid arthritis and chronic hepatitis B. *Rheumatology (Oxford)* 47:1838–1840
- Nagashima T, Maruyama A, Kamata Y et al (2011) Unchanged serum viral load and liver function during tocilizumab treatment in a patient with rheumatoid arthritis and hepatitis C virus infection. *Rheumatol Int* 32:2231–2232
- Tsuboi H, Tsujii A, Namei A et al (2011) A patient with rheumatoid arthritis treated with tocilizumab together with lamivudine prophylaxis after remission of infliximab-reactivated hepatitis B. *Mod Rheumatol* 21:701–705
- Kim PS, Ho GY, Prete PE et al (2012) Safety and efficacy of abatacept in eight rheumatoid arthritis patients with chronic hepatitis B. *Arthritis Care Res (Hoboken)* 64:1265–1268
- Ghrensias E, Meknian A, Rouaghe S, Granne N, Fain O (2012) Reactivation of resolved hepatitis B during rituximab therapy for rheumatoid arthritis. *Joint Bone Spine* 79:100–101
- Salman-Monte TC, Lisbona MP, García-Retortillo M, Maymóia J (2014) Reactivation of hepatitis virus B infection in a patient with

- rheumatoid arthritis after treatment with rituximab. *Reumatol Clin* 10(3):191–200
34. Germanidis G, Hytiroglou P, Zakalka M, Settas L (2012) Reactivation of occult hepatitis B virus infection, following treatment of refractory rheumatoid arthritis with abatacept. *J Hepatol* 56(6):1420–1421
  35. Vassilopoulos D, Calabrese LH (2012) Management of rheumatic disease with comorbid HBV or HCV infection. *Nat Rev Rheumatol* 8(6):348–357
  36. Jong EC, Freedman DO (2009) The immunocompromised traveler. In: CDC travelers' health yellow book. <http://wwwn.cdc.gov/travel/yellowbook/2010/chapter-8/immunocompromisedtraveler.aspx>
  37. Stuck AE, Minder CE, Frey FJ (1989) Risk of infectious complications in patients taking glucocorticosteroids. *Rev Infect Dis* 11:954–963
  38. Xuan D, Yiqi Y, Shao L, Wang J, Zhang W, Zou H (2014) Hepatitis reactivation in patients with rheumatic diseases after immunosuppressive therapy—a report of long-term follow-up of serial cases and literature review. *Clin Rheumatol* 33:577–586
  39. Bae JH, Sohn JH, Lee HS, Park HS, Hyun YS, Kim TY, Eun CS, Jeon YC, Han DS (2012) A fatal case of hepatitis B virus (HBV) reactivation during long-term, very-low-dose steroid treatment in an inactive HBV carrier. *Clin Mol Hepatol* 18(2):225–228
  40. Cheng J, Li JB, Sun QL, Li X (2011) Reactivation of hepatitis B virus after steroid treatment in rheumatic diseases. *J Rheumatol* 38(1):181–182. doi:10.3899/jrheum.100692
  41. Watanabe K, Takase K, Ohno S, Ideguchi H, Nozaki A, Ishigatsubo Y (2012) Reactivation of hepatitis B virus in a hepatitis B surface antigen-negative patient with rheumatoid arthritis treated with methotrexate. *Mod Rheumatol* 22(3):470–473
  42. Urata Y, Uesato R, Tanaka D, Kowatari K, Nitobe T, Nakamura Y, Motomura S (2011) Prevalence of reactivation of hepatitis B virus replication in rheumatoid arthritis patients. *Mod Rheumatol* 21(1):16–23
  43. Tan J, Zhou J, Zhao P, Wei J (2012) Prospective study of HBV reactivation risk in rheumatoid arthritis patients who received conventional disease-modifying antirheumatic drugs. *Clin Rheumatol* 31(8):1169–1175
  44. Tamori A, Koike T, Goto H, Wakitani S, Tada M, Morikawa H, Enomoto M, Inaba M, Nakatani T, Hino M, Kawada N (2011) Prospective study of reactivation of hepatitis B virus in patients with rheumatoid arthritis who received immunosuppressive therapy: evaluation of both HBsAg-positive and HBsAg-negative cohorts. *J Gastroenterol* 46(4):556–564
  45. Ito S, Nakazono K, Murasawa A, Mita Y, Hata K, Saito N, Kikuchi M, Yoshida K, Nakano M, Gejyo F (2001) Development of fulminant hepatitis B (precore variant mutant type) after the discontinuation of low-dose methotrexate therapy in a rheumatoid arthritis patient. *Arthritis Rheum* 44(2):339–342
  46. Nakamura J, Nagashima T, Nagatani K, Yoshio T, Iwamoto M, Minota S (2014) Reactivation of hepatitis B virus in rheumatoid arthritis patients treated with biological disease modifying antirheumatic drugs. *Int J Rheum Dis*. doi:10.1111/1756-185X.12359
  47. Lee YM, Kang M, Hwang JM, Lee S, Cho H, Kim YS (2010) Sulfasalazine induces apoptosis of HBx-expressing cells in an NF-kappaB-independent manner. *Virus Genes* 40(1):37–43
  48. Thong BY, Koh ET, Chng HH, Chow WC (2007) Outcomes of chronic hepatitis B infection in oriental patients with rheumatic diseases. *Ann Acad Med Singap* 36(2):100–105
  49. Narváez J, Rodríguez-Moreno J, Martínez-Aguilá MD, Clavaguera MT (1998) Severe hepatitis linked to B virus infection after withdrawal of low dose methotrexate therapy. *J Rheumatol* 25(10):2037–2038
  50. Hagiyaama H, Kubota T, Komano Y, Kurosaki M, Watanabe M, Miyasaka N (2004) Fulminant hepatitis in an asymptomatic chronic carrier of hepatitis B virus mutant after withdrawal of low-dose methotrexate therapy for rheumatoid arthritis. *Clin Exp Rheumatol* 22(3):375–376
  51. Mok MY, Ng WL, Yuen MF, Wong RW, Lau CS (2000) Safety of disease modifying anti-rheumatic agents in rheumatoid arthritis patients with chronic viral hepatitis. *Clin Exp Rheumatol* 18(3):363–368
  52. Helbling B, Reichen J (1994) Reactivation of hepatitis B following withdrawal of chloroquine. *Schweiz Med Wochenschr* 124(18):759–762
  53. Tonziello G, Pisaturo M, Sica A, Ferrara MG, Sagnelli C, Pasquale G, Sagnelli E, Guastafierro S, Coppola N (2013) Transient reactivation of occult hepatitis B virus infection despite lamivudine prophylaxis in a patient treated for non-Hodgkin lymphoma. *Infection* 41:225–229
  54. Garcia-Rodriguez MJ, Canales MA, Hernandez-Maraver D, Hernandez-Navarro F (2008) Late reactivation of resolved hepatitis B virus infection: an increasing complication post rituximab-based regimens treatment? *Am J Hematol* 83:673–675
  55. Sagnelli E, Pisaturo M, Martini S, Filippini P, Sagnelli C, Coppola N (2014) Clinical impact of occult hepatitis B virus infection in immunosuppressed patients. *World J Hepatol* 6(6):384–393
  56. Viganò M, Mangia G, Lampertico P (2014) Management of patients with overt or resolved hepatitis B virus infection undergoing rituximab therapy. *Expert Opin Biol Ther* 14(7):1019–1031
  57. Penn H (2006) Biologic therapies in autoimmune diseases. *Clin Med* 6:105
  58. Weaver AL (2003) Differentiating the new rheumatoid arthritis biologic therapies. *J Clin Rheumatol* 9:99–114
  59. Peterson JR, Hsu FC, Simkin PA et al (2003) Effect of tumor necrosis factor  $\alpha$  antagonism on serum transaminases and viraemia in patients with rheumatoid arthritis and chronic hepatitis C infection. *Ann Rheum Dis* 62:1078–1082
  60. Su F, Schneider RJ (1997) Hepatitis B virus Hbx protein sensitizes cells to apoptotic killing by tumor necrosis factor  $\alpha$ . *Proc Natl Acad Sci USA* 94:8744–8749
  61. Maini MK, Boni C, Lee CK (2000) The role of virus specific CD8<sup>+</sup> cells in liver damage and viral control during persistent hepatitis B virus infection. *J Exp Med* 191:1269–1280
  62. Millonig G, Kern M, Ludwiczek O et al (2006) Subfulminant hepatitis B after infliximab in Crohn's disease: need for HBV-screening? *World J Gastroenterol* 12:974–976
  63. Uhl EW, Moldawer LL, Busse WW et al (1998) Increased tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) gene expression in parainfluenza type 1 (Sendai) virus-induced bronchiolar fibrosis. *Am J Pathol* 152:513–522
  64. Lan JL, Chen YM, Hsieh TY, Chen YH, Hsieh ChW, Chen DY, Yang SS (2011) Kinetics of viral loads and risk of hepatitis B virus reactivation in hepatitis B core antibody-positive rheumatoid arthritis patients undergoing anti-tumour necrosis factor alpha therapy. *Ann Rheum Dis* 70:1719–1725
  65. Loomba R, Rowley A, Wesley R et al (2008) Systematic review: the effect of preventive lamivudine on hepatitis B reactivation during chemotherapy. *Ann Intern Med* 148:519–528
  66. Perez-Alvarez R, Diaz-Lagares C, Garcia-Hernandez F et al (2011) Hepatitis B virus (HBV) reactivation in patients receiving tumor necrosis factor (TNF)-targeted therapy: analysis of 257 cases. *Medicine (Baltimore)* 90(6):359–371
  67. Giles JT, Bathon JM (2004) Serious infections associated with anticytokine therapies in the rheumatic diseases. *J Intensive Care Med* 19:320–334
  68. Keane J, Gershon S, Wise SP et al (2001) Tuberculosis associated with infliximab, a tumor necrosis factor  $\alpha$ -neutralizing agent. *N Engl J Med* 345:1098–1103

69. Carrol BM, Bond IM (2008) Use of tumor necrosis factor- $\alpha$  inhibitors in patients with chronic hepatitis B infection. *Semin Arthritis Rheum* 38:208–217
70. Thimme R, Wieland S, Steiger C et al (2003) CD8 + cells mediate viral clearance and disease pathogenesis during acute hepatitis B virus infection. *J Virol* 77:68–76
71. Sakellariou GT, Chatzigiannis I (2007) Long term anti-TNF- $\alpha$  therapy for ankylosis spondylitis in two patients with chronic HBV infection. *Clin Rheumatol* 26:950–952
72. Maini RN, Breedveld FC, Kalden JR et al (1998) Therapeutic efficacy of multiple intravenous infusion of anti-tumor necrosis factor hepatitis  $\alpha$  monoclonal antibody combined with low-dose weekly methotrexate in rheumatoid arthritis. *Arthritis Rheum* 41:1552–1563
73. Cavazzana I, Ceribelli A, Cattaneo R, Franceschini F (2008) Treatment with etanercept in six patients with chronic hepatitis C infection and systemic autoimmune diseases. *Autoimmun Rev* 8:104–106
74. Furst DE, Keystone EC, Braun J et al (2011) Updated consensus statement on biological agents for the treatment of rheumatic diseases 2010. *Ann Rheum Dis* 70(Suppl 1):i2–i36
75. Nathan DM, Angus PW, Gibson PR (2006) Hepatitis B and C virus infections and anti-tumor necrosis factor- $\alpha$  therapy: guidelines for clinical approach. *J Gastroenterol Hepatol* 21:1366–1371
76. Wendling D, Di Martino V, Prati C et al (2009) Spondyloarthritis and chronic B hepatitis. Effect of anti-TNF therapy. *Joint Bone Spine* 76:308–311
77. Marzano A, Angelucci E, Andreone P et al (2007) Prophylaxis and treatment of hepatitis B in immunocompromised patients. *Dig Liver Dis* 39:397–408
78. Galeazzi M, Bellisai F, Giannitti C et al (2007) Safety of cyclosporin A in HCV-infected patients: experience with cyclosporin A in patients affected by rheumatological disorders and concomitant HCV infection. *Ann NY Acad Sci* 1110:544–549
79. Giannitti C, Benucci M, Caporali R et al (2009) Efficacy and safety of anti-TNF- $\alpha$  therapy combined with cyclosporine A in patients with rheumatoid arthritis and concomitant hepatitis C virus infection. *Int J Immunopathol Pharmacol* 22:543–546
80. Esteve i Comas M, Saro C, Gonzales-Huix F et al (2004) Chronic hepatitis B reactivation following infliximab therapy in Crohn's disease patients: need for primary prophylaxis. *Gut* 53:1363–1365
81. Michel M, Duvoux C, Hezode C et al (2003) Fulminant hepatitis after infliximab in patient with hepatitis B virus treated for an adult onset Still's disease. *J Rheumatol* 30:1624–1625
82. Caporali R, Bobbio-Pallavicini F, Atzeni F et al (2010) Safety of tumor necrosis factor alpha blockers in hepatitis B virus occult carriers (hepatitis B surface antigen negative/anti-hepatitis B core antigen positive) with rheumatic diseases. *Arthritis Care Res* 62:749–754
83. Ostuni P, Botsios C, Punzi L, Sfriso P, Todesco S (2003) Hepatitis B reactivation in a chronic hepatitis B surface antigen carrier with rheumatoid arthritis treated with infliximab and low dose methotrexate. *Ann Rheum Dis* 62:686–687
84. Chung SJ, Kim JK, Park MC, Park YB, Lee SK (2009) Reactivation of hepatitis B viral infection in inactive HBsAg carriers following anti-tumor necrosis factor- $\alpha$  therapy. *J Rheumatol* 36:2416–2420
85. Kim YJ, Bae SC, Sung YK et al (2010) Possible reactivation of potential hepatitis B virus occult infection by tumor necrosis factor- $\alpha$  blocker in the treatment of rheumatic diseases. *J Rheumatol* 37:346–350
86. Singh JA, Furst DE, Bharat A et al (2012) 2012 Update of the 2008 American College of rheumatology recommendations for the use of disease-modifying antirheumatic drugs and biologic agents in the treatment of rheumatoid arthritis. *Arthritis Care Res* 64:625–639
87. Zingarelli S, Frassi M, Bazzani C et al (2009) Use of tumor necrosis factor- $\alpha$ -blocking agents in hepatitis B virus-positive patients: reports of 3 cases and review of the literature. *J Rheumatol* 36:1189–1194