



Osteoimmunology: entwined regulation of integrated systems

Mary C. Nakamura^{1,2}

Published online: 22 October 2019

© This is a U.S. government work and not under copyright protection in the U.S.; foreign copyright protection may apply 2019

Osteoimmunology is the study of the close interrelationships between the skeletal and the immune systems that begin in development and continue as important functional regulators of both systems throughout life. Bone is a complex organ which both houses and influences immune cell development, differentiation, and function. The osteoclast, the important bone degrading cell, is a highly differentiated immune cell which is regulated by similar mechanisms as its myeloid cell relatives. Conversely, bone cells can influence immune cell development and immune responses, which can also have a significant impact during disease pathology. This issue focused on osteoimmunology that includes a set of new reviews that emphasize how regulation of immune function occurs in concert with the regulation of bone turnover, which has significant implications for the outcomes of bone density and disease due to immune activation during infection, autoimmunity, and aging.

Osteoclasts and macrophages have long been known to be unique differentiation states, both derived from the same myeloid progenitors [1, 2]; however, the review by Yang and Wan describes how macrophage polarization and osteoclasts can be viewed as competing differentiation outcomes [3]. This review delineates the important and opposing role of nuclear receptors in controlling osteoclast differentiation and polarized macrophage function. These molecular rheostats include PPAR γ which is a ligand-activated transcription factor that belongs to the nuclear hormone receptor superfamily that promotes osteoclast differentiation but suppresses proinflammatory macrophage activation

and inflammation. PPAR γ is also the target of the thiazolidinedione medications currently used for treatment of insulin resistance and type 2 diabetes by activating PPAR γ . Indeed, ligand activation of PPAR γ by rosiglitazone exacerbates osteoclast differentiation and use of pioglitazone can lead to the side effect of decreased bone density and increased fracture [4, 5].

Nuclear receptors are important regulators of both cellular differentiation and cellular metabolism. The review by Park-Min emphasizes the role of immunometabolism in osteoclast differentiation and function [6]. Immunometabolism in macrophages has previously been shown to have particular relevance to the study of intracellular infections [7]. Similar to the key metabolic differences identified in functional macrophage polarization states induced by cytokines [8, 9], osteoclast differentiation activates specific metabolic programs that are also required for their bone degradation function. During osteoclast multinucleation, upregulation of mitochondrial oxidative phosphorylation enzymes turns on and provides the energy needed for bone resorption. Just as targeted metabolic reprogramming of macrophages can markedly affect immune functions of macrophages, targeting the metabolic reprogramming of osteoclasts may be a useful strategy to modulate pathologic bone turnover as well.

Several other reviews in this issue focus on regulators of immune responses that have also been demonstrated to have significant roles in the regulation of osteoclasts and other bone cells. MicroRNAs (miRNAs) which are small non-coding RNAs with evolutionarily conserved sequences are expressed in various tissues and cells and have been shown to play a key role in regulating cellular differentiation, proliferation, and apoptosis [10]. In bacterial and viral infection and autoimmunity, miRNAs can regulate inflammatory responses particularly under states of cellular stress [10, 11]. The review by Inoue et al. [12] delineates miRNA regulation of osteoclast differentiation and function and the potential for therapeutic targeting of these pathways.

The review by Miller et al. [13] describes how TLR signals particularly those mediated by DAMP (damage-associated molecular patterns) ligands contribute to the pathogenesis of osteoarthritis. TLRs recognize both pathogen-associated

This article is a contribution to the special issue on Osteoimmunology - Guest Editor: Mary Nakamura

✉ Mary C. Nakamura
mary.nakamura@ucsf.edu

¹ Department of Medicine, Division of Rheumatology, University of California, San Francisco, 513 Parnassus Avenue, San Francisco, CA 94143, USA

² Arthritis/Immunology Section, San Francisco Veterans Administration Health Care System, 4150 Clement Street, 111R, San Francisco, CA 94121, USA

molecular patterns (PAMPS) multiple DAMPs, and ligand binding leads to activation of inflammation with cellular production of cytokines, chemokines, and proteases [14, 15]. Thus, TLRs play a significant role in both infection and sterile tissue injury. In this review, Miller et al. [13, 16] propose a novel model involving TLR effects on multiple cell types within the joint including connective tissue, bone, innate immune cells, and sensory neurons all interact to promote OA pathology and pain.

Interestingly, murine $\gamma\delta$ T cells regulate innate immune responses via multiple pathways including direct activation of TLR pathways in neutrophil and monocytes. As described in the review by Nguyen et al., $\gamma\delta$ T cells are non-conventional lymphocytes, which express T cell receptor (TCR) $\gamma\delta$ chains and have been demonstrated to play a role in skin and joint inflammation commonly observed in rheumatic diseases such as psoriatic arthritis [17]. $\gamma\delta$ T cells straddle the interface of innate and adaptive immunity as they influence the activation of innate cells and antigen presentation [18]. Discussed in this review are IL-17+ $\gamma\delta$ T cell subtypes seen in both human and mouse inflammatory arthritis that influence physiological bone remodeling, given that IL-17 induces RANKL from stromal cells as well as RANK receptor expression to direct osteoclast differentiation and function [19].

An additional pathway activated by TLR involves activation of inflammasomes. In the review by Alippe and Mbalaviele [20], the role of the inflammasome in the activation of bone degradation is discussed. Inflammasomes are intracellular multi-subunit protein complexes that assemble in response to a diverse range of pathogen-associated or danger-associated molecular patterns (PAMPs or DAMPs). The formation of the inflammasome multimeric complex sequentially leads to enzymatic activation of caspase-1 and proteolytic activation of interleukins 1 β and 18 (IL-1 β and IL-18). IL-1 β and IL-18 contribute to host defense against infections by activating innate cells such as monocytes, macrophages, dendritic cells, and neutrophils, but also influences T-helper 17 (Th17)- and Th1-mediated adaptive immune responses [21, 22]. The role for inflammasomes in bone is highlighted by the rare genetic disorders such as cryopyrin-associated periodic syndrome (CAPS) that are caused by mutations in genes that encode inflammasome components and involve systemic inflammation and bone pathologies that are successfully treated with IL-1 blockade [22–24]. However, also reviewed is evidence that inflammasomes are implicated in a wide range of diseases of bone loss driven by sterile and non-sterile inflammation including inflammatory arthritis, osteomyelitis, aging, and wear debris-induced osteolysis [20].

Finally, in the review by Terashima and Takayanagi [25], the complementary role of bone cell modulation of immune cell differentiation and function is discussed. Through the

maintenance of the bone marrow microenvironment, osteoblasts support hematopoietic stem cells and hematopoiesis; however, this effect is not critical to normal hematopoiesis [26]. Interestingly, the effect of bone cells on immune cells differs significantly during sepsis or disseminated acute bacterial infection. Sepsis induces rapid and marked bone loss through impaired osteoblastic bone formation, and Terashima et al. have shown that the depletion of osteoblasts is responsible for the reduced numbers of common lymphoid progenitors (CLPs) induced by sepsis, as well as the decrease in the number of peripheral T and B cells. Similar to sepsis, transient osteoblast ablation leads to a marked decrease in the CLP number and lymphopenia in the periphery, suggesting a role for osteoblasts in the regulation of lymphopoiesis. This is likely due to IL-7 production by osteoblast, as depletion of IL-7 from osteoblasts recapitulates the effect of osteoblast depletion on CLP numbers [25, 27]. Thus, bone cells directly contribute to sepsis-related immunocompromise and may be integrally linked to the high incidence of mortality. These studies suggest that osteoblasts could be a therapeutic target that could be of benefit to treatment of sepsis and potentially other infectious diseases [25].

Reviews in this issue highlight significant advances in our understanding of bone and immune cell interactions, highlight osteoimmunological contributions to pathologic disease, and identify potential therapeutic targets that affect both systems. The close bi-directional interactions highlight the need to direct future studies toward the study of bone during immune modulation of human disease and similarly toward the examination of the effects of treatments for bone disease on infection and immunity.

Funding information This work was supported by the National Institute of Arthritis, Musculoskeletal, and Skin Disorders grants AR070155, AR066735, and P30AR070155 funding the UCSF PREMIER program; National Institute of Aging grant AG04628; the Russell/Engleman Arthritis Center at UCSF; VA Merit Review BX002994; and the Department of Veteran's Affairs Health Care System.

References

1. Loutit JF, Nisbet NW (1982) The origin of osteoclasts. *Immunobiology* 1982161(3–4):193–203
2. Jacome-Galarza CE, Percin GI, Muller JT, Mass E, Lazarov T, Eitler J, Rauner M, Yadav VK, Crozet L, Bohm M, Loyher PL, Karsenty G, Waskow C, Geissmann F (2019) Developmental origin, functional maintenance and genetic rescue of osteoclasts. *Nature* 568:541–545
3. Yang D, Wan Y (2019) Molecular determinants for the polarization of macrophage and osteoclast. *Semin Immunopathol*. <https://doi.org/10.1007/s00281-019-00754-3>
4. Wei W, Wang X, Yang M, Smith LC, Dechow PC, Sonoda J, Evans RM, Wan Y (2010) PGC1 β mediates PPAR γ activation of osteoclastogenesis and rosiglitazone-induced bone loss. *Cell Metab* 11(6):503–516

5. Viscoli CM, Inzucchi SE, Young LH, Insogna KL, Conwit R, Furie KL, Gorman M, Kelly MA, Lovejoy AM, Kernan WN, for the IRIS Trial Investigators (2017) Pioglitazone and risk for bone fracture: safety data from a randomized clinical trial. *J Clin Endocrinol Metab* 102(3):914–922
6. Park-Min K (2019) Metabolic reprogramming in osteoclasts. *Semin Immunopathol*. <https://doi.org/10.1007/s00281-019-00757-0>
7. Russell DG, Huang L, VanderVen BC (2019) Immunometabolism at the interface between macrophages and pathogens. *Nat Rev Immunol* 19:291–304
8. O'Neill LA, Pearce EJ (2016) Immunometabolism governs dendritic cell and macrophage function. *J Exp Med* 213:15–23
9. Diskin C, Palsson-McDermott EM (2018) Metabolic modulation in macrophage effector function. *Front Immunol* 9:270
10. Drury RE, O'Connor D, Pollard AJ (2017) The clinical application of micromas in infectious disease. *Front Immunol* 8:1182. <https://doi.org/10.3389/fimmu.2017.01182>
11. Salvi V, Gianello V, Tiberio L, Sozzani S, Bosisio D (2019) Cytokine targeting by miRNAs in autoimmune disease. *Front Immunol* 10:15. <https://doi.org/10.3389/fimmu.2019.00015>
12. Inoue K, Nakano S, Zhao B (2019) Osteoclastic microRNAs and their translational potential in skeletal diseases. *Semin Immunopathol*. <https://doi.org/10.1007/s00281-019-00761-4>
13. Miller RE, Scanzello CR, Malfait A-M (2019) An emerging role for toll-like receptors at the neuroimmune interface in osteoarthritis. *Semin Immunopathol*. <https://doi.org/10.1007/s00281-019-00762-3>
14. Feldman N, Rotter-Maskowitz A, Okun E (2015) DAMPs as mediators of sterile inflammation in aging-related pathologies. *Ageing Res Rev* 24:29–39
15. Pelka K, Shibata T, Miyake K, Latz E (2016) Nucleic acid-sensing TLRs and autoimmunity: novel insights from structural and cell biology. *Immunol Rev* 269(1):60–75
16. Miller RE, Belmadani A, Ishihara S, Tran PB, Ren D, Miller RJ, Malfait AM (2015) A damage-associated molecular patterns generated in osteoarthritis directly excite murine nociceptive neurons through toll-like receptor 4. *Arthritis Rheum* 67(11):2933–2943
17. Nguyen CT, Maverakis E, Eberl M, Adamopoulos IE (2019) $\gamma\delta$ T cells in rheumatic diseases: from fundamental mechanisms to autoimmunity. *Semin Immunopathol*. <https://doi.org/10.1007/s00281-019-00752-5>
18. Schwacha MG, Rani M, Zhang Q, Nunez-Cantu O, Cap AP (2014) Mitochondrial damage-associated molecular patterns activate $\gamma\delta$ T cells. *Innate Immun* 20(3):261–268
19. Ito Y, Usui T, Kobayashi S, Iguchi-Hashimoto M, Ito H, Yoshitomi H, Nakamura T, Shimizu M, Kawabata D, Yukawa N, Hashimoto M, Sakaguchi N, Sakaguchi S, Yoshifuji H, Nojima T, Ohmura K, Fujii T, Mimori T (2009) $\gamma\delta$ T cells are the predominant source of interleukin-17 in 381 affected joints in collagen-induced arthritis, but not in rheumatoid arthritis. *Arthritis Rheum* 60 382(8):2294–2303
20. Alippe Y, Mbalaviele G (2019) Omnipresence of inflammasome activities in inflammatory bone diseases. *Semin Immunopathol*. <https://doi.org/10.1007/s00281-019-00753-4>
21. Shin JI, Lee KH, Joo YH, Lee JM, Jeon J, Jung HJ, Shin M, Cho S, Kim TH, Park S, Jeon BY, Jeong H, Lee K, Kang K, Oh M, Lee H, Lee S, Kwon Y, Oh GH, Kronbichler A (2019) Inflammasomes and autoimmune and rheumatic diseases: a comprehensive review. *J Autoimmun* 103:102299. <https://doi.org/10.1016/j.jaut.2019.06.010>
22. Qu C, Bonar SL, Hickman-Brecks CL, Abu-Amer S, McGeough MD, Peña CA, Broderick L, Yang C, Grimston SK, Kading J, Abu-Amer Y, Novack DV, Hoffman HM, Civitelli R, Mbalaviele G (2015) NLRP3 mediates osteolysis through inflammation-dependent and -independent mechanisms. *FASEB J* 29(4):1269–1279
23. Xiao J, Wang C, Yao JC, Alippe Y, Xu C, Kress D, Civitelli R, Abu-Amer Y, Kanneganti TD, Link DC, Mbalaviele G (2018) Gasdermin D mediates the pathogenesis of neonatal-onset multisystem inflammatory disease in mice. *PLoS Biol* 16(11):e3000047. <https://doi.org/10.1371/journal.pbio.3000047>
24. Dinarello CA, van der Meer JW (2013) Treating inflammation by blocking interleukin-1 in humans. *Semin Immunol* 25(6):469–484
25. Terashima A, Takayanagi H (2019) The role of bone cells in immune regulation during the course of infection. *Semin Immunopathol*. <https://doi.org/10.1007/s00281-019-00755-2>
26. Terashima A, Okamoto K, Nakashima T, Akira S, Ikuta K, Takayanagi H (2016) Sepsis-induced osteoblast ablation causes immunodeficiency. *Immunity* 44(6):1434–1443
27. Miyamoto K, Yoshida S, Kawasumi M, Hashimoto K, Kimura T, Sato Y, Kobayashi T, Miyauchi Y, Hoshi H, Iwasaki R, Miyamoto H, Hao W, Morioka H, Chiba K, Kobayashi T, Yasuda H, Penninger JM, Toyama Y, Suda T, Miyamoto T (2011) Osteoclasts are dispensable for hematopoietic stem cell maintenance and mobilization. *J Exp Med* 208(11):2175–2181

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.