#### REVIEW



# Tumour microenvironment as a predictive factor for immunotherapy in non-muscle-invasive bladder cancer

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#### Abstract

Bladder cancer (BC) can be divided into two subgroups depending on invasion of the muscular layer: non-muscle-invasive bladder cancer (NMIBC) and muscle-invasive bladder cancer (MIBC). Its aggressiveness is associated, inter alia, with genetic aberrations like losses of 1p, 6q, 9p, 9q and 13q; gain of 5p; or alterations in the p53 and p16 pathways. Moreover, there are reported metabolic disturbances connected with poor diagnosis—for example, enhanced aerobic glycolysis, gluconeogenesis or haem catabolism.

Currently, the primary way of treatment method is transurethral resection of the bladder tumour (TURBT) with adjuvant Bacillus Calmette–Guérin (BCG) therapy for NMIBC or radical cystectomy for MIBC combined with chemotherapy or immunotherapy. However, intravesical BCG immunotherapy and immune checkpoint inhibitors are not efficient in every case, so appropriate biomarkers are needed in order to select the proper treatment options. It seems that the success of immunotherapy depends mainly on the tumour microenvironment (TME), which reflects the molecular disturbances in the tumour. TME consists of specific conditions like hypoxia or local acidosis and different populations of immune cells including tumour-infiltrating lymphocytes, natural killer cells, neutrophils and B lymphocytes, which are responsible for shaping the response against tumour neoantigens and crucial pathways like the PD-L1/PD-1 axis.

In this review, we summarise holistically the impact of the immune system, genetic alterations and metabolic changes that are key factors in immunotherapy success. These findings should enable better understanding of the TME complexity in case of NMIBC and causes of failures of current therapies.

Keywords Bladder cancer · NMIBC · Microenvironment · Cancer phenotype · Immunotheraphy

# Introduction

In recent years, bladder cancer (BC) has become one of the most common cancers in the worldwide [1]. Almost 75% of patients with BC present with non-muscle-invasive disease (NMIBC) confined to the mucosa (stage Ta, carcinoma

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in situ [CIS]) or submucosa (stage T1); in younger patients, this percentage is higher [2, 3]. Despite a reduction in the incidence and mortality in some registries, in others an increase in the frequency has been noted, so BC remains a significant problem from the clinical and public health point of view [1, 4, 5]. The most important risk factor for BC is tobacco smoking as well as exposure to aromatic amines, polycyclic aromatic hydrocarbons and chlorinated hydrocarbons, causing almost 60% of new cases of the disease [2]. Chronic inflammation has been recognised as another important risk factor for BC [6-8]. The basic form of treatment in patients with lower stages of NMIBC is transurethral resection of the bladder tumour (TURBT) and adjuvant treatment. Previous studies have indicated that 48–61% of patients have recurrence of the disease and 7.4-14.2% of patients experience disease progression during 10 years [9-12]. There is also a group of patients with higher risk of progression to

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muscle-invasive bladder cancer (MIBC), reaching more than 50% after 10 years [12].

The treatment strategy in NMIBC depends on the risk groups determined by disease progression assessments. Multiple nomograms and models have been constructed in the past to predict the outcomes of NMIBC [10, 13, 14]. The currently recommended criteria to determine the risk of unfavourable outcomes according to the European Association of Urology (EAU) are presented in Table 1. According to the model, patients in the low-risk group should be treated with TURBT. The most effective form of adjuvant treatment in the intermediate- and high-risk groups of patients is adjuvant intravesical Bacillus Calmette-Guérin (BCG) immunotherapy with a maintenance course. Patients in the very-high-risk group should be treated with a radical cystectomy (RC) [15]. The standard treatment for patients with advanced urothelial BC is RC preceded by cisplatin-based neoadjuvant chemotherapy. Patients with more advanced disease are treated with palliative platin-containing combination chemotherapy with maintenance treatment with avelumab or palliative immunotherapy with pembrolizumab or atezolizumab [3]. Patients who are programmed death ligand 1 (PD-L1) positive and not eligible for cisplatin-based chemotherapy could receive pembrolizumab as a first-line treatment [16]. Pembrolizumab is also administered to patients with tumours that have relapsed after platinumbased therapy [17].

BCG intravesical treatment is associated with burdensome local and systemic side effects that could cause treatment stoppage. Besides, BCG infections after BCG instillations have been reported [18]. Serious side effects are encountered in < 5% of patients [19]; however, only 16-29% of patients are able to continue the full 3-year maintenance course of BCG immunotherapy due to the high frequency of local or systemic adverse events [20, 21]. Moreover, it has been estimated that almost one third of BCG-treated patients do not respond to the treatment [22]. Likewise, interruptions in BCG treatment may reduce its effectiveness. Some data indicate that patients who progress to MIBC have a worse prognosis than those who present with 'primary' muscle-invasive disease [23, 24]. From the oncological point of view, the optimal method of treatment in patients with BCG-unresponsive tumours is RC, but this method is connected with risks, morbidity and significant deterioration of quality of life (QoL) [14]. Taking this information into consideration, new, effective treatment options for high-risk patients are needed. A relatively new option to the landscape of treatment for BCG-unresponsive NMIBC is pembrolizumab, which was approved by the Food and Drug Administration (FDA) in 2020. There are ongoing clinical trials of other immune checkpoint inhibitors (ICIs) in the same setting [25–28]. Immunotherapy with ICIs is usually tolerable, but high frequencies of adverse events leading to discontinuation have been reported, so predictive biomarkers should be established [29] to assess which patients could achieve advantages from the treatment. PD-L1 expression is not enough to determine which patients could receive the benefit on the treatment [25, 30]. Investigators found that recurrence-free survival (RFS) correlated with high baseline stromal CD8<sup>+</sup> cells and high post-treatment fibroblast activation protein. Moreover, response to atezolizumab has been linked with inflamed phenotype and resistance to atezolizumab treatment has been detected in tumours with desert phenotype [31]. In case of MIBC patients, it seems that atezolizumab neoadjuvant treatment is effective

Table 1 The risk group definitions and risk factors for an unfavourable outcome according to the European association of urology (EAU)

Risk group	
Low risk	A primary, single, TaT1 LG/G1 tumour $< 3$ cm in diameter without CIS in a patient $\leq 70$ years
	A primary Ta LG/G1 tumour without CIS with at most one of the additional clinical risk factors
Intermediate risk	Patients without CIS who are not included in either the low-, high- or very-high-risk groups
High risk	All T1 HG/G3 without CIS, except those included in the very-high-risk group
	All patients with CIS, except those included in the very-high-risk group
	Stage, grade and additional clinical risk factors:
	Ta LG/G2 or T1G1, no CIS with all three risk factors
	Ta HG/G3 or T1 LG, no CIS with at least two risk factors
	T1G2, no CIS with at least one risk factor
Very high risk	Stage, grade and additional clinical risk factors:
	Ta HG/G3 and CIS with all three risk factors
	T1G2 and CIS with at least two risk factors
	T1 HG/G3 and CIS with at least one risk factor
	T1 HG/G3, no CIS with all three risk factors

CIS, carcinoma in situ

in circulating tumour DNA (ctDNA)-negative patients at baseline and after neoadjuvant therapy [32].

Tumour mutational (neoantigen) burden, the molecular subtype as well as immune gene expression profiling are considered possible biomarkers for predicting the response to ICIs; however, the negative predictive role of them is inadequate [33]. Molecular classification of BC as well as the metabolites might be designed to stratify prognostically relevant categories and to define the proper treatment options. Unfortunately, assessment of the end products of metabolic pathways, similarly to a proper appraisal of the molecular profile of the tumour, requires a specified condition and ideal markers should be easily detectable [34]. Moreover, implementation of molecular classification and metabolic biomarkers in clinical practice is limited due to the great complexity of the required technology, the high costs and the limited availability of this technology worldwide.

In this review, we try to determine new markers of high risk of progression to MIBC stage and BCG unresponsiveness. We analyse mainly the tumour microenvironment (TME) because it is connected to molecular aberrations and metabolic pathways in tumours and could influence the outcome. TME assessment is also easier and more accessible than assessment of molecular and metabolic disturbances, so it may represent a new prognostic tool that could be successfully applied in everyday clinical practice.

#### The molecular basics of bladder cancer

BC can be divided into two phenotypes based on whether it invades the muscle layer: NMIBC and MIBC [35–37]. Although CIS belongs to the NMIBC group, it has a high propensity for invasion and metastasis. A papillary tumour could also transform into a more aggressive phenotype. Despite the division according to phenotype, NMIBC tumours can also be classified into molecular subclasses, but differences in methodologies have resulted in several molecularly defined classifications.

Low-grade NMIBC tumours are often near diploid with loss of chromosome 9 and loss of heterozygosity of 11p. The other copy number changes including gains of 1q, 17 and 20q; amplifications of 11q; and loss of 10q. Moreover, up to 80% of low-grade NMIBC tumours show the fibroblast growth factor receptor 3 (*FGFR3*) gene mutationwith a constitutively active receptor tyrosine kinase–Ras pathway[37–39]. High-grade NMIBC tumours can be characterised by homozygous deletion of *CDKN2A* (encodes p16<sup>INK4a</sup>) [40]. The second group of BC include CIS and invasive tumours, which usually show alterations in the *TP53* and retinoblastoma (*RB*) genes and pathways [41]. In addition, chromosome 9 deletion is frequently reported as well as losses of 1p, 6q, 9p, 9q and 13q and gain of 5p [42]. Besides, a papillary tumour could transform to a more aggressive phenotype and is usually caused by the accumulation of alterations in the p53 and p16 pathways [35]. Comparison between NMIBC and MIBC mutation profiles has revealed lower overall mutation rates and more frequent mutations in *RHOB* and chromatin modifier genes in NMIBC [43].

According to Hedegaard et al., [44] NMIBC can be grouped into three major classes with different progressionfree survival (PFS) and clinical and histopathological features. Class 1 comprises low-risk tumours; NMIBC of high stage and grade, concomitant CIS and progression to MIBC are more frequently observed in higher classes. Dyrskjot et al. [45] determined that class 2 tumours show positive signatures for progression and include upregulated KPNA2, BIRC5, UBE2C, CDC25B, COL4A1, MSN and COL18A1 as well as downregulated COL4A3BP, MBNL2, NEK1, FABP4 and SKAP2. In addition, KRT20 expression associated with CIS lesions can be found in class 2 tumours. Class 3 tumours show some of the gene expression characteristics associated with MIBC (KRT5<sup>+</sup>, KRT14<sup>+</sup>, CD44<sup>+</sup>, KRT20<sup>-</sup> and PPARG<sup>-</sup>). Unfortunately, high GATA3 expression in class 3 tumours indicates that they should not be regarded as a precursor to MIBC. Moreover, additional proteins required to regulate differentiation are highly expressed in all classes and even higher in class 3, for which the authors expected lower expression. The authors found also that progressing tumours show class shifts from class 3 to class 2 during progression and only 1 of 24 MIBC tumours was classified as class 3 [44].

Hurst et al. [43] detected two subtypes of primary Ta tumours with differential risk of recurrence. They found that the high-risk Ta subtype had loss of 9q including *TSC1*, increased KI67 labelling index, upregulated glycolysis, DNA repair, mTORC1 signalling, features of the unfolded protein response and altered cholesterol homeostasis [43].

According to another NMIBC classification called the UROMOL algorithm, class 1 includes mainly low-grade Ta tumours with significantly higher FGFR3 gene expression than other groups. UROMOL class 2 tumours have the highest proliferation expression scores and are the most aggressive subtype. UROMOL class 3 is enriched in high-grade T1 tumours. The class 3 tumours have the highest expression of immune-related genes, whereas class 1 tumours have the lowest. Unfortunately, the molecular characteristic of the tumours does not agree with the histopathological characteristics of the tumours because class 1 comprises T1 tumours and high-grade tumours. Similarly, in class 3 almost one third of the tumours are Ta tumours. Besides, class 3 could also include low-grade tumours. According to the performed analyses, class 2 tumours were associated with worse RFS, but the subtype was not associated with recurrence rates in patients treated with BCG. Moreover, higher immune scores were also associated with improved RFS among the BCG-treated patients, but there was not a statistically significant difference in the immune score among the cohorts differ [46]. For this reason, we do not yet have the ability to determine precisely which tumour will progress to MIBC.

Besides genetic aberrations, another hallmark of cancer is energy metabolism reprogramming. The metabolic changes associated with cancer mainly concern changes in glucose and amino acid uptake. In addition, opportunistic modes of nutrient acquisition, increased nitrogen demand and using glycolysis to produce Krebs cycle intermediates and NADPH have been detected in cancer cells [47]. An important feature of BC metabolism is 'aerobic glycolysis' or the Warburg effect, leading to glucose addiction. Cancer cells modify their metabolism to produce large amounts of lactate, even under aerobic conditions. Aerobic glycolysis is less efficient than oxidative phosphorylation in terms of adenosine triphosphate (ATP) production: it can only produce two molecules of ATP, compared with 30 molecules for full glucose oxidation [48]. To meet their energy demands, cancer cells are forced to avidly take up glucose. Cancer cells obtain glucose from the environment, glycogenolysis of glycogen and gluconeogenesis, using lactate and glutamine as gluconeogenic precursors [49]. The increased glucose production is facilitated by upregulation of gluconeogenic enzymes that also favour cell proliferation [49]. The metabolic changes seem to be connected with genomic alterations, especially with loss of chromosome region 9q and tumour suppressor TSC1 and upregulated mTORC1 signalling as well as loss of *p53* and activating mutations in PIK3CA, RAS and AKT. Analyses performed on human urinary bladder cell lines have shown that the progression from a less to a more invasive stage is associated with lower expression of glucose transporters 1 (GLUT1) and phosphofructokinase-1 (PFK-1) in cell lines from a less invasive stage compared with a highly invasive stage, despite similar glucose consumption. Investigators found that BC cell lines differ in extracellular lactate up to ~eightfold across the cell line panel, a finding that suggest metabolic switch occurs via combinations of genetic mutationsin different tumours. They found higher lactate release rates in the mutant Ras BC cell lines and the lowest lactate release rate and largest ratio of oxygen consumed to lactate released associated with FGFR3-TACC3 fusions [50].

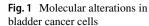
Less invasive BC cells use less pyruvate and produce low amounts of alanine while highly invasive BC cells need more. BC progression is also associated with increased production of lactate depending on decreased lactate dehydrogenase (LDH) expression. However, there are also data about higher rates of lactate release in Ta compared with T1 or T2 specimens and higher LDH-A expression in MIBC than NMIBC tumours [50, 51]. Besides, researchers found increased expression of lactate import transporter MCT1 in T1 and only some MIBC tumours compared with Ta tumours [50]. Early BC is also associated with galactose, starch and sucrose metabolism, whereas more advanced stages are characterised by changes in glycine, serine, threonine glycerophospholipid, arginine and proline metabolism [52]. In addition, NMIBC shows decreased eicosanoid metabolism compared with MIBC [53]. NAD<sup>+</sup> metabolism and haem catabolism are higher in MIBC than in NMIBC [53].

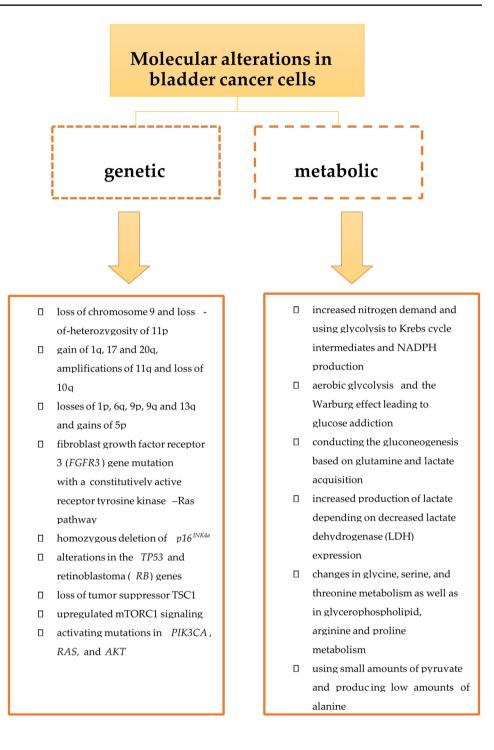
Indoleamine 2,3-dioxygenase-1 (IDO1) has been recognised as a one of the markers of invasiveness. Indeed, IDO1 expression is reduced significantly in invasive compared with non-invasive BC cells [54, 55].

In contrast to the above-mentioned data, the molecular classification of BC presented by Song et al. [56] does not clearly differentiate NMIBC and MIBC. According to the algorithm, low-grade NMIBC is usually included in class 1, characterised by decreased expression of genes involved in cell proliferation. Class 2, characterised by the downregulation of immune response pathways, includes both low-grade NMIBC and some MIBC. Class 3 contains both high-grade NMIBC and MIBC. Moreover, the expression of genes associated with BC prognosis, such as E2F1, FOXM1, CCNB1 and CCNE1, is observed in both the NMIBC and MIBC subgroups. Most MIBC tumours are classified into class 4, characterised by upregulation of genes implicated in extracellular matrix organisation along with strong activation of the immune response. In addition, PFS was different in three independent cohorts of patients with NMIBC, with a higher frequency of NMIBC progression in class 3 compared with the other classes. Class 3 tumours also have a high somatic mutation rate and alterations of genes involved in the DNA damage response that might indicate a potential benefit for immunotherapy [56]. Lopez-Beltran et al. [57] published similar results. In their division, NMIBC is classified more frequently as a luminal molecular subtype with the morphology of conventional urothelial carcinoma and usually low PD-L1 expression as well as high GATA3 or KRT20 messenger RNA (mRNA) levels. MIBC is usually classified as the basal molecular subtype with high aggressiveness, high PD-L1 expression and high KRT5 or KRT14 mRNA levels. They also reported a subgroup of Ta and MIBC tumours without GATA3, KRT20, KRT5 or KRT14 expression and with high PD-L1 expression. Although most of the luminal tumours have low PD-L1 expression and the basal subtype usually has high PD-L1 expression, both subgroups contain tumours with different patterns of PD-L1 expression [57]. Molecular and metabolic changes in BC cells are presented in Figs. 1 and 2.

#### The differences in cancer depending on the tumour microenvironment

Zheng et al. [58] confirmed that deregulation of the immune microenvironment promotes the malignant progression from

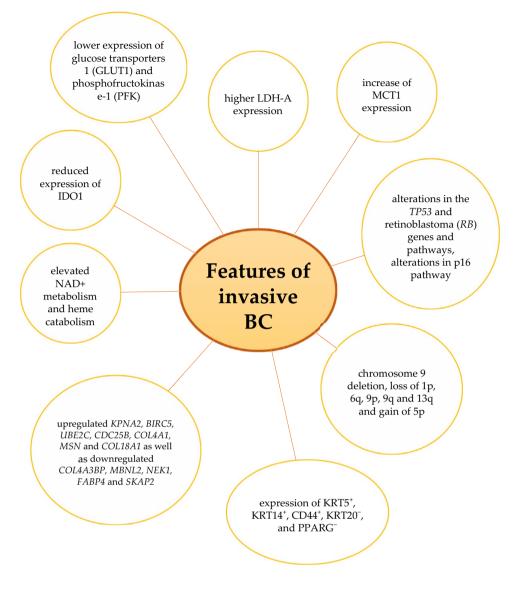




NMIBC to MIBC. They identified an immune prognostic signature that can stratify patients into different risk groups with distinct immunotherapeutic susceptibility, thus facilitating personalised immunotherapy [58]. Investigators reported higher proportions of macrophages, memory-activated CD4<sup>+</sup> T cells and activated natural killer (NK) in MIBC compared with NMIBC samples, and lower resting memory CD4<sup>+</sup> T cells in MIBC samples [59]. Peritumoural CD3<sup>+</sup> cells and CD83 dendritic cells were similar in HG NMIBC and MIBC

and were similarly expressed in HG NMIBC and MIBC [60]. The pattern of immune infiltration in tumours differs among patients. According to recently published data from patients with NMIBC, no immune cells were found in 0.7% of patients, focal infiltration in 42% of patients, mild infiltration in 35.7% of patients, moderate infiltration in 15.7% of patients and extensive infiltration in 6% of patients [61]. Figure. 3 (Supplementary figure) provides a summary of the immune component of the TME and its functions. Besides

**Fig. 2** The differences between non-invasive and invasive bladder cancer cells



of that, a morphological and an immunologic heterogeneity in bladder cancer samples were found and only 69% cancers had a pure urothelial carcinoma histology [62]. Moreover, investigators detected significantly higher PD-L1 protein expression on tumour cells and the tumour-infiltrating immune cells in the squamous differentiation as compared to urothelial carcinoma histology regions of 14 of 15 tumours.

# **Tumour-infiltrating lymphocytes**

Tumour-infiltrating lymphocytes (TILs) are located in the tumour stroma, notably the papillary axis, around tumour cells and exhibit diverse functions depending on their phenotype. In the stroma, TILs usually form lymphoid aggregates in the papillary axis and only rarely infiltrate the epithelial part of the tumour as individual cells [63]. In high-grade cT1N0M0 NMIBC tumours, the median level of TILs was 20%, with a range of 5-60% [64]. In some studies, authors have reported no association between CD3<sup>+</sup>, CD4<sup>+</sup> or CD8<sup>+</sup> TILs and tumour pathological T-stages or grades, but they have found a significant correlation with the multiplicity of tumours [65, 66]. However, in another study the baseline TIL level was significantly higher in patients with dipper invasion as the TIL infiltration was significantly higher in the T1b substage compared with the T1a substage [64]. Roumiguié et al. [30] reported that the expression level of CD3<sup>+</sup> and CD8<sup>+</sup> T lymphocytes in the tumour stroma is different between Ta and T1 stages, with deeper infiltration in T1 tumours [30]. In addition, intratumoural CD8<sup>+</sup> T cell counts were lower in high-grade NMINC than in MIBC [60]. These data suggest that tumour aggressiveness is associated with the immune system. Moreover, the TIL density may be a marker of disease aggressiveness.

#### Natural killer cells

NK cells exert their antitumour effects without prior antigen exposure and are CD3<sup>-</sup>CD56<sup>+</sup> cells. They constitute the major element of the innate immune system and play a crucial role in shaping the early immune response to tumours [67, 68]. According to published data, patients with NMIBC tumour  $\leq$  3 cm had a significantly higher percentage of infiltrating CD56<sup>+</sup> cells in the TME compared with patients with larger tumours [66].

#### **Regulatory T cells**

Regulatory T cells (Tregs) are mainly FoxP3<sup>+</sup> cells that suppress immune response to maintain homeostasis and self-tolerance [69]. Tregs suppress the proliferation and differentiation of T cells, and they can also suppress activities of differentiated CD4<sup>+</sup> and CD8<sup>+</sup> T cells as well as NK cells, B cells, macrophages and dendritic cells [69]. The percentage of intratumoural FOXP3<sup>+</sup> T cells correlates with clinicopathological stages and high tumour grade [70, 71]. Surprisingly, researchers found significantly more FoxP3<sup>+</sup> Tregs in low-grade tumours from female patients compared with tumours from male patients [72].

#### **Tertiary lymphoid structures**

Ectopic tertiary lymphoid structures (TLS) describe areas specialised in B cell maturation that arise due to chronic inflammation and persistent antigen exposure. They resemble the germinal centre located in secondary lymphoid organs. Ectopic TLS could be associated with cancer prognosis, but data about the correlation between the outcome and TLS are contradictory. In ovarian, colorectal, breast and lung cancers, intratumoural B cells have been linked with a favourable prognosis, but in melanoma, prostate, renal cell and hepatocellular carcinoma, intratumoural B cells have been negatively correlated with the outcome [72–75]. TLS have also been detected in NMIBC samples. Koti et al. [75] reported TLS in 25% of the analysed NMIBC Ta low-grade tumours and 75% of the analysed MIBC cases. TLS identifiable by investigators contain CD20<sup>+</sup> B cells and CD21<sup>+</sup> follicular dendritic cell networks surrounded by CD3<sup>+</sup> and CD8<sup>+</sup> T cells. Furthermore, the authors highlighted the organisation of CD20<sup>+</sup> B cells in the immune component of the tumour and identified B cell clusters in 75% of tumours from the NMIBC Ta low-grade group, and in 81% of tumours from the MIBC group [76]. The authors speculated that the degree of TLS formation and its maturity may be associated with aggressiveness of the disease and the severity of stromal inflammatory response but the small number of analysed cases limits this considerations.

#### **Neutrophils**

Neutrophils are known as the first line of defence in the innate immune system. They have the ability to respond to multiple factors that regulate inflammation and the immune system as well as to modulate the activities of neighbouring cells [77]. Research suggests that tumour-associated neutrophils (TANs) have various antitumour properties, including direct cytotoxicity towards tumour cells and inhibition of metastasis. TANs are able to promote the angiogenic switch and stimulate tumour cell motility, migration and invasion, so they are connected to tumour progression [78]. Neutrophils can be polarised into either an antitumoural (N1) or a protumoural (N2) phenotype, each with distinct functions. N1 neutrophils induce cytotoxicity, mediating tumour destruction when N2 neutrophils support tumour progression. In the early phase of cancer development, neutrophils exert anticancer activity by reactive oxygen species (ROS) that cause tumour cell lysis and by the production of co-stimulatory molecules that enhance the proliferation of CD4<sup>+</sup> and CD8<sup>+</sup> T lymphocytes. In more advanced stages, neutrophils could take part in cancer progression by releasing growth-stimulating signals, matrix-degrading proteases and angiogenic factors [65]. The elevated level of tumourinfiltrating neutrophils (TINs) is significantly associated with the worst tumour T-stages and grades of patients [65].

#### **Tumour-associated macrophages**

Similarly to TANs, tumour-associated macrophages (TAMs) can be divided into two functionally different polarisation states, namely M1 and M2. M1 macrophages activate the adaptive immune system cells and can express nitric oxide synthase (iNOS), ROS and the cytokine interleukin 12 (IL-12), and they can destroy target cells. M2 macrophages promote angiogenesis, tissue reconstruction and tumourigenesis. BC cells also induce the polarisation of tissue-resident and reactive macrophages, potentially influencing tumour progression and treatment response [72]. Moreover, M1 and M2 macrophages have a high degree of plasticity and thus can be converted into each other upon TME changes [79]. Activated macrophages promote carcinogenesis through the expression of growth factors and matrix proteases, and they promote angiogenesis by suppressing the antitumoural immune response. Dufresne et al. [80] described that proinflammatory M1 macrophages should suppress tumour growth; instead, anti-inflammatory M2 macrophages, via production of IL-10 and other soluble factors, suppress the antitumoural effects of M1 macrophages [80].

The main antigens connected with macrophages are CD68 and CD163. CD68<sup>+</sup> cells (Mtot) have been found in all tested NMIBC specimens [81, 82]. CD163<sup>+</sup> TAMs have been found in the papillary axis, lymphoid aggregates

and the tumour stroma as well as in tumour islets. CD163<sup>+</sup> TAMs are mostly distributed around the microvasculature of the tumour stroma. Moreover, in some cases there is CD163 staining in the cytoplasm and membrane of tumour cells [83]. The mean expression level of tumour-infiltrating immune cell subsets is similar in women and men [84]. CD163<sup>+</sup> TAMs recognised as M2 have been found in the tumour stroma and tumour islets, mainly distributed around the microvasculature or between tumour cells [70, 85]. There are also data that CD68<sup>+</sup> TAM infiltration is significantly higher in the lamina propria without invasion compared with the neoplastic urothelium [84]. Moreover, there is positive CD163 staining in the cytoplasm and membrane of tumour cells[66].

Researchers have reported a significant correlation between CD68 and FOXP3 and a significant inverse correlation between CD68 and the CD4/CD8 ratio [84, 85]. In addition, TAM infiltration significantly correlates with counts of IL-6<sup>+</sup> cancer cells; IL-6 is one of the major proinflammatory cytokines in the TME [70].

In some cases, the proportion of CD68<sup>+</sup> cells correlates with stage and is higher in the group of patients with deeper muscle invasion. However, there are also reports that CD163<sup>+</sup> TAMs are not associated with the tumour stage, grade or lymphovascular invasion (LVI) [62, 81–84]. Xue at al. [86] found that macrophages and M2 macrophages are found significantly more frequently in high-grade compared with low-grade BC tumours: they constitute the largest proportion of the immune component[70, 87].

#### **B** cells

B cells express clonally differentiated immunoglobulin (Ig) receptors on their cell surface that recognise specific antigenic epitopes and are capable of producing a single species of antibody, with a unique antigen-binding site [88]. The most popular markers of B cells are CD19 and CD20 [89]. CD79 is also expressed almost exclusively on B cells [90]. It has been detected that more B cells were detected in BCa tissues than adjacent normal bladder tissues [91]. CD79a<sup>+</sup> B cell infiltration is higher in the epithelial and stromal compartments of high-grade tumours than in those of low-grade tumours from both sexes [72]. Researchers have also detected that CD20 expression was significantly increased in MIBC than in high-grade NMIBC [60].

#### Programmed death ligand 1

PD-L1 is a membrane protein expressed by cancer cells as well as some immune cells. Based on published studies, 26–46% of NMIBC cases express PD-L1 [63, 93–95]. According to published data, PD-L1 expression is more frequently detected on infiltrating immune cells than on cancer cells, and the degree of expression on immune cells differs among tumours. PD-L1 expression is absent in 26% of patients, focal in 39% of patients, mild in 25% of patients, moderate in 8% of patients and extensive in 1% of patients [62, 94, 95]. Wankowicz et al. [61] reported that only 4.6% of NMIBC cases presented PD-L1 expression on the cancer cell surface; however, other authors have detected significantly higher PD-L1 expression on the surface of cancer cells compared with immune cells [30, 62, 93]. Moreover, PD-L1 expression in the TME is higher in T1 than in Ta tumours [30, 93]. Investigators have also detected significantly higher expression of the immune checkpoint genes CTLA4, PDCD1, LAG3 and ICOS and PD-L1 protein expression in high-grade compared with low-grade tumours [73, 96]. Likewise, tissues collected from females showed higher expression of analysed genes than samples from males [72].

Eich et al. [97] reported the highest mean tumour cell expression of PD-L1 in clone E1L3N in LGTa tumours and invasive carcinomas defined as pT1; they found significantly lower expression in higher stages. HGTa tumours also show lower expression than LGTa tumours. In peritumoural lymphocytes, invasive tumours have higher PD-L1 expression than non-invasive tumours, such as CIS and LGTa and HGTa tumours. Moreover, HGTa tumours show markedly higher PD-L1 expression on lymphocytes than CIS and LGTa tumours [94]. Kates et al. [90] reported no PD-L1 expression on CIS samples.

PD-L1 correlates with immune infiltration in the TME. Fifty per cent of patients with dense immune infiltrate show PD-L1 expression on tumour cells (TC) compared with only 10% of patients with weak immune infiltration. In case of immune cells, researchers found PD-L1 expression in 47.2% of patients with a weak immune infiltrate versus 95.2% of patients with a dense infiltrate [96].

The differences in cancer phenotype depending on peripheral blood mononuclear cells Investigators have also assessed peripheral blood mononuclear cells(PBMC) in patients with high-risk NMIBC defined as T1, high grade, CIS or multiple recurrent large low-grade Ta tumours. They have found that the phenotype of PBMCs from patients with NMIBC is significantly different from healthy donors (HD) but the proportion of total circulating CD4<sup>+</sup> and CD8<sup>+</sup> T cells is similar between patients with cancer and HD. They also detected that the proportion of CD8<sup>+</sup> T cells is significantly higher in BCG responders than in BCG non-responders [98]. Audenet et al. [98] found that the ratio of circulating NK cells and the expression of Tim-3 and TIGIT in PBMC in patients with high-risk NMIBC is significantly higher compared with HD. The frequency of peripheral blood Tregs correlates with phases of cancer progression: the Treg frequency in pT1 tumours is higher than in pTa tumours.

Moreover, the fraction of CD4<sup>+</sup>FOXP3<sup>+</sup> T cells is significantly higher in the TIL compartment compared with PBMCs [99]. Patients with the highest percentages of peritumoural PD-L1<sup>+</sup> cells show a significantly lower number of peripheral blood lymphocytes [95]. The expression of programmed cell death protein 1 (PD-1) is low in PBMCs from both patients with high-risk NMIBC and HD; however, PD-1 expression is significantly higher on CD4<sup>+</sup> and CD8<sup>+</sup> T cells in patients with NMIBC [98].

## The non-immune components of the tumour microenvironment

The tumour microenvironment contains not only immune cells but also epithelium, extracellular matrix and cancerassociated fibroblasts (CAFs). It has been detected that fibroblast activation protein (FAP) expression which acts as a surrogate marker for CAFs was significantly higher in HG T1 tumours that progress to MIBC than in tumours that non-progress [100]. High expression of FAP and CD90 expressed by fibroblasts as well as platelet-derived growth factor receptor beta (PDGFRb) were furthermore correlated with higher grade of bladder tumour [101]. The other microenvironmental component connected with carcinogenesis is CD73 expression. CD73 (or ecto-5'-nucleotidase) is a cell surface protein that takes part in extracellular purinergic signalling by catalysing the hydrolysis of adenosine monophosphate (AMP) into adenosine and phosphate and, besides of it, CD73 inhibits T cell-mediated immune responses, the extravasation of leukocytes, increases neoangiogenesis, and improves barrier functions of epithelium and endothelium [102]. The CD73 is expressed by fibroblasts as well as by epithelium cells. On stromal fibroblasts, CD73 expression was found in only 22% of the patients in the NMIBC cohort. In case of endothelial cells, CD73 expression is correlated with tumour grade, T-category and CIS in NMIBC as the majority of grade 3/high-grade tumours had lost CD73 from the epithelium. Also basal cell layer of epithelium (BCL) CD73 expression correlated significantly with the tumour grade. In addition, CD73 expression on lymphocytes in NMIBC correlated with these same variables and lymphovascular invasion (LVI). CD73 positive stromal fibroblasts in NMIBC did not correlate with any of the parameters studied. Moreover, intratumoural lymphatic vessels were common in pT2 tumours, whereas only a few of pTa specimens showed intratumoural lymphatic vessels [103]. The density of lymphatic vessels corelated with higher pT stage, higher grade and sessile growth patterns. It has been also detected that circulating endothelial cells (CECs) is associated with higher tumour stage and grade [104].

# The influence of cellular disturbances in cancer cells on the immune component of the tumour

Cancer development and progression are associated with modifications in cell metabolism in order to supply energy for cell growth and proliferation. These modifications include increased oxygen consumption, the depletion of nutrients and the generation of reactive nitrogen and oxygen intermediates. The Warburg effect is the preferential use aerobic glycolysis and lactate fermentation as opposed to oxidative phosphorylation despite the presence of oxygen and fully functional mitochondria in order to supply the necessary production of energy, lipids, proteins and nucleic acids. It is one of crucial features of BC cells, similarly to other solid tumours.

This modification leads to changes in the activation of metabolic pathways and modifies the TME but, as mentioned before, it provides less efficient ATP production. For this reason, cancer cells have to increase glucose and glutamine uptake from the microenvironment by increasing expression of GLUT1 on their surface during the progression of cancer invasiveness [105, 106]. The increased glucose uptake from the extracellular surface leads to metabolic competition between effector T cells and tumour cells [107]. Because differentiated CD8<sup>+</sup> T cells show increased glucose-dependent metabolism compared with naïve cells, glucose deprivation negatively influences effector functions in CD8<sup>+</sup> T cells, resulting in impaired effector functions and possibly limited response to immune checkpoint therapy [107, 108]. Moreover, if glucose uptake is limited, proapoptotic Bcl-2 family members become activated, promoting cell death [109]. Researchers recently found that restricting glucose consumption in T cells might be minimised by inosine of fatty acid metabolism, but additional studies in this field are needed [110, 111]. In addition, human bladder transitional cell carcinoma cell line has upregulated PD-L1 expression by glutamine deprivation by the EGFR/MEK/ERK/c-Jun signalling pathway. Furthermore, PD-L1 upregulation and MEK/ERK/c-Jun pathway activation were reduced after glutamine recovery in vitro and in vivo [112].

The increased lactate production and its excessive extracellular concentration lead to acidification and, thereby, to the increased expression of hypoxia-inducible factor (HIF) target genes such as IL-8 and vascular endothelial growth factor (VEGF), which are proangiogenic factors [113–117]. Acidification of the TME also induces the expression of hyaluronic acid from tumour-associated fibroblasts [114, 118]. Moreover, the acidification contributes to decreased CD3<sup>+</sup>, CD4<sup>+</sup> and CD8<sup>+</sup> TIL infiltration, anergy and decreased T cell proliferation and

activation after antigen stimulation [119-122]. NK cells and neutrophils also show lower infiltration and decreased function due to reduced expression of granzyme B, perforin and some activating receptors such as NKp46, and by enhanced expression of myeloid-derived suppressor cells (MDSC) in the acidic environment [119, 123–126]. Exposure to lactate inhibits tumour necrosis factor (TNF) secretion and ROS production of monocytes and reduces the expression of pro-inflammatory cytokines [127, 128]. The high lactate concentration in the TME disturbs monocyte migration, reducing their effectiveness, impedes dendritic cell maturation and inhibits antigen presentation of dendritic cells. These changes lead to the acquisition of tumour-associated dendritic cell phenotypes that favour tumour progression [119, 129, 130]. Lactate is also one of the factors that contributes to switch inflammatory M1 macrophage polarisation towards the immunosuppressive M2 with a protumourigenic phenotype [117, 131, 132]. In contrast, Tregs do not seem to be sensitive to lactate and acidification [119, 133].

An important part of cancer metabolism is tryptophan catabolism into N-formyl-kynurenine, which is necessary to production of the energy cofactor NAD<sup>+</sup>. The first step of the catabolic pathway is mediated by IDO encoded by the IDO1 and IDO2 genes on human chromosome 8p11 and induced by pro-inflammatory cytokines such as interferon (IFN-). In a tumour, IDO can be detected in cancer cells, lymphocytes, dendritic cells, macrophages and others. Its leads to tryptophan deprivation that, similarly to glucose deprivation, disturbs immune cell functions [105].T, B and NK cells are very sensitive to tryptophan deficiency. They become anergic and show reduced proliferation as well as to increased sensitivity to Fas-mediated apoptosis due to the tryptophan shortage and the toxicity of tryptophan catabolites [134–139]. Tryptophan deprivation is also connected with the decreased expression of CD3<sup>+</sup> cells in the TME and with the polarisation of CD4<sup>+</sup> T cells into Tregs [140–142]. Moreover, IDO is committed to induce a tolerogenic phenotype of naïve dendritic cells by modifying their antigen-presenting ability due to the inhibition of the production of IL-12 and increased secretion of IL-10 and transforming growth factor beta (TGF- $\beta$ ) [143, 144]. Macrophages are also sensitive to tryptophan deficiency because one of its metabolites, 3-HAA, inhibits nuclear factor kappa B (NF-kB) and iNOS expression and, thereby, suppresses secretion of NO, one of the important factors necessary to eliminate tumour cells [145].

Immune cells compete with cancer cells for serine uptake as it is one of the important element of T cell activity upon its activation [146]. Cancer cells also consume more serine due to its usefulness in nucleotide production, methylation processes and NADH/NADPH pool renewal [147, 148]. The increased uptake of serine from the local environment might contribute to T cell function impairment.

A common feature of tumour tissues is hypoxia caused by oxygen diffusion limitations leading to HIF-1 $\alpha$  stabilisation. HIF-1 $\alpha$  promotes tumour neoangiogenesis as well as aerobic glycolysis by LDH-A and pyruvate dehydrogenase kinase 1 (PDK1) expression [149, 150]. Hypoxia suppresses T cell function and enhances Treg recruitment[150–153]. Moreover, HIF-1 $\alpha$  stimulates protumoural polarisation in TAMs by, inter alia, upregulated neuropilin-1 (NRP-1) and acidosis connected with anaerobic glycolysis and increased concentrations of lactic acid in the hypoxic TME. TAMs might increase T cell proliferation and cytotoxicity inhibition under hypoxic conditions and acidosis compared with T cell apoptosis promotion through the PD-L1/PD-1 pathway [154–156].

# The influence of the microenvironment on the outcome

BC tumours show abundant immune infiltration [157]. The leukocyte proportion of the tumour stromal fraction varies across immune subtypes of the tumours and is associated with overall survival (OS) and the progression-free interval (PFI). However, the association between the density and composition of immune infiltration and the outcome is unclear.

According to data published by Thorsson et al., [157] BC should be mainly described as C1 with elevated expression of angiogenic genes, a high proliferation rate and a Th2 cell bias to the adaptive immune infiltrate. A high proportion of BC tumours belongs to the C2 subtype characterised by high M1/M2 macrophage polarisation and a strong CD8 infiltration. Only a small portion of BC belongs to the C3 subtype with high Th17 and Th1 expression. According to the performed analyses, the C2 and C1 subtypes have less favourable outcomes than the C3 subtype despite having a substantial immune component. Moreover, a more pronounced lymphocyte signature is associated with improved outcome in C1 and C2 subtypes [157].

Investigators have found that higher CD3<sup>+</sup> and CD8<sup>+</sup> infiltration correlates with longer survival without the cancer in patients with NMIBC [63, 65]. He et al. [158] reported similar results: tumour-infiltrating immune cells vary depending on risk scores and the proportions of CD8<sup>+</sup> T cells, activated memory CD4<sup>+</sup> as well as T follicular helper cells, all of which are significantly higher in the low-risk group with a better prognosis [158]. Roumiguié et al.[30] found that the CD3<sup>+</sup>/CD8<sup>+</sup> T cell ratio in the TME is prognostic for disease-free survival (DFS) [30]. On the contrary, the authors of another study reported that the proportion of CD8<sup>+</sup> T cells in peripheral blood is significantly higher in BCG responders than in BCG non-responders [98]. Investigators have also found that patients with a higher CD4<sup>+</sup> T cell density in the TME have significantly shorter OS than patients with lower infiltration [150]. Despite the lack of a significant correlation between CD4<sup>+</sup> density in the TME and RFS, the 10-year RFS of patients with a higher CD4<sup>+</sup> T density was 1.7 times shorter than patients with a CD4<sup>+</sup> T density below the median [159]. Unfortunately, other data did not confirm this dependence, mainly regarding the range of CD4<sup>+</sup> TILs [63, 64]. Shorter OS was also correlated with high density of CD66b<sup>+</sup> neutrophils in NMIBC microenvironment [160]. Another group reported that the level of TME-infiltrating NK cells is significantly higher in the lowrisk NMIBC group than in high-risk group [158]. Unfortunately, there is also a report about no significant difference in infiltration by CD56<sup>+</sup> cells between patients with different outcomes, so it is possible that NK cells are engaged in the tumour elimination mainly in the early phase of tumour development due to development of specific immunosuppressive mechanisms during tumour progression [63].

Data about the connection between Tregs and the outcome of NMIBC are inconsistent, but the percentage of Tregs in the immune infiltration could be used as a predictor of tumour recurrence, progression and OS [70, 71, 158]. According to Murai et al. [71], the median RFS was 20 months for patients with high percentages of Foxp3<sup>+</sup> T cells and 113 months for patients with lower infiltration [71]. On the other hand, Eich et al. [97] found no association between Tregs and tumour recurrence [97], so additional studies are needed to validate the FOXP3<sup>+</sup> T cell function in BC.

The published data also indicate that high TAM infiltration is significantly associated with shorter OS [83, 87, 159]. High TAM counts have also been associated with shorter RFS in patients with NMIBC [70, 81, 82, 84]. Stromal CD79a<sup>+</sup> cell infiltration also influences RFS, but this dependence was confirmed only in univariate analysis [72].

An elevated level of TINs and a higher density of CD79a<sup>+</sup> B cells have also been independently correlated with shorter RFS [65, 72]. Also a significant association between higher CD20<sup>+</sup> cells density and worse disease-specific survival (DSS) was found; similarly, a trend towards shorter OS and DFS was observed [60]. It could be connected with BCa metastasis promotion by tumour-infiltrating B cells confirmed by Ou et al [91]. Moreover, a recently published study indicated the negative prognostic effect of a high preoperative neutrophil-to-lymphocyte ratio (NLR) [13, 161, 162]. Krpina et al. [63] reported no significant difference in infiltration of CD68<sup>+</sup> and CD20<sup>+</sup> cells between patients with an unfavourable outcome and those who did not develop recurrence [63].

Increased *CD274* mRNA expression (encodes PD-L1) has been reported to be a significant predictor of better RFS, mainly in pT1 tumours [46, 96, 97, 163, 164]. PFS

and cancer-specific survival could also be correlated with PD-L1 expression [163]. Unfortunately, no differences in the expression pattern of PD-L1 have been found between BCG responders and non-responders with NMIBC [93, 95]. Similarly, there is no correlation between tumour PD-L1 expression or tumour-infiltrating immune cells PD-L1 expression and the outcome in patients with NMIBC [60].

In addition, the outcome is dependent also from nonimmune component of the tumour microenvironment. Performed analyses indicated an association between shorter survival time and high expression for the stroma markers such as FAP and PDGFRb [100]. It seems also that patients with Ta stage and high PDGFRa expression and T1 stage patients with high ASMA expression had a worse 5-year overall survival [100]. In addition, in MIBC the presence of FAP is connected with worse CSS [100]. In case of CD73, the total epithelium expression in the NMIBC was associated with favourable PFS, whereas CD73-positive stromal fibroblasts associated with poor PFS [102]. Investigators found also that CEC served as an unfavourable predictor of RFS in low-risk NMIBC patients treated with TURBT [104].

# The influence of the immune system on the outcome in Bacillus Calmette– Guérin-treated patients

There are also data indicating that assessment of the immune system might be useful to estimate BCG treatment efficacy. Kates et al. [92] reported that a considerable group of BCG non-responders show pre-treatment co-localisation of PD-L1 in areas with a high density of CD8<sup>+</sup> cells and a low density of CD4<sup>+</sup> T cells, whereas BCG-responsive tumours are rich in CD8<sup>+</sup> and CD4<sup>+</sup> T cells and show almost no PD-L1 expression [92]. Moreover, assessment of the tuberculininduced frequencies and functionalities of cytokine-expressing CD4<sup>+</sup> T cells before and after BCG immunotherapy might be used as a marker for BCG treatment effectiveness before beginning induction. Investigators have detected that the higher frequencies of IFNy-producing CD4<sup>+</sup> T cells and higher amount of secreted IFNy before the induction treatment are characteristic of patients who do not show tumour recurrence at 6 months. The amount of secreted IL-2 differentiates the two groups of patients [165]. TAMs are associated with worse RFS in patients treated with intravesical BCG [166]. Moreover, in a meta-analysis published in 2018, the authors found that pre-treatment blood-based NLR is associated with an increased risk of disease recurrence and progression in BCG-treated patients with NMIBC after TURBT [167]. Investigators have also reported that a high NLR assessed within 180 days of BCG therapy could predict recurrence of the diseases [168].

There have been inconsistent data regarding how PD-L1 expression correlates with the BC outcome. The results have been dependent on the assessment methods and antibodies. Roumiguié et al. [30] found that PD-L1 expression in TC correlates with DFS and is higher in patients with disease recurrence. They found a significant correlation between PD-L1 expression and DFS when using the E1N3L antibody. However, when using the SP142/SP263 and 28.8 antibodies, they found higher PD-L1 expression in the unfavourable group but it was not significantly associated with DFS. Unfortunately, PD-L1 expression in the TME is not a biomarker for DFS[30]. Kates et al. [92] reported higher PD-L1 expression (for the SP142 and 22C3 antibodies) in BCG non-responders compared with BCG responders. Moreover, PD-L1<sup>+</sup> tissues from non-responders showed very low expression of CD4<sup>+</sup> T cells; however, among PD-L1<sup>-</sup> tissues from non-responders, CD4<sup>+</sup> T cells were common (60% for the Sp142 antibody, 50% for the 22C3 antibody). In addition, all PD-L1<sup>+</sup> samples had evidence of PD-L1 and CD8 co-localisation with an increased density of CD8<sup>+</sup> cells in areas of PD-L1 expression [91]. These data suggest that the long-term outcome could depend on the immune system and may play an important role in promoting BC recurrence and progression.

#### The influence of the immune system on the outcome in immune checkpoint inhibitor-treated patients

ICI treatment has emerged as an important therapeutic approach for patients with BC [169–172]. Unfortunately, only some patients who receive ICI treatment show benefits.

Song et al. [56] found that BC (NMIBC and MIBC) could be divided into four subgroups depending on their molecular and clinical characteristics. Tumours included in class 3 show a higher frequency of NMIBC progression and a higher somatic mutation rate compared with the other classes. In patients with class 3 tumours, there is a higher response rate to atezolizumab with a superior survival rate than in the other classes, indicating that patients with class 3 tumours might have a better response to ICI treatment [56]. There are other data about the connection between the TME elements and the response to ICI treatment: investigators have found that intratumoural CD4<sup>+</sup> and CD8<sup>+</sup> T cells correlate with the response to ICI treatment[173–176]. For urothelial cancer, M1 macrophages, CD8<sup>+</sup> T cells, activated memory CD4<sup>+</sup> T cells, and Tfh cells are significantly associated with response, while resting memory CD4<sup>+</sup> T cells are significantly associated with a lack of response. Moreover, infiltrated CD8<sup>+</sup> T cells are associated with OS [173]. There is a correlation between intraepithelial CD8+ T cell infiltration and pathological response to neoadjuvant atezolizumab treatment in MIBC; however, no significant correlation between PD-L1 expression on either immune cells or tumour cells and outcome after neoadjuvant atezolizumab treatment was found [31]. Besides of them, cytotoxic T cell transcriptional signature was significantly increased in responders compared to nonresponders and patients who relapsed [31]. Unfortunately, in the NABUCCO trial the investigators found no association between baseline CD8<sup>+</sup> T cell density and response to ipilimumab+nivolumab [177]. Moreover, there are a lack of biomarkers predictive of NMIBC response to ICI. Recently, it has been found that the outcome on atezolizumab adjuvant treatment after tumour resection in MIBC is depending on the circulating tumour DNA (ctDNA). Patients positive for circulating ctDNA had higher DFS and OS in atezolizumab arm, and no difference was found between groups who were negative for ctDNA [31]. In addition, conversion from ctDNA positive to ctDNA negative was detected in 18.2% of patients who were positive for ctDNA at the beginning of atezolizumab treatment compared with 3.8% of patients who were positive for ctDNA in the observation arm and patients with ctDNA conversion during the atezolizumab treatment had superior DFS compared with those who remained positive for ctD-NAt. Investigators found also that higher benefits from the atezolizumab treatment were connected with higher tumour mutational burden (TMB) and PD-L1 expression and worse outcomes detected in patients with high angiogenesis-related gene expression; however, in the ABACUS trial the pCR rate was not increased in TMB-high tumours so further studies are needed [31, 177].

Investigators found also that neoadjuvant atezolizumab treatment caused in MIBC a significant increase in intraepithelial CD8, PD-L1, FAP and granzyme B expression [31]. In addition, an increase in intra-epithelial CD8 levels occurred only in responding tumours, and FAP expression decrease in responders during the therapy. The taxonomy changes based on molecular differences occurred in 64.1% patients and 93.3% responsive tumours could be classified as 'infiltrated' after treatment, with increased immune infiltrate, angiogenesis and decreased cell cycle signatures; how-ever, changes to immune phenotypes occurred with therapy only in 18% patients.

## Conclusions

The landscape of immunotherapy has changed in recent years for BC. In metastatic BC, ICIs are currently recommended as a second-line treatment in case of progression after chemotherapy, as well as a first-line treatment in PD-L1-positive patients who are not eligible for cisplatin-based combination chemotherapy. ICIs are also indicated as a maintenance followed by cisplatin-based or carboplatinbased first-line chemotherapy. In NMIBC, ICIs are approved for treatment of BCG-unresponsive tumours. Despite the high efficacy of ICIs in some patients with BC, the clinical application of ICIs is restricted by its limited efficacy in some treated patients, meaningful adverse events and high costs. To optimise the obtainable treatment options and to individualise the treatment strategy, adequate biomarkers should be identified and utilised. Even though several molecular and metabolic biomarkers have been associated with the progression to the muscle layer, no biomarkers have been identified that are able to robustly select patients with NMIBC for a specific 'aggressive' treatment that prevents MIBC progression. Based on the published studies, TME and immune cell infiltration assessments have shown promise as a predictors of cancer progression and treatment response. These findings will hopefully reveal biomarkers that are easily evaluated by immunohistochemistry on routine pathology specimens, providing widely available and cost-effective tools for personalised medicine. Further understanding of tumour biology is a mandatory step to find a signature that can be used to choose appropriate treatments with both conventional and novel agents.

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#### Declarations

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#### References

- Sung H, Ferlay J, Siegel RL et al (2021) Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 71(3):209–249. https://doi.org/10.3322/caac.21660
- Compérat E, Larré S, Roupret M et al (2015) Clinicopathological characteristics of urothelial bladder cancer in patients less than 40 years old. Virchows Arch 466(5):589–594. https://doi.org/10. 1007/s00428-015-1739-2
- M Babjuk, M Burger, E Compérat, et al. (2021) EAU Guidelines. Edn. presented at the EAU annual congress milan. (EAU Guidelines Office, Arnhem, 2021) ISBN 978-94-92671-13-4.
- 4. http://onkologia.org.pl, dostęp z dnia 24.02.2022
- 5. https://seer.cancer.gov dostęp z dnia 24.02.2022
- Gakis G (2014) The role of inflammation in bladder cancer. Adv Exp Med Biol 816:183–196. https://doi.org/10.1007/ 978-3-0348-0837-8\_8
- Balkwill F, Mantovani A (2001) Inflammation and cancer: back to Virchow? Lancet 357(9255):539–545. https://doi.org/10.1016/ S0140-6736(00)04046-0
- Sui X, Lei L, Chen L, Xie T, Li X (2017) Inflammatory microenvironment in the initiation and progression of bladder cancer. Oncotarget 8(54):93279–93294. https://doi.org/10.18632/oncot arget.21565
- Alsheikh A, Mohamedali Z, Jones E, Masterson J, Gilks CB (2001) Comparison of the WHO/ISUP classification and cytokeratin 20 expression in predicting the behavior of low-grade papillary urothelial tumors. World/Health organization/International society of urologic pathology. Mod Pathol 14(4):267–272. https://doi.org/10.1038/modpathol.3880300
- Sylvester RJ, van der Meijden AP, Oosterlinck W et al (2006) Predicting recurrence and progression in individual patients with stage Ta T1 bladder cancer using EORTC risk tables: a combined analysis of 2596 patients from seven EORTC trials. Eur Urol 49(3):466–465. https://doi.org/10.1016/j.eururo.2005.12.031. (Discussion 475–477)
- Zieger K, Wolf H, Olsen PR, Hojgaard K (2000) Long-term follow-up of noninvasive bladder tumours (stage Ta): recurrence and progression. BJU Int 85(7):824–828. https://doi.org/ 10.1046/j.1464-410x.2000.00547.x
- Sylvester RJ, Rodríguez O, Hernández V et al (2021) European Association of Urology (EAU) prognostic factor risk groups for non-muscle-invasive bladder cancer (NMIBC) incorporating the WHO 2004/2016 and WHO 1973 classification systems for grade: an update from the EAU NMIBC guidelines panel. Eur Urol 79(4):480–488. https://doi.org/10.1016/j.eururo.2020.12. 033
- Chung JW, Kim JW, Lee EH et al (2021) Prognostic significance of the neutrophil-to-lymphocyte ratio in patients with nonmuscle invasive bladder cancer treated with intravesical Bacillus Calmette-Guérin and the relationship with the CUETO scoring model. Urol J. https://doi.org/10.22037/uj.v18i.6765
- Babjuk M, Burger M, Compérat EM et al (2019) European association of urology guidelines on non-muscle-invasive bladder cancer (TaT1 and Carcinoma In Situ)–2019 update. Eur Urol 76(5):639–657. https://doi.org/10.1016/j.eururo.2019.08.016
- EAU Guidelines. Edn. Presented at the EAU Annual Congress Amsterdam 2022. (EAU Guidelines Office, Arnhem, 2022) ISBN 978-94-92671-16-5
- 16. Powles T, Csőszi T, Özgüroğlu M et al (2021) Pembrolizumab alone or combined with chemotherapy versus chemotherapy as first-line therapy for advanced urothelial carcinoma (KEY-NOTE-361): a randomised, open-label, phase 3 trial. Lancet

Oncol 22(7):931–945. https://doi.org/10.1016/S1470-2045(21) 00152-2

- 17. Fradet Y, Bellmunt J, Vaughn DJ et al (2019) Randomized phase III KEYNOTE-045 trial of pembrolizumab versus paclitaxel, docetaxel, or vinflunine in recurrent advanced urothelial cancer: results of >2 years of follow-up. Ann Oncol 30(6):970–976. https://doi.org/10.1093/annonc/mdz127
- Larsen ES, Nordholm AC, Lillebaek T, Holden IK, Johansen IS (2019) The epidemiology of bacille Calmette-Guérin infections after bladder instillation from 2002 through 2017: a nationwide retrospective cohort study. BJU Int 124(6):910–916. https://doi. org/10.1111/bju.14793
- van der Meijden AP, Sylvester RJ, Oosterlinck W et al (2003) Maintenance Bacillus Calmette-Guerin for Ta T1 bladder tumors is not associated with increased toxicity: results from a European organisation for research and treatment of cancer genito-urinary group phase III trial. Eur Urol 44(4):429–434. https://doi.org/10. 1016/s0302-2838(03)00357-9
- Lamm DL, Blumenstein BA, Crissman JD et al (2000) Maintenance bacillus Calmette-Guerin immunotherapy for recurrent TA, T1 and carcinoma in situ transitional cell carcinoma of the bladder: a randomized Southwest oncology group study. J Urol 163:1124–1129
- 21. Sylvester RJ, Brausi MA, Kirkels WJ et al (2010) Long-term efficacy results of EORTC genito-urinary group randomized phase 3 study 30911 comparing intravesical instillations of epirubicin, bacillus Calmette-Guérin, and bacillus Calmette-Guérin plus isoniazid in patients with intermediate- and high-risk stage Ta T1 urothelial carcinoma of the bladder. Eur Urol 57(5):766–773. https://doi.org/10.1016/j.eururo.2009.12.024
- Kikuchi E, Hayakawa N, Fukumoto K, Shigeta K, Matsumoto K (2020) Bacillus Calmette-Guérin-unresponsive non-muscleinvasive bladder cancer: its definition and future therapeutic strategies. Int J Urol 27(2):108–116. https://doi.org/10.1111/iju. 14153
- Moschini M, Sharma V, Dell'oglio P et al (2016) Comparing long-term outcomes of primary and progressive carcinoma invading bladder muscle after radical cystectomy. BJU Int 117(4):604–610. https://doi.org/10.1111/bju.13146
- Schrier BP, Hollander MP, van Rhijn BW, Kiemeney LA, Witjes JA (2004) Prognosis of muscle-invasive bladder cancer: difference between primary and progressive tumours and implications for therapy. Eur Urol 45(3):292–296. https://doi.org/10.1016/j.eururo.2003.10.006
- Pfail JL, Katims AB, Alerasool P, Sfakianos JP (2021) Immunotherapy in non-muscle-invasive bladder cancer: current status and future directions. World J Urol 39(5):1319–1329. https://doi. org/10.1007/s00345-020-03474-8
- Roviello G, Catalano M, Santi R et al (2021) Immune checkpoint inhibitors in urothelial bladder cancer: state of the art and future perspectives. Cancers (Basel) 13(17):4411. https://doi.org/10. 3390/cancers13174411
- Brahmer JR, Tykodi SS, Chow LQ et al (2012) Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. N Engl J Med 366(26):2455–2465. https://doi.org/10.1056/NEJMo a1200694
- Topalian SL, Hodi FS, Brahmer JR et al (2012) Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. N Engl J Med 366(26):2443–2454. https://doi.org/10.1056/NEJMoa1200 690
- Bellmunt J, Hussain M, Gschwend JE et al (2021) IMvigor010 study group adjuvant atezolizumab versus observation in muscleinvasive urothelial carcinoma (IMvigor010): a multicentre, openlabel, randomised, phase 3 trial. Lancet Oncol 22(4):525–537. https://doi.org/10.1016/S1470-2045(21)00004-8

- Roumiguié M, Compérat E, Chaltiel L et al (2021) PD-L1 expression and pattern of immune cells in pre-treatment specimens are associated with disease-free survival for HR-NMIBC undergoing BCG treatment. World J Urol 39(11):4055–4065. https://doi.org/10.1007/s00345-020-03329-2
- Powles T, Kockx M, Rodriguez-Vida A et al (2019) Clinical efficacy and biomarker analysis of neoadjuvant atezolizumab in operable urothelial carcinoma in the ABACUS trial. Nat Med 25(11):1706–1714. https://doi.org/10.1038/s41591-019-0628-7
- 32. Szabados B, Kockx M, Assaf ZJ et al (2022) Final results of neoadjuvant Atezolizumab in Cisplatin-ineligible patients with muscle-invasive urothelial cancer of the bladder. Eur Urol 82(2):212–222. https://doi.org/10.1016/j.eururo.2022.04.013
- Kim HS, Seo HK (2018) Immune checkpoint inhibitors for urothelial carcinoma. Investig Clin Urol 59(5):285–296. https:// doi.org/10.4111/icu.2018.59.5.285
- 34. Amara CS, Vantaku V, Lotan Y, Putluri N (2019) Recent advances in the metabolomic study of bladder cancer. Expert Rev Proteomics 16(4):315–324. https://doi.org/10.1080/14789 450.2019.1583105
- Mitra AP (2016) Molecular substratification of bladder cancer: moving towards individualized patient management. Ther Adv Urol 8(3):215–233. https://doi.org/10.1177/1756287216638981
- Wu XR (2005) Urothelial tumorigenesis: a tale of divergent pathways. Nat Rev Cancer 5(9):713–725. https://doi.org/10.1038/ nrc1697
- Knowles MA (2006) Molecular subtypes of bladder cancer: Jekyll and Hyde or chalk and cheese? Carcinogenesis 27(3):361– 373. https://doi.org/10.1093/carcin/bgi310
- van Rhijn BW, van der Kwast TH, Vis AN et al (2004) FGFR3 and P53 characterize alternative genetic pathways in the pathogenesis of urothelial cell carcinoma. Cancer Res 64(6):1911– 1914. https://doi.org/10.1158/0008-5472.can-03-2421
- 39. Bakkar AA, Wallerand H, Radvanyi F et al (2003) FGFR3 and TP53 gene mutations define two distinct pathways in urothelial cell carcinoma of the bladder. Cancer Res 63(23):8108–8112
- 40. Orlow I, LaRue H, Osman I et al (1999) Deletions of the INK4A gene in superficial bladder tumors association with recurrence. Am J Pathol 155(1):105–113. https://doi.org/10.1016/S0002-9440(10)65105-X
- Mitra AP, Datar RH, Cote RJ (2006) Molecular pathways in invasive bladder cancer: new insights into mechanisms, progression, and target identification. J Clin Oncol 24(35):5552–5564. https:// doi.org/10.1200/JCO.2006.08.2073
- Knowles MA (2006) Molecular subtypes of bladder cancer: Jekyll and Hyde or chalk and cheese? Carcinogenesis 27(3):361– 373. https://doi.org/10.1093/carcin/bgi310
- Hurst CD, Alder O, Platt FM et al (2017) Genomic subtypes of non-invasive bladder cancer with distinct metabolic profile and female gender bias in KDM6A mutation frequency. Cancer Cell 32(5):701-715.e7. https://doi.org/10.1016/j.ccell.2017.08.005
- Hedegaard J, Lamy P, Nordentoft I et al (2016) Comprehensive transcriptional analysis of early-stage urothelial carcinoma. Cancer Cell 30:27–42. https://doi.org/10.1016/j.ccell.2016.05.004
- 45. Dyrskjøt L, Reinert T, Novoradovsky A et al (2012) Analysis of molecular intra-patient variation and delineation of a prognostic 12-gene signature in non-muscle invasive bladder cancer; technology transfer from microarrays to PCR. Br J Cancer 107:1392–1398. https://doi.org/10.1038/bjc.2012.412
- 46. Damrauer JS, Roell KR, Smith MA et al (2021) Identification of a novel inflamed tumor microenvironment signature as a predictive biomarker of Bacillus Calmette-Guérin immunotherapy in nonmuscle-invasive bladder cancer. Clin Cancer Res 27(16):4599– 4609. https://doi.org/10.1158/1078-0432.CCR-21-0205

- Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. Cell 144(5):646–674. https://doi.org/10.1016/j.cell. 2011.02.013
- Vander Heiden MG, Cantley LC, Thompson CB (2009) Understanding the Warburg effect: the metabolic requirements of cell proliferation. Science 324(5930):1029–1033. https://doi.org/10. 1126/science.1160809
- Afonso J, Santos LL, Longatto-Filho A, Baltazar F (2020) Competitive glucose metabolism as a target to boost bladder cancer immunotherapy. Nat Rev Urol 17(2):77–106. https://doi.org/10. 1038/s41585-019-0263-6
- Burns JE, Hurst CD, Knowles MA, Phillips RM, Allison SJ (2021) The Warburg effect as a therapeutic target for bladder cancers and intratumoral heterogeneity in associated molecular targets. Cancer Sci 112(9):3822–3834. https://doi.org/10.1111/ cas.15047
- 51. Conde VR, Oliveira PF, Nunes AR et al (2015) The progression from a lower to a higher invasive stage of bladder cancer is associated with severe alterations in glucose and pyruvate metabolism. Exp Cell Res 335(1):91–98. https://doi.org/10.1016/j.yexcr.2015.04.007
- Kouznetsova VL, Kim E, Romm EL, Zhu A, Tsigelny IF (2019) Recognition of early and late stages of bladder cancer using metabolites and machine learning. Metabolomics 15(7):94. https://doi.org/10.1007/s11306-019-1555-9
- Sahu D, Lotan Y, Wittmann B, Neri B, Hansel DE (2017) Metabolomics analysis reveals distinct profiles of nonmuscleinvasive and muscle-invasive bladder cancer. Cancer Med 6(9):2106–2120. https://doi.org/10.1002/cam4.1109
- Kim D, Kim JM, Kim JS, Kim S, Kim KH (2020) Differential expression and clinicopathological significance of HER2, indoleamine 2,3-dioxygenase and PD-L1 in urothelial carcinoma of the bladder. J Clin Med 9(5):1265. https://doi.org/10. 3390/jcm9051265
- 55. Santos HJSP, Matheus LHG, Silva A et al (2022) Indoleamine 2,3-dioxygenase-1 expression is changed during bladder cancer cell invasion. Int J Tryptophan Res 15:11786469211065612. https://doi.org/10.1177/11786469211065612
- 56. Song B, Kim S, Mun J et al (2019) Identification of an immunotherapy-responsive molecular subtype of bladder cancer. EBioMedicine 50:238–245. https://doi.org/10.1016/j.ebiom. 2019.10.058
- Lopez-Beltran A, Blanca A, Cimadamore A, Gogna R, Montironi R, Cheng L (2021) Molecular classification of bladder urothelial carcinoma using NanoString-based gene expression analysis. Cancers (Basel) 13(21):5500. https://doi.org/10.3390/ cancers13215500
- Zheng Z et al (2020) Dysregulation of the immune microenvironment contributes to malignant progression and has prognostic value in bladder cancer. Front Oncol 10:542492. https:// doi.org/10.3389/fonc.2020.542492
- 59. Zheng Y, Mao S, Zhang W et al (2020) Dysregulation of the immune microenvironment contributes to malignant progression and has prognostic value in bladder cancer. Front Oncol 10:542492. https://doi.org/10.3389/fonc.2020.542492
- Viveiros N, Flores BC, Lobo J et al (2022) Detailed bladder cancer immunoprofiling reveals new clues for immunotherapeutic strategies. Clin Transl Immunol 11(9):e1402. https:// doi.org/10.1002/cti2.1402
- Wankowicz SAM, Werner L, Orsola A et al (2017) Differential expression of PD-L1 in high grade T1 vs muscle invasive bladder carcinoma and its prognostic implications. J Urol 198(4):817–823. https://doi.org/10.1016/j.juro.2017.04.102
- 62. Warrick JI, Hu W, Yamashita H et al (2022) FOXA1 repression drives lineage plasticity and immune heterogeneity in

bladder cancers with squamous differentiation. Nat Commun 13(1):6575. https://doi.org/10.1038/s41467-022-34251-3

- Krpina K, Babarović E, Jonjić N (2015) Correlation of tumorinfiltrating lymphocytes with bladder cancer recurrence in patients with solitary low-grade urothelial carcinoma. Virchows Arch 467(4):443–448. https://doi.org/10.1007/ s00428-015-1808-6
- 64. Rouanne M, Betari R, Radulescu C et al (2019) Stromal lymphocyte infiltration is associated with tumour invasion depth but is not prognostic in high-grade T1 bladder cancer. Eur J Cancer 108:111–119. https://doi.org/10.1016/j.ejca.2018.12.010
- 65. Liu K, Zhao K, Wang L, Sun E (2018) The prognostic values of tumor-infiltrating neutrophils, lymphocytes and neutrophil/ lymphocyte rates in bladder urothelial cancer. Pathol Res Pract 214(8):1074–1080. https://doi.org/10.1016/j.prp.2018.05.010
- 66. Krpina K, Babarović E, Španjol J, Đorđević G, Maurer T, Jonjić N (2016) Correlation of tumor-associated macrophages and NK cells with bladder cancer size and T stage in patients with solitary low-grade urothelial carcinoma. Wien Klin Wochenschr 128(7–8):248–252. https://doi.org/10.1007/s00508-015-0907-3
- Abel AM, Yang C, Thakar MS, Malarkannan S (2018) Natural killer cells: development, maturation, and clinical utilization. Front Immunol 9:869. https://doi.org/10.3389/fimmu.2018.01869
- Paul S, Lal G (2017) The molecular mechanism of natural killer cells function and its importance in cancer immunotherapy. Front Immunol 8:1124. https://doi.org/10.3389/fimmu.2017.01124
- Sakaguchi S, Yamaguchi T, Nomura T, Ono M (2008) Regulatory T cells and immune tolerance. Cell 133(5):775–787. https://doi. org/10.1016/j.cell.2008.05.009
- 70. Miyake M, Tatsumi Y, Gotoh D et al (2017) Regulatory T cells and tumor-associated macrophages in the tumor microenvironment in non-muscle invasive bladder cancer treated with intravesical Bacille Calmette-Guérin: a long-term follow-up study of a Japanese cohort. Int J Mol Sci 18(10):2186. https://doi.org/10. 3390/ijms18102186
- Murai R, Itoh Y, Kageyama S et al (2018) Prediction of intravesical recurrence of non-muscle-invasive bladder cancer by evaluation of intratumoral Foxp3<sup>+</sup> T cells in the primary transurethral resection of bladder tumor specimens. PLoS One 13(9):e0204745. https://doi.org/10.1371/journal.pone.0204745
- Chenard S, Jackson C, Vidotto T et al (2021) Sexual dimorphism in outcomes of non-muscle-invasive bladder cancer: a role of CD163<sup>+</sup> macrophages, B cells, and PD-L1 immune checkpoint. Eur Urol Open Sci 29:50–58. https://doi.org/10.1016/j.euros. 2021.05.002
- Dieu-Nosjean MC, Antoine M, Danel C et al (2008) Long-term survival for patients with non-small-cell lung cancer with intratumoral lymphoid structures. J Clin Oncol 26(27):4410–4417. https://doi.org/10.1200/JCO.2007.15.0284
- Finkin S, Yuan D, Stein I et al (2015) Ectopic lymphoid structures function as microniches for tumor progenitor cells in hepatocellular carcinoma. Nat Immunol 16(12):1235–1244. https:// doi.org/10.1038/ni.3290
- Quinn DI, Shore ND, Egawa S, Gerritsen WR, Fizazi K (2015) Immunotherapy for castration-resistant prostate cancer: progress and new paradigms. Urol Oncol 33(5):245–260. https://doi.org/ 10.1016/j.urolonc.2014.10.009
- Koti M, Xu AS, Ren KYM et al (2017) Tertiary lymphoid structures associate with tumour stage in urothelial bladder cancer. Bladder Cancer 3(4):259–267. https://doi.org/10.3233/ BLC-170120
- Rosales C (2018) Neutrophil: a cell with many roles in inflammation or several cell types? Front Physiol 9:113. https://doi.org/ 10.3389/fphys.2018.00113

- Shaul ME, Fridlender ZG (2019) Tumour-associated neutrophils in patients with cancer. Nat Rev Clin Oncol 16(10):601–620. https://doi.org/10.1038/s41571-019-0222-4
- Pan Y, Yu Y, Wang X, Zhang T (2020) Tumor-associated macrophages in tumor immunity. Front Immunol 11:583084. https:// doi.org/10.3389/fimmu.2020.583084
- Dufresne M, Dumas G, Asselin E, Carrier C, Pouliot M, Reyes-Moreno C (2011) Pro-inflammatory type-1 and anti-inflammatory type-2 macrophages differentially modulate cell survival and invasion of human bladder carcinoma T24 cells. Mol Immunol 48(12–13):1556–1567. https://doi.org/10.1016/j.molimm.2011. 04.022
- Suriano F, Santini D, Perrone G et al (2013) Tumor associated macrophages polarization dictates the efficacy of BCG instillation in non-muscle invasive urothelial bladder cancer. J Exp Clin Cancer Res 32(1):87. https://doi.org/10.1186/1756-9966-32-87
- 82. Takayama H, Nishimura K, Tsujimura A et al (2009) Increased infiltration of tumor associated macrophages is associated with poor prognosis of bladder carcinoma in situ after intravesical bacillus Calmette-Guerin instillation. J Urol 181(4):1894–1900. https://doi.org/10.1016/j.juro.2008.11.090
- 83. Wang LA, Yang B, Tang T et al (2020) Correlation of APE1 with VEGFA and CD163<sup>+</sup> macrophage infiltration in bladder cancer and their prognostic significance. Oncol Lett 20(3):2881–2887. https://doi.org/10.3892/ol.2020.11814
- 84. Pichler R, Fritz J, Zavadil C, Schäfer G, Culig Z, Brunner A (2016) Tumor-infiltrating immune cell subpopulations influence the oncologic outcome after intravesical Bacillus Calmette-Guérin therapy in bladder cancer. Oncotarget 7(26):39916–39930. https://doi.org/10.18632/oncotarget.9537
- Lima L, Oliveira D, Tavares A et al (2014) The predominance of M2-polarized macrophages in the stroma of low-hypoxic bladder tumors is associated with BCG immunotherapy failure. Urol Oncol 32(4):449–457. https://doi.org/10.1016/j.urolonc.2013.10. 012
- Xue Y, Tong L, LiuAnwei Liu F et al (2019) Tumor-infiltrating M2 macrophages driven by specific genomic alterations are associated with prognosis in bladder cancer. Oncol Rep 42(2):581– 594. https://doi.org/10.3892/or.2019.7196
- Hanada T, Nakagawa M, Emoto A, Nomura T, Nasu N, Nomura Y (2000) Prognostic value of tumor-associated macrophage count in human bladder cancer. Int J Urol 7(7):263–269. https:// doi.org/10.1046/j.1442-2042.2000.00190.x
- Alberts B, Johnson A, Lewis J et al (2002) Molecular biology of the cell, 4th edn. Garland Science, New York
- Sanz I, Wei C, Jenks SA et al (2019) Challenges and opportunities for consistent classification of human B cell and plasma cell populations. Front Immunol 10:2458. https://doi.org/10.3389/ fimmu.2019.02458
- Chu PG, Arber DA (2001) CD79: a review. Appl Immunohistochem Mol Morphol 9(2):97–106. https://doi.org/10.1097/00129 039-200106000-00001
- Ou Z, Wang Y, Liu L et al (2015) Tumor microenvironment B cells increase bladder cancer metastasis via modulation of the IL-8/androgen receptor (AR)/MMPs signals. Oncotarget 6(28):26065–26078. https://doi.org/10.18632/oncotarget.4569
- 92. Kates M, Matoso A, Choi W et al (2020) Adaptive immune resistance to intravesical BCG in non-muscle invasive bladder cancer: implications for prospective BCG unresponsive trials. Clin Cancer Res 26(4):882–891. https://doi.org/10.1158/1078-0432.CCR-19-1920
- 93. Aydin AM, Baydar DE, Hazir B, Babaoglu B, Bilen CY (2020) Prognostic significance of pre- and post-treatment PD-L1 expression in patients with primary high-grade non-muscle-invasive bladder cancer treated with BCG immunotherapy. World J Urol 38(10):2537–2545. https://doi.org/10.1007/s00345-019-03065-2

- 94. Hashizume A, Umemoto S, Yokose T et al (2018) Enhanced expression of PD-L1 in non-muscle-invasive bladder cancer after treatment with Bacillus Calmette-Guerin. Oncotarget 9(75):34066–34078. https://doi.org/10.18632/oncotarget.26122
- 95. Martínez R, Tapia G, De Muga S et al (2019) Combined assessment of peritumoral Th1/Th2 polarization and peripheral immunity as a new biomarker in the prediction of BCG response in patients with high-risk NMIBC. Oncoimmunology 8(8):1602460. https://doi.org/10.1080/2162402X.2019.1602460
- 96. Blinova E, Buzdin A, Enikeev D et al (2020) Prognostic role of FGFR3 expression status and tumor-related microRNAs level in association with PD-L1 expression in primary luminal nonmuscular invasive bladder carcinoma. Life (Basel) 10(11):305. https://doi.org/10.3390/life10110305
- Eich ML, Chaux A, Guner G et al (2019) Tumor immune microenvironment in non-muscle-invasive urothelial carcinoma of the bladder. Hum Pathol 89:24–32. https://doi.org/10.1016/j.humpa th.2019.04.003
- Audenet F, Farkas AM, Anastos H, Galsky MD, Bhardwaj N, Sfakianos JP (2018) Immune phenotype of peripheral blood mononuclear cells in patients with high-risk non-muscle invasive bladder cancer. World J Urol 36(11):1741–1748. https:// doi.org/10.1007/s00345-018-2359-7
- 99. Winerdal ME, Krantz D, Hartana CA et al (2018) Urinary bladder cancer Tregs suppress MMP2 and potentially regulate invasiveness. Cancer Immunol Res 6(5):528–538. https://doi. org/10.1158/2326-6066.CIR-17-0466
- 100. Muilwijk T, Akand M, Daelemans S et al (2021) Stromal marker fibroblast activation protein drives outcome in T1 nonmuscle invasive bladder cancer. PLoS One 16(9):e0257195. https://doi.org/10.1371/journal.pone.0257195
- 101. Mezheyeuski A, Segersten U, Leiss LW et al (2020) Fibroblasts in urothelial bladder cancer define stroma phenotypes that are associated with clinical outcome. Sci Rep 10(1):281. https:// doi.org/10.1038/s41598-019-55013-0
- 102. Koivisto MK, Tervahartiala M, Kenessey I et al (2019) Celltype-specific CD73 expression is an independent prognostic factor in bladder cancer. Carcinogenesis 40(1):84–92. https:// doi.org/10.1093/carcin/bgy154
- 103. Bolenz C, Auer M, Ströbel P et al (2013) The lymphatic system in clinically localized urothelial carcinoma of the bladder: morphologic characteristics and predictive value. Urol Oncol 31(8):1606–1614. https://doi.org/10.1016/j.urolonc.2012.02. 012
- 104. Yang X, Lv J, Zhou Z et al (2022) Clinical application of circulating tumor cells and circulating endothelial cells in predicting bladder cancer prognosis and neoadjuvant chemosensitivity. Front Oncol 3(11):802188. https://doi.org/10.3389/fonc.2021. 802188
- 105. Domblides C, Lartigue L, Faustin B (2019) Control of the antitumor immune response by cancer metabolism. Cells 8(2):104. https://doi.org/10.3390/cells8020104
- 106. Reis H, Tschirdewahn S, Szarvas T, Rübben H, Schmid KW, Grabellus F (2011) Expression of GLUT1 is associated with increasing grade of malignancy in non-invasive and invasive urothelial carcinomas of the bladder. Oncol. Lett. 2(6):1149–1153. https:// doi.org/10.3892/ol.2011.394
- 107. Renner K, Bruss C, Schnell A et al (2019) Restricting glycolysis preserves T cell effector functions and augments checkpoint therapy. Cell Rep 29(1):135-150.e9. https://doi.org/10.1016/j. celrep.2019.08.068
- Cham CM, Driessens G, O'Keefe JP, Gajewski TF (2008) Glucose deprivation inhibits multiple key gene expression events and effector functions in CD8<sup>+</sup> T cells. Eur J Immunol 38(9):2438– 2450. https://doi.org/10.1002/eji.200838289

- Maciver NJ, Jacobs SR, Wieman HL, Wofford JA, Coloff JL, Rathmell JC (2008) Glucose metabolism in lymphocytes is a regulated process with significant effects on immune cell function and survival. J Leukoc Biol 84(4):949–957. https://doi.org/ 10.1189/jlb.0108024
- 110. Wang T, Gnanaprakasam JNR, Chen X et al (2020) Inosine is an alternative carbon source for CD8<sup>+</sup>-T-cell function under glucose restriction. Nat Metab 2(7):635–647. https://doi.org/10.1038/ s42255-020-0219-4
- 111. Zhang Y, Kurupati R, Liu L et al (2017) Enhancing CD8<sup>+</sup> T cell fatty acid catabolism within a metabolically challenging tumor microenvironment increases the efficacy of melanoma immunotherapy. Cancer Cell 32(3):377-391.e9. https://doi.org/10.1016/j. ccell.2017.08.004
- 112. Wang L, Xu T, Yang X et al (2021) Immunosuppression induced by glutamine deprivation occurs via activating PD-L1 transcription in bladder cancer. Front Mol Biosci 8:687305. https://doi. org/10.3389/fmolb.2021.687305
- 113. Baryła M, Semeniuk-Wojtaś A, Róg L, Kraj L, Małyszko M, Stec R (2022) Oncometabolites-a link between cancer cells and tumor microenvironment. Biology (Basel) 11(2):270. https://doi.org/10. 3390/biology11020270
- 114. Walenta S, Mueller-Klieser WF (2004) Lactate: mirror and motor of tumor malignancy. Semin Radiat Oncol 14(3):267–274. https://doi.org/10.1016/j.semradonc.2004.04.004
- 115. Polet F, Feron O (2013) Endothelial cell metabolism and tumour angiogenesis: glucose and glutamine as essential fuels and lactate as the driving force. J Intern Med 273(2):156–165. https://doi. org/10.1111/joim.12016
- 116. Fukumura D, Xu L, Chen Y, Gohongi T, Seed B, Jain RK (2001) Hypoxia and acidosis independently up-regulate vascular endothelial growth factor transcription in brain tumors in vivo. Cancer Res 61(16):6020–6024
- 117. Végran F, Boidot R, Michiels C, Sonveaux P, Feron O (2011) Lactate influx through the endothelial cell monocarboxylate transporter MCT1 supports an NF-κB/IL-8 pathway that drives tumor angiogenesis. Cancer Res 71(7):2550–2560. https://doi. org/10.1158/0008-5472.CAN-10-2828
- Rudrabhatla SR, Mahaffey CL, Mummert ME (2006) Tumor microenvironment modulates hyaluronan expression: the lactate effect. J Invest Dermatol 126(6):1378–1387. https://doi.org/10. 1038/sj.jid.5700255
- 119. Lacroix R, Rozeman EA, Kreutz M, Renner K, Blank CU (2018) Targeting tumor-associated acidity in cancer immunotherapy. Cancer Immunol Immunother 67(9):1331–1348. https://doi.org/ 10.1007/s00262-018-2195-z
- Shime H, Yabu M, Akazawa T et al (2008) Tumor-secreted lactic acid promotes IL-23/IL-17 proinflammatory pathway. J Immunol 180(11):7175–7183. https://doi.org/10.4049/jimmunol.180.11. 7175
- 121. Fischer K, Hoffmann P, Voelkl S et al (2007) Inhibitory effect of tumor cell-derived lactic acid on human T cells. Blood 109(9):3812–3819. https://doi.org/10.1182/ blood-2006-07-035972
- 122. Calcinotto A, Filipazzi P, Grioni M et al (2012) Modulation of microenvironment acidity reverses anergy in human and murine tumor-infiltrating T lymphocytes. Cancer Res 72(11):2746–2756. https://doi.org/10.1158/0008-5472.CAN-11-1272
- 123. Husain Z, Huang Y, Seth P, Sukhatme VP (2013) Tumor-derived lactate modifies antitumor immune response: effect on myeloidderived suppressor cells and NK cells. J Immunol 191(3):1486– 1495. https://doi.org/10.4049/jimmunol.1202702
- Lardner A (2001) The effects of extracellular pH on immune function. J Leukoc Biol 69(4):522–530
- 125. Loeffler DA, Juneau PL, Masserant S (1992) Influence of tumour physico-chemical conditions on interleukin-2-stimulated

lymphocyte proliferation. Br J Cancer 66(4):619–622. https://doi. org/10.1038/bjc.1992.326

- 126. Severin T, Müller B, Giese G et al (1994) pH-dependent LAK cell cytotoxicity. Tumour Biol 15(5):304–310. https://doi.org/10. 1159/000217905
- Dietl K, Renner K, Dettmer K et al (2010) Lactic acid and acidification inhibit TNF secretion and glycolysis of human monocytes. J Immunol 184(3):1200–1209. https://doi.org/10.4049/jimmunol. 0902584
- Peter K, Rehli M, Singer K, Renner-Sattler K, Kreutz M (2015) Lactic acid delays the inflammatory response of human monocytes. Biochem Biophys Res Commun 457(3):412–418. https:// doi.org/10.1016/j.bbrc.2015.01.005
- Gottfried E, Kunz-Schughart LA, Ebner S et al (2006) Tumorderived lactic acid modulates dendritic cell activation and antigen expression. Blood 107(5):2013–2021. https://doi.org/10.1182/ blood-2005-05-1795
- 130. Goetze K, Walenta S, Ksiazkiewicz M, Kunz-Schughart LA, Mueller-Klieser W (2011) Lactate enhances motility of tumor cells and inhibits monocyte migration and cytokine release. Int J Oncol 39(2):453–463. https://doi.org/10.3892/ijo.2011.1055
- Colegio OR, Chu NQ, Szabo AL et al (2014) Functional polarization of tumour-associated macrophages by tumour-derived lactic acid. Nature 513(7519):559–563. https://doi.org/10. 1038/nature13490
- 132. Zhao Y, Wang D, Xu T et al (2015) Bladder cancer cells reeducate TAMs through lactate shuttling in the microfluidic cancer microenvironment. Oncotarget 6(36):39196–39210. https://doi.org/10.18632/oncotarget.5538
- 133. Michalek RD, Gerriets VA, Jacobs SR et al (2011) Cutting edge: distinct glycolytic and lipid oxidative metabolic programs are essential for effector and regulatory CD4<sup>+</sup> T cell subsets. J Immunol 186(6):3299–3303. https://doi.org/10.4049/ jimmunol.1003613
- 134. Frumento G, Rotondo R, Tonetti M, Ferrara GB (2011) T cell proliferation is blocked by indoleamine 2,3-dioxygenase. Transplant Proc 33(1–2):428–430. https://doi.org/10.1016/ s0041-1345(00)02078-9
- Mellor AL, Keskin DB, Johnson T, Chandler P, Munn DH (2002) Cells expressing indoleamine 2,3-dioxygenase inhibit T cell responses. J Immunol 168(8):3771–3776. https://doi.org/ 10.4049/jimmunol.168.8.3771
- 136. Cheong JE, Sun L (2018) Targeting the IDO1/TDO2-KYN-AhR pathway for cancer immunotherapy - challenges and opportunities. Trends Pharmacol Sci 39(3):307–325. https:// doi.org/10.1016/j.tips.2017.11.007
- Lee GK, Park HJ, Macleod M et al (2002) Tryptophan deprivation sensitizes activated T cells to apoptosis prior to cell division. Immunology 107(4):452–460. https://doi.org/10.1046/j. 1365-2567.2002.01526.x
- 138. Terness P, Bauer TM, Röse L et al (2002) Inhibition of allogeneic T cell proliferation by indoleamine 2,3-dioxygenaseexpressing dendritic cells: mediation of suppression by tryptophan metabolites. J Exp Med 196(4):447–457. https://doi. org/10.1084/jem.20020052
- 139. Della Chiesa M, Carlomagno S, Frumento G et al (2006) The tryptophan catabolite L-kynurenine inhibits the surface expression of NKp46- and NKG2D-activating receptors and regulates NK-cell function. Blood 108(13):4118–4125. https://doi.org/ 10.1182/blood-2006-03-006700
- 140. Brandacher G, Perathoner A, Ladurner R et al (2006) Prognostic value of indoleamine 2,3-dioxygenase expression in colorectal cancer: effect on tumor-infiltrating T cells. Clin Cancer Res 12(4):1144–1151. https://doi.org/10.1158/1078-0432. CCR-05-1966

- 141. Munn DH, Sharma MD, Baban B et al (2005) GCN2 kinase in T cells mediates proliferative arrest and anergy induction in response to indoleamine 2,3-dioxygenase. Immunity 22(5):633–642. https://doi.org/10.1016/j.immuni.2005.03.013
- 142. Fallarino F, Grohmann U, You S et al (2006) The combined effects of tryptophan starvation and tryptophan catabolites down-regulate T cell receptor zeta-chain and induce a regulatory phenotype in naive T cells. J Immunol 176(11):6752– 6761. https://doi.org/10.4049/jimmunol.176.11.6752
- 143. Ravishankar B, Liu H, Shinde R et al (2015) The amino acid sensor GCN2 inhibits inflammatory responses to apoptotic cells promoting tolerance and suppressing systemic autoimmunity. Proc Natl Acad Sci USA. 112(34):10774–10779. https://doi.org/10.1073/pnas.1504276112
- 144. Fallarino F, Grohmann U, Puccetti P (2012) Indoleamine 2,3-dioxygenase: from catalyst to signaling function. Eur J Immunol 42(8):1932–1937. https://doi.org/10.1002/eji.20124 2572
- 145. Wang XF, Wang HS, Wang H et al (2014) The role of indoleamine 2,3-dioxygenase (IDO) in immune tolerance: focus on macrophage polarization of THP-1 cells. Cell Immunol 289(1–2):42– 48. https://doi.org/10.1016/j.cellimm.2014.02.005
- 146. Ma EH, Bantug G, Griss T et al (2017) Serine is an essential metabolite for effector T cell expansion. Cell Metab 25(2):345– 357. https://doi.org/10.1016/j.cmet.2016.12.011
- 147. Possemato R, Marks KM, Shaul YD et al (2011) Functional genomics reveal that the serine synthesis pathway is essential in breast cancer. Nature 476(7360):346–350. https://doi.org/10. 1038/nature10350
- Maddocks OD, Berkers CR, Mason SM et al (2013) Serine starvation induces stress and p53-dependent metabolic remodelling in cancer cells. Nature 493(7433):542–546. https://doi.org/10. 1038/nature11743
- 149. Kim JW, Tchernyshyov I, Semenza GL, Dang CV (2006) HIF-1-mediated expression of pyruvate dehydrogenase kinase: a metabolic switch required for cellular adaptation to hypoxia. Cell Metab 3(3):177–185. https://doi.org/10.1016/j.cmet.2006.02.002
- 150. Semenza GL, Jiang BH, Leung SW et al (1996) Hypoxia response elements in the aldolase A, enolase 1, and lactate dehydrogenase A gene promoters contain essential binding sites for hypoxia-inducible factor 1. J Biol Chem 271(51):32529–32537. https://doi.org/10.1074/jbc.271.51.32529
- 151. Kono K, Salazar-Onfray F, Petersson M et al (1996) Hydrogen peroxide secreted by tumor-derived macrophages down-modulates signal-transducing zeta molecules and inhibits tumorspecific T cell-and natural killer cell-mediated cytotoxicity. Eur J Immunol 26(6):1308–1313. https://doi.org/10.1002/eji.18302 60620
- 152. Hansen W, Hutzler M, Abel S et al (2012) Neuropilin 1 deficiency on CD4<sup>+</sup>Foxp3<sup>+</sup> regulatory T cells impairs mouse melanoma growth. J Exp Med 209(11):2001–2016. https://doi.org/10. 1084/jem.20111497
- 153. Deng B, Zhu JM, Wang Y et al (2013) Intratumor hypoxia promotes immune tolerance by inducing regulatory T cells via TGFβ1 in gastric cancer. PLoS One 8(5):e63777. https://doi.org/10. 1371/journal.pone.0063777
- 154. Doedens AL, Stockmann C, Rubinstein MP et al (2010) Macrophage expression of hypoxia-inducible factor-1 alpha suppresses T-cell function and promotes tumor progression. Cancer Res 70(19):7465–7475. https://doi.org/10.1158/0008-5472. CAN-10-1439
- He Z, Zhang S (2021) Tumor-associated macrophages and their functional transformation in the hypoxic tumor microenvironment. Front Immunol 12:741305. https://doi.org/10.3389/fimmu. 2021.741305

- 156. Shan T, Chen S, Chen X et al (2020) M2-TAM subsets altered by lactic acid promote T-cell apoptosis through the PD-L1/PD-1 pathway. Oncol Rep 44(5):1885–1894. https://doi.org/10.3892/ or.2020.7767
- 157. Thorsson V, Gibbs DL, Brown SD et al (2019) The immune landscape of cancer. Immunity 51(2):411–412. https://doi.org/ 10.1016/j.immuni.2019.08.004.Erratumfor:Immunity.2018;48(4) ,812-830.e14
- 158. He Y, Wu Y, Liu Z et al (2021) Identification of signature genes associated with invasiveness and the construction of a prognostic model that predicts the overall survival of bladder cancer. Front Genet 12:694777. https://doi.org/10.3389/fgene.2021.694777
- 159. Zhang Q, Hao C, Cheng G et al (2015) High CD4<sup>+</sup> T cell density is associated with poor prognosis in patients with non-muscleinvasive bladder cancer. Int J Clin Exp Pathol 8(9):11510–11516
- 160. Yang M, Wang B, Hou W et al (2022) Negative effects of stromal neutrophils on T cells reduce survival in resectable urothelial carcinoma of the bladder. Front Immunol 13:827457. https://doi. org/10.3389/fimmu.2022.827457
- Vartolomei MD, Ferro M, Cantiello F et al (2018) Validation of neutrophil-to-lymphocyte ratio in a multi-institutional cohort of patients with T1G3 non-muscle-invasive bladder cancer. Clin Genitourin Cancer 16(6):445–452. https://doi.org/10.1016/j.clgc. 2018.07.003
- 162. Yıldız HA, Değer MD, Aslan G (2021) Prognostic value of preoperative inflammation markers in non-muscle invasive bladder cancer. Int J Clin Pract 75(6):e14118. https://doi.org/10.1111/ ijcp.14118
- 163. Breyer J, Wirtz RM, Otto W et al (2018) High PDL1 mRNA expression predicts better survival of stage pT1 non-muscleinvasive bladder cancer (NMIBC) patients. Cancer Immunol Immunother 67(3):403–412. https://doi.org/10.1007/ s00262-017-2093-9
- 164. Kubon J, Sikic D, Eckstein M et al (2020) Analysis of CXCL9, PD1 and PD-L1 mRNA in stage T1 non-muscle invasive bladder cancer and their association with prognosis. Cancers (Basel) 12(10):2794. https://doi.org/10.3390/cancers12102794
- 165. Jallad S, Thomas P, Newport MJ, Kern F (2018) Baseline cytokine profiles of tuberculin-specific CD4<sup>+</sup> T cells in nonmuscle-invasive bladder cancer may predict outcomes of BCG immunotherapy. Cancer Immunol Res 6(10):1212–1219. https:// doi.org/10.1158/2326-6066.CIR-18-0046
- 166. Kardoust Parizi M, Shariat SF, Margulis V, Mori K, Lotan Y (2011) Value of tumour-infiltrating immune cells in predicting response to intravesical BCG in patients with non-muscle-invasive bladder cancer: a systematic review and meta-analysis. BJU Int 127(6):617–625. https://doi.org/10.1111/bju.15276
- 167. Vartolomei MD, Porav-Hodade D, Ferro M et al (2018) Prognostic role of pretreatment neutrophil-to-lymphocyte ratio (NLR) in patients with non-muscle-invasive bladder cancer (NMIBC) A systematic review and meta-analysis. Urol Oncol 36(9):389–399. https://doi.org/10.1016/j.urolonc.2018.05.014
- 168. Plasek J, Weissert J, Downs T, Richards K, Ravvaz K (2021) Clinicopathological criteria predictive of recurrence following Bacillus Calmette-Guérin therapy initiation in non-muscle-invasive bladder cancer: retrospective cohort study. JMIR Cancer 7(2):e25800. https://doi.org/10.2196/25800
- 169. Vuky J, Balar AV, Castellano D et al (2020) Long-term outcomes in KEYNOTE-052: phase II study investigating first-line pembrolizumab in cisplatin-ineligible patients with locally advanced or metastatic urothelial cancer. J Clin Oncol 38(23):2658–2666. https://doi.org/10.1200/JCO.19.01213
- 170. van der Heijden MS, Loriot Y, Durán I et al (2021) Atezolizumab versus chemotherapy in patients with platinum-treated locally advanced or metastatic urothelial carcinoma: a long-term overall survival and safety update from the phase 3 IMvigor211 clinical

trial. Eur Urol 80(1):7–11. https://doi.org/10.1016/j.eururo.2021. 03.024

- 171. Balar AV, Kamat AM, Kulkarni GS et al (2021) Pembrolizumab monotherapy for the treatment of high-risk non-muscle-invasive bladder cancer unresponsive to BCG (KEYNOTE-057), an open-label, single-arm, multicentre, phase 2 study. Lancet Oncol 22(7):919–930. https://doi.org/10.1016/S1470-2045(21)00147-9
- 172. Bajorin DF, Witjes JA, Gschwend JE et al (2021) Adjuvant nivolumab versus placebo in muscle-invasive urothelial carcinoma. N Engl J Med 384(22):2102–2114. https://doi.org/10. 1056/NEJMoa2034442
- 173. Liu R, Yang F, Yin JY, Liu YZ, Zhang W, Zhou HH (2021) Influence of tumor immune infiltration on immune checkpoint inhibitor therapeutic efficacy: a computational retrospective study. Front Immunol 12:685370. https://doi.org/10.3389/fimmu. 2021.685370
- 174. Giraldo NA, Becht E, Pagès F et al (2015) Orchestration and prognostic significance of immune checkpoints in the microenvironment of primary and metastatic renal cell cancer. Clin

Cancer Res 21(13):3031–3040. https://doi.org/10.1158/1078-0432.CCR-14-2926

- 175. Sade-Feldman M, Yizhak K, Bjorgaard SL et al (2018) Defining T cell states associated with response to checkpoint immunotherapy in melanoma. Cell 175(4):998-1013.e20. https://doi.org/ 10.1016/j.cell.2018.10.038
- Tumeh PC, Harview CL, Yearley JH et al (2014) PD-1 blockade induces responses by inhibiting adaptive immune resistance. Nature 515(7528):568–571. https://doi.org/10.1038/nature13954
- 177. Powles T, Assaf ZJ, Davarpanah N et al (2021) ctDNA guiding adjuvant immunotherapy in urothelial carcinoma. Nature 595(7867):432-437. https://doi.org/10.1038/ s41586-021-03642-9

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