



Engineering of the unfolded protein response pathway in *Pichia pastoris*: enhancing production of secreted recombinant proteins

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Abstract

Folding and processing of proteins in the endoplasmic reticulum (ER) are major impediments in the production and secretion of proteins from *Pichia pastoris* (*Komagataella* sp.). Overexpression of recombinant genes can overwhelm the innate secretory machinery of the *P. pastoris* cell, and incorrectly folded proteins may accumulate inside the ER. To restore proper protein folding, the cell naturally triggers an unfolded protein response (UPR) pathway, which upregulates the expression of genes coding for chaperones and other folding-assisting proteins (e.g., Kar2p, Pdi1, Ero1p) via the transcription activator Hac1p. Unfolded/misfolded proteins that cannot be repaired are degraded via the ER-associated degradation (ERAD) pathway, which decreases productivity. Co-expression of selected UPR genes, along with the recombinant gene of interest, is a common approach to enhance the production of properly folded, secreted proteins. Such an approach, however, is not always successful and sometimes, protein productivity decreases because of an unbalanced UPR. This review summarizes successful chaperone co-expression strategies in *P. pastoris* that are specifically related to overproduction of foreign proteins and the UPR. In addition, it illustrates possible negative effects on the cell's physiology and productivity resulting from genetic engineering of the UPR pathway. We have focused on *Pichia*'s potential for commercial production of valuable proteins and we aim to optimize molecular designs so that production strains can be tailored to suit a specific heterologous product.

Key points

- Chaperones co-expressed with recombinant genes affect productivity in *P. pastoris*.
- Enhanced UPR may impair strain physiology and promote protein degradation.
- Gene copy number of the target gene and the chaperone determine the secretion rate.

Keywords *Pichia pastoris* · Productivity of recombinant protein production · Folding and secretion · Unfolded protein response (UPR) · Chaperone · Co-expression strategy

Introduction

The methylotrophic yeast *Pichia pastoris* (*Komagataella phaffii*) is an established platform for applied research, specifically for the biotechnological production of a wide range of recombinant proteins. These include various intracellular, membrane and surface-displayed proteins and, most importantly, recombinant proteins that are secreted in large quantities (Cereghino and Cregg 2000; Daly and Hearn 2005; Gasser et al. 2013; Emmerstorfer et al. 2014; Spohner et al. 2015). The ability of *P. pastoris* to efficiently secrete recombinant proteins of unparalleled high quality (i.e., correctly folded, and post-translationally modified, without contamination from other proteins) makes this yeast an appropriate host for the industrial production of biopharmaceuticals or

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commercially valuable enzymes. Protein secretion is a multi-step process, involving various cellular compartments (Fig. 1). After the transcription of its recombinant gene in the nucleus, the protein is synthesized, folded, and post-translationally modified in the endoplasmic reticulum (ER). From there, it is translocated in COPII vesicles to the Golgi apparatus (Antonny and Schekman 2001), where the post-translational modifications are finalized. The protein is then packed and shipped in a vesicle towards its destination, which, in the case of proteins intended for secretion, is towards the cell membrane. The vesicles fuse with the cell membrane and the protein is finally released to the extracellular environment (Puxbaum et al. 2015). It was found that in *P. pastoris*, recombinant proteins aimed for secretion, localized in the ER, are inherited during cell division to buds, from whence exocytosis of these soluble proteins predominantly occurs (Puxbaum et al. 2016).

Information about secretory mechanisms in *P. pastoris* are still mainly based on knowledge derived from the model yeast *S. cerevisiae*. Nevertheless, information about the *P. pastoris* cell factory has advanced over the last decade due to whole genome characterization (De Schutter et al. 2009), available

omics analyses (Zahl et al. 2017), and the development of novel tools facilitating genomic engineering, such as CRISPR-Cas9 technology (Weninger et al. 2016; Raschmanová et al. 2018; Weninger et al. 2018). Based on genomic comparisons of different yeast species and mammals, it was shown that some patterns of *P. pastoris*'s secretory pathway resemble those of mammalian cells rather than those of *S. cerevisiae* (Delic et al. 2013). For example, structural organization of the Golgi compartment differs in *S. cerevisiae* and *P. pastoris*; the Golgi apparatus in *P. pastoris* is arranged in stacks and embedded in a ribosome-excluding matrix, which is similar to mammalian and plant cells (Rossanese et al. 1999; Mogelsvang et al. 2003). Also, some patterns of response to ER stress observed in *P. pastoris* resemble those of mammalian cells (Graf et al. 2008). These results indicate that valid conclusions for *P. pastoris* cannot be generally drawn from the model species *S. cerevisiae*. More intensive basic research on the secretory pathway and its bottlenecks in *P. pastoris* is needed, to effectively optimize the production/secretion of recombinant proteins by this host.

Because of the complex character of the secretory pathway, optimization of productivity of secreted proteins is

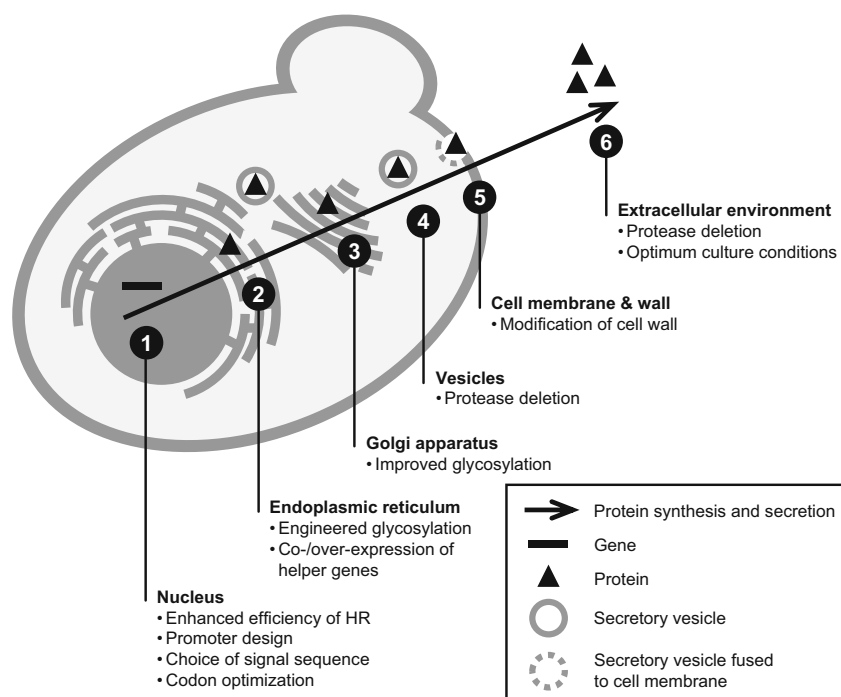


Fig. 1 Approaches to enhance recombinant protein secretion in *P. pastoris*. The production and secretion of recombinant protein can be enhanced by different approaches, aimed at different stages of the recombinant protein's production and secretion. By improving the rate of homologous recombination (HR), the integration of (multiple) expression cassettes is enhanced. The expression level of the heterologous gene is determined by the promoter used, and processing and secretion of the protein can be improved by its codon optimization and the choice of a suitable secretion signal sequence, respectively. Correct glycosylation can be ensured in glycoengineered production

strains, and folding or building of disulfide bridges might be enhanced by co-expressing chaperone or other helper genes. The intracellular proteolytic degradation of the recombinant proteins can be avoided by deletion of genes encoding proteases. The release of the proteins to the extracellular environment may be enhanced by modifications of the cell membrane and cell wall. Stability of the secreted protein in the extracellular environment is preserved by the choice of appropriate cultivation conditions (pH, temperature) and can be improved by the deletion of genes encoding secreted proteases

challenging and often requires a combinatorial approach. The right optimization strategy seems not to be generally predictable, even for proteins of similar structures and properties (Obst et al. 2017), so, unfortunately, it must be designed for each protein specifically. Production/secretion might be generally optimized at various levels: the expression cassette (promoter engineering, secretion signal sequences, codon optimization etc.), the host strain (co-/overexpression of chaperone genes or genes of other folding-assisting proteins, co-expression of transcription- and translation-enhancing elements, disruption of protease genes, modification of cell wall properties etc.), the cultivation conditions (pH, temperature), and the bioprocess strategy (specific growth rate of biomass etc.) (Marx et al. 2006; Emmerstorfer et al. 2014; Looser et al. 2015; Barrero et al. 2018; Gidijala et al. 2018; Zepeda et al. 2018; Duan et al. 2019; Fischer and Glieder 2019; Liu et al. 2019; Naranjo et al. 2019) (Fig. 1).

An extensive effort has been mounted to increase protein secretion by co-expression of different folding factor genes involved in the UPR pathway. However, this strategy has not been successful in all cases and rather, has been applied on an ad hoc basis. In this review, we have analyzed published data on co-expression strategies, with the aim of identifying the best strategy to enhance recombinant protein production/secretion in *P. pastoris*. Importantly, we also point out the undesirable effects on strain physiology and production, potentially resulting from the co-expression of folding factor genes, i.e., an unbalancing of the UPR pathway by its genetic engineering.

Secretion bottlenecks and unfolded protein response (UPR) in *P. pastoris*

Proteins intended for secretion enter the lumen of the ER through the Sec61 protein-translocation channel (Marsalek et al. 2019). Integral membrane proteins (except for peroxisomal and mitochondrial membrane proteins) also enter the secretory pathway, starting in the ER (Emmerstorfer et al. 2014). In the lumen of the ER, post-translational modifications and folding take place. Correct folding of the proteins is ensured by folding-assisting proteins such as chaperones or foldases (Zimmermann et al. 2011; Delic et al. 2013), and only correctly folded proteins may leave the ER and proceed through the secretory pathway. The formation of disulfide bonds (Damasceno et al. 2012), protein folding (Helenius et al. 1992), and/or the transport of folded proteins out of the ER (Love et al. 2012) are considered to be the rate-limiting steps of the secretory pathway in *P. pastoris*, as previously shown for recombinant human serum albumin (Shen et al. 2012; Puxbaum et al. 2016) and *Rhizopus chinensis* lipase (Sha et al. 2013b), or suggested for bovine lactoferrin (Sun et al. 2019), penicillin-G-acylase (Borčinová et al. 2020), or

peptidoglycan recognition protein (Yang et al. 2016). Inappropriate cultivation conditions or high levels of production of recombinant proteins (Gasser et al. 2007a), especially those that are surface-displayed, as well as membrane or complex secreted proteins, may overwhelm the folding capacity of the ER, where-upon misfolded/unfolded proteins begin to accumulate in the lumen of the ER. These proteins cause stress to the cell and trigger the UPR (Graf et al. 2008), a signaling cascade aimed at reducing the level of incorrectly folded proteins in the ER, and thus eliminating the stress. The UPR results in upregulation of the expression of genes encoding chaperones and foldases, proteins ensuring correct post-translational modifications, and genes encoding proteins involved in protein translocation and ER quality control (Gasser et al. 2007a; Vogl et al. 2014). At the same time, the expression of many genes involved in protein synthesis is downregulated (Vogl et al. 2014). If the proteins fail to fold correctly, they are translocated back to the cytosol, ubiquitinated, and degraded by the ER-associated degradation (ERAD) pathway (Xie and Ng 2010). Upregulation of ERAD may also be a way to decrease the protein load on the ER if the secretory capacity of the cell is exceeded (Zahrl et al. 2018).

The regulatory mechanisms of UPR were first studied and extensively described in *S. cerevisiae* (Cox and Walter 1996). The key components of the UPR pathway are the following: the kinase/RNase Ire1p, the transcription factor Hac1p, and the chaperone Kar2p, which is a yeast homologue of the mammalian BiP (Casagrande et al. 2000). The Kar2p chaperone resides in the lumen of the ER and, under non-stress conditions, associates with the luminal domain of the monomeric Ire1p. As soon as unfolded proteins occur in the lumen of the ER, Kar2p dissociates from the luminal domain of Ire1p to assist with proper folding (Sidrauski and Walter 1997; Okamura et al. 2000). When Kar2p unbinds, Ire1p assembles into dimers, which results in its phosphorylation and activation of the RNase function of the cytosolic domain of Ire1p (Papa et al. 2003; Kimata et al. 2007). In *S. cerevisiae*, it was demonstrated that besides Kar2p dissociation, there is an additional mechanism of Ire1p activation, based on a direct interaction of unfolded proteins with clustered Ire1p (Kimata et al. 2004, 2007). The RNase domain of Ire1p then non-conventionally (i.e., spliceosome-independent) splices the *HAC1* pre-mRNA (*HAC1^u* mRNA) into its mature form (*HAC1ⁱ* mRNA) (Cox and Walter 1996). The *HAC1* pre-mRNA is targeted to Ire1p via a stem-loop structure within the 3' UTR of the pre-mRNA (Aragon et al. 2009; Kohno 2010). After the excision of the intron from the *HAC1* mRNA, the two exons are joined by tRNA ligase, encoded by the *RLG1* gene (Sidrauski et al. 1996). The mature *HAC1ⁱ* mRNA is translated to the protein Hac1p, which is translocated to the nucleus where it acts as a transcription activator recognizing the so-called UPRE (unfolded protein response element) sequence, and initiates the transcription of

UPR-associated genes in the nucleus (Travers et al. 2000). Besides genes of ER chaperones and proteins involved in folding in *P. pastoris*, Hac1p also induces genes encoding cytosolic chaperones, and genes involved in translation, ribosome biogenesis, organelle biosynthesis, intracellular membrane expansion, protein glycosylation, and translocation (Graf et al. 2008; Guerfal et al. 2010). The UPR was also shown to play an important role in regulating lipid metabolism in *P. pastoris* (Zhang et al. 2016; Adelantado et al. 2017), and to affect the cytosolic redox balance, because redox processes in the ER are counterbalanced by redox processes in the cytosol (Delic et al. 2012). Imbalanced redox processes enhance the likely development of folding-related diseases (e.g., Alzheimer's or Parkinson's).

UPR regulation by Ire1 and Hac1 is highly phylogenetically conserved in eukaryotes and is the main pathway that responds to ER-stress (Bernales et al. 2006; Ron and Walter 2007). Nevertheless, there are variations in the molecular mechanism, and the physiological and stress-responsive roles of the UPR between different yeast species (Hernández-Elvira et al. 2018). The differences in UPR between *P. pastoris* and *S. cerevisiae* include the sequence of UPRE (Mori et al. 1996; Guerfal et al. 2010), the regulation of *HAC1* mRNA splicing (Guerfal et al. 2010; Baumann et al. 2011; Fauzee et al. 2020; Raschmanová et al., in preparation), the length of the *HAC1* intron (Mori et al. 2000; Guerfal et al. 2010), and the role of UPR genes in inositol biosynthesis (Chang et al. 2004; Raschmanová et al., in preparation). Recently, it was reported that the basal level of ER stress (i.e., without external stressing stimuli) in *P. pastoris* is higher than in *S. cerevisiae*, likely due to the enhanced passage of endogenous N-glycosylated proteins through the ER and the secretory pathway (Fauzee et al. 2020). It becomes evident that information about the UPR cannot be solely adopted from *S. cerevisiae*, and more basic research in this area is needed for *P. pastoris* in order to engineer the UPR pathway effectively in terms of increasing productivity. Moreover, general knowledge on chaperones involved in membrane protein folding is limited in yeasts (Emmerstorfer et al. 2014).

In *P. pastoris*, intracellular retention or aggregation of recombinant proteins, or even their intracellular degradation, was observed, accompanied by the upregulation of UPR and/or ERAD (Table 1). Intracellular degradation of the protein may account for up to 60% of the total product (Pfeffer et al. 2011). Recombinant proteins triggering the UPR are of different types, including various secreted proteins: antibody fragments (Gasser et al. 2006, 2007a; Khatri et al. 2011; Pfeffer et al. 2011; Delic et al. 2012; Pfeffer et al. 2012), human interleukin (Zhong et al. 2014), human serum albumin (Aw et al. 2017), different secreted enzymes (Resina et al. 2007; Tawde and Freimuth 2012; Lin et al. 2013; Sha et al. 2013a; Raschmanová et al. 2019), membrane proteins (Vogl et al. 2014), and enhanced green fluorescent protein (EGFP)

(Liu et al. 2014). For example, human serum albumin (HSA) is considered to be a well-secreted protein by *P. pastoris*, as grams per liter of secreted HSA can be obtained (Kobayashi et al. 2000), while heterodimeric antibody fragments are typically produced in only milligrams per liter (Gasser et al. 2006). Yet, both were shown to upregulate the UPR (Table 1). To assign secretion capability (good vs. poor) of the recombinant protein, a combination of several characteristics should be considered (Raschmanová et al. 2019): titer (e.g., grams of secreted protein per liter), specific productivity (e.g., grams of secreted protein per liter and per gram of biomass), intracellular protein accumulation/degradation, and physiological state of the cells (e.g., compromised growth, proportion of non-viable cells). However, all these characteristics are rarely assessed and described in the available literature. Typically therefore, the distinction between a well and poorly secreted protein is based solely on the titer achieved, i.e., extracellular protein concentration.

It seems that the secretion is not predominantly determined by the origin of the protein, in the sense of being naturally secreted or cytosolic; typically, intracellular proteins can also be successfully secreted by *P. pastoris*, e.g., human catalase (0.55 g per liter) (Shi et al. 2007). Rather, the ease of secretion and UPR upregulation seem to be the result of combined effects of the strength of expression (i.e., promoter), gene copy number, protein thermostability, and cultivation conditions used. In all the cases listed in Table 1, the heterologous genes were expressed from strong promoters, either the constitutive glyceraldehyde phosphate dehydrogenase (*GAP*) promoter, or the methanol inducible alcohol oxidase 1 (*AOX1*) promoter. Generally, the higher the copy number of the heterologous gene, the more pronounced was the UPR (Table 1). In the study performed by Love et al. (2012), increasing the copy number of genes expressed from *P_{AOX1}* led to decreased rates of secretion for three proteins with different folding complexities: EGFP, and glycosylated and aglycosylated versions of a human Fc fragment. Nevertheless, there are also proteins whose secretion increases with high-copy number (Huang et al. 2017). The relationship between protein thermostability, secretion, and UPR/ERAD was studied (Whyteside et al. 2011). These authors showed that the production of mutationally destabilized variants of human lysozyme led to higher UPR and ERAD levels, and the protein was retained intracellularly, i.e., poorly secreted and targeted for degradation, more so than the stable variant of lysozyme (Whyteside et al. 2011). Cultivation conditions such as specific growth rate of biomass, temperature, or osmolarity regulate the UPR. An increased specific growth rate of biomass upregulated the UPR, while proteolytic degradation of secretory proteins (ERAD) was downregulated (Rebnegger et al. 2014). Reduction of the cultivation temperature from 30 to 20°C upregulated UPR (Zhong et al. 2014), which probably led to a more rapid processing of the recombinant product in the ER,

Table 1 ER-stress during production of recombinant proteins in *P. pastoris*

Recombinant protein	Promoter	Intracellular accumulation/degradation	ER-stress	References
Secreted proteins				
Antibody Fab fragment	P_{GAP}	Accumulation	Expression of <i>KAR2</i> , <i>PDII</i> , <i>ROT2</i> , <i>ERO1</i> , calnexin, <i>SEC31</i> , <i>SEC53</i> ↑	(Gasser et al. 2006; Gasser et al. 2007a)
Antibody Fab3H6 fragment	P_{GAP}	Degradation	Expression of <i>KAR2</i> ↑ 3.5-fold, <i>LHS1</i> ↑ 1.6-fold, proteolytic activity ↑ by more than 20%	(Pfeffer et al. 2011; Pfeffer et al. 2012)
Anti-HIV antibody 2F5 Fab fragment	P_{GAP}	Not analyzed	Expression of <i>KAR2</i> , <i>ERO1</i> , <i>PDII</i> , <i>HAC1</i> ↑ 2–3-fold	(Delic et al. 2012)
Single-chain antibody fragment (scFv)	P_{AOXI}	Not analyzed	Expression of <i>KAR2</i> ↑ app. 1.6-fold, <i>PDII</i> unchanged	(Khatri et al. 2011)
Hepatitis B virus surface antigen (8 copies)	P_{AOXI}	Potential degradation (ERAD)	Content of Pdi, ERAD proteins ↑	(Vanz et al. 2012)
Anti-CD3 immunotoxin	P_{AOXI} or P_{GAP}	No, but slow secretion	Content of Kar2p ↑ 1.5–3-fold	(Liu et al. 2005)
Synovial sarcoma X break point 2	P_{AOXI}	Accumulation	Expression of <i>KAR2</i> ↑ 2.3–3.5-fold	(Huang et al. 2010)
Human interleukin-10 (different copy number: 1, 5, 10; different temperature: 20°C or 30°C)	P_{AOXI}	Accumulation (both immature and mature protein)	20°C: Expression of <i>HAC1</i> ↑ app. 1.5-fold, <i>KAR2</i> ↑ app. 1.9-fold, <i>ERO1</i> ↑ app. 2.4-fold (compared to 30°C) 5-copy strain: Expression of <i>HAC1</i> ↑ app. 2.5-fold, <i>KAR2</i> ↑ app. 2.5-fold, <i>ERO1</i> ↑ app. 3.8-fold Higher level of ER-phagy at 30°C than at 20°C	(Zhang et al. 2014)
Human serum albumin, nine different clones (all single copy)	P_{AOXI}	Not analyzed	Expression of <i>HAC1</i> , <i>KAR2</i> , and <i>PDII</i> ↑, ↓ or unchanged, depending on the clone	(Aw et al. 2017)
Rabies virus glycoprotein (2–8 copies)	P_{AOXI}	Accumulation	Expression of <i>HAC1</i> ↑ up to 3-fold (8-copy strain), <i>PDII</i> ↑ up to 2.8-fold (7-copy strain), <i>KAR2</i> ↑ up to 2.8-fold (7-copy strain), ERAD genes <i>HRDI</i> and <i>CDC48</i> unchanged	(Ben Azoun et al. 2016a)
Rabies virus glycoprotein (1–8 copies)	P_{GAP}	Accumulation	Expression of <i>HAC1</i> ↑ up to 5.1-fold, <i>PDII</i> ↑ up to 4.8-fold, <i>KAR2</i> ↑ up to 5.1-fold, <i>HRDI</i> ↑ up to 2.3-fold, <i>CDC48</i> ↑ up to 1.5-fold (highest for 8-copy strain)	(Ben Azoun et al. 2016b)
Rabies virus glycoprotein (1, 2, 3, 5, or 10 copies)	P_{AOXI}	Degradation	Expression of <i>HAC1</i> ↑ up to 4.3, <i>IRE1</i> ↑ up to 3.6-fold, <i>PDII</i> ↑ up to 3.1-fold, <i>KAR2</i> ↑ up to 3.6-fold, <i>HRDI</i> ↑ up to 7.5-fold, <i>CDC48</i> ↑ up to 6.1-fold (highest for 10-copy strain)	(Ben Azoun et al. 2017)
Porcine insulin precursor (PIP)	P_{AOXI}	Not analyzed	6-copy strain: Expression of <i>KAR2</i> ↑ 1.68-fold, <i>PDII</i> ↑ 1.43-fold, 18-copy strain: Expression of <i>KAR2</i> ↑ 5.78-fold, <i>PDII</i> ↑ 2.14-fold	(Zhu et al. 2011)
Insulin precursor	P_{AOXI}	Accumulation less than 10%	Amount of UPR- and ERAD-proteins (<i>Kar2p</i> , <i>Pdi</i>) ↓	(Vanz et al. 2014)
Human lysozyme (nine mutational variants with different stability)	P_{AOXI}	Accumulation (the lower stability, the higher amount)	Expression of <i>HAC1</i> ↑ up to 6-fold, <i>KAR2</i> ↑ up to 7.5-fold, <i>PDII</i> ↑ up to 5-fold, <i>DER1</i> ↑ up to 1.8-fold, <i>HRD3</i> ↑ up to 1.6-fold, <i>SEC61</i> ↑ up to 1.2-fold The lower stability, the higher increase	(Whyteside et al. 2011)
Human lysozyme (variant prone to intracellular aggregation 156T and misfolded but secretable variant T70N)	P_{AOXI}	Aggregation (20–30% in case of T70N, up to 60% in case of 156T)	Expression of <i>KAR2</i> and <i>PDII</i> ↑ 2-fold transiently	(Hesketh et al. 2013)
Human trypsinogen	P_{AOXI} or P_{GAP}	Accumulation	Content of Kar2p ↑ up to 4-fold	

Table 1 (continued)

Recombinant protein	Promoter	Intracellular accumulation/degradation	ER-stress	References
Human trypsinogen	P_{AOXI}	Not analyzed	Expression of <i>PDI1</i> , <i>HAC1</i> , <i>ERO1</i> , etc. ↑	
Human trypsinogen (1, 2, or 3 copies)	P_{AOXI}	Not analyzed	Expression of <i>HAC1</i> , <i>KAR2</i> , <i>PDI</i> , ERAD genes ↑ 3-copy strain: Expression of <i>HAC1</i> ↑ 3–4-fold higher than in 1-copy strain	(Delic et al. 2012)
Porcine trypsinogen	P_{GAP}	Not analyzed	>1-copy strain: Expression of <i>KAR2</i> , <i>ERO1</i> , <i>PDI1</i> , <i>HAC1</i> ↑ 2–3-fold	(Wang et al. 2017)
Prolyl endopeptidase	P_{AOXI}	Degradation	Expression of <i>HAC1</i> ↑ up to app. 4.7-fold, <i>KAR2</i> ↑ up to app. 5-fold, and <i>PDI1</i> ↑ up to 5.7-fold	(Liu et al. 2014)
Enhanced green fluorescent protein (1–6 copies)	P_{AOXI}	Accumulation (in strains with 4 and 5 copies)	Expression of <i>KAR2</i> ↑ app. 1.5–5.5-fold, <i>PDI</i> ↑ app. 1.5–4-fold. The highest increase in 4-copy strain	(Resima et al. 2007)
<i>Rhizopus oryzae</i> lipase	P_{FLDI}	Not analyzed	Shake flasks: Expression of <i>KAR2</i> ↑ app. 4-fold, <i>PDI</i> ↑ app. 5-fold	
<i>Rhizopus chinensis</i> lipase (1, 3, 5, or 6 copies)	P_{AOXI}	No	Bioreactors: Expression of <i>KAR2</i> ↑ from 0 to 2.5 fmol mg ⁻¹ total RNA, <i>PDI</i> ↑ from 0 to 2 fmol mg ⁻¹ total RNA	(Sha et al. 2013a)
<i>Arabidopsis</i> modular cellulases AtGH9C1 (C1) and AtGH9C2 (C2) and their truncated versions	P_{AOXI}	Accumulation (truncated versions)	Expression of <i>ERO1</i> ↑ up to 1.7-fold, <i>PDI1</i> ↑ up to 3.7-fold (highest for 6-copy strain)	(Tawde and Freimuth 2012)
Xylanase A from <i>Bacillus halodurans</i> (1 or 4 copies)	P_{AOXI}	Not analyzed specifically (only total intracellular protein), probably no degradation	1-copy strain: Expression of <i>HAC1</i> ↑ app. 1.5-fold, <i>ERO1</i> unchanged, <i>KAR2</i> ↓ app. 0.5-fold, <i>CNE1</i> ↓ app. 0.4-fold, levels of proteins involved in folding and stress response ↓	(Lin et al. 2013)
<i>Escherichia coli</i> penicillin G acylase (<i>EcPGA</i>), <i>Candida antarctica</i> lipase B (<i>CaLB</i>), <i>Thermomyces lanuginosus</i> xylanase A (<i>TIXymA</i>)	P_{AOXI}	Accumulation in case of <i>EcPGA</i> (50–70%)	4-copy strain: Expression of <i>HAC1</i> ↑ 2.2-fold, <i>KAR2</i> ↑ 1.9-fold, <i>ERO1</i> ↑ 1.5-fold, <i>CNE1</i> ↑ 1.9-fold, chaperone content ↑	(Raschmanová et al. 2019)
Membrane proteins			Activity of P_{KAR2} ↑ (60% cells producing <i>EcPGA</i> , 35% cells producing <i>CaLB</i> , and 30% cells producing <i>TIXymA</i>), expression of <i>KAR2</i> ↑ up to 5.6-fold in case of <i>EcPGA</i> production	
Alternative oxidase from <i>P. pastoris</i> (<i>PpAodp</i>), human CMP-Sia transporter (<i>HsCstp</i>), copper transporter Ctr3 from <i>S. cerevisiae</i> (<i>ScCtr3p</i>), all linked to GFP	P_{AOXI}	---	Expression of genes involved in stress response (<i>PpAodp</i> , <i>HsCstp</i> , <i>ScCtr3p</i>) and protein folding (<i>PpAodp</i>) ↑	(Vogl et al. 2014)

decreased levels of immature forms of the protein, and increased product yield (Zhong et al. 2014).

Enhancing protein secretion by overexpression of UPR genes

A possible strategy to enhance production and secretion of a recombinant protein is to co-express a chaperone gene or other genes involved in the UPR, assuming that the co-expressed partner will assist and ensure correct protein folding. Nevertheless, upregulation of the UPR is beneficial only in the cases where protein folding, rather than its passage through the secretory pathway, becomes rate-limiting (Love et al. 2012). In *P. pastoris*, increased expression or secretion of many different recombinant proteins resulted from co-expression of the following: the ER-chaperone Kar2p or protein disulfide isomerase Pdi1 (Inan et al. 2006; Damasceno et al. 2007; Sallada et al. 2019), enzymes involved in the ER redox control and oxidative stress such as Ero1, Gpx1, Aha1, or Ypt6 (Sha et al. 2013c; Ben Azoun et al. 2016a; Huangfu et al. 2016; Sallada et al. 2019), the UPR transcription factor Hac1p (Guerfal et al. 2010; Vogl et al. 2014; Li et al. 2015; Krainer et al. 2016; Huang et al. 2017; Han et al. 2020; Liu et al. 2020), the kinase/RNase Ire1p (Yu et al. 2020), or new co-chaperones (Huangfu et al. 2016) (Table 2). Glycosylation activity was also increased (Moon et al. 2015) or product homogeneity and processing of the secretion α -factor were improved (Guerfal et al. 2010). Recently, three novel folding factors, Mpd1p (member of the PDI family), Pdi2p (protein disulfide isomerase), and Sil1p (nucleotide exchange factor for the ER luminal Hsp70 chaperone Kar2p), were characterized and their genes co-expressed in *P. pastoris* (Duan et al. 2019). In this work, only Sil1p improved the specific extracellular activity and the secretion ratio of one out of three recombinant proteins tested (Duan et al. 2019).

In the vast majority of published works, the helper gene, as well as the target gene of interest, was expressed from the classic strong *Pichia* promoters, *GAP* or *AOX1* (Table 2). When co-expressing 1, 2, 4, 6, 8, or 11 copies of *HAC1* from the *AOX1* promoter and additional 4, 6, 9, 10, 13, 17 copies of *HAC1* from the *GAP* promoter along with the raw-starch hydrolyzing enzyme, α -amylase, the best improvement of product concentration was reached with 6 copies of *HAC1* expressed from the *AOX1* promoter and 17 copies of *HAC1* expressed from the *GAP* promoter (Huang et al. 2017). In another work, the effect of *HAC1* overexpression on heterologous protein levels was stronger when *HAC1* was expressed from the inducible *AOX1* promoter than from the constitutive *GAP* promoter (Guerfal et al. 2010). As shown recently, it might also be beneficial to examine alternative promoters. The yield of bovine lactoferrin was improved by 109.5% by *HAC1ⁱ* expressed from a novel methanol-inducible promoter

P₀₅₄₇, while it decreased when using the *GAP* promoter (Sun et al. 2019). Recently, the UPR-inducible *PDII* promoter, whose strength was found to be equivalent to 20–25% of the *GAP* promoter and 4.5–5% of the *AOX1* promoter, was used for moderate expression of the *Candida antarctica* lipase B gene (Prattipati et al. 2020).

Improved expression/secretion was also affected by the copy number of the recombinant gene (Lin et al. 2013; Yang et al. 2016; Sallada et al. 2019; Huang et al. 2020), as well as of the co-expressed helper gene (Yang et al. 2016; Huang et al. 2017). For example, while the amount of secreted xylanase A from *Bacillus halodurans* increased 1.4-fold in a 4-copy strain by the co-expression of *HAC1*, it was not changed in a co-expressing strain containing only one copy of the xylanase A gene (Lin et al. 2013). A similar trend was observed for the production of secreted *Rhizomucor miehei* lipase; overexpression of *PDII* led to enhanced activity (2-fold) in a 4-copy strain, whereas activity in the strain carrying two copies of the lipase gene remained unchanged (Huang et al. 2020). In a *P. pastoris* strain producing hydrophobin HFBI, co-expression of *KAR2* increased expression of hydrophobin 14-fold in a 1-copy strain, 9.8-fold in a 2-copy strain, and 22-fold in a 3-copy strain (Sallada et al. 2019). Co-expression of other helper genes, *PDII* and *ERO1*, only increased the expression of hydrophobin in the 3-copy strain (7.8-fold and 30-fold, respectively) (Sallada et al. 2019). Another example was the co-expression of *PDII* in *P. pastoris* strains containing low-, medium-, and high-copy numbers of the porcine peptidoglycan recognition protein gene (Yang et al. 2016). Improvements in the amount of secreted product were more significant the higher the copy number, i.e., unchanged, 2.8-fold higher, and 5-fold higher in the low-, medium-, and high-copy strains, respectively (Yang et al. 2016). These results indicate that co-expression of helper UPR genes is particularly helpful, or more pronounced, in strains containing higher copy numbers of the heterologous gene. In the end, this can lead to higher secretion by strains containing a high-copy number of the heterologous gene than by the low-copy strains, which originally, i.e., without the co-expressed chaperone, secreted more product (Yang et al. 2016). The co-expression of multiple copies of the chaperone genes improved the secretion of porcine peptidoglycan recognition protein (high-copy strain) (Yang et al. 2016) or α -amylase from *Geobacillus* sp. (Huang et al. 2017).

In addition, the origin (i.e., the homologue used) of the co-expressed helper gene is important for the extent of its effect on recombinant protein secretion (Bankefa et al. 2018). While the specific activity of β -galactosidase produced with *P. pastoris* was the most improved by co-expression of *HAC1* from *Trichoderma reesei* (by 81%), in the case of β -mannanase, the best co-expression partner was the *HAC1* homologue from *Homo sapiens* (improvement of 49%), and for glucose oxidase, the co-expression of *P. pastoris HAC1* worked the best

Table 2 Examples of co-expression strategies involving UPR genes to improve production/secretion of recombinant proteins in *P. pastoris*

Recombinant protein (secreted, if not stated otherwise)	Co-expressed helper gene	Promoter for recombinant/helper gene expression	Production/secretion-related effect(s)	References
Human parathyroid hormone	<i>Sc*</i> <i>PDII</i>	<i>P_{AOXI}/P_{AOXI}</i>	Secretion ↑ app. 3-fold	(Vad et al. 2005)
Antibody Fab fragment	<i>ScHAC1ⁱ</i>	<i>P_{AOXI}</i> or <i>P_{GAP}/P_{GAP}</i>	Secretion ↑ 1.3-fold	(Gasser et al. 2006)
Antibody Fab fragment	<i>ScPDI</i>	<i>P_{AOXI}</i> or <i>P_{GAP}/P_{GAP}</i>	Secretion ↑ 1.9-fold	(Gasser et al. 2006)
Antibody Fab fragment	<i>ScPDII</i> , <i>ScERO1</i> , <i>ScKAR2</i> , or <i>ScHAC1</i>	<i>P_{GAP}/P_{GAP}</i>	Productivity ↑ 1.7-fold (<i>PDII</i>), ↑ 1.4-fold (<i>ERO1</i>), 1.5-fold (<i>KAR2</i>), 1.5-fold (<i>HAC1</i>)	(Gasser et al. 2007b)
A33 single-chain antibody fragment	<i>Pp*</i> <i>KAR2</i> and/or <i>PpPDI</i>	<i>P_{AOXI}/P_{AOXI}</i>	Secretion ↑ 3-fold (<i>KAR2</i>), no effect (<i>PDI</i>), no effect (<i>KAR2</i> and <i>PDI</i>)	(Damasceno et al. 2007)
Interleukin-2-human serum albumin fusion protein	<i>PDII</i> , <i>KAR2</i> , <i>ERO1</i>	<i>P_{AOXI}/P_{GAP}</i>	Secretion level ↑ 2.2-fold (<i>PDII</i>), 1.9-fold (<i>KAR2</i>), 2.3-fold (<i>ERO1</i>)	(Guan et al. 2016)
Interleukin-1 receptor antagonist-human serum albumin fusion protein (high-copy number)	<i>PpPDI</i> , <i>PpKAR2</i>	<i>P_{AOXI}/P_{AOXI}</i>	Yield ↑ 2.4–3.7-fold (different copy numbers of <i>PDII</i>), ↓ (different copy numbers of <i>KAR2</i>)	(Shen et al. 2012)
Human granulocyte-colony stimulating factor	<i>ScKAR2</i> and/or <i>ScPDI</i>	<i>P_{AOXI}/P_{GAP}</i>	Activity ↑ 5.6-fold (<i>ScKAR2</i>), 4-fold (<i>ScPDI</i>), 6.5-fold (<i>ScKAR2</i> and <i>ScPDI</i>)	(Zhang et al. 2006)
Mouse interferon-γ, human interferon-β, human thrombomodulin, human erythropoietin (all surface-displayed proteins)	<i>PpHAC1ⁱ</i>	<i>P_{AOXI}/P_{AOXI}</i> or <i>P_{GAP}</i>	<i>P_{AOXI}</i> -expressed <i>HAC1ⁱ</i> : Expression of human thrombomodulin ↑ 1.9-fold, mL-10 ↑ 2.2-fold, <i>Trypanosoma cruzi</i> trans-sialidase ↑ 2.1-fold, other proteins ↓, homogeneity and processing of the α-mating factor of adenosine A2A receptor improved	(Guerfal et al. 2010)
mL- ³ -10, <i>Trypanosoma cruzi</i> trans-sialidase protein			<i>P_{GAP}</i> -expressed <i>HAC1ⁱ</i> : Little or no improvement of production	
Adenosine A2A receptor (membrane protein)			Expression of <i>Sc</i> -Ctr3p unchanged, <i>HsCstp</i> ↑ 2.1-fold, <i>HsCtri1p</i> ↑ 1.7-fold, <i>OsCstp</i> ↑ 1.5-fold	(Vogl et al. 2014)
Human CMP-Sia transporter (<i>HsCstp</i>), copper transporter Ctr3 from <i>S. cerevisiae</i> (<i>ScCtri3p</i>), rice (<i>Oryza sativa</i>) CMP-Sia transporter (<i>OsCstp</i>), human copper transporter Ctr1 (<i>HsCtri1p</i>), all linked to GFP (membrane proteins)	<i>PpHAC1ⁱ</i>	<i>P_{AOXI}/P_{AOXI}</i>	Expression of <i>Sc</i> -Ctr3p unchanged, <i>HsCstp</i> ↑ 2.1-fold, <i>HsCtri1p</i> ↑ 1.7-fold, <i>OsCstp</i> ↑ 1.5-fold	
<i>Necator americanus</i> secretory protein (different copy numbers)	<i>PpPDII</i>	<i>P_{AOXI}/P_{AOXI}</i>	Secretion ↑ app. 4–8-fold	(Inan et al. 2006)
Rabies virus glycoprotein	<i>PDII</i> , <i>ERO1</i> , <i>GPXI</i> , <i>GLRI</i> , or <i>YAPI</i>	<i>P_{AOXI}/P_{GAP}</i>	Level ↑ up to 9.6-fold (<i>PDII</i>), ↑ app. 3-fold (<i>ERO1</i>), ↑ 8.2-fold (<i>GPXI</i>), unchanged (<i>GLRI</i> , <i>YAPI</i>)	(Ben Azoun et al. 2016a)
Rabies virus glycoprotein	<i>PDII</i> , <i>ERO1</i> , <i>GPXI</i> , <i>GLRI</i> , or <i>YAPI</i>	<i>P_{GAP}/P_{GAP}</i>	Expression ↑ up to 15-fold (<i>PDII</i>), ↑ 4-fold (<i>ERO1</i>), ↑ 9-fold (<i>GPXI</i>), ↑ 1.7-fold (<i>GLRI</i>), unchanged (<i>YAPI</i>)	(Ben Azoun et al. 2016b)
Porcine peptidoglycan recognition protein (low-, medium-, high-copy)	<i>PDII</i> and/or <i>KAR2</i>	<i>P_{AOXI}/P_{GAP}</i>	Amount in medium-copy strain ↑ up to app. 2.8-fold (<i>PDII</i>), high-copy strain ↑ up to app. 5-fold (<i>PDII</i>), unchanged in low-copy strain (<i>PDII</i>). Amount unchanged or ↓ (<i>KAR2</i>)	(Yang et al. 2016)
Hydrophobin HFBI (1–3-copies)	<i>PpKAR2</i> , <i>PpPDII</i> , <i>PpERO1</i>	<i>P_{AOXI}/P_{AOXI}</i>	Expression in 1-copy strain ↑ 14-fold (<i>KAR2</i>), insignificant change (<i>PDII</i> , <i>ERO1</i>) Expression in 2-copy strain ↑ 9.8-fold (<i>KAR2</i>), insignificant change (<i>PDII</i> , <i>ERO1</i>)	(Sallada et al. 2019)

Table 2 (continued)

Recombinant protein (secreted, if not stated otherwise)	Co-expressed helper gene	Promoter for recombinant/helper gene expression	Production/secretion-related effect(s)	References
Bovine lactoferrin (2 copies)	<i>PpHACIⁱ</i>	<i>P_{AOXI}/P_{GAP}</i> ⁹⁴ <i>P_{AOXI}/P₀₅₄₇</i> <i>P_{AOXI}/P_{AOXI}</i>	Expression in 3-copy strain ↑ 22-fold (<i>KAR2</i>), ↑ 7.8-fold (<i>PDI</i>), 30-fold (<i>EROI</i>) Yield ↓ by 20.9% Yield ↑ by 109.5% Lysozyme activity ↑ by 21.3%	(Sun et al. 2019) (Liu et al. 2020) (He et al. 2020)
Human lysozyme (4 copies)	<i>PpHACIⁱ</i>	<i>P_{AOXI}/P_{GAP}</i>	Activity ↑ (<i>PpEROI</i> , <i>PpPDI</i> , <i>PpEROI</i> + <i>PpPDI</i>), ↓ (<i>PpKAR2</i>)	(Delic et al. 2012)
Human lysozyme (6 copies)	<i>PpKAR2</i> , <i>PpEROI</i> , <i>PpPDI</i>	<i>P_{AOXI}/P_{GAP}</i>	Titer ↑ 2-fold (<i>PpPDI</i>), unchanged (<i>PpEROI</i>)	(Resina et al. 2007)
Porcine trypsinogen	<i>PpPDI</i> or <i>PpEROI</i>	<i>P_{GAP}/P_{GAP}</i>	Extracellular activity ↑ 1.5-fold	(Resina et al. 2009)
<i>Rhizopus oryzae</i> lipase	<i>ScHACI</i>	<i>P_{FLD}/P_{GAP}</i>	Specific productivity ↑ 3-fold	(Sha et al. 2013c)
<i>Rhizopus oryzae</i> lipase	<i>ScHACI</i>	<i>P_{FLD}/P_{GAP}</i>	Enzyme yield ↑ by 30%	(Samuel et al. 2013)
<i>Rhizopus chinensis</i> lipase	<i>PpEROI</i> and <i>PpPDI</i> (simultaneously)	<i>P_{AOXI}/P_{AOXI}</i>	Activity ↓ 0.7-fold	(Huang et al. 2020)
<i>Candida antarctica</i> lipase B	<i>PpKAR2</i>	<i>P_{AOXI}/P_{AOXI}</i>	Activity in 2-copy strain unchanged (<i>PDI</i>), ↓ by app. 33% (<i>EROI</i>)	(Huang et al. 2020)
<i>Rhizomucor miehei</i> lipase (2 or 4 copies)	<i>PpPDI</i> , <i>PpEROI</i>	<i>P_{AOXI}/P_{AOXI}</i>	Activity in 4-copy strain ↑ 2-fold (<i>PDI</i>), ↓ by app. 12% (<i>EROI</i>)	(Lan et al. 2016)
Lipase MAS1 from marine <i>Sireptomyces</i> sp.	<i>PpPDI</i> , <i>PpHACI</i> , <i>PpKAR2</i>	<i>P_{AOXI}/P_{AOXI}</i>	Activity ↑ 1.7-fold (<i>PDI</i>), ↑ slightly (<i>HACI</i> , <i>KAR2</i>)	(Liu et al. 2013)
α-glucosidase from <i>Aspergillus niger</i>	<i>PpPDI</i>	<i>P_{AOXI}/P_{AOXI}</i>	Concentration unchanged or ↓	(Lin et al. 2013)
Xylanase A from <i>Bacillus halodurans</i>	<i>HACI</i>	<i>P_{AOXI}/P_{AOXI}</i>	Amount ↑ 1.4-fold (4-copy strain), unchanged (1-copy strain)	(Li et al. 2015)
Phytase from <i>Citrobacter amalonaticus</i>	<i>HACIⁱ</i>	<i>P_{AOXI}</i> (engineered)/ <i>P_{AOXI}</i>	Concentration ↑ 1.4-fold	(Krainer et al. 2016)
Horseradish peroxidase	<i>HACIⁱ</i>	<i>P_{AOXI}/P_{AOXI}</i> or <i>P_{HXXI}</i> (bidirectional)	Specific activity ↑	(Huangfu et al. 2016)
β-glucuronidase	<i>AHAI</i> , <i>SBAI</i> , <i>SISI</i> , <i>YPT6</i>	<i>P_{AOXI}/P_{GAP}</i>	Specific activity ↑ 1.9-fold (<i>AHAI</i>), ↑ 1.6-fold (<i>SBAI</i>), ↑ 1.4-fold (<i>SISI</i>), ↑ 1.8-fold (<i>YPT6</i>), ↑ 2.3 (<i>AHAI</i> with <i>YPT6</i>), altered glycosylation	(Huangfu et al. 2016)
Endo-β-1,4-xylanase	<i>AHAI</i> , <i>YPT6</i>	<i>P_{AOXI}/P_{GAP}</i>	Specific activity ↑ 2-fold (<i>AHAI</i>), ↑ app. 2.5-fold (<i>YPT6</i>)	(Huangfu et al. 2016)
Phospholipase C-Y from <i>Bacillus cereus</i>	<i>HACI</i> , <i>PDI</i> , <i>KAR2</i>	<i>P_{AOXI}/P_{AOXI}</i> (attenuated)	Titer ↑ 6.2-fold (<i>HACI</i>), unchanged (<i>PDI</i> , <i>KAR2</i>)	(Elena et al. 2016)
Raw-starch hydrolyzing α-amylase	<i>HACIⁱ</i>	<i>P_{AOXI}/P_{AOXI}</i> or <i>P_{GAP}</i>	Concentration ↑ up to 7.2-fold (6 copies of <i>HACIⁱ</i> from <i>P_{AOXI}</i>), ↑ 12.1-fold (+ other 17 copies of <i>HACIⁱ</i> from <i>P_{GAP}</i>)	(Huangfu et al. 2017)
<i>Starmarella bombicola</i> lactone esterase	<i>HACIⁱ</i>	<i>P_{AOXI}/P_{AOXI}</i>	Concentration ↑ 1.8-fold	(De Waele et al. 2018)
β-galactosidase from <i>A. oryzae</i> (Lac), β-mannanase from <i>Bacillus</i> (Man), glucose oxidase from <i>A. niger</i> (Gox) (all codon-optimized for <i>P. pastoris</i>)	<i>HACI</i> homologues: <i>PpHACI</i> , <i>ScHACI</i> , <i>Tr⁸⁶HACI</i> or <i>Hs⁸⁶XBP1</i>	<i>P_{AOXI}/P_{AOXI}</i>	Specific activity of Lac ↑ by 75% (<i>PpHACI</i>), ↑ by 57% (<i>ScHACI</i>), ↑ by 81% (<i>Tr⁸⁶HACI</i>), ↓ by 62% (<i>Hs⁸⁶XBP1</i>) Specific activity of Man ↑ by 8% (<i>Tr⁸⁶HACI</i>), ↑ by 49% (<i>Hs⁸⁶XBP1</i>), ↓ by 3% (<i>PpHACI</i>), ↓ by 41% (<i>ScHACI</i>)	(Bankefa et al. 2018)

Table 2 (continued)

Recombinant protein (secreted, if not stated otherwise)	Co-expressed helper gene	Promoter for recombinant/helper gene expression	Production/secretion-related effect(s)	References
Yeast-enhanced green fluorescent protein (yEGFP), β -galactosidase (Gal), cephalosporin C acylase (SECA)	<i>PDI1</i> , <i>KAR2</i> , <i>HAC1</i> , <i>MPD1</i> , <i>PDI2</i> , <i>SIL1</i>	P_{AOXI}/P_{AOXI}	Specific activity of Cox \uparrow by 13% (<i>PpHAC1</i>), \uparrow by 10% (<i>ScHAC1</i>), \uparrow by 5% (<i>HsXBPI</i>), \downarrow by 3% (<i>TrHAC1</i>) Specific extracellular fluorescence of yEGFP \uparrow by 26% (<i>PDI1</i>), \uparrow by 14% (<i>KAR2</i>), \uparrow by 99% (<i>HAC1</i>), \downarrow (others), secretion ratio* ⁷ unchanged (all) Extracellular production of Gal \uparrow slightly (<i>PDI2</i> , <i>KAR2</i>), \downarrow dramatically (others) Extracellular production of SECA \uparrow 3-fold (<i>SIL1</i> , <i>HAC1</i>), \downarrow or unchanged (others), secretion ratio \uparrow 2.7-fold (<i>SIL1</i>) and 3.2-fold (<i>HAC1</i>)	(Duan et al. 2019)
<i>Pseudomonas aeruginosa</i> elastase	<i>HAC1</i>	P_{AOXI}/P_{AOXI}	Activity \uparrow 1.8–3.9-fold. Negligible effect on N-glycosylation	(Han et al. 2020)
<i>Zobellia k-carrageenase</i>	<i>KAR2</i> , <i>ERO1</i> , <i>PDI</i> , <i>YAPI</i> , <i>AHA1</i> , <i>YPT6</i> , <i>PRX1</i> , <i>RPN4</i> , <i>IRE1</i>	P_{AOXI}/P_{AOXI}	Enzymatic activity unchanged (<i>KAR2</i> , <i>PDI</i>), \uparrow 1.24–1.35-fold (all others)	(Yu et al. 2020)

*¹ *Saccharomyces cerevisiae*, *² *Pichia pastoris*, *³ mouse interleukin, *⁴ novel methanol-inducible promoter (Xu et al. 2018), *⁵ *Trichoderma reesei*, *⁶ *Homo sapiens*, *⁷ secreted to total protein amount

(improvement of 13%). These results indicate that the native Hac1p or its homologue from a closely related species does not necessarily have to be the best option generally for enhancing the secretion of any protein (Bankefa et al. 2018).

An alternative strategy, based on regulating/engineering the UPR, which may improve protein secretion in *P. pastoris*, is inhibition of the proteasome, including ERAD (Pfeffer et al. 2012). However, recent research revealed that the disruption of proteasomal and ERAD components did not increase the secretion of an antibody fragment produced by *P. pastoris* and the authors proposed that the protein was probably degraded prior to entering the secretory pathway (Zahrl et al. 2018). Another approach enhancing recombinant protein production might be deletion of certain chaperones; in *S. cerevisiae*, deletion of *CNE1*, encoding the yeast homologue of mammalian calnexin and calreticulin, increased the production of human transferrin receptor (Prinz et al. 2003). In another review, the strategy of improving the production of recombinant G-protein coupled receptors (GPCR) in yeasts by addition of GPCR-specific ligands or chemical chaperones, such as DMSO, histidine, or glycerol, was discussed (Emmerstorfer et al. 2014). These chemical chaperones are involved in, e.g., gene regulation, modulating ER/Golgi transport, cell wall integrity, membrane permeability, stabilizing protein conformation, or supposedly acting as antioxidants (Emmerstorfer et al. 2014). Other engineering approaches to improve secretion by *P. pastoris* are reviewed elsewhere (Ahmad et al. 2014; Puxbaum et al. 2015; Fischer and Glieder 2019).

Pitfalls of engineering the UPR

It is apparent from the published studies that the effect of co-expressed factors is product-specific (Table 2); in some cases, the production/secretion of the recombinant proteins was unchanged (Damasceno et al. 2007; Delic et al. 2012; Liu et al. 2013; Vogl et al. 2014; Ben Azoun et al. 2016a; Ben Azoun et al. 2016b; Elena et al. 2016; Duan et al. 2019), and sometimes it was even reduced (Liu et al. 2013; Yang et al. 2016; Bankefa et al. 2018; Duan et al. 2019; Sun et al. 2019). For example, the secretion of A33 single-chain antibody fragment was increased by *KAR2* co-expression but was not changed by the co-expression of *PDI* or simultaneous co-expression of *KAR2* and *PDI* (Damasceno et al. 2007). In contrast, *PDI* co-expression increased secretion levels of an antibody Fab fragment (Gasser et al. 2006), *Necator americanus* secretory protein (different copy numbers) (Inan et al. 2006), or porcine trypsinogen (Delic et al. 2012).

A decrease in protein secretion in *P. pastoris* was reported for different membrane- and surface-displayed proteins after co-expression of *PpHAC1*¹ from P_{AOXI} (Guerfal et al. 2010), α -glucosidase from *Aspergillus niger* after co-expression of

PpPDI1 from *P_{AOXI}* (Liu et al. 2013), bovine lactoferrin after co-expression of *PpHAC1ⁱ* from *P_{GAP}* (by 20.9%) (Sun et al. 2019), porcine peptidoglycan recognition protein after co-expression of *KAR2* from *P_{GAP}* (Yang et al. 2016), or *Candida antarctica* lipase B after co-expression of *KAR2* from *P_{AOXI}* (0.7-fold) (Samuel et al. 2013). These negative effects might be attributed to the use of a strong promoter for the co-expression of the UPR gene, which induces the UPR to an inappropriately high level and results in elevated ERAD, re-translocation of the protein to the cytosol and its subsequent degradation (Guerfal et al. 2010; Liu et al. 2013). The overexpression of *KAR2* increased the intracellular insoluble fraction of a recombinant peptidoglycan recognition protein, and the prolonged retention of the protein in the ER probably led to its degradation via ERAD (Yang et al. 2016). Moreover, excess Kar2p molecules in the ER, caused by *KAR2* overexpression, might — even in the presence of unfolded proteins — lead to sustained association of Kar2p with Ire1p, and thus prevent activation of Ire1p and subsequent upregulation of the UPR (Samuel et al. 2013). The efficiency of UPR regulation is also determined by the source of the overexpressed *HAC1* (Bankefa et al. 2018); the specific activity of β -galactosidase from *A. oryzae* was decreased in the case of co-expression of the *Homo sapiens* homologue of *HAC1* from *P_{AOXI}* (by 62%), and the specific activity of β -mannanase from *Bacillus* was decreased (by 41%) after co-expression of the *S. cerevisiae* homologue of *HAC1* from *P_{AOXI}* (Bankefa et al. 2018). In the case of β -mannanase, it was shown that overexpression of *ScHAC1* had little or even a negative effect on the expression of chaperones, compared to the *HAC1* homologue from *Homo sapiens*, which also increased the specific β -mannanase activity (Bankefa et al. 2018).

It is important to keep in mind that overexpression of the UPR genes affects the UPR balance and other cellular processes. Overexpression of *PDI1* in *P. pastoris* producing an antibody fragment (Fab) enhanced the secretion rate of Fab, but did not reduce the UPR stress (Gasser et al. 2007a). In addition, the constitutive expression of *HAC1* activated ERAD (Guerfal et al. 2010). Prolonged activation of the UPR can result in so-called ER-phagy, when parts of the ER are removed to relieve the ER stress and remove the misfolded proteins (Kruse et al. 2006). In addition, a sustained activation of UPR can impair cellular growth, as reported for different yeasts (Cox et al. 1993; Kawahara et al. 1997; Chawla et al. 2011; Cheon et al. 2011; Miyazaki et al. 2013; Moon et al. 2015). In *P. pastoris*, slower growth was observed in the case of co-expression of *HAC1* in strains producing xylanase A from *Bacillus halodurans* or human lysozyme (Lin et al. 2013; Liu et al. 2020), of *PDI1* in a strain producing α -glucosidase from *Aspergillus niger* (Liu et al. 2013), or of *ERO1* in a strain producing *Rhizomucor miehei* lipase (Huang et al. 2020). Other authors reported a decreased (by 27%) maximum specific growth rate (μ_{max}) of a *P. pastoris*

strain producing β -galactosidase, as a result of the co-expression of *KAR2*, but a comparable final cell density (Duan et al. 2019). Co-expression of *PDI1* increased the final cell concentration by 35% but did not affect the μ_{max} of that strain. The growth rate of a *P. pastoris* strain producing cephalosporin C acylase was not affected by the co-expression of *PDI1*, but was decreased by the co-expression of folding factors *HAC1*, *KAR2*, *MPD1*, *PDI2*, and *SIL1*, with *SIL1* having the most detrimental effect: μ_{max} was decreased by 39% (Duan et al. 2019). Nevertheless, in other works, no negative effect of overexpression of *PDI1* and/or *KAR2* on the growth of cells was observed (Damasceno et al. 2007; Guan et al. 2016), and the co-expression of *HAC1ⁱ* was even reported to enhance cellular growth (Han et al. 2020). These results suggest that the effect of the co-expressed gene on a strain's physiology and growth has to be determined individually for each product. In this context, it is important to note that it is not quite correct to evaluate the effect of the co-expressed helper gene on protein production/secretion only by comparing protein concentrations or activities. Knowing that co-expression might influence the strain's growth characteristics, it is essential to also assess the biomass growth. To evaluate the effect of the co-expression strategy, protein to biomass yields (mass of protein produced per mass of biomass) or specific productivities (mass of protein produced per mass of biomass per hour) should be compared, instead of only protein mass (mass of protein produced) or concentrations (mass of protein per liter). This is, however, usually not taken into account (Table 2).

Outcomes: Recommendations for co-expression strategies

The correct folding and rate of secretion of a recombinant protein are affected by the strength of expression of its gene, gene copy number (Love et al. 2012), thermostability of the protein (Whyteside et al. 2011), and cultivation conditions (Rebnecker et al. 2014; Zhong et al. 2014). It seems that the combination of these effects can outweigh the effect of the protein's origin (cytosolic vs. secreted) and character with respect to its folding and secretion complexity. Low gene copy number (Love et al. 2012; Yang et al. 2016), increased thermostability of the protein (Whyteside et al. 2011), and decreased cultivation temperature (Zhong et al. 2014) can enhance the folding and secretion rate by alleviating the UPR.

Based on the currently available information, it seems that the effect of a co-expressed folding partner on recombinant protein secretion cannot be predicted a priori. The most suitable folding partner must be verified experimentally for each individual product. According to the literature search summarized in Table 2, the most frequently used co-expression partner genes employed to promote recombinant protein secretion

in *P. pastoris* were as follows: *HAC1* encoding a transcription factor of UPR genes, *PDII* encoding a protein disulfide isomerase, and *KAR2* encoding an ER chaperone. In the case of *HAC1*, the use of different promoters (P_{GAP} , P_{AOX1} , P_{0547} , and P_{HTX1}) for its expression, different copy numbers, or different homologues was investigated, which makes *HAC1* the best so far described co-expression partner in *P. pastoris*. As summarized in Table 2, overexpression of yeast *HAC1* (i.e., the homologue from *P. pastoris* or *S. cerevisiae*) enhanced recombinant protein production/secretion in approx. 60% of reported cases (as reported in the literature) and co-expression of *PDII* and *KAR2* improved protein production/secretion in approx. 73% and 53% of the published cases, respectively. However, such a broad brush view should be taken with care: The number of publications describing overexpression of *PDII* and *KAR2* was lower than those reporting *HAC1* co-expression (Table 2). We acknowledge that the number of unpublished results, either negative or positive, is uncertain. Nevertheless, this purely statistical view should be helpful given the wide scientific interest in the UPR-topic.

There were only a few studies where the effects of *HAC1*, *PDII*, and *KAR2* co-expression were compared for the same product. For the antibody Fab fragment and lipase from marine *Streptomyces* sp., *PDII* co-expression resulted in a greater increase in secreted product than *HAC1* and *KAR2* co-expression (Gasser et al. 2007b; Lan et al. 2016), while the secreted amount of yeast-enhanced green fluorescent protein (yEGFP) was increased the most significantly by *HAC1* co-expression (Duan et al. 2019). The extracellular production of cephalosporin C acylase was improved by *HAC1* co-expression, but not by *PDI* or *KAR2* (Duan et al. 2019). Due to the low number of studies comparing the effect of *HAC1*, *PDII*, and *KAR2* co-expression, it is not possible to draw general conclusions about which co-expression partner would be the most suitable for any particular recombinant protein. Additionally, it cannot be concluded whether constitutive or inducible expression of the co-expressed helper gene would be more suitable, as both were shown to result in improved, but also unchanged or reduced secretion of recombinant proteins. There is a lack of literature describing the effect of chaperone gene co-expression on the production of membrane and surface-displayed proteins; only Hac1p was tested as a helper, and this improved the production of only some proteins (Guerfal et al. 2010; Vogl et al. 2014). It might be beneficial to employ promoters alternative to P_{GAP} and P_{AOX1} for expression of the helper gene, including weak to moderate promoters for a better fine-tuning of the UPR. Failed co-expression strategies were, in some cases, attributed to the UPR having been upregulated to inappropriately high levels by the co-expression of the UPR genes from strong *GAP* or *AOX1* promoters, which might have resulted in increased ERAD (Guerfal et al. 2010; Liu et al. 2013). It is also necessary to note that there might be many failed co-expression

experiments in *P. pastoris* that were never published, but which might actually shed more light on the UPR mechanism and expression fine-tuning.

The literature search for co-expression strategies that employ a UPR-involved gene to enhance recombinant protein production in *P. pastoris* led to the following recommendations:

- Consider the copy number of the heterologous gene of interest. Folding stress can be reduced, thus secretion enhanced, by reducing the copy number of the heterologous gene (Love et al. 2012, Yang et al. 2016). However, co-expression of a helper UPR gene might reverse this trend, resulting in more enhanced secretion in strains with a higher copy number of the heterologous gene than in low-copy number strains (Yang et al. 2016).
- Use a combinatorial approach to optimize the co-expression strategy. Try different co-expression helper genes and promoters (also weak ones) for their expression, different copy numbers of the helper gene, different homologues of the helper gene, simultaneous co-expression of multiple helper genes etc. (Fig. 2).
- If an extensive combinatorial approach is not feasible, as a minimum we recommend examining several different co-expression partners; this might improve the chances of an unknown bottleneck in protein processing in the ER being overcome. We suggest the co-expression of the following: (1) $HAC1^i$ as the transcription factor upregulating the entire UPR, thus also increasing the expression of genes of chaperones, foldases, and others; (2) *PDI* and/or *ERO1*, which are involved in the formation of disulfide bonds and oxidative stress in the ER; and (3) the ER luminal chaperone *KAR2* that assists in correct protein folding.
- For “one-shot” scenarios, when testing of several different co-expression partners is not feasible, we suggest using $HAC1^i$ as a co-expression partner, since it was shown to improve the secretion of different types of proteins including antibody fragments, transporter proteins, lysozyme, a broad range of hydrolytic enzymes, and enhanced the production of a surface-displayed protein (Table 2). However, if the protein of interest is rich in disulfide bridges (Sha et al. 2013c; Guan et al. 2016), the co-expression of *PDII* or *ERO1* might be preferred (Gasser et al. 2007b; Guan et al. 2016).
- Along with protein titer/productivity, we recommend assessing the effect of the co-expressed helper gene on the strain’s physiology and growth of the production strain (Raschmanová et al. 2019), as these, and thus overall productivity and robustness of a bioprocess may be impaired. A negative effect on biomass growth was reported for all of the three most frequently used co-expression partners, Hac1p, Pdi1, and Kar2p. When evaluating protein production/secretion, it is reasonable to calculate the specific

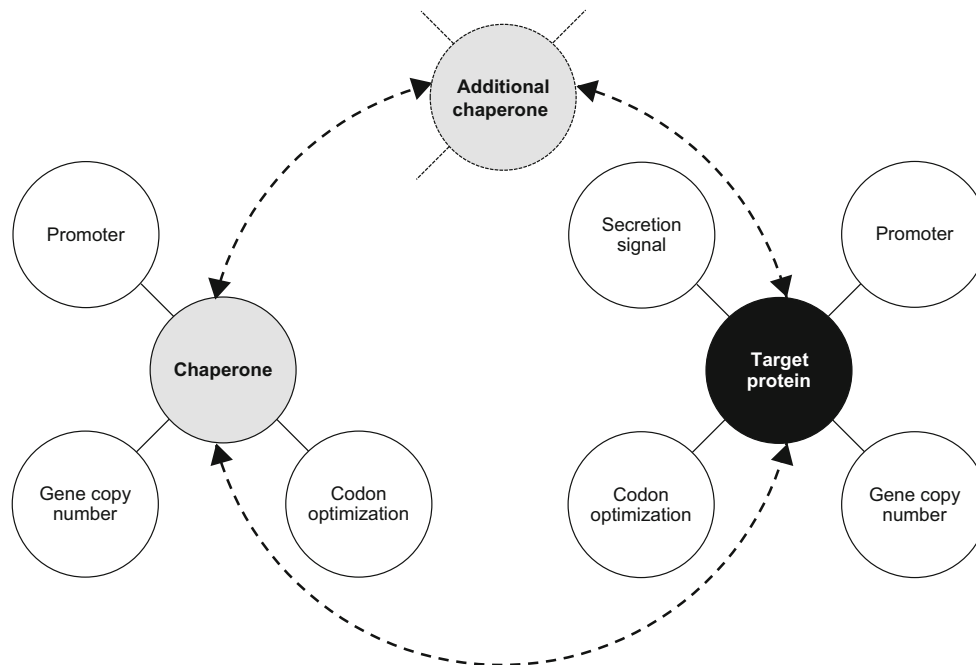


Fig. 2 Combinatorial map of a co-expression strategy for genes of the target protein together with chaperones in *P. pastoris*. Co-expression of a chaperone gene is a possible method to enhance production/secretion of a target recombinant protein, in addition to the choice of appropriate promoter and secretion signal sequences, codon optimization, and optimized copy number of the gene. Various chaperone genes with

codon-optimized sequences and optimized gene copy numbers should be considered and tested with promoters of different strengths (strong, moderate, weak). The promoter used to control expression of the target gene might be different from those used with chaperones. The recombinant protein also needs a secretion signal, but not the chaperone which acts within the cell

productivity (mass of product produced per mass of biomass per hour), which reflects the effect of the co-expressed gene on product secretion, as well as on biomass growth. For difficult-to-secrete proteins, it is useful to assess the proportion of secreted as well as intracellularly retained protein (Duan et al. 2019; Borčinová et al. 2020), and calculate a secretion ratio, i.e., the ratio of secreted to total protein, as a relevant characteristic differentiating between the effect of the co-expressed gene on total production, versus its secretion (Duan et al. 2019).

Conclusions and outlook

A commonly used strategy to boost folding and protein processing in the ER, and thus to overcome secretory bottlenecks in *P. pastoris*, is the overexpression of genes encoding proteins involved in the UPR, such as the transcription activator of UPR genes, Hac1p, or chaperones and foldases, e.g., Kar2p, Pdi1, or Ero1p. In this review, we comprehensively analyzed the successes and failures of such co-expression strategies in *P. pastoris*. Currently, as basic research about the UPR in *P. pastoris* is limited and no general instructions that guarantee enhanced protein secretion can be followed, it is necessary to design and optimize a co-expression strategy for each individual product, since different proteins may benefit from different

levels of UPR activity. Nevertheless, we have summarized recommendations on the best practices for co-expression strategies. In terms of future prospects for recombinant protein production and secretion, the application of novel folding-factors and promoters weaker than the classic P_{GAP} and P_{AOXI} for their co-expression could promote folding and secretion of diverse recombinant proteins that require fine-tuning of the UPR.

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Upon invitation from the AMAB journal (by Prof. Alexander Steinbüchel), all authors jointly shaped the texts to create a conceptual framework for a better understanding of the rationale behind strain construction.

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Declarations

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References

- Adelantado N, Tarazona P, Grillitsch K, García-Ortega X, Monforte S, Valero F, Feussner I, Daum G, Ferrer P (2017) The effect of hypoxia on the lipidome of recombinant *Pichia pastoris*. Microb Cell Factories 16:86. <https://doi.org/10.1186/s12934-017-0699-4>
- Ahmad M, Hirz M, Pichler H, Schwab H (2014) Protein expression in *Pichia pastoris*: recent achievements and perspectives for heterologous protein production. Appl Microbiol Biotechnol 98:5301–5317. <https://doi.org/10.1007/s00253-014-5732-5>
- Antonny B, Schekman R (2001) ER export: public transportation by the COPII coach. Curr Opin Cell Biol 13:438–443. [https://doi.org/10.1016/s0955-0674\(00\)00234-9](https://doi.org/10.1016/s0955-0674(00)00234-9)
- Aragon T, van Anken E, Pincus D, Serafimova IM, Korennykh AV, Rubio CA, Walter P (2009) Messenger RNA targeting to endoplasmic reticulum stress signalling sites. Nature 457:736–749. <https://doi.org/10.1038/nature07641>
- Aw R, Barton GR, Leak DJ (2017) Insights into the prevalence and underlying causes of clonal variation through transcriptomic analysis in *Pichia pastoris*. Appl Microbiol Biotechnol 101:5045–5058. <https://doi.org/10.1007/s00253-017-8317-2>
- Bankefa OE, Wang M, Zhu T, Li Y (2018) Hac1p homologues from higher eukaryotes can improve the secretion of heterologous proteins in the yeast *Pichia pastoris*. Biotechnol Lett 40:1149–1156. <https://doi.org/10.1007/s10529-018-2571-y>
- Barrero JJ, Casler JC, Valero F, Ferrer P, Glick BS (2018) An improved secretion signal enhances the secretion of model proteins from *Pichia pastoris*. Microb Cell Factories 17:161. <https://doi.org/10.1186/s12934-018-1009-5>
- Baumann K, Dato L, Graf AB, Frascotti G, Dragosits M, Porro D, Mattanovich D, Ferrer P, Branduardi P (2011) The impact of oxygen on the transcriptome of recombinant *S. cerevisiae* and *P. pastoris* - a comparative analysis. BMC Genomics 12:218. <https://doi.org/10.1186/1471-2164-12-218>
- Ben Azoun S, Belhaj AE, Göngrich R, Gasser B, Kallel H (2016a) Molecular optimization of rabies virus glycoprotein expression in *Pichia pastoris*. Microb Biotechnol 9:355–368. <https://doi.org/10.1111/1751-7915.12350>
- Ben Azoun S, Belhaj AE, Kallel H (2016b) Rabies virus glycoprotein enhanced expression in *Pichia pastoris* using the constitutive GAP promoter. Biochem Eng J 113:77–85. <https://doi.org/10.1016/j.bej.2016.05.013>
- Ben Azoun S, Ben Zakour M, Sghaier S, Kallel H (2017) Expression of rabies virus glycoprotein in the methylotrophic yeast *Pichia pastoris*. Biotechnol Appl Biochem 64:50–61. <https://doi.org/10.1002/bab.1471>
- Bernales S, Papa FR, Walter P (2006) Intracellular signaling by the unfolded protein response. Annu Rev Cell Dev Biol 22:487–508. <https://doi.org/10.1146/annurev.cellbio.21.122303.120200>
- Borčinová M, Raschmanová H, Zamora I, Looser V, Marešová H, Hirsch S, Kyslík P, Kovar K (2020) Production and secretion dynamics of prokaryotic Penicillin G acylase in *Pichia pastoris*. Appl Microbiol Biotechnol 104:5787–5800. <https://doi.org/10.1007/s00253-020-10669-x>
- Casagrande R, Stern P, Diehn M, Shamu C, Osario M, Zú M, Brown PO, Ploegh H (2000) Degradation of proteins from the ER of *S. cerevisiae* requires an intact unfolded protein response pathway. Mol Cell 5(4):729–735. [https://doi.org/10.1016/s1097-2765\(00\)80251-8](https://doi.org/10.1016/s1097-2765(00)80251-8)
- Cereghino JL, Cregg JM (2000) Heterologous protein expression in the methylotrophic yeast *Pichia pastoris*. FEMS Microbiol Rev 24:45–66. <https://doi.org/10.1111/j.1574-6976.2000.tb00532.x>
- Chang HJ, Jesch SA, Gaspar ML, Henry SA (2004) Role of the unfolded protein response pathway in secretory stress and regulation of INO1 expression in *Saccharomyces cerevisiae*. Genetics 168:1899–1913. <https://doi.org/10.1534/genetics.104.032961>
- Chawla A, Chakrabarti S, Ghosh G, Niwa M (2011) Attenuation of yeast UPR is essential for survival and is mediated by IRE1 kinase. J Cell Biol 193:41–50. <https://doi.org/10.1083/jcb.201008071>
- Cheon SA, Jung KW, Chen YL, Heitman J, Bahn YS, Kang HA (2011) Unique evolution of the UPR pathway with a novel bZIP transcription factor, HxL1, for controlling pathogenicity of *Cryptococcus neoformans*. PLoS Pathog 7:e1002177. <https://doi.org/10.1371/journal.ppat.1002177>
- Cox JS, Walter P (1996) A novel mechanism for regulating activity of a transcription factor that controls the unfolded protein response. Cell 87:391–404. [https://doi.org/10.1016/s0092-8674\(00\)81360-4](https://doi.org/10.1016/s0092-8674(00)81360-4)
- Cox JS, Shamu CE, Walter P (1993) Transcriptional induction of genes encoding endoplasmic reticulum resident proteins requires a transmembrane protein kinase. Cell 73:1197–1206. [https://doi.org/10.1016/0092-8674\(93\)90648-A](https://doi.org/10.1016/0092-8674(93)90648-A)
- Daly R, Hearn MT (2005) Expression of heterologous proteins in *Pichia pastoris*: a useful experimental tool in protein engineering and production. J Mol Recognit 18:119–138. <https://doi.org/10.1002/jmr.687>
- Damasceno LM, Anderson KA, Ritter G, Cregg JM, Old LJ, Batt CA (2007) Cooverexpression of chaperones for enhanced secretion of a single-chain antibody fragment in *Pichia pastoris*. Appl Microbiol Biotechnol 74:381–389. <https://doi.org/10.1007/s00253-006-0652-7>
- Damasceno LM, Huang CJ, Batt CA (2012) Protein secretion in *Pichia pastoris* and advances in protein production. Appl Microbiol Biotechnol 93:31–39. <https://doi.org/10.1007/s00253-011-3654-z>
- De Schutter K, Lin YC, Tiels P, Van Hecke A, Glinka S, Weber-Lehmann J, Rouze P, Van de Peer Y, Callewaert N (2009) Genome sequence of the recombinant protein production host *Pichia pastoris*. Nat Biotechnol 27:561–566. <https://doi.org/10.1038/nbt.1544>
- De Waele S, Vandenberghe I, Laukens B, Planckaert S, Verweire S, Van Bogaert INA, Soetaert W, Devreese B, Ciesielska K (2018) Optimized expression of the *Starmerella bombicola* lactone esterase in *Pichia pastoris* through temperature adaptation, codon-

- optimization and co-expression with *HAC1*. Protein Expr Purif 143: 62–70. <https://doi.org/10.1016/j.pep.2017.10.016>
- Delic M, Rebnegger C, Wanka F, Puxbaum V, Haberhauer-Troyer C, Hann S, Kollensperger G, Mattanovich D, Gasser B (2012) Oxidative protein folding and unfolded protein response elicit differing redox regulation in endoplasmic reticulum and cytosol of yeast. Free Radic Biol Med 52:2000–2012. <https://doi.org/10.1016/j.freeradbiomed.2012.02.048>
- Delic M, Valli M, Graf AB, Pfeiffer M, Mattanovich D, Gasser B (2013) The secretory pathway: exploring yeast diversity. FEMS Microbiol Rev 37:872–914. <https://doi.org/10.1111/1574-6976.12020>
- Duan G, Ding L, Wei D, Zhou H, Chu J, Zhang S, Qian J (2019) Screening endogenous signal peptides and protein folding factors to promote the secretory expression of heterologous proteins in *Pichia pastoris*. J Biotechnol 306:193–202. <https://doi.org/10.1016/j.jbiotec.2019.06.297>
- Elena C, Ravasi P, Cerminati S, Peiru S, Castelli ME, Menzella HG (2016) *Pichia pastoris* engineering for the production of a modified phospholipase C. Process Biochem 51:1935–1944. <https://doi.org/10.1016/j.procbio.2016.08.022>
- Emmerstorfer A, Wriessnegger T, Hirz M, Pichler H (2014) Overexpression of membrane proteins from higher eukaryotes in yeasts. Appl Microbiol Biotechnol 98(18):7671–7698. <https://doi.org/10.1007/s00253-014-5948-4>
- Fauzee YNBM, Taniguchi N, Ishiwata-Kimata Y, Takagi H, Kimata Y (2020) The unfolded protein response in *Pichia pastoris* without external stressing stimuli. FEMS Yeast Res 7:foaa053. <https://doi.org/10.1093/femsyr/foaa053>
- Fischer JE, Glieder A (2019) Current advances in engineering tools for *Pichia pastoris*. Curr Opin Biotechnol 59:175–181. <https://doi.org/10.1016/j.copbio.2019.06.002>
- Gasser B, Maurer M, Gach J, Kunert R, Mattanovich D (2006) Engineering of *Pichia pastoris* for improved production of antibody fragments. Biotechnol Bioeng 94:353–361. <https://doi.org/10.1002/bit.20851>
- Gasser B, Maurer M, Rautio J, Sauer M, Bhattacharyya A, Saloheimo M, Penttila M, Mattanovich D (2007a) Monitoring of transcriptional regulation in *Pichia pastoris* under protein production conditions. BMC Genomics 8:18. <https://doi.org/10.1186/1471-2164-8-179>
- Gasser B, Sauer M, Maurer M, Stadlmayr G, Mattanovich D (2007b) Transcriptomics-based identification of novel factors enhancing heterologous protein secretion in yeasts. Appl Environ Microbiol 73: 6499–6507. <https://doi.org/10.1128/AEM.01196-07>
- Gasser B, Prielhofer R, Marx H, Maurer M, Nocon J, Steiger M, Puxbaum V, Sauer M, Mattanovich D (2013) *Pichia pastoris*: protein production host and model organism for biomedical research. Future Microbiol 8:191–208. <https://doi.org/10.2217/fmb.12.133>
- Gidijala L, Uthoff S, Kampen SJ, Steinbüchel A, Verhaert RMD (2018) Presence of protein production enhancers results in significantly higher methanol-induced protein production in *Pichia pastoris*. Microb Cell Factories 17:112. <https://doi.org/10.1186/s12934-018-0961-4>
- Graf A, Gasser B, Dragosits M, Sauer M, Leparic GG, Tuchler T, Kreil DP, Mattanovich D (2008) Novel insights into the unfolded protein response using *Pichia pastoris* specific DNA microarrays. BMC Genomics 9:390. <https://doi.org/10.1186/1471-2164-9-390>
- Guan B, Chen F, Su S, Duan Z, Chen Y, Li H, Jin J (2016) Effects of co-overexpression of secretion helper factors on the secretion of a HSA fusion protein (IL2-HSA) in *Pichia pastoris*. Yeast 33:587–600. <https://doi.org/10.1002/yea.3183>
- Guerfal M, Ryckaert S, Jacobs PP, Ameloot P, Van Craenenbroeck K, Derycke R, Callewaert N (2010) The *HAC1* gene from *Pichia pastoris*: characterization and effect of its overexpression on the production of secreted, surface displayed and membrane proteins. Microb Cell Factories 9:49. <https://doi.org/10.1186/1475-2859-9-49>
- Han M, Wang W, Zhou J, Gong X, Xu C, Li Y, Li Q (2020) Activation of the unfolded protein response via co-expression of the *HAC1i* gene enhances expression of recombinant elastase in *Pichia pastoris*. Biotechnol Bioprocess Eng 25:302–307. <https://doi.org/10.1007/s12257-019-0381-2>
- He H, Wu S, Mei M, Ning J, Li C, Ma L, Zhang G, Yi L (2020) A combinatorial strategy for effective heterologous production of functional human lysozyme in *Pichia pastoris*. Front Bioeng Biotechnol 8:118. <https://doi.org/10.3389/fbioe.2020.00118>
- Helenius A, Marquardt T, Braakman I (1992) The endoplasmic reticulum as a protein-folding compartment. Trends Cell Biol 2:227–231. [https://doi.org/10.1016/0962-8924\(92\)90309-B](https://doi.org/10.1016/0962-8924(92)90309-B)
- Hernández-Elvira M, Torres-Quiroz F, Escamilla-Ayala A, Domínguez-Martin E, Escalante R, Kawasaki L, Ongay-Larios L, Coria R (2018) The unfolded protein response pathway in the yeast *Kluyveromyces lactis*. A comparative view among yeast species. Cells 7:106. <https://doi.org/10.3390/cells7080106>
- Hesketh AR, Castrillo JL, Sawyer T, Archer DB, Oliver SG (2013) Investigating the physiological response of *Pichia (Komagataella) pastoris* GS115 to the heterologous expression of misfolded proteins using chemostat cultures. Appl Microbiol Biotechnol 97:9747–9762. <https://doi.org/10.1007/s00253-013-5186-1>
- Huang CJ, Anderson KA, Damasceno LM, Ritter G, Old LJ, Batt CA (2010) Improved secretion of the cancer-testis antigen SSX2 in *Pichia pastoris* by deletion of its nuclear localization signal. Appl Microbiol Biotechnol 86:243–253. <https://doi.org/10.1007/s00253-009-2275-2>
- Huang M, Gao Y, Zhou X, Zhang Y, Cai M (2017) Regulating unfolded protein response activator HAC1p for production of thermostable raw-starch hydrolyzing A-amylase in *Pichia pastoris*. Bioprocess Biosyst Eng 40:341–350. <https://doi.org/10.1007/s00449-016-1701-y>
- Huang J, Zhao Q, Chen L, Zhang C, Bu W, Zhang X, Zhang K, Yang Z (2020) Improved production of recombinant *Rhizomucor miehei* lipase by coexpressing protein folding chaperones in *Pichia pastoris*, which triggered ER stress. Bioengineered 11:375–385. <https://doi.org/10.1080/21655979.2020.1738127>
- Huangfu J, Xu Y, Li C, Li J (2016) Overexpressing target helper genes enhances secretion and glycosylation of recombinant proteins in *Pichia pastoris* under simulated microgravity. J Ind Microbiol Biotechnol 43:1429–1439. <https://doi.org/10.1007/s10295-016-1817-8>
- Inan M, Aryasomayajula D, Sinha J, Meagher MM (2006) Enhancement of protein secretion in *Pichia pastoris* by overexpression of protein disulfide isomerase. Biotechnol Bioeng 93:771–778. <https://doi.org/10.1002/bit.20762>
- Kawahara T, Yanagi H, Yura T, Mori K (1997) Endoplasmic reticulum stress-induced mRNA splicing permits synthesis of transcription factor Hac1p/Ern4p that activates the unfolded protein response. Mol Biol Cell 8:1845–1862. <https://doi.org/10.1091/mbc.8.10.1845>
- Khatri NK, Gocke D, Trentmann O, Neubauer P, Hoffmann F (2011) Single-chain antibody fragment production in *Pichia pastoris*: benefits of prolonged pre-induction glycerol feeding. Biotechnol J 6: 452–462. <https://doi.org/10.1002/biot.201000193>
- Kimata Y, Oikawa D, Shimizu Y, Ishiwata-Kimata Y, Kohno K (2004) A role for BiP as an adjuvant for the endoplasmic reticulum stress-sensing protein Ire1. J Cell Biol 167:445–456. <https://doi.org/10.1083/jcb.200405153>
- Kimata Y, Ishiwata-Kimata Y, Ito T, Hirata A, Suzuki T, Oikawa D, Takeuchi M, Kohno K (2007) Two regulatory steps of ER-stress sensor Ire1 involving its cluster formation and interaction with unfolded proteins. J Cell Biol 179:75–86. <https://doi.org/10.1083/jcb.200704166>
- Kobayashi K, Kuwae S, Ohya T, Ohda T, Ohya M, Ohi H, Tomomitsu K, Ohmura T (2000) High-level expression of recombinant human serum albumin from the methylotrophic yeast *Pichia pastoris* with

- minimal protease production and activation. *J Biosci Bioeng* 89:55–61. [https://doi.org/10.1016/S1389-1723\(00\)88050-0](https://doi.org/10.1016/S1389-1723(00)88050-0)
- Kohno K (2010) Stress-sensing mechanisms in the unfolded protein response: similarities and differences between yeast and mammals. *J Biochem* 147:27–33. <https://doi.org/10.1093/jb/mvp196>
- Krainer FW, Gerstmann MA, Darnhofer B, Birner-Gruenberger R, Glieder A (2016) Biotechnological advances towards an enhanced peroxidase production in *Pichia pastoris*. *J Biotechnol* 233:181–189. <https://doi.org/10.1016/j.jbiotec.2016.07.012>
- Kruse KB, Brodsky JL, McCracken AA (2006) Autophagy: an ER protein quality control process. *Autophagy* 2:135–137. <https://doi.org/10.4161/auto.2.2.2388>
- Lan D, Qu M, Yang B, Wang Y (2016) Enhancing production of lipase MAS1 from marine *Streptomyces* sp. strain in *Pichia pastoris* by chaperones co-expression. *Electron J Biotechnol* 22:62–67. <https://doi.org/10.1016/j.ejbt.2016.06.003>
- Li C, Lin Y, Zheng X, Pang N, Liao X, Liu X, Huang Y, Liang S (2015) Combined strategies for improving expression of *Citrobacter amalonaticus* phytase in *Pichia pastoris*. *BMC Biotechnol* 15:88. <https://doi.org/10.1186/s12896-015-0204-2>
- Lin XQ, Liang SL, Han SY, Zheng SP, Ye YR, Lin Y (2013) Quantitative iTRAQ LC-MS/MS proteomics reveals the cellular response to heterologous protein overexpression and the regulation of *HAC1* in *Pichia pastoris*. *J Proteome* 91:58–72. <https://doi.org/10.1016/j.jprot.2013.06.031>
- Liu YY, Woo JH, Neville DM Jr (2005) Overexpression of an anti-CD3 immunotoxin increases expression and secretion of molecular chaperone BiP/Kar2p by *Pichia pastoris*. *Appl Environ Microbiol* 71:5332–5340. <https://doi.org/10.1128/AEM.71.9.5332-5340.2005>
- Liu X, Wu D, Wu J, Chen J (2013) Optimization of the production of *Aspergillus niger* alpha-glucosidase expressed in *Pichia pastoris*. *World J Microbiol Biotechnol* 29:533–540. <https://doi.org/10.1007/s11274-012-1207-y>
- Liu H, Qin Y, Huang Y, Chen Y, Cong P, He Z (2014) Direct evaluation of the effect of gene dosage on secretion of protein from yeast *Pichia pastoris* by expressing EGFP. *J Microbiol Biotechnol* 24:144–151. <https://doi.org/10.4014/jmb.1308.08010>
- Liu WC, Inwood S, Gong T, Sharma A, Yu LY, Zhu P (2019) Fed-batch high-cell-density fermentation strategies for *Pichia pastoris* growth and production. *Crit Rev Biotechnol* 39:258–271. <https://doi.org/10.1080/07388551.2018.1554620>
- Liu J, Han Q, Cheng Q, Chen Y, Wang R, Li X, Liu Y, Yan D (2020) Efficient expression of human lysozyme through the increased gene dosage and co-expression of transcription factor Hac1p in *Pichia pastoris*. *Curr Microbiol* 77:846–854. <https://doi.org/10.1007/s00284-019-01872-9>
- Looser V, Bruhlmann B, Bumbak F, Stenger C, Costa M, Camattari A, Fotiadis D, Kovar K (2015) Cultivation strategies to enhance productivity of *Pichia pastoris*: a review. *Biotechnol Adv* 33:1177–1193. <https://doi.org/10.1016/j.biotechadv.2015.05.008>
- Love KR, Politano TJ, Panagiotou V, Jiang B, Stadheim TA, Love JC (2012) Systematic single-cell analysis of *Pichia pastoris* reveals secretory capacity limits productivity. *PLoS One* 7:e37915. <https://doi.org/10.1371/journal.pone.0037915>
- Marsalek L, Puxbaum V, Buchetics M, Mattanovich D, Gasser B (2019) Disruption of vacuolar protein sorting components of the HOPS complex leads to enhanced secretion of recombinant proteins in *Pichia pastoris*. *Microb Cell Factories* 18:119. <https://doi.org/10.1186/s12934-019-1155-4>
- Marx H, Sauer M, Resina D, Vai M, Porro D, Valero F, Ferrer P, Mattanovich D (2006) Cloning, disruption and protein secretory phenotype of the *GAS1* homologue of *Pichia pastoris*. *FEMS Microbiol Lett* 264:40–47. <https://doi.org/10.1111/j.1574-6968.2006.00427.x>
- Miyazaki T, Nakayama H, Nagayoshi Y, Kakeya H, Kohno S (2013) Dissection of Ire1 functions reveals stress response mechanisms uniquely evolved in *Candida glabrata*. *PLoS Pathog* 9:20. <https://doi.org/10.1371/journal.ppat.1003160>
- Mogelsvang S, Gomez-Ospina N, Soderholm J, Glick BS, Staehelin LA (2003) Tomographic evidence for continuous turnover of Golgi cisternae in *Pichia pastoris*. *Mol Biol Cell* 14:2277–2291. <https://doi.org/10.1091/mbc.E02-10-0697>
- Moon HY, Cheon SA, Kim H, Agaphonov MO, Kwon O, Oh DB, Kim JY, Kang HA (2015) *Hansenula polymorpha* Hac1p is critical to protein N-glycosylation activity modulation, as revealed by functional and transcriptomic analyses. *Appl Environ Microbiol* 81:6982–6993. <https://doi.org/10.1128/aem.01440-15>
- Mori K, Kawahara T, Yoshida H, Yanagi H, Yura T (1996) Signalling from endoplasmic reticulum to nucleus: transcription factor with a basic-leucine zipper motif is required for the unfolded protein-response pathway. *Genes Cells* 1:803–817. <https://doi.org/10.1046/j.1365-2443.1996.d01-274.x>
- Mori K, Ogawa N, Kawahara T, Yanagi H, Yura T (2000) mRNA splicing-mediated C-terminal replacement of transcription factor Hac1p is required for efficient activation of the unfolded protein response. *Proc Natl Acad Sci U S A* 97:4660–4665. <https://doi.org/10.1073/pnas.050010197>
- Naranjo CA, Jivan AD, Vo MN, de Sa Campos KH, Deyarmin JS, Hekman RM, Uribe C, Hang A, Her K, Fong MM, Choi JJ, Chou C, Rabara TR, Myers G, Moua P, Thor D, Risser DD, Vierra CA, Franz AH, Lin-Cereghino J, Lin-Cereghino GP (2019) Characterization of the role of BGS13 in the secretory mechanism of *Pichia pastoris*. *Appl Environ Microbiol* 24:e01615–e01619. <https://doi.org/10.1128/aem.01615-19>
- Obst U, Lu TK, Sieber V (2017) A modular toolkit for generating *Pichia pastoris* secretion libraries. *ACS Synth Biol* 6:1016–1025. <https://doi.org/10.1021/acssynbio.6b00337>
- Okamura K, Kimata Y, Higashio H, Tsuru A, Kohno K (2000) Dissociation of Kar2p/BiP from an ER sensory molecule, Ire1p, triggers the unfolded protein response in yeast. *Biochem Biophys Res Commun* 279:445–450. <https://doi.org/10.1006/bbrc.2000.3987>
- Papa FR, Zhang C, Shokat K, Walter P (2003) Bypassing a kinase activity with an ATP-competitive drug. *Science* 302:1533–1537. <https://doi.org/10.1126/science.1090031>
- Pfeffer M, Maurer M, Kollensperger G, Hann S, Graf AB, Mattanovich D (2011) Modeling and measuring intracellular fluxes of secreted recombinant protein in *Pichia pastoris* with a novel 34S labeling procedure. *Microb Cell Factories* 10:47. <https://doi.org/10.1186/1475-2859-10-47>
- Pfeffer M, Maurer M, Stadlmann J, Grass J, Delic M, Altmann F, Mattanovich D (2012) Intracellular interactome of secreted antibody Fab fragment in *Pichia pastoris* reveals its routes of secretion and degradation. *Appl Microbiol Biotechnol* 93:2503–2512. <https://doi.org/10.1007/s00253-012-3933-3>
- Prattipati M, Ramakrishnan K, Sankaranarayanan M (2020) *Pichia pastoris* protein disulfide isomerase (PDI1) promoter for heterologous protein production and its sequence characterization. *Enzym Microb Technol* 140:109633. <https://doi.org/10.1016/j.enzmictec.2020.109633>
- Prinz B, Stahl U, Lang C (2003) Intracellular transport of a heterologous membrane protein, the human transferrin receptor, in *Saccharomyces cerevisiae*. *Int Microbiol* 6:49–55. <https://doi.org/10.1007/s10123-003-0100-9>
- Puxbaum V, Mattanovich D, Gasser B (2015) Quo vadis? The challenges of recombinant protein folding and secretion in *Pichia pastoris*. *Appl Microbiol Biotechnol* 99:2925–2938. <https://doi.org/10.1007/s00253-015-6470-z>
- Puxbaum V, Gasser B, Mattanovich D (2016) The bud tip is the cellular hot spot of protein secretion in yeasts. *Appl Microbiol Biotechnol* 100:8159–8168. <https://doi.org/10.1007/s00253-016-7674-6>

- Raschmanová H, Weninger A, Glieder A, Kovar K, Vogl T (2018) Implementing CRISPR-Cas technologies in conventional and non-conventional yeasts: current state and future prospects. *Biotechnol Adv* 36:641–665. <https://doi.org/10.1016/j.biotechadv.2018.01.006>
- Raschmanová H, Zamora I, Borčinová M, Meier P, Weninger A, Mächler D, Glieder A, Melzoch K, Knejzlík Z, Kovar K (2019) Single-cell approach to monitor the unfolded protein response during biotechnological processes with *Pichia pastoris*. *Front Microbiol* 10:335. <https://doi.org/10.3389/fmicb.2019.00335>
- Rebnegger C, Graf AB, Valli M, Steiger MG, Gasser B, Maurer M, Mattanovich D (2014) In *Pichia pastoris*, growth rate regulates protein synthesis and secretion, mating and stress response. *Biotechnol J* 9:511–525. <https://doi.org/10.1002/biot.201300334>
- Resina D, Bollok M, Khatri NK, Valero F, Neubauer P, Ferrer P (2007) Transcriptional response of *P. pastoris* in fed-batch cultivations to *Rhizopus oryzae* lipase production reveals UPR induction. *Microb Cell Factories* 6:11. <https://doi.org/10.1186/1475-2859-6-21>
- Resina D, Maurer M, Cos O, Arnau C, Carnicer M, Marx H, Gasser B, Valero F, Mattanovich D, Ferrer P (2009) Engineering of bottlenecks in *Rhizopus oryzae* lipase production in *Pichia pastoris* using the nitrogen source-regulated *FLDI* promoter. *New Biotechnol* 25:396–403. <https://doi.org/10.1016/j.nbt.2009.01.008>
- Ron D, Walter P (2007) Signal integration in the endoplasmic reticulum unfolded protein response. *Nat Rev Mol Cell Biol* 8:519–529. <https://doi.org/10.1038/nrm2199>
- Rossanese OW, Soderholm J, Bevis BJ, Sears IB, O'Connor J, Williamson EK, Glick BS (1999) Golgi structure correlates with transitional endoplasmic reticulum organization in *Pichia pastoris* and *Saccharomyces cerevisiae*. *J Cell Biol* 145:69–81. <https://doi.org/10.1083/jcb.145.1.69>
- Sallada ND, Harkins LE, Berger BW (2019) Effect of gene copy number and chaperone coexpression on recombinant hydrophobin HFBI biosurfactant production in *Pichia pastoris*. *Biotechnol Bioeng* 116:2029–2040. <https://doi.org/10.1002/bit.26982>
- Samuel P, Prasanna Vadhana AK, Kamatchi R, Antony A, Meenakshisundaram S (2013) Effect of molecular chaperones on the expression of *Candida antarctica* lipase B in *Pichia pastoris*. *Microbiol Res* 168:615–620. <https://doi.org/10.1016/j.micres.2013.06.007>
- Sha C, Yu XW, Li F, Xu Y (2013a) Impact of gene dosage on the production of lipase from *Rhizopus chinensis* CCTCC M201021 in *Pichia pastoris*. *Appl Biochem Biotechnol* 169:1160–1172. <https://doi.org/10.1007/s12010-012-0050-9>
- Sha C, Yu XW, Lin NX, Zhang M, Xu Y (2013b) Enhancement of lipase r27RCL production in *Pichia pastoris* by regulating gene dosage and co-expression with chaperone protein disulfide isomerase. *Enzym Microb Technol* 53:438–443. <https://doi.org/10.1016/j.enzmictec.2013.09.009>
- Sha C, Yu XW, Zhang M, Xu Y (2013c) Efficient secretion of lipase r27RCL in *Pichia pastoris* by enhancing the disulfide bond formation pathway in the endoplasmic reticulum. *J Ind Microbiol Biotechnol* 40:1241–1249. <https://doi.org/10.1007/s10295-013-1328-9>
- Shen Q, Wu M, Wang HB, Naranmandura H, Chen SQ (2012) The effect of gene copy number and co-expression of chaperone on production of albumin fusion proteins in *Pichia pastoris*. *Appl Microbiol Biotechnol* 96:763–772. <https://doi.org/10.1007/s00253-012-4337-0>
- Shi X-L, Feng M-Q, Shi J, Shi Z-H, Zhong J, Zhou P (2007) High-level expression and purification of recombinant human catalase in *Pichia pastoris*. *Protein Expr Purif* 54:24–29. <https://doi.org/10.1016/j.pep.2007.02.008>
- Sidrauski C, Walter P (1997) The transmembrane kinase Ire1p is a site-specific endonuclease that initiates mRNA splicing in the unfolded protein response. *Cell* 90:1031–1039. [https://doi.org/10.1016/S0092-8674\(00\)80369-4](https://doi.org/10.1016/S0092-8674(00)80369-4)
- Sidrauski C, Cox JS, Walter P (1996) tRNA ligase is required for regulated mRNA splicing in the unfolded protein response. *Cell* 87:405–413. [https://doi.org/10.1016/S0092-8674\(00\)81361-6](https://doi.org/10.1016/S0092-8674(00)81361-6)
- Spohner SC, Müller H, Quitmann H, Czermak P (2015) Expression of enzymes for the usage in food and feed industry with *Pichia pastoris*. *J Biotechnol* 202:118–134. <https://doi.org/10.1016/j.jbiotec.2015.01.027>
- Sun J, Jiang J, Zhai X, Zhu S, Qu Z, Yuan W, Wang Z, Wei C (2019) Coexpression of Kex2 endopeptidase and Hac1 transcription factor to improve the secretory expression of bovine lactoferrin in *Pichia pastoris*. *Biotechnol Bioprocess Eng* 24:934–941. <https://doi.org/10.1007/s12257-019-0176-5>
- Tawde MD, Freimuth P (2012) Toxic misfolding of *Arabidopsis* cellulases in the secretory pathway of *Pichia pastoris*. *Protein Expr Purif* 85:211–217. <https://doi.org/10.1016/j.pep.2012.08.009>
- Travers KJ, Patil CK, Wodicka L, Lockhart DJ, Weissman JS, Walter P (2000) Functional and genomic analyses reveal an essential coordination between the unfolded protein response and ER-associated degradation. *Cell* 101:249–258. [https://doi.org/10.1016/S0092-8674\(00\)80835-1](https://doi.org/10.1016/S0092-8674(00)80835-1)
- Vad R, Nafstad E, Dahl LA, Gabrielsen OS (2005) Engineering of a *Pichia pastoris* expression system for secretion of high amounts of intact human parathyroid hormone. *J Biotechnol* 116:251–260. <https://doi.org/10.1016/j.jbiotec.2004.12.004>
- Vanz AL, Lunsdorf H, Adnan A, Nimtz M, Gurrakonda C, Khanna N, Rinas U (2012) Physiological response of *Pichia pastoris* GS115 to methanol-induced high level production of the Hepatitis B surface antigen: catabolic adaptation, stress responses, and autophagic processes. *Microb Cell Factories* 11:103. <https://doi.org/10.1186/1475-2859-11-103>
- Vanz AL, Nimtz M, Rinas U (2014) Decrease of UPR- and ERAD-related proteins in *Pichia pastoris* during methanol-induced secretory insulin precursor production in controlled fed-batch cultures. *Microb Cell Factories* 13:23. <https://doi.org/10.1186/1475-2859-13-23>
- Vogl T, Thallinger GG, Zellnig G, Drew D, Cregg JM, Glieder A, Freigassner M (2014) Towards improved membrane protein production in *Pichia pastoris*: general and specific transcriptional response to membrane protein overexpression. *New Biotechnol* 31:538–552. <https://doi.org/10.1016/j.nbt.2014.02.009>
- Wang XD, Jiang T, Yu XW, Xu Y (2017) Effects of UPR and ERAD pathway on the prolyl endopeptidase production in *Pichia pastoris* by controlling of nitrogen source. *J Ind Microbiol Biotechnol* 44:1053–1063. <https://doi.org/10.1007/s10295-017-1938-8>
- Weninger A, Hatzl AM, Schmid C, Vogl T, Glieder A (2016) Combinatorial optimization of CRISPR/Cas9 expression enables precision genome engineering in the methylotrophic yeast *Pichia pastoris*. *J Biotechnol* 235:139–149. <https://doi.org/10.1016/j.jbiotec.2016.03.027>
- Weninger A, Fischer JE, Raschmanová H, Kniely C, Vogl T, Glieder A (2018) Expanding the CRISPR/Cas9 toolkit for *Pichia pastoris* with efficient donor integration and alternative resistance markers. *J Cell Biochem* 119:3183–3198. <https://doi.org/10.1002/jcb.26474>
- Whyteside G, Alcocer MJ, Kumita JR, Dobson CM, Lazarou M, Pleass RJ, Archer DB (2011) Native-state stability determines the extent of degradation relative to secretion of protein variants from *Pichia pastoris*. *PLoS One* 6:e22692. <https://doi.org/10.1371/journal.pone.0022692>
- Xie W, Ng DTW (2010) ERAD substrate recognition in budding yeast. *Semin Cell Dev Biol* 21:533–539. <https://doi.org/10.1016/j.semcdb.2010.02.007>
- Xu N, Zhu J, Zhu Q, Xing Y, Cai M, Jiang T, Zhou M, Zhang Y (2018) Identification and characterization of novel promoters for recombinant protein production in yeast *Pichia pastoris*. *Yeast* 35:379–385. <https://doi.org/10.1002/yea.3301>

- Yang J, Lu Z, Chen J, Chu P, Cheng Q, Liu J, Ming F, Huang C, Xiao A, Cai H, Zhang L (2016) Effect of cooperation of chaperones and gene dosage on the expression of porcine PGLYRP-1 in *Pichia pastoris*. *Appl Microbiol Biotechnol* 100:5453–5465. <https://doi.org/10.1007/s00253-016-7372-4>
- Yu Y, Liu Z, Chen M, Yang M, Li L, Mou H (2020) Enhancing the expression of recombinant κ -carrageenase in *Pichia pastoris* using dual promoters, co-expressing chaperones and transcription factors. *Biocatal Biotransfor* 38(10):1–10. <https://doi.org/10.1080/10242422.2019.1655001>
- Zahl RJ, Peña DA, Mattanovich D, Gasser B (2017) Systems biotechnology for protein production in *Pichia pastoris*. *FEMS Yeast Res* 17:fox068. <https://doi.org/10.1093/femsyr/fox068>
- Zahl RJ, Mattanovich D, Gasser B (2018) The impact of ERAD on recombinant protein secretion in *Pichia pastoris* (*Syn Komagataella* spp.). *Microbiology (Reading)* 164:453–463. <https://doi.org/10.1099/mic.0.000630>
- Zepeda AB, Pessoa A, Farias JG (2018) Carbon metabolism influenced for promoters and temperature used in the heterologous protein production using *Pichia pastoris* yeast. *Braz J Microbiol* 49:119–127. <https://doi.org/10.1016/j.bjm.2018.03.010>
- Zhang W, Zhao HL, Xue C, Xiong XH, Yao XQ, Li XY, Chen HP, Liu ZM (2006) Enhanced secretion of heterologous proteins in *Pichia pastoris* following overexpression of *Saccharomyces cerevisiae* chaperone proteins. *Biotechnol Prog* 22:1090–1095. <https://doi.org/10.1021/bp060019r>
- Zhang M, Yu Q, Liang C, Zhang B, Li M (2016) Lipid homeostasis is involved in plasma membrane and endoplasmic reticulum stress in *Pichia pastoris*. *Biochem Biophys Res Commun* 478:777–783. <https://doi.org/10.1016/j.bbrc.2016.08.024>
- Zhong YJ, Yang L, Guo YG, Fang F, Wang D, Li R, Jiang M, Kang WY, Ma JJ, Sun J, Xiao WH (2014) High-temperature cultivation of recombinant *Pichia pastoris* increases endoplasmic reticulum stress and decreases production of human interleukin-10. *Microb Cell Factories* 13:10. <https://doi.org/10.1186/s12934-014-0163-7>
- Zhu T, Guo M, Zhuang Y, Chu J, Zhang S (2011) Understanding the effect of foreign gene dosage on the physiology of *Pichia pastoris* by transcriptional analysis of key genes. *Appl Microbiol Biotechnol* 89:1127–1135. <https://doi.org/10.1007/s00253-010-2944-1>
- Zimmermann R, Eyrich S, Ahmad M, Helms V (2011) Protein translocation across the ER membrane. *Biochim Biophys Acta Biomembr* 1808:912–924. <https://doi.org/10.1016/j.bbamem.2010.06.015>

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