



Head and neck squamous cell carcinoma: a potential therapeutic target for the Wnt signaling pathway

Khosrow Siamak Houschyar¹ · Mimi R. Borrelli² · Susanne Rein³ · Christian Tapking⁴ · Daniel Popp⁵ · Alen Palackic⁵ · Behrus Puladi⁶ · Mark Ooms⁶ · Madeline Houschyar⁷ · Ludwik K. Branski⁸ · Laurenz Schmitt¹ · Ali Modabber⁶ · Albert Rübber¹ · Frank Hölzle⁶ · Amir S. Yazdi¹

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Abstract

Squamous cell carcinoma (SCC) of the head and neck region accounts for 3% of all tumors worldwide. The incidence is higher in men, with most carcinomas found in the oral cavity. At the point of initial diagnosis, distant metastases are rare. The Wnt signaling pathway is critically involved in cell development and stemness and has been associated with SCC. Understanding precisely how Wnt signaling regulates SCC progression and how it can, therefore, be modulated for the therapeutic benefit has enormous potential in the treatment of head and neck SCC. In this review, we will describe the underlying mechanisms of Wnt signaling and outline how Wnt signaling controls cellular processes both in homeostasis and in the development and progression of SCC.

Level of evidence: Not gradable.

Keywords Stem cells · Carcinoma · Wnt signaling

Introduction

Over 85% of cancers in the head and neck area are squamous cell carcinomas (HNSCCs), and the oral cavity is the most common location. [1] More than 500,000 new HNSCCs are diagnosed every year, placing them amongst the top ten cancers worldwide. [2] Etiologically, HNSCCs result from mutations in the DNA of keratinocytes. [3] Mutated keratinocytes are at risk of becoming pre-malignant or malignant cells with the potential for unregulated growth. [4] Once growth is autologous, HNSCCs develop. Growth of cancerous cells beyond the basement membrane leads to metastases in the lymph nodes, bones, brain, liver, and other organs. [5]

Mutations to keratinocytes can occur spontaneously, but often follow exposure to mutagenic substances. Mutagens may be chemicals (e.g., tobacco and alcohol), physical, or microbiological (e.g., viruses like human papillomavirus (HPV) or Epstein-Barr virus (EBV)). [6, 7] Radiation exposure, immunosuppression, and poor oral hygiene are all risk factors for HNSCC in the mouth and throat. [8] Worldwide, 25% of oral SCC is associated with tobacco consumption, and 7–19% is associated with alcohol consumption. [9] Combined consumption of alcohol and tobacco increases the risk of developing mouth or throat SCC sevenfold compared

✉ Khosrow Siamak Houschyar
Khosrow-Houschyar@gmx.de

¹ Department of Dermatology and Allergology, RWTH University Hospital Aachen, Pauwelsstraße 30, 52074 Aachen, Germany

² Division of Plastic and Reconstructive Surgery, Department of Surgery, Stanford School of Medicine, Stanford, CA 94305, USA

³ Department of Plastic and Hand Surgery-Burn Center, Klinikum St. Georg, Leipzig, Germany

⁴ Department of Hand, Plastic and Reconstructive Surgery, Burn Trauma Center, BG Trauma Center Ludwigshafen, University of Heidelberg, Heidelberg, Germany

⁵ Division of Plastic, Aesthetic and Reconstructive Surgery, Department of Surgery, Medical University of Graz, Graz, Austria

⁶ Department of Oral and Maxillofacial Surgery, University Hospital RWTH, Aachen, Germany

⁷ Institute of Agricultural and Nutrition Sciences, Martin Luther University of Halle, Wittenberg, Germany

⁸ Department of Surgery, Shriners Hospitals for Children-Galveston, University of Texas Medical Branch, 815 Market Street, Galveston, TX 77550, USA

to the sole exposition to one of the noxious agents. [10] This synergistic effect may relate to how alcohol facilitates penetration of carcinogens into the oral mucosa as well as how chronic alcohol consumption extends the carcinogen-mucosal surface contact time due to reduced salivary gland function. [11]

Recent advances in sequencing technology and characterization of cancer genomes highlighted the involvement of the Wnt signaling pathway in human cancers including SCCs. [12] The Wnt signaling pathway is a complex and ancient signal transduction pathway critical in regulating cellular stemness and development throughout embryology and adult life. Consequently, Wnt signaling is ubiquitous to all cells, tissues, and organ systems and has been highly conserved across evolution, dating back to anaerobic metazoans. [13] A considerable amount of cancer research has centered on Wnt signaling; many of the critical protein regulators are defined, and recent genetic and biochemical studies have further identified novel Wnt pathway components and their functions [14, 15] including Wnt secretory machinery, Wnt co-receptors, components of the β -catenin destruction complex, and nuclear co-factors. [16] This review aims to discuss the involvement of Wnt signaling in the maintenance of stemness in cancer stem cells (CSC) with particular reference to HNSCC. We will then discuss

the different therapeutic and diagnostic options for HNSCCs and shed light on the most promising therapeutic agent for the HNSCC treatment to date.

Wnt signaling pathways

Wnt-mediated signal transduction processes regulate basic processes that determine the functionality or dysfunction of numerous cell types. [17] Specifically, Wnt signaling is a critical regulator of embryonic development and the creation of embryonic cell patterns, as well as the differentiation, proliferation, invasion, polarity, and apoptosis of cells in adult organisms. [18, 19] Disorders of the Wnt signaling can result in various diseases, including cancer and osteoporosis. [20, 21]

To date, at least three different intracellular Wnt signaling pathways have been described; (1) the canonical Wnt/ β -catenin signaling pathway, (2) the non-canonical pathways Wnt/PCP (planar cell polarity), and (3) the Wnt/ Ca^{2+} signaling pathway. [22] Common to all three pathways are the Wnt proteins, which function as ligands to specific cell surface receptors (Frizzled receptors or Fzds), which trigger intracellular signaling cascades (Fig. 1). The term “canonical Wnts” came from the observation that strongly transforming Wnts transmit signals via β -catenin. [23] At first, it was assumed

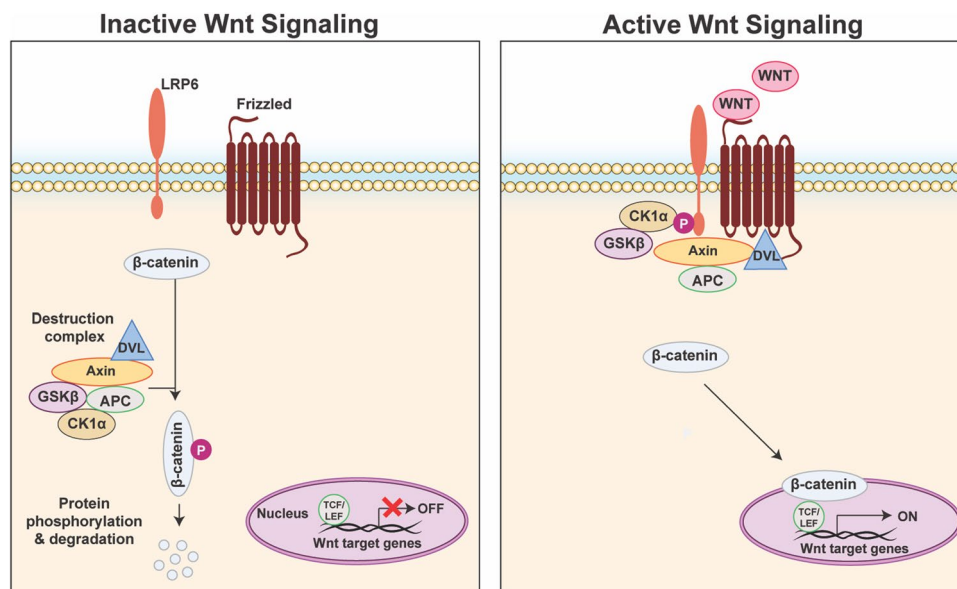


Fig. 1 Canonical Wnt signaling. *Left:* In the inactive state, Wnt ligands are absent, and intracellular β -catenin is phosphorylated by the destruction complex (a complex of Axin, APC, GSK3 β , casein kinase (CK1 α)). Phosphorylated β -catenin is then ubiquitinated and targeted for proteasomal degradation. The absence of intranuclear β -catenin allows the TCF (lymphoid enhancer factor)/LEF (lymphoid enhancer factor) proteins to promote histone deacetylases which leads to repression of target genes. *Right:* In the active state, secreted Wnt ligands bind Frizzled (Fzd) receptors and lipoprotein receptor-related

protein (LRP) co-receptors leading to LRP phosphorylation by CK1 α and GSK3 β , and recruitment of disheveled (Dvl) proteins. The accumulation of Dvl proteins leads to their polymerization and activation which in turn inactivates the destruction complex. The cytoplasmic β -catenin is free to accumulate and translocate to the nucleus where it associates with LEF and TCF and recruits histone-modifying co-activators to promote gene expression and activation of numerous cellular processes

that all other Wnts signaled via β -catenin-independent signaling pathways and hence were termed “non-canonical Wnts.” [24] However, the *canonical* Wnt3a can also signal independent of β -catenin by activating Rho and Rho-kinase. [25] In addition, under certain conditions, the non-canonical Wnt5a activates both a β -catenin-dependent and a β -catenin-independent signaling pathway. [26] Consequently, the classic distinction between canonical and non-canonical Wnts can no longer be strictly maintained.

The canonical Wnt/ β -catenin signaling pathway primarily regulates the developmental trajectory of cells and the invasiveness and proliferation potential of CSCs. Binding of Wnts to Fzd receptors destabilizes a degradation complex that initiates the breakdown of β -catenin typically. Intracellular β -catenin consequently accumulates and translocates to the cell nucleus and acts as a transcription factor for various Wnt/ β -catenin target genes. [16]

The two non-canonical signal pathways cannot be completely separated from one another. Although both pathways have been extensively studied in *Drosophila* and *Xenopus*, little is known about the function of these pathways in mammals [27]. It is known that the Wnt/PCP pathway participates in the control of cell polarity and cell movement during gastrulation. [28] In the Wnt/PCP signaling pathway, Wnt-Fzd binding activates disheveled (Dvl). Activated Dvl, in turn, can transmit the signal in two independent ways. [22] This is how Daam1 enables the complex formation of Dvl and Rho, a G protein, followed by the activation of Rho kinase (RhoK) via Rac, another G protein, Dvl leads in parallel to the activation of JNK (JUN N-terminal kinase). [29]

In the Wnt/ Ca^{2+} signaling pathway, Wnt-Fzd binding leads to an increased intracellular Ca^{2+} concentration and activates heterotrimeric G proteins, which in turn activate phospholipase C and phosphodiesterase. [30] The high intracellular Ca^{2+} and activated G proteins also activate the calcium/calmodulin-dependent protein kinase II (CaMKII) and the protein kinase C. [31] It is suspected that the Wnt/ Ca^{2+} signaling pathway regulates cell proliferation and migration and also functions to inhibit the Wnt/ β -catenin pathway. CamKII and TAK1- activate NLK mitogen-activated protein kinase (NLK-MAPK), which then phosphorylates the T-cell factor (TCF) and thereby prevents the binding of the β -catenin/TCF complex to the DNA and thus prevents transcription. [32]

The molecular genetics of tumor development

The genetic changes predispose to tumor initiation and progression can occur as relatively small sequence changes to the DNA or chromosomal deviations. [33] The sequence changes affect two classes of genes: the proto-oncogenes and the tumor suppressor genes. [34]

Proto-oncogenes code for proteins that stimulate cell proliferation or progression through the cell cycle. They are converted into oncogenes by amplification of the gene locus, chromosomal translocation, or point mutations, i.e., to activated forms (gain of function), which results in uncontrolled growth of the cells. [35] The mutations in the proto-oncogenes are dominant, which means that the mutation in one of the two alleles is sufficient for the activation of the proto-oncogene. [36] Today, more than 100 known proto-oncoproteins have been described, including growth factors (e.g., PDGF, Wnt-1), their receptors (e.g., EGF receptors), proteins of signal transduction (e.g., Ras proteins, Src kinase), and transcription factors (e.g., c-Jun, c-Fos, c-Myc). [37]

Tumor suppressor genes, in contrast, have an inhibitory effect on cell growth, [38] and thus mutations that impair tumor suppressor function predispose to unregulated cell growth (i.e., cancer). Tumor suppressor gene function may be altered following faulty chromosome or chromatid division during cell division, mitotic recombination, or due to point mutations or deletions. [39] However, the functioning of both tumor suppressor gene alleles must be lost before tumorigenesis occurs; [40] this is referred to as a “loss of heterozygosity” (LOH). Some individuals inherited a mutated allele through the germline, and are thus at an increased risk for LOH and developing cancer. For example, individuals with familial adenomatous polyposis (FAP) syndrome are born with a mutated APC gene and show increased risk of colon cancer. [41] An alternative mechanism of tumor suppressor gene inactivation is epigenetic modification. Various studies have shown that the promoters of different tumor suppressor genes are often methylated in tumors but are free of methylation in healthy tissues. [42] The methylation of the DNA affects 5'-CG-3' dinucleotides (CpG), which are located in the promoter regions of the genes, [43] and methylation is thus involved in the regulation of gene expression and chromosome condensation. [44] Today, about 30 tumor suppressor genes with different functions are known, including APC, PTEN, SASH1, and p21CIP1/WAF1. [45] One of the most extensively studied tumor suppressor genes is p53, which is reported to be mutated in 50% of all cancer cells. p53 functions to either block the cell cycle or induce apoptosis, and is a transcription factor with an important role in maintaining genomic integrity, hence its name the “guardian of the genome.” [46]

In addition to the classic oncogenes and tumor suppressor genes, there is a large number of additional “tumor genes” that act as modulators of tumor development or that play an essential role in clinical practice as diagnostic and prognostic tumor markers. [47] The chromosomal aberrations lead to aneuploidies — deviations from the normal diploid chromosome set. The changes can affect both the number of chromosomes and the structure of the chromosomes and lead

to chromosomal instability (CIN). [48] Genetic instability within the cell can also arise at the nucleotide sequence level (MIN, microsatellite instability). In this case, the tumor cell chromosome set is usually diploid. [49] Furthermore, gene function can also be modified at an epigenetic level, which recent research has increasingly highlighted. [50]

The importance of the Wnt signaling during the Embryonic development as well as in regeneration and tumor development

Cancer is not a single disease, but a term for many different types of disease. It is a genetic disease that is based on the uncontrolled growth of specific cells. Cancer is now the second leading cause of death after cardiovascular disease in western countries and the third leading cause of adult deaths in developing countries. [51] Tumor formation is a multi-stage process in which the physiological control of cell proliferation, cell differentiation, and cell–cell interactions are gradually lost. [52] Apart from the inherited forms, most tumors are caused by somatic mutations. [53] The majority of tumors are presumably monoclonal, meaning they arise from a *single cell* through the accumulation of several genetic and epigenetic changes through proliferation and clonal selection. [54] Tumors start as benign growths of cells maintained in differentiated states exhibition organization to their tissue architecture. As tumorigenesis progresses, additional genetic changes accumulate and lead to the formation of a malignant tumor comprised of poorly differentiated cells, which acquired the ability to penetrate the neighboring tissues and metastasize to distant organs. [55] Six changes in the physiology of the cell define the malignant phenotype: (1) independence from growth-promoting signals, (2) insensitivity to growth-inhibiting signals, (3) resistance to apoptosis, (4) unlimited multiplication potential, (5) persistent angiogenesis, and (6) ability to invade tissue and metastasize. [56]

The Wnt/ β -catenin signaling regulates the development and function of many tissues and organs throughout embryonic development and in adult life. [57, 58] The Wnt pathways also interact with many other important signaling pathways, such as the transforming growth factor- β (TGF- β)/Bmp-, Fgf-, Notch-, and hedgehog signaling pathways, which influence each other either in series or in parallel. [19, 59] Even at a very early stage of embryonic development, before gastrulation, Wnt β -catenin signaling is critical for the induction of the mesoderm and the direction the correct formation of the embryonic body axis, [19] as highlighted by experiments where injection of Wnt agonists (Lithium Chloride or Wnt1 mRNA) induced a second body axis in *Xenopus* embryos. [60] The Wnt/ β -catenin signaling pathway is also essential for the development of the primitive endoderm, formation of the head region, and development

of many organs, as highlighted by various mouse models. [61] Activation of the Wnt/ β -catenin signaling pathway is also necessary to maintain the pluripotency of embryonic stem (ES) cells and for the process of reprogramming differentiated fibroblasts into pluripotent stem cells (iPS). [62]

In certain organs, such as the pituitary gland, dorsal spinal cord, and bone, the Wnt/ β -catenin pathway controls the formation, maintenance, and/or specification of progenitor cells. [63] For example, β -catenin/LEF/TCF induces the expression of *Pitx2* in the pituitary gland, which is responsible for the formation of *Pit1*-expressing progenitor cells. [64] During bone development, β -catenin controls the differentiation of osteo-chondrogenic progenitor cells into osteoblasts. [65] The importance of the maintenance of progenitor cells is also observed in the development of the dorsal spinal cord, [63] where Wnt/ β -catenin signaling activation promotes the proliferation of *Olig3*-expressing progenitor cells and to differentiate excess *Foxd3* and *Isl1/2*-expressing interneurons (dI2 and 3 regions). [66] In contrast, the 0 mutation of β -catenin leads to the absence of progenitor cells expressing *Olig3*, and the loss of *Olig3* prevents the formation of dI2- and dI3 neurons. [67] The combination of *Olig3* null mutation and Stabilization of β -catenin causes the loss of the dI2 and dI3 neurons and the increase in *Lbx1*-expressing neurons (dI4-dI6) while maintaining the proliferative effect on the progenitor cells. [68] These results indicate that Wnt/ β -catenin in the dorsal spinal cord is important for the formation and maintenance of *Olig3*-expressing progenitor cells, but not for differentiation into dI2 and dI3 neurons. [68]

In continuously renewing tissues and organs, the Wnt/ β -catenin signaling pathway is responsible for the self-renewal of multipotent tissue stem cells. [69, 70] The intestinal stem cells of the intestinal crypts which continually renew the intestinal epithelium require active Wnt/ β -catenin signaling for their proliferative function. [71] Newly formed intestinal cells lose the ability to divide, and the Wnt/ β -catenin signaling pathway is switched off as they differentiate into terminal cells. [16] However, mutations that result in permanent activation of the Wnt pathway (e.g., involving APC or β -catenin genes) lead to the development of intestinal polyps, characteristic of early stages of colon cancer. [72] On the other hand, in the case of degenerative diseases of the intestinal mucosa, such as Crohn's disease, there is assumed to be insufficient Wnt/ β -catenin activity. [73]

The skin is another organ that continuously renews throughout life. In embryology, Wnt signaling directs the embryonic ectoderm to differentiate into the epithelium by promoting production of keratin and blocking fibroblast growth factor (FGR). [74] Wnt signaling also inducts formation of skin appendages, especially hair follicles, by orchestrating dynamic signaling between the epidermis and dermis. [75] In adult life, Wnt/ β -catenin signaling continues to

control hair follicle cycling by coordinating the differentiation of follicular stem cells into hair cells in the bulges of the hair follicle. [76] Within hair follicle stem cells, β -catenin is found within the nucleus during anagen (growth phase), but in the cell membrane during telogen (rest phase), and is thus activation of the Wnt pathway is thought to induce the onset of anagen from telogen. [77] Wnt signaling is then further thought to control cell fate within hair follicles. [78] In addition, Wnt/ β -catenin is inhibited by the Bmp signaling pathway during the hair cycle — an important criterion for the oscillating renewal and differentiation of the hair follicle stem cells. [79] The signal pathway can induce the development of hair follicle tumors. It could also be shown that cancer stem cells from ras-induced epidermal tumors require active β -catenin for their self-maintenance, which illustrates the great importance of Wnt/ β -catenin for the cancer stem cells of the skin. [80] β -catenin is also involved in epidermal proliferation and maintenance of the epidermal stem cell population, as shown in lineage-tracing work in mice. [81, 82] However, interestingly, loss of β -catenin leads to hair loss but has little impact on the integrity of the epidermis. [83] Wnt/ β -catenin signaling also helps to orchestrate cutaneous wound healing. *In vitro* work has shown that human keloid keratinocytes stimulate fibroblasts to secrete R-Spondin2, a Wnt agonist which promotes epidermal proliferation and is thought to lead to the thickened epidermis characteristic of keloid scars. [84] Further work has also shown that TGF- β , a potent regulator of fibrosis, induces activation of β -catenin within dermal fibroblasts, and β -catenin staining is increased in hypertrophic scars and keloid, compared to normal skin. [85] Regarding skin cancer, and specifically SSCs, chromosomes containing WNT and FZD genes are amplified in SSCs in genomic hybridization studies. [86] Messenger RNA levels of Wnt ligands and receptors are increased, and

levels of Wnt inhibitors are decreased, further implicating role for Wnt activation in SSC development. [87–89]

Together, these findings highlight the critical roles of the canonical Wnt signaling pathway in cell regulation in normal functioning and in the development of cancer through altering the regulation of cell expansion and survival of mature cells and stem cells by their direct and/or indirect target genes (Table 1).

Wnt/ β -catenin signaling as a therapeutic target for HNSCC

Head and neck tumors are treated with surgery, radiotherapy, and chemotherapy. [90] Early-stage cancers are often resected entirely with adjuvant radiotherapy and/or without chemotherapy. Cancer too large to be removed can be debulked and are typically treated with a combination of radiotherapy and chemotherapy. Progress in all three therapeutic areas has been small and the average 5-year survival for patients with head and neck cancer remains at 61%. [91]

Although numerous Wnt receptors and Wnt signaling components are highly expressed in head and neck cancers, [13] there has been hesitation to target β -catenin therapeutically due to its structural role in apical junctions (AJs) and its functional role in CSC self-renewal. [16, 70] However, numerous Wnt inhibitors have been investigated for their antitumor effects in both preclinical and clinical trials. [92] The most promising candidates are inhibitors of porcupine, an acyl-transferase and critical protein in the Wnt/ β -catenin cascade. For example, the porcupine inhibitor LGK974 can disrupt HNSCC growth and reduce distant metastases. [93, 94] Recent insight into the specific function of β -catenin within the cell nucleus, however, has supported its potential as a druggable target; [95] antibodies which target Wnt-1

Table 1 Elevated expression of upstream and downstream of Wnt signaling pathway genes and their associated diseases are reported

Gene transduction level	Gene	Disease (cell type)	Associated biological gene function
Upstream	WNT FZD	Head and neck squamous cell carcinoma (cancer stem cell)	Self-renewal
Downstream	C-MYC AXIN2 LEF1 OCT14 NANOG SOX2 MMP7 TWIST ABCB1	Colon cancer (cancer stem cell) Breast cancer (cancer stem cell) Non-disease (embryonic stem cell) Head and neck squamous cell carcinoma (cancer stem cell) Non-disease (embryonic stem cell) Oral squamous cell carcinoma (cancer cell) Breast cancer (immortalized human breast epithelial cell) Head and neck squamous cell carcinoma (cancer cell) Colon cancer, neuroblastoma (cancer cell)	Metastasis Drug resistance

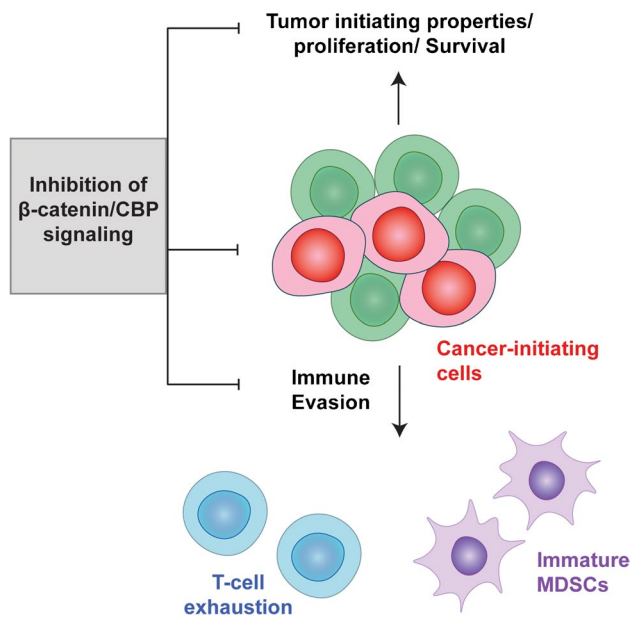


Fig. 2 Therapeutic targeting the β -catenin/cAMP-responsive element-binding protein (CBP) axis for head and neck squamous cell carcinoma (HNSCC). It is believed that inhibition of the association between β -catenin and CBP can deplete tumor cancer stem cells (CSCs), stimulate epithelial cell differentiation, deplete the exhausted T cells, immature heterogeneous cells of the myeloid lineage, and/or myeloid-derived suppressor cells (MDSCs) involved in immune evasion

and inhibit cell proliferation can decrease tumor burden, [12, 96] and β -catenin/CBP axis inhibitors can intercept oncogenic activities in the tumor niche. Axitinib is a small-molecular inhibitor of nuclear β -catenin which works by stabilizing ubiquitin ligase SHPRH (SNF2, histone linker, PHD, and RNIG finger domain-containing helicase), and thus decreasing the availability of nuclear β -catenin. [97] The association of β -catenin with fibrosis and immune cell depletion suggests that targeting the β -catenin-CBP interaction will have intratumoral inhibitory effects (Fig. 2). It is likely that targeting oncogenic pathways in combination with Wnt/ β -catenin signaling may be more effective than monotherapies and standard radiation therapy/chemotherapy for the treatment of HNSCC. For example, combining β -catenin/CBP axis inhibitors with immune checkpoint blockades may effectively deplete aggressive CSCs and promote anticancer immune cell function.[98]

Conclusions and future directions

Preclinical and early-phase clinical trials have highlighted the clinical utility of targeting the canonical Wnt signaling pathway to disrupt the malignant potential of

CSCs. Cancer is a complex disease and Wnt signaling is ubiquitous, thus therapeutic manipulation a challenge. Specifically, since Wnt signaling regulates many essential activities within every cell during homeostasis, it is critical to develop inhibitors specific to cancer cells to avoid unfavorable side effects. The major hurdles facing the development of Wnt pathway antagonists include limited efficacy, non-specific binding, and undetermined therapeutic windows. Many current molecules suffer low bioavailability as single-agent treatments and may benefit from combination treatments which facilitate drug delivery. Ongoing work is focused on exploiting advances in computational and system biology methodologies to map signaling networks and cellular metabolism in different tumor cell populations. Combining this knowledge with emerging epigenome editing technologies can further elucidate the causal relationships between cellular epigenetic landscapes and regulatory processes in order to better understand how β -catenin pathways are deregulated in HNSCC.

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Data availability Please contact author for data requests.

Declarations

Ethics approval and consent to participate Ethical approval was not required for this study.

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References

1. Heroiu AC, Danciu C, Popescu C (2013) Multiple cancers of the head and neck. *Amaltea Medical, Editura Magister*
2. Hashim D, Genden E, Posner M, Hashibe M, Boffetta P (2019) Head and neck cancer prevention: from primary prevention to impact of clinicians on reducing burden. *Ann Oncol* 30(5):744–756
3. Dotto GP, Rustgi AK (2016) Squamous cell cancers: a unified perspective on biology and genetics. *Cancer Cell* 29(5):622–637
4. Jimson S, Murali S, Zunt SL, Goldblatt LI, Srinivasan M (2016) Epithelial expression of keratinocytes growth factor in oral pre-cancer lesions. *Dent Res J* 13(3):199
5. Sanderson RJ, Ironside JA (2002) Squamous cell carcinomas of the head and neck. *BMJ* 325(7368):822–827. <https://doi.org/10.1136/bmj.325.7368.822>
6. Hamasni FM, El Hajj F (2016) Expression of bone morphogenetic protein-2 and histological differentiation of oral squamous cell carcinomas. *Asian Pac J Cancer Prev: APJCP* 17(12):5243
7. Glastonbury CM (2020) Head and neck squamous cell cancer: approach to staging and surveillance. *Diseases of the Brain, Head and Neck, Spine 2020–2023*. Springer. 215–222
8. Scully C, Porter S (2000) ABC of oral health. *Oral cancer*. *Bmj* 321(7253):97–100. <https://doi.org/10.1136/bmj.321.7253.97>
9. Goldstein BY, Chang SC, Hashibe M, La Vecchia C, Zhang ZF (2010) Alcohol consumption and cancers of the oral cavity and pharynx from 1988 to 2009: an update. *Eur J Cancer Prev* 19(6):431–465. <https://doi.org/10.1097/CEJ.0b013e32833d936d>
10. Dal Maso L, Torelli N, Biancotto E et al (2016) Combined effect of tobacco smoking and alcohol drinking in the risk of head and neck cancers: a re-analysis of case-control studies using bi-dimensional spline models. *Eur J Epidemiol* 31(4):385–393. <https://doi.org/10.1007/s10654-015-0028-3>
11. Marcotte H, Lavoie MC (1998) Oral microbial ecology and the role of salivary immunoglobulin A. *Microbiol Mol Biol Rev* 62(1):71–109
12. Zhan T, Rindtorff N, Boutros M (2017) Wnt signaling in cancer. *Oncogene* 36(11):1461–1473. <https://doi.org/10.1038/ncr.2016.304>
13. Kahn M (2014) Can we safely target the WNT pathway? *Nat Rev Drug Discovery* 13(7):513–532
14. Tang W, Dodge M, Gundapaneni D, Michnoff C, Roth M, Lum L (2008) A genome-wide RNAi screen for Wnt/beta-catenin pathway components identifies unexpected roles for TCF transcription factors in cancer. *Proc Natl Acad Sci U S A* 105(28):9697–9702. <https://doi.org/10.1073/pnas.0804709105>
15. van Amerongen R, Nusse R (2009) Towards an integrated view of Wnt signaling in development. *Development* 136(19):3205–3214. <https://doi.org/10.1242/dev.033910>
16. MacDonald BT, Tamai K, He X (2009) Wnt/beta-catenin signaling: components, mechanisms, and diseases. *Dev Cell* 17(1):9–26. <https://doi.org/10.1016/j.devcel.2009.06.016>
17. Clevers H, Nusse R (2012) Wnt/ β -catenin signaling and disease. *Cell* 149(6):1192–1205
18. Yang K, Wang X, Zhang H et al (2016) The evolving roles of canonical WNT signaling in stem cells and tumorigenesis: implications in targeted cancer therapies. *Lab Invest* 96(2):116–136. <https://doi.org/10.1038/labinvest.2015.144>
19. Houschyar KS, Tapking C, Borrelli MR et al (2018) Wnt pathway in bone repair and regeneration - what do we know so far. *Front Cell Dev Biol* 6:170. <https://doi.org/10.3389/fcell.2018.00170>
20. Luo J, Chen J, Deng ZL et al (2007) Wnt signaling and human diseases: what are the therapeutic implications? *Lab Invest* 87(2):97–103. <https://doi.org/10.1038/labinvest.3700509>
21. De Santis M, Di Matteo B, Chisari E et al (2018) The role of Wnt pathway in the pathogenesis of OA and its potential therapeutic implications in the field of regenerative medicine. *Biomed Res Int* 2018:7402947. <https://doi.org/10.1155/2018/7402947>
22. Komiya Y, Habas R (2008) Wnt signal transduction pathways. *Organogenesis* 4(2):68–75. <https://doi.org/10.4161/org.4.2.5851>
23. Amin N, Vincan E (2012) The Wnt signaling pathways and cell adhesion. *Front Biosci (Landmark Ed)* 17:784–804. <https://doi.org/10.2741/3957>
24. van Amerongen R (2012) Alternative Wnt pathways and receptors. *Cold Spring Harb Perspect Biol* 4(10). <https://doi.org/10.1101/cshperspect.a007914>
25. Garcia de Herreros A, Dunach M (2019) Intracellular signals activated by canonical Wnt ligands independent of GSK3 inhibition and beta-catenin stabilization. *Cells* 8(10). <https://doi.org/10.3390/cells8101148>
26. Kang MI, Baker AR, Dextras CR, Cabarcas SM, Young MR, Colburn NH (2012) Targeting of noncanonical Wnt5a signaling by AP-1 blocker dominant-negative Jun when it inhibits skin carcinogenesis. *Genes Cancer* 3(1):37–50. <https://doi.org/10.1177/1947601912448820>
27. Ding X, Liu J, Zheng L, et al (2019) Genome-wide identification and expression profiling of wnt family genes in the silkworm, *Bombyx mori*. *Int J Mol Sci* 20(5). <https://doi.org/10.3390/ijms20051221>
28. Yang Y, Mlodzik M (2015) Wnt-Frizzled/planar cell polarity signaling: cellular orientation by facing the wind (Wnt). *Annu Rev Cell Dev Biol* 31:623–646. <https://doi.org/10.1146/annurev-cellbio-100814-125315>
29. Amano M, Nakayama M, Kaibuchi K (2010) Rho-kinase/ROCK: A key regulator of the cytoskeleton and cell polarity. *Cytoskeleton (Hoboken)* 67(9):545–554. <https://doi.org/10.1002/cm.20472>
30. Kohn AD, Moon RT (2005) Wnt and calcium signaling: beta-catenin-independent pathways. *Cell Calcium* 38(3–4):439–446. <https://doi.org/10.1016/j.ceca.2005.06.022>
31. Shifman JM, Choi MH, Mihalas S, Mayo SL, Kennedy MB (2006) Ca²⁺/calmodulin-dependent protein kinase II (CaMKII) is activated by calmodulin with two bound calciums. *Proc Natl Acad Sci U S A* 103(38):13968–13973. <https://doi.org/10.1073/pnas.0606433103>
32. Ishitani T, Kishida S, Hyodo-Miura J et al (2003) The TAK1-NLK mitogen-activated protein kinase cascade functions in the Wnt-5a/Ca(2+) pathway to antagonize Wnt/beta-catenin signaling. *Mol Cell Biol* 23(1):131–139. <https://doi.org/10.1128/mcb.23.1.131-139.2003>
33. Asatryan AD, Komarova NL (2016) Evolution of genetic instability in heterogeneous tumors. *J Theor Biol* 396:1–12. <https://doi.org/10.1016/j.jtbi.2015.11.028>
34. Osada H, Takahashi T (2002) Genetic alterations of multiple tumor suppressors and oncogenes in the carcinogenesis and progression of lung cancer. *Oncogene* 21(48):7421–7434. <https://doi.org/10.1038/sj.onc.1205802>
35. Felsner DW (2004) Oncogenes as therapeutic targets. *Semin Cancer Biol* 1(1)
36. Wang LH, Wu CF, Rajasekaran N, Shin YK (2018) Loss of tumor suppressor gene function in human cancer: an overview. *Cell Physiol Biochem* 51(6):2647–2693. <https://doi.org/10.1159/000495956>
37. Sever R, Brugge JS (2015) Signal transduction in cancer. *Cold Spring Harb Perspect Med* 5(4). <https://doi.org/10.1101/cshperspect.a006098>
38. Krug U, Ganser A, Koeffler HP (2002) Tumor suppressor genes in normal and malignant hematopoiesis. *Oncogene* 21(21):3475–3495. <https://doi.org/10.1038/sj.onc.1205322>

39. Symington LS, Rothstein R, Lisby M (2014) Mechanisms and regulation of mitotic recombination in *Saccharomyces cerevisiae*. *Genetics* 198(3):795–835. <https://doi.org/10.1534/genetics.114.166140>
40. Rivlin N, Brosh R, Oren M, Rotter V (2011) Mutations in the p53 tumor suppressor gene: important milestones at the various steps of tumorigenesis. *Genes Cancer* 2(4):466–474. <https://doi.org/10.1177/1947601911408889>
41. Leoz ML, Carballal S, Moreira L, Ocana T, Balaguer F (2015) The genetic basis of familial adenomatous polyposis and its implications for clinical practice and risk management. *Appl Clin Genet* 8:95–107. <https://doi.org/10.2147/tacg.s51484>
42. Ng JM, Yu J (2015) Promoter hypermethylation of tumour suppressor genes as potential biomarkers in colorectal cancer. *Int J Mol Sci* 16(2):2472–2496. <https://doi.org/10.3390/ijms16022472>
43. Vinson C, Chatterjee R (2012) CG methylation. *Epigenomics* 4(6):655–663. <https://doi.org/10.2217/epi.12.55>
44. Miller JL, Grant PA (2013) The role of DNA methylation and histone modifications in transcriptional regulation in humans. *Subcell Biochem* 61:289–317. https://doi.org/10.1007/978-94-007-4525-4_13
45. Morris LG, Chan TA (2015) Therapeutic targeting of tumor suppressor genes. *Cancer* 121(9):1357–1368. <https://doi.org/10.1002/cncr.29140>
46. Aubrey BJ, Strasser A, Kelly GL (2016) Tumor-suppressor functions of the TP53 pathway. *Cold Spring Harb Perspect Med* 6(5). <https://doi.org/10.1101/cshperspect.a026062>
47. Si W, Shen J, Zheng H, Fan W (2019) The role and mechanisms of action of microRNAs in cancer drug resistance. *Clin Epigenetics* 11(1):25. <https://doi.org/10.1186/s13148-018-0587-8>
48. Sansregret L, Swanton C (2017) The role of aneuploidy in cancer evolution. *Cold Spring Harb Perspect Med* 7(1). <https://doi.org/10.1101/cshperspect.a028373>
49. Pikor L, Thu K, Vucic E, Lam W (2013) The detection and implication of genome instability in cancer. *Cancer Metastasis Rev* 32(3–4):341–352. <https://doi.org/10.1007/s10555-013-9429-5>
50. Sherwood V, Leigh IM (2016) WNT signaling in cutaneous squamous cell carcinoma: a future treatment strategy? *J Invest Dermatol* 136(9):1760–1767
51. Ma X, Yu H (2006) Global burden of cancer. *Yale J Biol Med* 79(3–4):85–94
52. Peto R (2016) Epidemiology, multistage models, and short-term mutagenicity tests. *Int J Epidemiol* 45(3):621–637. <https://doi.org/10.1093/ije/dyv199>
53. Luzzatto L (2011) Somatic mutations in cancer development. *Environ Health* 10(Suppl 1):S12. <https://doi.org/10.1186/1476-069x-10-s1-s12>
54. Podlaha O, Riester M, De S, Michor F (2012) Evolution of the cancer genome. *Trends Genet* 28(4):155–163. <https://doi.org/10.1016/j.tig.2012.01.003>
55. Valastyan S, Weinberg RA (2011) Tumor metastasis: molecular insights and evolving paradigms. *Cell* 147(2):275–292. <https://doi.org/10.1016/j.cell.2011.09.024>
56. Fouad YA, Aanei C (2017) Revisiting the hallmarks of cancer. *Am J Cancer Res* 7(5):1016–1036
57. Houschyar KS, Momeni A, Pyles MN, Maan ZN, Whittam AJ, Siemers F (2015) Wnt signaling induces epithelial differentiation during cutaneous wound healing. *Organogenesis* 11(3):95–104. <https://doi.org/10.1080/15476278.2015.1086052>
58. Bastakoty D, Young PP (2016) Wnt/beta-catenin pathway in tissue injury: roles in pathology and therapeutic opportunities for regeneration. *Faseb J* 30(10):3271–3284. <https://doi.org/10.1096/fj.201600502R>
59. Guo X, Wang XF (2009) Signaling cross-talk between TGF-beta/BMP and other pathways. *Cell Res* 19(1):71–88. <https://doi.org/10.1038/cr.2008.302>
60. Kiecker C, Niehrs C (2001) A morphogen gradient of Wnt/beta-catenin signalling regulates anteroposterior neural patterning in *Xenopus*. *Development* 128(21):4189–4201
61. Munoz-Descalzo S, Hadjantonakis AK, Arias AM (2015) Wnt/ss-catenin signalling and the dynamics of fate decisions in early mouse embryos and embryonic stem (ES) cells. *Semin Cell Dev Biol* 47–48:101–109. <https://doi.org/10.1016/j.semcdb.2015.08.011>
62. Miki T, Yasuda SY, Kahn M (2011) Wnt/beta-catenin signaling in embryonic stem cell self-renewal and somatic cell reprogramming. *Stem Cell Rev Rep* 7(4):836–846. <https://doi.org/10.1007/s12015-011-9275-1>
63. Sokol SY (2011) Maintaining embryonic stem cell pluripotency with Wnt signaling. *Development* 138(20):4341–4350. <https://doi.org/10.1242/dev.066209>
64. Potok MA, Cha KB, Hunt A et al (2008) WNT signaling affects gene expression in the ventral diencephalon and pituitary gland growth. *Dev Dyn* 237(4):1006–1020. <https://doi.org/10.1002/dvdy.21511>
65. Houschyar KS, Tapking C, Duscher D, et al (2019) Regulation of bone metabolism by the Wnt signaling pathway. *Handchir Mikrochir Plast Chir* 51(4):309–318. Regulation des Knochenstoffwechsels durch den Wnt-Signalweg. <https://doi.org/10.1055/a-0642-1830>
66. Cohen ED, Wang Z, Lepore JJ et al (2007) Wnt/beta-catenin signaling promotes expansion of Isl-1-positive cardiac progenitor cells through regulation of FGF signaling. *J Clin Invest* 117(7):1794–1804. <https://doi.org/10.1172/jci31731>
67. Grigoryan T, Wend P, Klaus A, Birchmeier W (2008) Deciphering the function of canonical Wnt signals in development and disease: conditional loss- and gain-of-function mutations of beta-catenin in mice. *Genes Dev* 22(17):2308–2341. <https://doi.org/10.1101/gad.1686208>
68. Zechner D, Muller T, Wende H et al (2007) Bmp and Wnt/beta-catenin signals control expression of the transcription factor Olig3 and the specification of spinal cord neurons. *Dev Biol* 303(1):181–190. <https://doi.org/10.1016/j.ydbio.2006.10.045>
69. Houschyar KS, Tapking C, Puladi B, et al (2020) Wnt signaling in cutaneous wound healing. *Handchir Mikrochir Plast Chir* 52(2):151–158. Wnt-Signalwege bei kutaner Wundheilung. <https://doi.org/10.1055/a-1017-3600>
70. Kim YM, Kahn M (2014) The role of the Wnt signaling pathway in cancer stem cells: prospects for drug development. *Res Rep Biochem* 4:1–12. <https://doi.org/10.2147/rrbc.s53823>
71. Mah AT, Yan KS, Kuo CJ (2016) Wnt pathway regulation of intestinal stem cells. *J Physiol* 594(17):4837–4847. <https://doi.org/10.1113/jp271754>
72. Novellasedemunt L, Antas P, Li VS (2015) Targeting Wnt signaling in colorectal cancer. A review in the theme: cell signaling: proteins, pathways and mechanisms. *Am J Physiol Cell Physiol* 309(8):C511–21. <https://doi.org/10.1152/ajpcell.00117.2015>
73. Blagodatski A, Poteryaev D, Katanaev VL (2014) Targeting the Wnt pathways for therapies. *Mol Cell Ther* 2:28. <https://doi.org/10.1186/2052-8426-2-28>
74. Wilson S, Rydström A, Trimborn T et al (2001) The status of Wnt signalling regulates neural and epidermal fates in the chick embryo. *Nature* 411(6835):325–330
75. Fu J, Hsu W (2013) Epidermal Wnt controls hair follicle induction by orchestrating dynamic signaling crosstalk between the epidermis and dermis. *J Invest Dermatol* 133(4):890–898
76. Houschyar KS, Borrelli MR, Tapking C, et al (2020) Molecular mechanisms of hair growth and regeneration: current

- understanding and novel paradigms. *Dermatology* 1–10. <https://doi.org/10.1159/000506155>
77. Greco V, Chen T, Rendl M et al (2009) A two-step mechanism for stem cell activation during hair regeneration. *Cell Stem Cell* 4(2):155–169. <https://doi.org/10.1016/j.stem.2008.12.009>
 78. Lim X, Nusse R (2013) Wnt signaling in skin development, homeostasis, and disease. *Cold Spring Harb Perspect Biol* 5(2):a008029
 79. Wu P, Zhang Y, Xing Y et al (2019) The balance of Bmp6 and Wnt10b regulates the telogen-anagen transition of hair follicles. *Cell Commun Signal* 17(1):16. <https://doi.org/10.1186/s12964-019-0330-x>
 80. Li J, Ji L, Chen J, Zhang W, Ye Z (2015) Wnt/beta-catenin signaling pathway in skin carcinogenesis and therapy. *Biomed Res Int* 2015:964842. <https://doi.org/10.1155/2015/964842>
 81. Choi YS, Zhang Y, Xu M et al (2013) Distinct functions for Wnt/ β -catenin in hair follicle stem cell proliferation and survival and interfollicular epidermal homeostasis. *Cell Stem Cell* 13(6):720–733
 82. Jensen KB, Collins CA, Nascimento E et al (2009) Lrig1 expression defines a distinct multipotent stem cell population in mammalian epidermis. *Cell Stem Cell* 4(5):427–439
 83. Ray S, Foote HP, Lechler T (2013) beta-Catenin protects the epidermis from mechanical stresses. *J Cell Biol* 202(1):45–52. <https://doi.org/10.1083/jcb.201212140>
 84. Chua AW, Ma D, Gan SU et al (2011) The role of R-spondin2 in keratinocyte proliferation and epidermal thickening in keloid scarring. *J Invest Dermatol* 131(3):644–654
 85. Madoka S (2006) Upregulation of the Wnt/ β -catenin pathway induced by transforming growth factor- β in hypertrophic scars and keloids. *Acta Derm Venereol* 86:300–307
 86. Popp S, Waltering S, Herbst C, Moll I, Boukamp P (2002) UV-B-type mutations and chromosomal imbalances indicate common pathways for the development of Merkel and skin squamous cell carcinomas. *Int J Cancer* 99(3):352–360
 87. Ra SH, Li X, Binder S (2011) Molecular discrimination of cutaneous squamous cell carcinoma from actinic keratosis and normal skin. *Mod Pathol* 24(7):963–973
 88. Haider AS, Peters SB, Kaporis H et al (2006) Genomic analysis defines a cancer-specific gene expression signature for human squamous cell carcinoma and distinguishes malignant hyperproliferation from benign hyperplasia. *J Invest Dermatol* 126(4):869–881
 89. Watt S, Pourreyron C, Purdie K et al (2011) Integrative mRNA profiling comparing cultured primary cells with clinical samples reveals PLK1 and C20orf20 as therapeutic targets in cutaneous squamous cell carcinoma. *Oncogene* 30(46):4666–4677
 90. Yeh SA (2010) Radiotherapy for head and neck cancer. *Semin Plast Surg* 24(2):127–136. <https://doi.org/10.1055/s-0030-1255330>
 91. Rutkowski T (2014) The role of tumor volume in radiotherapy of patients with head and neck cancer. *Radiat Oncol* 9:23. <https://doi.org/10.1186/1748-717x-9-23>
 92. Zhang X, Hao J (2015) Development of anticancer agents targeting the Wnt/ β -catenin signaling. *Am J Cancer Res* 5(8):2344
 93. Krishnamurthy N, Kurzrock R (2018) Targeting the Wnt/beta-catenin pathway in cancer: Update on effectors and inhibitors. *Cancer Treat Rev* 62:50–60
 94. Liu J, Pan S, Hsieh MH et al (2013) Targeting Wnt-driven cancer through the inhibition of Porcupine by LGK974. *Proc Natl Acad Sci* 110(50):20224–20229
 95. Shang S, Hua F, Hu ZW (2017) The regulation of beta-catenin activity and function in cancer: therapeutic opportunities. *Oncotarget* 8(20):33972–33989. <https://doi.org/10.18632/oncotarget.15687>
 96. de Sousa EMF, Vermeulen L (2016) Wnt Signaling in cancer stem cell biology. *Cancers (Basel)* 8(7). <https://doi.org/10.3390/cancers8070060>
 97. Qu Y, Gharbi N, Yuan X et al (2016) Axitinib blocks Wnt/ β -catenin signaling and directs asymmetric cell division in cancer. *Proc Natl Acad Sci* 113(33):9339–9344
 98. Pai SG, Carneiro BA, Mota JM et al (2017) Wnt/beta-catenin pathway: modulating anticancer immune response. *J Hematol Oncol* 10(1):101. <https://doi.org/10.1186/s13045-017-0471-6>

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