



# Green turtle diet is dominated by seagrass in the Western Indian Ocean except amongst gravid females

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## Abstract

Green turtles (*Chelonia mydas*) are key herbivores of tropical and subtropical neritic habitats and play a major role in structuring seagrass meadows. We present the first detailed assessment of green turtle diet in the Western Indian Ocean using the gut contents of salvaged animals from three atolls in the Republic of Seychelles separated from each other by 400–825 km: Cosmoledo (adults,  $n = 12$ ), Farquhar (adults,  $n = 33$ ; immature,  $n = 1$ ) collected in 1982–1983; and Desroches (immatures,  $n = 8$ ) in 2016–2018. We report the first comparison of the diets of gravid females ( $n = 17$ ), males ( $n = 26$ ) and non-breeding females ( $n = 2$ ) at sites providing both foraging and breeding habitat. Seagrass (mostly *Thalassodendron ciliatum*) dominated the diet, accounting for 95% of the mean gut content biomass for males and non-breeding females but only 58% for gravid females, alongside relatively large amounts of substrate (14%) and macroalgae (13%). Satellite tracking of post-nesting green turtles from Chagos Archipelago in 2016 located foraging sites at Farquhar Atoll that coincided with capture locations of 26 of the 33 adult turtles sampled there in 1983. In situ surveys of those sites in 2018 revealed extensive nearly monospecific beds of *T. ciliatum*. The prominence of seagrass in the diet of green turtles and connectivity between foraging and nesting habitats throughout the region illustrate the need to conserve and monitor seagrass habitats of the Western Indian Ocean especially in the context of changing green turtle population densities.

## Introduction

Green turtles (*Chelonia mydas*) are marine ecosystem engineers that structure seagrass meadows through their foraging activities, often increasing its complexity (Coleman and Williams 2002) or, conversely, reducing species composition (Kelkar et al. 2013a; Lal et al. 2010) and production rates

(Kelkar et al. 2013b). There is increasing evidence that this functioning helps to provide seagrass habitats with greater resilience to anthropogenic impacts (Christianen et al. 2012). Changes in green turtle density at foraging sites will, thus, have a consequence for seagrass ecosystem health. Due to numerous anthropogenic threats and population declines, the green turtle had been listed as globally endangered on the IUCN Red List (Seminoff 2004), but long-term population recovery, thanks to successful conservation strategies such as protection of turtles and their habitats at nesting sites, has led to long-term population recovery. This is reflected in recent and ongoing IUCN downlisting of various green turtle subpopulations around the world (e.g., to “least concern” for the Hawaiian and South Atlantic subpopulations (Chaloupka and Pilcher 2019; Broderick and Patricio 2019) and to Vulnerable for the North Indian Ocean subpopulation (Mancini et al. 2019) and, in literature, documenting of increasing population trends globally (Chaloupka et al. 2008; Seminoff et al. 2015; Mazaris et al. 2017), including upward trends in the Western Indian Ocean (Bourjea et al. 2007; Lauret-Stepler et al. 2007; Mortimer et al. 2011).

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Upon leaving the nesting beach as hatchlings, young green turtles occupy open ocean pelagic habitats (Bjorndal 1997). Post-hatchling green turtles are believed to spend their first 3–5 years in open ocean pelagic habitat where they attain carapace lengths of 25–35 cm feeding primarily on jellies and salps and then recruit to neritic habitats (Reich et al. 2007). Ontogenetic dietary change is complex and varies regionally (e.g., Cardona et al. 2010; Fukuoka et al. 2019). Typically, the diet is omnivorous during the early pelagic stages and becomes primarily herbivorous after recruitment to the neritic zone (e.g., Howell et al. 2016; Vélez-Rubio et al. 2016; Burgett et al. 2018). In the Central North Pacific, however, some individuals up to 70 cm curved carapace length (CCL) forage pelagically on a primarily carnivorous diet (Parker et al. 2011). At some neritic foraging habitats, seagrass predominates in the diet (e.g., Mortimer 1981; Vander Zanden et al. 2013); while at other sites, macroalgae is the primary food source (e.g., Garnett et al. 1985; Prior et al. 2016). Even terrestrial plant material, especially mangrove leaves and propagules (or seeds), can feature prominently in green turtle diets (Arthur et al. 2009; Limpus and Limpus 2000; Nagaoka et al. 2012). Contrary to the long-held belief that green turtles are strictly herbivorous, however, animal matter ranging from accidentally ingested small hydrozoans and bryozoans that encrust plant material to purposefully consumed sponges (Mortimer 1981) and significant amounts of gelatinous macrozooplankton (e.g., Amorocho and Reina 2007; Burkholder et al. 2011; Fukuoka et al. 2019; González Carman et al. 2014; Quiñones et al. 2010) have been reported in the diet of both juvenile and adult green turtles.

Food selection may correlate with differences in intestinal microflora (Bjorndal 1980), which varies as turtles transition from pelagic to neritic habitats (Campos et al. 2018; Price 2016) and with state of health (Ahasan et al. 2017). It follows that intestinal microflora typical of individuals that feed on seagrass likely differs from that of algivores and they are less efficient at digesting algae and vice versa (Bjorndal et al. 1991), but diets comprising large amounts of both seagrass and algae are also sometimes reported (e.g., López-Mendilaharsu et al. 2005; Shimada et al. 2014; Whiting et al. 2014). Stable isotope studies have been used to show that individuals in the same area can have long-term dietary preferences (Thomson et al. 2018). On the other hand, abrupt shifts between algae, seagrass and mangrove diets have been observed by examining the guts of individual green turtles (Brand et al. 1999; Arthur et al. 2009) and may simply reflect the food items available that yield the highest nutritional value with minimal search and handling costs (Bjorndal 1997).

Foraging patterns of adult turtles can be disrupted by reproduction. Appropriate forage is often lacking during long-distance migrations between feeding and breeding

grounds (Carr 1975) and at some sites is not available in the vicinity of the nesting beach (e.g., at Tortuguero, Costa Rica and Ascension Island, South Atlantic Ocean). Guts of migrating green turtles captured between seagrass pastures in Nicaragua and the Tortuguero nesting beach contained only small quantities of low grade forage (Mortimer 1982) as did those of nesting females examined at Tortuguero (Meylan 1978), which contained water hyacinth debris and flotsam deposited in the river mouth (Mortimer 1982). At Ascension, the guts of females were empty; while in Northern Cyprus, a nesting area where there is abundant seagrass, females were found to have guts packed with seagrass (Hays et al. 2002). Breeding Ascension turtles sometimes move in to eat refuse dumped from ships or shore (Carr et al. 1974) suggesting that their fast may not be voluntary. At Raine Island Australia, analysis of the gut contents of 101 nesting females killed on shore by heat stress found that 60% of the guts were completely empty, and the remainder showed evidence of only intermittent foraging, mostly on calcareous algae and animal matter including sea anemones and jellyfish (Tucker and Read 2001). The authors did not describe availability of forage near Raine Island, so it is unclear whether the restricted food intake was determined by habitat, or as Bjorndal (1997) suggests, that occupation of visceral space by female reproductive tissues may preclude or limit continuous feeding. To date, however, no published studies have evaluated the feeding behaviour of both breeding males and females at nesting sites where abundant forage was available.

Here, we investigate green turtle foraging ecology at three Western Indian Ocean coral reef atolls in the Republic of Seychelles, in the context of what is known globally about the diet of immature and adult green turtles (Esteban et al. 2019 Mar Biol in review). Specifically, we (1) examine green turtle gut contents (oesophagus and stomach) from adult male turtles and breeding and non-breeding females, and we compare the amount and relative importance of food items encountered; (2) examine the oesophageal contents of 8 otherwise apparently healthy immature green turtles found dead at Desroches atoll; and (3) assess the habitat at locations where adult turtles were captured in 1982–1983 and which satellite tracking later identified as green turtle foraging habitat in 2016. By combining these results, the study will improve knowledge of green turtle habitat requirements in the Western Indian Ocean to help guide regional conservation management.

## Materials and methods

### Study site and sample collection

This study focuses on the diet composition of green turtles at three sites (Cosmoledo, Farquhar and Desroches atolls)

(Fig. 1) in the outer islands of the Republic of Seychelles. All three atolls host the following life stages of green turtles: adult males and females that mate and lay eggs at the atolls (Mortimer 1984); adult females that nest at distant sites but use the atoll as adult foraging habitat during intervals between nesting seasons (Bourjea et al. 2015; Christiansen et al. 2017; GC Hays and N Esteban unpubl data); and immature and subadult turtles (> 35 cm carapace length) that use the atoll as developmental habitat. The atolls probably also provide adult foraging habitat for males that breed at distant sites, but this is not yet confirmed. All three atolls feature expanses of seagrass habitat inside and outside their fringing reefs. These are dominated by the following species: Cosmoledo, *Thalassodendron ciliatum*, *Thalassia hemprichii*, *Cymodocea rotundata*, and *Cymodocea serrulata*; Farquhar, *T. ciliatum* and *T. hemprichii*; and Desroches, *T. ciliatum*, *T. hemprichii*, and *Halophila ovalis*, as well as *Syringodium isoetifolium* (Kalugina-Gutnik et al. 1992; JA Mortimer and M Morgan unpubl data). Associations of seagrass and macroalgae occur, the most common involving the calcareous algae, *Halimeda* spp (see Kalugina-Gutnik et al. 1992 for descriptions and list of species).

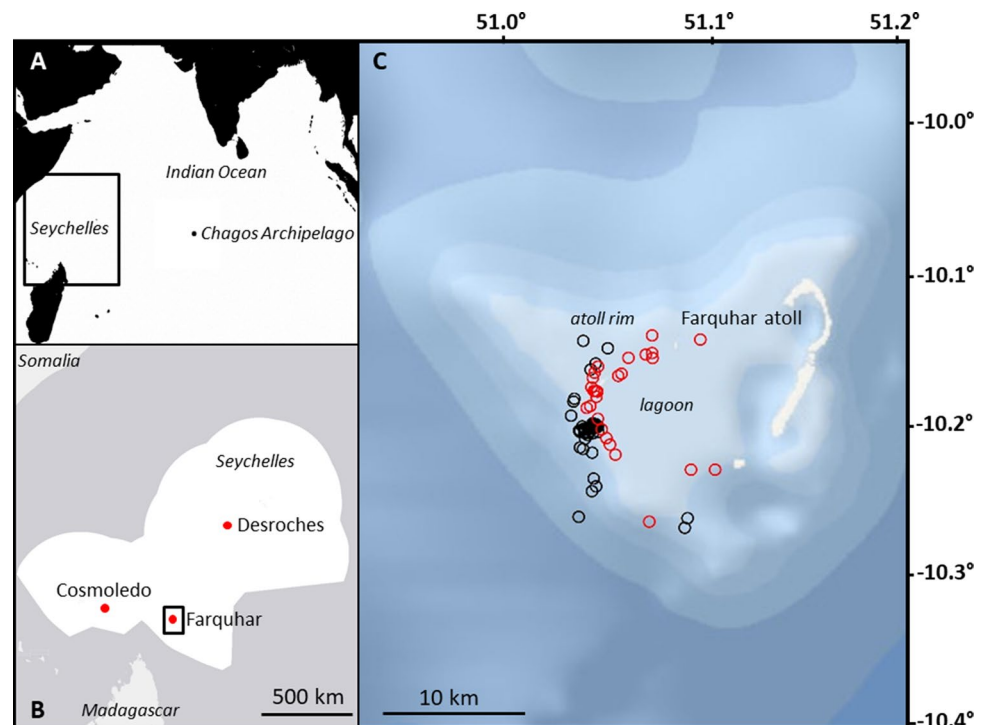
During 1982–1983, green turtles were hunted at Cosmoledo and Farquhar atolls for human consumption. Adult males were legally harpooned at sea, usually within or just beyond the reef crest of the atoll. Some males were kept alive and transported (~ 800–1100 km) to the human population centres of the Inner Islands, while others were slaughtered on site and their meat was salted (Mortimer 1984). In

addition, nesting females whose fatty meat was considered superior to that of the males were occasionally taken (illegally) from the nesting beach for local consumption. No turtles were killed specifically to provide samples for this study.

The digestive tract of each turtle was dissected on site after slaughter and all food items were removed from the oesophagus and upper cardiac region of the stomach at the point where food had not yet been modified by digestion (defined as gut throughout this study). In all, 46 samples (Cosmoledo,  $n = 12$ ; Farquhar,  $n = 34$ ) were collected along with the following data: location of capture (marked on Admiralty Chart), CCL (cm), and sex (Table 1). All the males were adults with long tails (Mortimer 1984) and many of them had been harpooned while mating. Reproductive (gravid) females were identified by ovarian follicles > 2 cm (Owens 1980), but most breeding females were captured on the nesting beach. The samples were initially fixed in 5% formalin in sea water for 24 h and subsequently transferred to 70% ethanol for preservation before analysis. An absence of indeterminate slurry suggests that our samples were well preserved despite their long storage.

During 2016–2018, at Desroches atoll, Island Conservation Society (ICS) personnel collected gut content samples from dead immature green turtles stranded on the beach ( $n = 8$ ). They all appeared to be in good health prior to stranding and their guts were packed full of food (seagrass). Cause of death is unknown but may have resulted from drowning subsequent to entanglement in fishing gear. CCL was measured and a small food sample from a single point

**Fig. 1** The location of study sites: **a** Location of Republic of Seychelles in the Western Indian Ocean. **b** Three atolls (indicated by red dots) within the Seychelles Exclusive Economic Zone (shown in white) where samples were collected. **c** Comparison of capture locations of green turtles sampled in 1982–1983 (red open circles) with Fastloc-GPS locations (black open circles) indicating core feeding sites (within and outside of the atoll rim) of post-nesting female green turtle satellite tracked from Diego Garcia atoll in Chagos Archipelago (location shown on Fig. 1a) to Farquhar atoll (location indicated by black box on Fig. 1b) in 2015–2016. Bathymetry map source: Esri, DeLorme, GEBCO, NOAA NGDC, and other contributors



**Table 1** Capture dates, locations and details (number, size, sex) of green turtles harvested in Republic of Seychelles

Location	Latitude and Longitude	Dates sampled	Number			Size Range (CCL, cm)	Mean $\pm$ SE (cm)
			M	F	U		
Cosmoledo	9.7088°S 47.5153°E	19 Jan–25 May 1982	4	8	–	93–117	109 $\pm$ 2.19
Farquhar	10.1881°S 51.1566°E	10 May–7 Oct 1983	23	10	1	72–115	103 $\pm$ 1.28
Desroches	5.6912°S 53.6671°E	Jan 2016–Jan 2018	–	–	8	48.5–71	63 $\pm$ 2.44

*M* male, *F* female, *U* unknown; immature

along the full oesophagus was collected from each, frozen, and stored in 70% ethanol prior to analysis.

### Diet analysis

Processing and identification of samples took place at Swansea University. Samples were filtered, blotted dry, and wet mass measured to the nearest 0.01 g. The 1982–1983 Cosmoledo/Farquhar samples were subsampled, but the 2016–2018 Desroches samples were analysed fully given their small size. A stratified sub-sampling protocol was followed to ensure representation of the whole sample: (1) large items were removed from sample for identification, (2) the remaining sample was mixed, spread out evenly and split into 10 equal subsamples in a gridded tray, (3) a random number generator was used to randomly select each subsample for analysis to reduce bias, (4) for the first five samples, 50% of the sample was analysed, 10% at a time. No new dietary items were found after identifying specimens in the first two 10% subsamples. Remaining samples: two subsamples were analysed (20% of the original sample size).

Within each gut sample, dietary items were identified to the lowest taxonomic level using a dissecting microscope (Olympus SZX61, 0.7–11.5x with an SDF PLAPO 1XPF objective lens) and available identification guides, literature and websites (Lanyon 1986; Kalugina-Gutnik et al. 1992; Waycott et al. 2004; WILD Singapore, 2016). To gain a positive identification of sponges, a small fragment was dissolved in sodium hypochlorite (NaOCl) to check for the presence of spicules. Photomicrographs were taken of all dietary items (Olympus UC30 3.2 Microscope Digital Camera attachment; Olympus cellSens imaging software) for subsequent verification of species. Dry mass of each taxon was weighed to the nearest 0.1 mg after oven drying for 24 h at 40 °C.

### Identifying and ground-truthing foraging locations at Farquhar Atoll

Two sets of data were used to identify green turtle foraging locations at Farquhar atoll. The first entailed recording onto Admiralty Charts the locations where 27 green turtles were captured in 1982–1983 and then transferring those points to Google Earth. The second comprised day-time

surfacing locations of a green turtle that had been equipped with a Fastloc-GPS Argos satellite tag after nesting on the island of Diego Garcia (7.42°S, 72.45°E) in the Chagos Archipelago (Esteban et al. 2017), and then tracked to its foraging grounds at Farquhar in 2016 (Fig. 1a, b). Fastloc-GPS locations obtained from 4 or more satellites and with residual values of  $<35$  were examined (see Dujon et al. 2014 for a description of Fastloc-GPS accuracy). The tracking data were analysed using ArcGIS 10.3 to identify locations ( $n=98$ ) visited during local daytime [i.e., those positions recorded between 0900 and 1700 h local time (UTC + 4 h)].

During March 2016 and March 2018, as many of the daytime Fastloc-GPS location points as possible were surveyed to assess benthic habitats. On 25 March 2016, 14 points situated outside the rim of the atoll were assessed in terms of depth and visible characteristics of the benthos, using a glass-bottomed bucket and the fathometer built into the small research skiff. Two years later, on 22–23 March 2018, benthic habitat at a sub-sample of Fastloc-GPS daytime locations ( $n=15$ ), this time inside the lagoon, was ground-truthed and benthic features of the seagrass habitats were recorded using Seagrass-Watch monitoring protocols ([www.seagrasswatch.org](http://www.seagrasswatch.org)) for a distance of at least 20 m from each location. Substrate type was assessed by placing quadrats (0.25 m<sup>2</sup>;  $n = 52$ , mean = 3.47 quadrats, range = 1–5 per site) at intervals of at least 10 m apart (estimated using fin kicks) and recording relative cover of broad benthic groups (seagrass, hard coral, soft coral, sponge, macroalgae, rock, rubble, dead coral, sand) and species composition. Care was taken to search under the seagrass canopy for any additional species. Depths at those sites were measured from the boat using a hand-held depth finder (HawkEye DT1H). Separate qualitative surveys of seagrass habitat were also conducted at various points around the inside of the lagoon to determine what other seagrass species and habitats occur at Farquhar.

### Data analysis

Univariate analyses were performed in R version 3.3.3 (R Core Team 2017). To understand the relative importance of various species in the diet, the percent frequency of occurrence, percent biomass and an Index of Relative Importance (IRI) were calculated for each diet item/species (see von Brandis et al. 2014 for equation).

To account for differences in gut fullness, the biomass of diet species was standardised. Diet items/species biomass % by location, sex, and breeding status (of females based on status of the ovaries) were presented using the ggplot2 package (Wickham 2016). All means are presented with  $\pm 1$  SE. We conducted analysis of differences in green turtle diet composition by sex, breeding status and location using a two-way nested ANOSIM in Primer v7 (Clarke and Warwick 1994).

Diet composition data (percentages based on biomass) were arcsine-square-root-transformed for analysis. A Mann–Whitney  $U$  was used to investigate the difference between the biomass of seagrass consumed by “gravid females” and by “males and non-breeding females”. A one-way ANOVA was used to investigate the difference in gut weight between “gravid females”, “non-breeding females” and “males”. The Tukey test was used to compare mean gut content wet weight of males, gravid females, and non-breeding females.

## Results

### Diet analysis

Green turtle diet was primarily plant based in the Seychelles. At Cosmoledo and Farquhar atolls, seagrass represented an average of 81% of the diet (SE = 4%;  $n = 46$ ). In 37 individuals (80.4%) seagrass, mostly *Thalassodendron ciliatum*, dominated; in 1 turtle (2.2%) macroalgae dominated; and 8 turtles (17.4%) were characterised by mixed diets. No significant difference ( $p = 0.605$ ) was evident between the diets of males ( $n = 26$ ) and non-breeding females ( $n = 2$ ); so, data for these two groups were pooled for subsequent analysis. The diet of non-breeding females was not considered separately due to the low sample size.

Dietary items were categorised into 8 dietary component groups: seagrass, macroalgae, sponges, bryozoans, cnidarians (mostly epiphytic), miscellaneous invertebrates, substrate, and marine debris (Table 2). Seagrass accounted for the biomass of almost all of the gut contents analysed of adult males and non-breeding females ( $95 \pm 2\%$ ; mean  $\pm$  SE;  $n = 28$ ), but only  $58 \pm 8\%$  (mean  $\pm$  SE;  $n = 17$ ) of the diet of gravid breeding females (Fig. 3a, Table 2). This difference was statistically significant (Mann–Whitney  $U$  test,  $U = 62$ ,  $N1 = 28$ ,  $N2 = 17$ ,  $p < 0.0001$ ). When adult males ( $n = 26$ ) and non-breeding females ( $n = 2$ ) were considered separately, seagrass accounted for  $95 \pm 2\%$  and  $100 \pm 0\%$  (mean  $\pm$  SE) of the diet, respectively. In contrast, the gravid female diet included relatively large amounts of substrate ( $14 \pm 5\%$ ; mean  $\pm$  SE) and macroalgae ( $13 \pm 3\%$ ; mean  $\pm$  SE). *Halimeda* spp were disproportionately abundant (35%) in gravid females ( $n = 6$ ), but absent from the guts of adult males and non-breeding females. Cnidarians (mainly epiphytic hydrozoans) and sponges occurred frequently but with low biomass across all turtle groups.

In terms of frequency of occurrence, seagrass was encountered in all turtles (100%). In male and non-breeding females, epiphytic cnidarians (75%), macroalgae (43%), sponge (39%), and marine debris (32%) were all ingested frequently, with other dietary components less so. In the diet of gravid females, most dietary components occurred with relatively high frequency: macroalgae (82%), sponges (82%), substrate (71%), bryozoans (65%), cnidarians (41%) and marine debris (35%).

The IRI of dietary items included 26 dietary items: 15 at Cosmoledo, 25 at Farquhar and 7 at Desroches (Table 3). Seagrass was the most important dietary item at all sites and amongst all groups of turtles. This was mostly *T. ciliatum* but in some cases *T. hemprichii*. Small amounts of *Halodule uninervis* and *Halophila stipulacea*

**Table 2** Key diet composition of male and non-breeding female ( $n = 28$ ) and gravid female ( $n = 17$ ) green turtles in Seychelles. Frequency of occurrence (FO) of dietary components present in turtles (1982–1983) and relative biomass of dietary components found in the gut contents

Diet groups	Male and non-breeding female					Gravid female				
	FO	Relative proportion of biomass				FO	Relative proportion of biomass			
		Mean	$\pm$ SE	Min	Max		Mean	$\pm$ SE	Min	Max
Seagrasses	100	95	2	40	100	100	58	8	1	100
Macroalgae	43	2	2	0	57	82	15	3	0	35
Sponges	39	2	1	0	24	82	7	2	0	29
Bryozoans	4	*	*	0	0	65	3	1	0	20
Cnidarians	75	*	*	0	0	41	*	*	0	2
Invertebrates <sup>a</sup>	7	*	*	0	1	12	*	*	0	0
Substrate	4	*	*	0	0	71	15	6	0	84
Marine debris <sup>b</sup>	32	*	*	0	11	35	1	1	0	19

\* <0.1 mg (trace)

<sup>a</sup>Gastropoda, Crustacea, Polychaeta, Holothuroidea

<sup>b</sup>Plastic, glass, other

were also recorded. These results indicate consistency in green turtle diet at three sites separated from each other by 400–825 km in the Western Indian Ocean. Gravid female diets had lower IRI seagrass scores than those of male, non-breeding female, and immature turtles. *Halimeda* spp featured prominently in the diet of gravid females at Cosmoledo (IRI = 15), while calcium carbonate substrate was prominent in the diet of gravid females at Farquhar (IRI = 12).

Differences in gut content biomass between “breeding females” and “males and non-breeding females” were statistically significant by a two-way nested ANOSIM (global  $r=0.569$ ,  $p<0.01$ ) (Fig. 2). Diet compositions at Cosmoledo and Farquhar were similar (global  $r=0$ ,  $p=0.667$ ). Gravid female gut contents were 53% less in volume and more varied (seven dietary component groups including macroalgae, calcium carbonate substrate, seagrass) compared

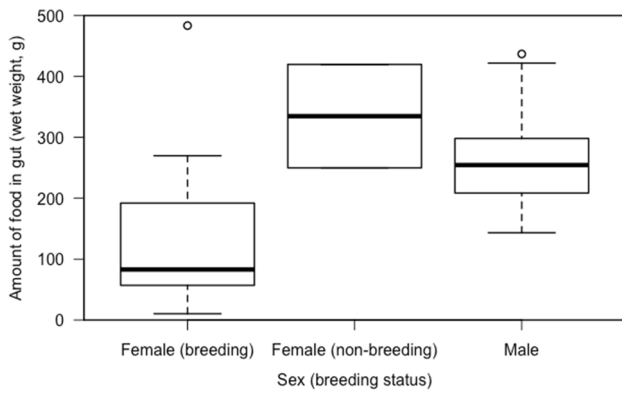
to male and non-breeding female guts (only three dietary component groups found  $>0.1$  mg despite food availability) (Table 2). Wet weights of the gut samples differed significantly (ANOVA,  $F_{2,42}=11.51$ ,  $p<0.001$ ) by sex and female breeding status. Breeding female ( $n=17$ ) gut biomass was significantly lower than those of males ( $p<0.001$ ;  $n=26$ ) and non-breeding females ( $p<0.05$ ;  $n=2$ ) (Fig. 2).

Figure 3 compares the contributions made by the eight dietary component groups and by individual species of seagrass to the diets of breeding females, males, and non-breeding females at Cosmoledo and Farquhar. Seagrass was the most abundant dietary component in all turtle groups (Fig. 3a) and at both sites (Fig. 3c). *T. ciliatum* was the dominant seagrass in the diets of both males and breeding females, but *T. hemprichii* was dominant in the diets of non-breeding females ( $n=2$ ) (Fig. 3b).

**Table 3** Index of Relative Importance (IRI) of dietary items in gut contents of green turtles captured in 1982–1983 at Cosmoledo atoll (C) ( $n=7$  gravid females (G);  $n=5$  males and non-breeding females (O) and Farquhar atoll (F) [ $n=10$  gravid females (G);  $n=23$  males and non-breeding females (O);  $n=1$  immature (I)], and in 2016–2018 at Desroches atoll (D) [ $n=8$  immature (I)]

Kingdom Phylum	Genus species	IRI (sites)					
		C		F			D
		G	O	G	O	I	I
Plantae							
Tracheophyta	<i>Thalassodendron ciliatum</i>	51	40	58	74	88	92
	<i>Halodule uninervis</i>	<1	<1	–	–	–	–
	<i>Thalassia hemprichii</i>	4	48	<1	4	–	–
	<i>Halophila stipulacea</i>	–	–	–	<1	–	–
Chlorophyta	<i>Halimeda</i> spp	15	–	<1	–	–	<1
	<i>Caulerpa</i> spp	–	–	2	<1	12	–
	<i>Caulerpa serrulata</i>	–	–	<1	–	–	–
Rhodophyta	x	–	–	–	<1	–	–
	<i>Hypnea esperi</i>	<1	–	–	<1	–	–
	<i>Heterosiphonia</i> spp	–	–	<1	<1	–	–
Phaeophyta	<i>Turbinaria</i> sp	–	–	1	–	–	–
Macroalgae	x	3	–	1	1	–	<1
Animalia							
Sponges	x	2	<1	6	1	–	4
Cnidaria	x	–	<1	–	<1	–	–
	x	<1	<1	<1	<1	–	–
Bryozoa	x	2	<1	1	–	–	<1
Echinodermata	x	<1	–	<1	–	–	–
Annelida	x	–	–	–	<1	–	<1
Crustacea	x	–	–	–	<1	–	–
	<i>Ocypode cordimanus</i>	–	–	–	<1	–	–
Mollusca	x	–	–	–	<1	–	–
Other							
Glass		<1	–	–	<1	–	–
Plastic		<1	<1	<1	<1	<1	–
Debris		–	–	<1	<1	–	–
Shell		<1	–	<1	–	–	–
Substrate		3	–	12	<1	–	<1

x unidentified species, – none

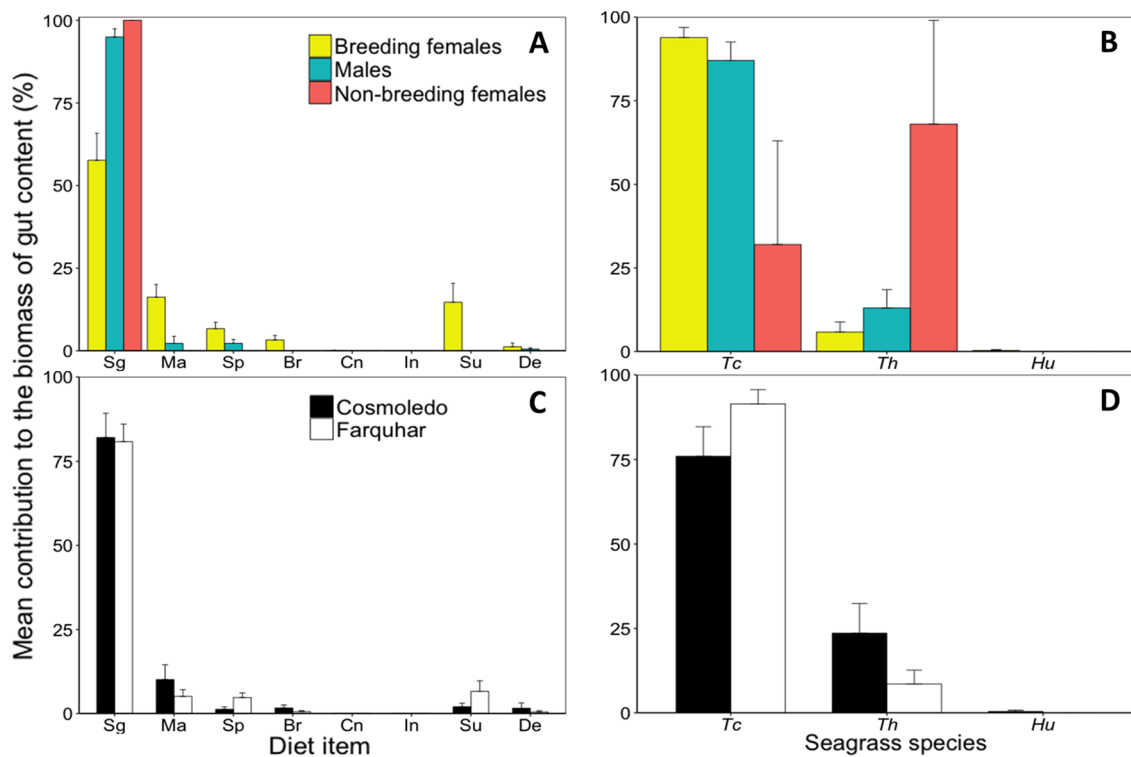


**Fig. 2** Comparison of the amount of food found in the guts of breeding females, non-breeding females, and males. Wet weight (g) was recorded for each gut sample. Bold horizontal lines indicate median, boxes delineate upper and lower quartiles and whiskers define range. Outliers are plotted as separate points

In terms of biomass, *T. ciliatum* was the most abundant seagrass in the guts of turtles at both Farquhar (92%) and Cosmoledo (76%), followed by *T. hemprichii* which was relatively more abundant in guts from Cosmoledo (23%) than Farquhar (8%) (Fig. 3d).

**Foraging habitat assessment at Farquhar Atoll**

Capture locations of 27 turtles sampled at Farquhar in 1982–1983 are shown by red circles in Fig. 1c. For safety, fishermen rarely operated outside the atoll rim; so, most capture locations are concentrated along the western perimeter of the atoll inside the lagoon, especially near the shallow entrance to the lagoon where turtles (but not boats) can pass at high tide. The daytime locations of the green turtle equipped with a Fastloc-GPS Argos satellite tag are indicated by black circles ( $n = 99$ ) in Fig. 1c. The majority of these locations overlap and are close to the atoll rim ( $n = 93$ ); whilst only a few locations are at a mean distance of 1.0 km from the atoll rim ( $SE = 6.44$ , range = 0.49–2.58 km;  $n = 6$ ). These locations span a range of depths both inside and outside the lagoon. Sites outside the atoll rim, surveyed in



**Fig. 3** Comparison of green turtle diets by sex and female breeding status [breeding females (yellow;  $n = 17$ ); non-breeding females (red;  $n = 2$ ); males (blue;  $n = 26$ )] and site [Cosmoledo (black;  $n = 12$ ); Farquhar (white;  $n = 34$ )]. Relative contribution of eight dietary component groups by **a** sex and female breeding status and by **c** site. Relative contribution of three seagrass species [*Thalassodendron ciliatum* (*Tc*), *Thalassia hemprichii* (*Th*) and *Halodule uninervis* (*Hu*)]

by **b** sex and female breeding status and by **d** site. Mean values are shown with positive standard error bars. Dietary component groups: seagrasses (Sg), macroalgae (Ma), sponges (Sp), bryozoans (Br), cnidaria (Cn), invertebrates (In), substrate (Su) and debris (De). *Halophila stipulacea* not included, as weight of the fragment ( $n = 1$ ) was negligible

March 2016, were relatively deep (mean = 36.6 m; SE = 7.5; range = 8.8–90.5 m;  $n = 14$ ) and, in most cases, the seabed was not visible through the glass-bottomed bucket. Sites within the lagoon, surveyed in March 2018, were generally shallow (mean = 2.7 m; SE = 0.2;  $n = 15$ ). Seagrass was present at 100% of those sites with a high mean seagrass cover (76.9%; SE = 3.9;  $n = 13$ ). *T. ciliatum* was the only seagrass species recorded, with a mean canopy height of 25.8 cm (SE = 1.3). Macroalgae were recorded at only four sites (mean cover = 0.7%; SE = 0.5;  $n = 14$ ). Substrate colonised by seagrass included rocks, rubble, dead coral, sponge and sand. Seagrass communities observed elsewhere around the Farquhar lagoon were also dominated by *T. ciliatum*, but included small patches of *T. hemprichii*, which in most cases appeared to have been cropped by turtles.

## Discussion

At sites in the Republic of Seychelles, we report an almost exclusive diet of seagrass for green turtles. Seagrass biomass represented 95% of the diet of the adults sampled in our study ( $n = 28$ ), excluding the gravid females ( $n = 17$ ). This figure is amongst the highest reported in the world (Esteban et al. 2019 *Mar Biol* in review). All countries in the Western Indian Ocean host extensive seagrass meadows (Gullström et al. 2002); so, it is likely that seagrass is an important diet component for green turtles throughout this largely understudied region. Other comparable sites in the Western Indian Ocean, where seagrass exceeds 90% of the diet, include the United Arab Emirates (Hasbún et al. 2000), Yemen (Hirth et al. 1973) and Aldabra atoll in Seychelles (Frazier 1971). A high prevalence of seagrass has also been reported in Oman (Ross 1985). These findings have been supported by recent studies in Mayotte, Western Indian Ocean where green turtles graze on an assemblage of eight seagrass species commonly found there (Ballorain et al. 2010).

The most abundant seagrass species recorded in the guts of the Seychelles turtles at Cosmoledo, Farquhar and Desroches atolls were *T. ciliatum* and *T. hemprichii*, which is not surprising as these were the dominant seagrass species recorded at those sites by Kalugina-Gutnik et al. (1992). *T. ciliatum* is particularly abundant in the outer islands of Seychelles, forming extensive monospecific meadows, especially in deeper localities on sandy bottoms, coral reefs and sand-covered rocks (Short et al. 2010). Early studies of green turtles at Aldabra atoll in Seychelles recorded *T. ciliatum* as the primary diet component along with small amounts of *T. hemprichii* and macroalgae (Frazier 1971). The importance of different seagrass species for green turtle diet in Farquhar atoll suggests a relative preference for *T. hemprichii* (over *T. ciliatum*) given its scarcity in seagrass habitats (current study). Although *T. ciliatum* formed the

bulk of the turtle diet, anecdotal accounts from Seychellois turtle hunters (Mortimer, pers comm) indicate that turtles prefer other seagrass species if they are available. Neutral Detergent Fibre (NDF) is a measurement of most structural components in plant cells and it is possible that NDF in *T. ciliatum* is higher than other seagrasses, indicating increased difficulty for digestion, although data for *T. ciliatum* are not available (Trevathan-Tackett et al. 2017). Elsewhere in the Western Indian Ocean, the importance of *T. ciliatum* to the diet is variable: at Mayotte it does not feature prominently in the diet (Ballorain et al. 2010); while green turtles certainly forage on *T. ciliatum* in the monospecific seagrass meadows in the Great Chagos Bank (Esteban et al. 2018).

It is noteworthy that some daytime locations for tracked turtles were deep, being up to 90 m. The foraging site in Farquhar atoll is in the vicinity of a very steep drop-off (< 1 km between atoll rim and 500 m depth; Stoddart and Poore 1970) which is a relatively small area for a proficient swimmer with daytime home ranges of 19 km<sup>2</sup> (95% KUD, SD = 15.78,  $n = 6$ ) in the Seychelles (Christiansen et al. 2017). Likewise, we have shown previously that turtles forage in relatively deep areas on the Great Chagos Bank (29 m) where in situ surveys with divers and drop-down cameras have confirmed extensive seagrass beds (Esteban et al. 2018). Taken together, these findings add to the growing evidence that, in the Indian Ocean, seagrass may be found much deeper than previously assumed. In support of this conclusion, Kalugina-Gutnik et al. (1992) used SCUBA to survey distribution of seagrass and macroalgae in the Seychelles, and reported seagrass growing to depths of 37 m; and, on the Saya de Malha Bank, the EAF-Nansen Leg 2.1 (May 2018) expedition also recorded *T. ciliatum* at 37 m and *Halophila decipiens* at 70 m (Sundy Ramah, pers. comm.). We report that foraging sites of one satellite tracked individual in areas of extensive nearly monospecific beds of *T. ciliatum* in 2016 coincided with capture locations of > 75% of turtles captured in 1983, indicating long-term health of seagrass habitat in Farquhar atoll but possibly underrepresenting the overall foraging habitat size and distribution around the atoll.

Our regional case study presents evidence that the diet of gravid females differs from that of adult males (breeding and non-breeding) and non-breeding females. We present the first findings that compare the diets of breeding males and gravid female green turtles at sites where abundant benthic forage occurs adjacent to nesting beaches. Previous studies that recorded relatively empty guts and low-quality diets of gravid female green turtles (Carr et al. 1974; Mortimer 1982; Tucker and Read 2001) did not confirm whether reproductive status or forage availability caused the females to fast. At both Cosmoledo and Farquhar, the food present in the gut of gravid females weighed significantly less and was also more variable than that of both adult males and



non-breeding females. Due to the low sample size of non-breeding females, this observation clearly deserves further investigation.

We conclude that the restricted and unusual diets of gravid female green turtles are not driven by forage availability but more likely by a combination of other factors. During the inter-nesting period, green turtles often exhibit restricted movements between clutch depositions (Esteban et al. 2017). It may be that resting and conserving energy in locations inaccessible to the unwanted attentions of courting males and predators while the next egg clutch matures may be more important than foraging. It may also be that the maturing clutches of eggs take up so much volume inside a turtle that there is no room for a full gut. Clutch size scales with body size in sea turtles, suggesting that sea turtles lay the largest clutch of eggs they can fit inside their bodies (Hays and Speakman 1991). Another possibility is that gravid females have peculiar dietary needs. The guts of males and non-breeding females foraging on the seagrass flats of Nicaragua (Mortimer 1981) and those of adult males and non-breeding females in the present study contained little or no calcareous matter. In contrast, relatively large amounts of calcareous material (including the calcareous algae (*Halimeda spp.*), and/or calcareous substrates) were ingested by gravid females in the present study and at Raine Island (Tucker and Read 2001), by gravid green turtles migrating to the Tortuguero nesting grounds (Mortimer 1981, 1982), by gravid hawksbill turtles (*Eretmochelys imbricata*) in the Caribbean (Meylan 1984) and by gravid desert tortoises (*Gopherus agassizii*) (Marlow and Tollestrup 1982). Tucker and Read (2001) hypothesised that ingested calcareous material might supply calcium for eggshells, restore depleted calcium reserves in breeding females, or neutralise stomach acid in fasting animals. This explanation is reasonable given that gravid female green turtles may lay up to ten egg clutches at two-week intervals during a single nesting season (Esteban et al. 2017) and the eggs of each clutch need to be shelled on site just prior to being laid. Given the upward trends in green turtle populations in the Western Indian Ocean, there is a need to conserve and monitor seagrass habitats in the context of changing green turtle population densities. Green turtle foraging locations can be indicators of healthy ecosystems that warrant inclusion in MPA proposals (Scott et al. 2012). The Republic of Seychelles is currently developing a comprehensive Marine Spatial Plan (Government of Seychelles 2017) and has committed protection for up to 30% of its territorial waters to ensure representative species and habitats have long-term protection to improve resilience of coastal ecosystems in a changing climate. Cosmoledo, Farquhar and Desroches atolls all feature prominently in this zoning plan. In the samples we analysed from the Seychelles, it is noteworthy that while the proportion of seagrass in the diet varied across

individuals, all individuals had consumed at least some seagrass. This observation adds support for the use of green turtles as indicators of seagrass habitats, reiterating the value of remotely tracking green turtles to help identify hitherto unknown seagrass beds (Hays et al. 2018). Finally, regional satellite tracking studies (Hays et al. 2010, 2014; Bourjea et al. 2015; Christiansen et al. 2017) combined with green turtle foraging ecology studies help resource managers to identify critical habitats in need of long-term protection and enable a better understanding of foraging hotspots and regional connectivity.

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**Author contributions** NE, JAM and GCH conceived the study. JAM led all fieldwork in Seychelles. GCH and NE carried out the satellite tracking study in 2015–16. HJS carried out all laboratory analysis with input from NE, RFKU and JAM. HJS led the data analysis with contributions from all authors. HJS, JAM and NE led the writing of the manuscript with contributions from all authors.

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## Compliance with ethical standards

**Conflict of interest** The authors declare that they have no conflict of interest.

**Ethical approval** In the Seychelles, no turtles were killed specifically to provide samples for this study. Permits for attachment of transmitters to green turtles in the Chagos Archipelago were issued by the Commissioner for the British Indian Ocean Territory (BIOT). Fieldwork was

approved by the Swansea University Ethics Committee and the BIOT Scientific Advisory Group (SAG) of the FCO.

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