

Versatile synthesis of probes for high-throughput enzyme activity screening

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Abstract Mass spectrometry based technologies are promising as generalizable high-throughput assays for enzymatic activity. In one such technology, a specialized enzyme substrate probe is presented to a biological mixture potentially exhibiting enzymatic activity, followed by an in situ enrichment step using fluororous interactions and nanostructure-initiator mass spectrometry. This technology, known as *Nimzyme*, shows great potential but is limited by the need to synthesize custom substrate analogs. We describe a

synthetic route that simplifies the production of these probes by fashioning their perfluorinated invariant portion as an alkylating agent. This way, a wide variety of compounds can be effectively transformed into enzyme activity probes. As a proof of principle, a chloramphenicol analog synthesized according to this methodology was used to detect chloramphenicol acetyltransferase activity in cell lysate. This verifies the validity of the synthetic strategy employed and constitutes the first reported application of *Nimzyme* to a non-carbohydrate-active enzyme. The simplified synthetic approach presented here may help advance the application of mass spectrometry to high-throughput enzyme activity determination.

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Introduction

Given the biochemical and biotechnological significance of enzymes, high-throughput assays for the detection of enzymatic activity are in high demand. High-throughput enzyme activity assays are critical in the drug and biomarker discovery processes and can be a valuable tool for functional gene annotation. They figure prominently in directed evolution experiments, where libraries of enzyme variants are screened for improved substrate specificity, thermal stability, or other beneficial properties.

High-throughput enzyme activity assays can help alleviate experimental bottlenecks, but few generally applicable technologies are currently available. Some enzymatic reactions can be coupled to a spectroscopic readout by employing chromogenic substrates or by indirectly measuring product formation via a coupled assay or biosensor. Such assays are fast, but applicable to only the narrow range

of biochemical transformations for which methods have been developed. On the other hand, mass spectrometry-based assays are more universal, but require lengthy chromatographic separations to avoid spectral complexity and ionization suppression, drastically reducing experimental throughput (Fig. 1a).

Nimzyme is a nanostructure-initiator mass spectrometry (NIMS)-based analytical technique that can detect enzymatic activity in complex mixtures such as crude cell lysate [1–3]. It circumvents time-intensive chromatographic separations by means of an in situ fluorophilic affinity purification (Fig. 1b). This, in combination with acoustic sample deposition [4], shows potential as a high-throughput enzyme activity assay.

Currently, two hurdles limit the applications of *Nimzyme*. First, the technique requires the chemical synthesis of a specialized analog for each substrate of interest [5]. Second, all reported applications of *Nimzyme* thus far have been on enzymes acting on carbohydrates [1–4]. Hence, the generalizability of the technique has yet to be demonstrated.

Here, we report a synthetic route towards *Nimzyme* probes in which the substrate moiety is introduced towards the end of the synthesis. Accordingly, the invariant portion of these compounds needs to be synthesized only once. By fashioning this fragment as an alkylating agent, a variety of synthetically and biologically significant molecules can be readily transformed into *Nimzyme* probes. We also show that a chloramphenicol analog synthesized according to this methodology is effective in detecting chloramphenicol acetyltransferase (CAT) activity in crude cell lysate.

Experimental

General synthetic remarks and materials

Moisture-free conditions were employed only where indicated. Reagents and solvents were purchased from TCI America, ChemPep, Alfa Aesar, BDH, and Sigma-Aldrich,

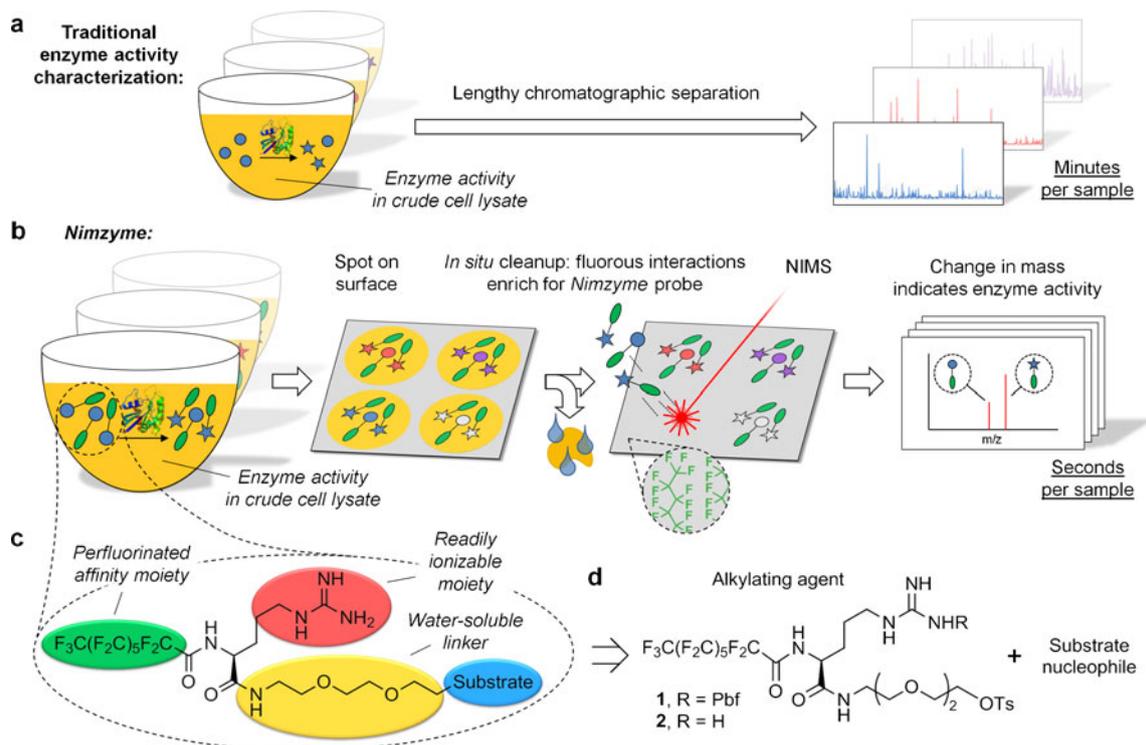


Fig. 1 Overview of the *Nimzyme* technology. **a** Performing protein purification on a large number of samples is usually prohibitive. Hence, high-throughput activity assays are typically performed in whole cells or in cell lysate, which requires separation by chromatography before mass spectrometric analysis. **b** Workflow of the high-throughput *Nimzyme* enzyme activity assay as described in ref. [2]. Enzyme substrate analogs harboring perfluoroalkyl moieties are incubated in a complex mixture exhibiting enzymatic activity (e.g., crude cell lysate). The mixture is quenched and spotted (e.g., using acoustic deposition) on a NIMS chip coated with a perfluorinated initiator substance. The surface is subjected to aqueous washes, allowing cell debris to be rinsed away while fluorophilic interactions with the chip

surface retain the analytes. Subsequent NIMS, performed on a MALDI-TOF mass spectrometer, generates mass spectra where the ratio of substrate to product mass speaks is a measure of enzyme activity. **c** Generalized anatomy of a *Nimzyme* substrate. Substrate analogs that can be employed in *Nimzyme* require a perfluorinated moiety to allow for in situ cleanup and an ionizable group to improve NIMS sensitivity. **d** The retrosynthetic strategy employed in this manuscript. The invariable part of these substrates is synthesized once, in the form of tosylate alkylating agents **1** and **2**. These can react with a wide variety of nucleophiles to provide *Nimzyme*-amenable substrate analogs. Key: *Ts* = *p*-toluenesulfonyl; *Pbf* = 2,2,4,6,7-pentamethyldihydrobenzofuran-5-sulfonyl

anhydrous whenever possible, and were used as received unless otherwise indicated. *FluoroFlash* fluorosolid-phase extraction cartridges were purchased from Fluorous Technologies Incorporated.

Design of alkylating agents

A generalized structure of Nimzyme-amenable substrate analogs as prepared in this manuscript is shown in Fig. 1c. A perfluoroalkanoyl moiety imparts the required fluorophilic character to allow for on-chip purification, and an arginine moiety ensures high ionization efficiency in NIMS [1]. Avoiding reliance on the substrate moiety to impart ionizability also allows for direct comparison of NIMS peak heights to determine relative abundance of chemical species. A tri(ethylene glycol) linker was installed to improve enzyme–substrate accessibility and enhance the probe's solubility in water.

Nucleophilic substitution chemistry was chosen to couple substrates of interest to the probe's invariant portion because many biologically relevant substrates possess nucleophilic functional groups. Hence, we incorporated a *p*-toluenesulfonate ester (tosylate) leaving group into the invariant portion of the Nimzyme probe, resulting in alkylating agents 1 and 2 (Fig. 1d).

Synthesis of alkylating agents

A synthetic route to 1 and 2 was devised, which starts from readily available starting materials. A synthetic scheme, detailed synthetic protocols, and spectral data of all intermediates are reported in the Electronic Supplementary Material (Scheme S1 and S2 and Figs. S1–S11). Briefly, 2-(2-(2-aminoethoxy)ethoxy)ethanol—obtained by means of a Gabriel synthesis from 2-(2-(2-chloroethoxy)ethoxy)ethanol—was coupled to Fmoc-Arg(Pbf)-OH using conventional solution-phase peptide synthesis methodology. The Fmoc group was removed with diethylamine and the resulting amine was acylated with perfluorooctanoyl chloride under Schotten–Baumann conditions. The resulting alcohol was tosylated to afford 1 in 26 % yield with respect to Fmoc-Arg(Pbf)-OH (four steps). Lastly, the Pbf protecting group was removed with 90:10 TFA/MeOH to afford 2.

Alkylation reactions

The alkylation and deprotection reactions were monitored by NIMS of 0.1 μ L of the reaction mixture dissolved in 10 μ L methanol. Fluorous solid-phase extraction (F-SPE) was performed on *FluoroFlash* 2 g cartridges as follows: The cartridges were preconditioned with 1 mL *N,N*-dimethylformamide (DMF) and 7 mL 80:20 MeOH/H₂O, loaded with the crude alkylation reaction mixture, washed with 7 mL 80:20 MeOH/H₂O, and eluted with

10 mL MeOH. Each F-SPE cartridge was used only once. Possible product isomerism was ruled out by LC/MS (see ESM). Because the quantities prepared here are too small to be accurately weighed, their yields were determined relative to a known concentration of Nz-OMe (12 in the ESM) by mixing them in a 1:1 ratio and determining the ratio of NIMS peak intensities. 12 was chosen as an internal standard because its mass does not overlap with any of the reagents, products, or possible side products and can reasonably be assumed to ionize similarly to 3 through 11. For amine nucleophiles (products 5, 6, and 11), using the hydrochloride form led to the formation of what was presumed to be “Nz(Pbf)-Cl” (NIMS calc'd for [M+H]⁺ 972.2; found 972.1; characteristic 3:1 M+H:M+H+2 ratio). Hence, we either purchased or generated the free base form of these substrates. Representative reactions using alkylating agents 1 and 2 are described in detail below. All others are described in the ESM; ‘Nz’=2-(2-(*N*^α-perfluorooctanoylargininamidoethoxy)ethoxy)ethyl (structural formula shown in Fig. 2a)

9—Ibuprofen Nz ester

Into a borosilicate test tube with stir bar, 5.7 mg ibuprofen sodium salt (25 μ mol, 50 eq), 10 μ L 50 mM Nz-OTs (2) in DMF, and 490 μ L DMF were added. The mixture was stirred at 70 °C for 3 h. After cooling to room temperature (RT), 5 mL 1 M aqueous NaOH was added, the resulting solution was extracted with 3 \times 5 mL chloroform, and each of the extracts was in turn washed with another 5 mL 1 M aqueous NaOH. A few crystals of NH₃Cl were added to the chloroform extracts, which were filtered and evaporated in vacuo to yield a colorless residue which was taken up in MeOH 78 % yield.

11—(1*R*,2*R*)-*N*-Nz-1-(4-nitrophenyl)propane-1,3-diol

Into a borosilicate test tube with stir bar, 50 μ L 100 mM Nz(Pbf)-OTs (1) in DMF, 200 μ L more DMF, and 53.1 mg (250 μ mol, 50 eq) (1*R*,2*R*)-2-amino-1-(4-nitrophenyl)-1,3-propanediol were added. The mixture was stirred at 110 °C for 2 h, cooled to RT, and F-SPE purified. The eluent was evaporated in vacuo, redissolved in 2 mL 90:10 TFA/MeOH, and left to stir at RT for 12 h. The TFA/MeOH was evaporated under a gentle stream of nitrogen, leaving a white residue which was taken up in MeOH 45 % yield.

NIMS surface fabrication

The production of NIMS chips has been described elsewhere [6]. Briefly, a silicon wafer is cleaned thoroughly with methanol, followed by anodic etching with 25 % hydrofluoric acid (*w/v*) in ethanol in a custom-made Teflon etching chamber using a current of 2.4 A for 15 min. Next, the chips are coated by

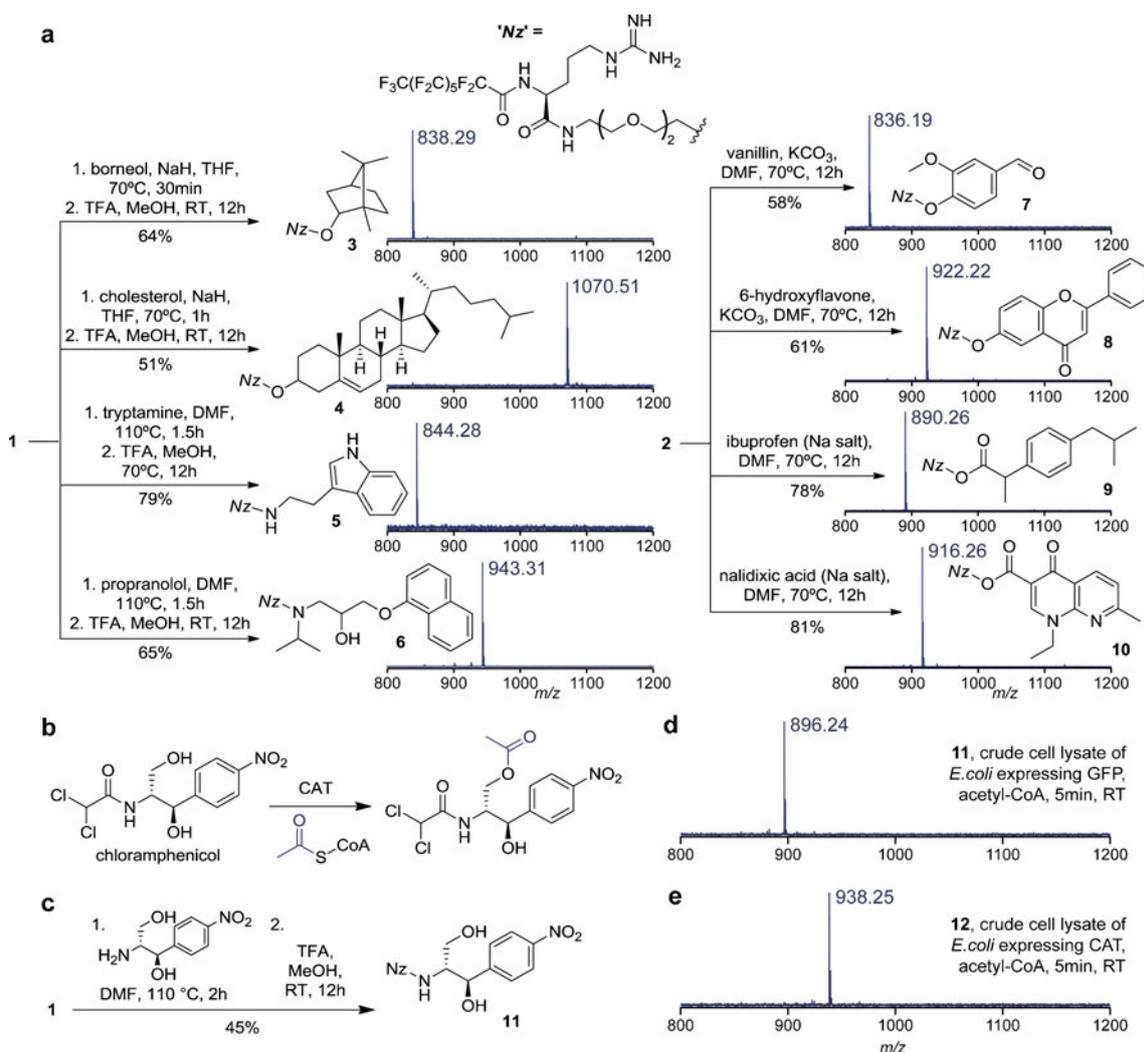


Fig. 2 Convenient synthesis of Nimzyme probes and their suitability for the detection of CAT activity in cell lysate. **a** Synthesis of Nimzyme substrates using tosylates **1** and **2** as alkylating agents, and NIMS spectra of the products purified only by F-SPE (for **1**), or liquid–liquid extraction (for **2**). Yields reported are combined yields for alkylation, purification, and (for **1**) deprotection. **b** CAT catalyzes the *O*³-acetylation of chloramphenicol. **c** Chloramphenicol analog **11** was synthesized through the alkylation of (1*R*,2*R*)-2-amino-1-(4-nitrophenyl)-1,3-

propanediol with **1** according to the methodology shown. **d** Exposure of **11** to control lysate followed by the Nimzyme workflow shown in Fig. 1b shows a clean mass peak corresponding to **11** (expected *M*+*H*, *m/z*=896.26). **e** Exposure of **11** to lysate of *E. coli* having expressed CAT, followed by Nimzyme, shows a mass shift of exactly one acetyl unit relative to **11** (expected *M*+*H*, *m/z*=938.26). Key: THF = tetrahydrofuran; TFA = trifluoroacetic acid; DMF = *N,N'*-dimethylformamide

adding the perfluorinated initiator liquid bis(heptadecafluoro-1,1,2,2-tetrahydrodecyl)tetramethyl-disiloxane for 20 min. Excess initiator is blown off with nitrogen.

Enzymatic activity assay

Overnight cultures of *Escherichia coli* DH1 harboring either pBbB8k-GFP or pBbB8k-CAT (the construction of these is described in the ESM) were diluted 1:10 in LB with 50 μg mL⁻¹ kanamycin, and grown for 1 h at 37 °C, after which they were induced with 0.2 % (*w/v*) arabinose. After 2 h of growth, 0.5 mL of the culture was centrifuged at ~12,000×*g* for 1 min, the pellet was resuspended in 0.25 mL aqueous 50 mM

sodium phosphate (pH 7.5), and sonicated for 30 s. Acetyl-CoA was added to a final concentration of 2 mM and **11** to a final concentration of 0.5 mM. The mixture was mixed and incubated at room temperature for 5 min and subsequently quenched with an equal volume of methanol. A 0.3-μL droplet of the mixture was spotted onto a NIMS chip and the excess liquid removed 5 s later by touching it with a Kimwipe (Kimberly-Clark). The spotted area was washed with 2 × 1 μL deionized water. The NIMS chip was taped to a modified standard matrix-assisted laser desorption/ionization (MALDI) plate, which was then loaded into an Applied Biosystems 4800 MALDI time of flight (TOF)/TOF mass spectrometer. Agilent ESI-L Low Concentration Tuning Mix

was spotted nearby on the NIMS chip to allow for mass calibration of the instrument. Spectra were acquired in manual mode and positive polarity.

Results

Alkylation reactions

We investigated the capability of 1 and 2 to react with a number of biologically relevant nucleophiles (Fig. 2a). It was found that 1 was able to alkylate alcohols by means of a Williamson ether synthesis, forming Nimzyme probes 3 and 4. Amines were likewise readily alkylated by 1 to form 5 and 6, provided the free base form was used. We used excess nucleophile to drive the reactions and to avoid over-alkylation of amines. The alkylated intermediates could be recovered using F-SPE, circumventing cumbersome chromatographic purifications. Subsequent deprotection of the Pbf group affords the desired Nimzyme probes in good yield. Alkylating agent 2 was found to react readily with carboxylate salts and phenolates to directly form Nimzyme probes 7–10. We were unable to find conditions under which these could be purified by F-SPE, and hence, we chose to remove excess nucleophile using basic aqueous washes.

Detection of chloramphenicol acetyltransferase activity

To verify that the substrate analogs synthesized according to the described methodology could be used to detect enzymatic activity in a Nimzyme assay, we synthesized chloramphenicol analog 11 as a probe for CAT activity (Fig. 2b, c). CAT catalyzes the transfer of an acetyl group from acetyl-CoA to O^3 of chloramphenicol [7]. When 11 is exposed to a control *E. coli* cell lysate, Nimzyme reveals a clean mass peak corresponding to this substrate (Fig. 2d). When instead 11 is exposed to lysate from *E. coli* overexpressing CAT, the peak shifts by 42 mass units, as expected from monoacetylation (Fig. 2e). This result verifies the validity of the synthetic strategy employed and furthermore comprises the first reported application of Nimzyme to a non-carbohydrate-active enzyme.

Discussion

In summary, we present the synthesis of a pair of alkylating agents and conditions under which they can be used to transform a wide range of structures into Nimzyme probes. The alkylation reactions and subsequent purifications are undemanding, such that an inexperienced chemist can perform them with minimal effort. One probe synthesized according to this methodology, a chloramphenicol analog, could serve to detect CAT activity in cell lysate, verifying the synthetic

approach and broadening the scope of the Nimzyme technology. The other synthesized probes were not tested in enzymatic reactions in this work.

The strategy described here allows for the construction of libraries of Nimzyme substrates without the need to repeatedly re-synthesize the invariable portion of the probes. By streamlining access to compatible enzyme substrates, and by showing that Nimzyme can be applied to non-carbohydrate-active enzymes, we hope to have cleared two of the barriers to the widespread adoption of this technology for high-throughput enzyme characterization. Certain drawbacks remain to be solved, such as the possibility that the modifications required of substrates to be compatible with Nimzyme affect enzyme–substrate binding interactions. While this has been shown not to be the case for β -glycosidases [2], enzymes with sufficiently buried active sites may or may not accept the probes presented here as substrates. Work to overcome these shortcomings is currently ongoing.

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References

1. Northen TR, Lee JC, Hoang L, Raymond J, Hwang DR, Yannone SM, Wong CH, Siuzdak G (2008) A nanostructure-initiator mass spectrometry-based enzyme activity assay. *Proc Natl Acad Sci USA* 105:3678–3683
2. Reindl W, Deng K, Gladden JM, Cheng G, Wong A, Singer SW, Singh S, Lee JC, Yao CH, Hazen TC, Singh AK, Simmons BA, Adams PD, Northen TR (2011) Colloid-based multiplexed screening for plant biomass-degrading glycoside hydrolase activities in microbial communities. *Energy Environ Sci* 4:2884–2893
3. Deng K, George KW, Reindl W, Keasling JD, Adams PD, Lee TS, Singh AK, Northen TR (2012) Encoding substrates with mass tags to resolve stereospecific reactions using Nimzyme. *Rapid Commun Mass Spectrom* 26:611–615
4. Greving M, Cheng X, Reindl W, Bowen B, Deng K, Louie K, Nyman M, Cohen J, Singh A, Simmons B, Adams P, Siuzdak G, Northen T (2012) Acoustic deposition with NIMS as a high-throughput enzyme activity assay. *Anal Bioanal Chem* 403:707–711
5. Fernandez-Arrojo L, Guazzaroni ME, Lopez-Cortes N, Beloqui A, Ferrer M (2010) Metagenomic era for biocatalyst identification. *Curr Opin Biotechnol* 21:725–733
6. Woo HK, Northen TR, Yanes O, Siuzdak G (2008) Nanostructure-initiator mass spectrometry: a protocol for preparing and applying NIMS surfaces for high-sensitivity mass analysis. *Nat Protoc* 3:1341–1349
7. Shaw WV (1984) Bacterial resistance to chloramphenicol. *Br Med Bull* 40:36–41