

Carbon nanoparticles in lateral flow methods to detect genes encoding virulence factors of Shiga toxin-producing *Escherichia coli*

P. Noguera · G. A. Posthuma-Trumpie · M. van Tuil ·
F. J. van der Wal · A. de Boer · A. P. H. A. Moers ·
A. van Amerongen

Received: 7 July 2010 / Revised: 7 October 2010 / Accepted: 12 October 2010 / Published online: 28 October 2010
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Abstract The use of carbon nanoparticles is shown for the detection and identification of different Shiga toxin-producing *Escherichia coli* virulence factors (*vt1*, *vt2*, *eae* and *ehxA*) and a 16S control (specific for *E. coli*) based on the use of lateral flow strips (nucleic acid lateral flow immunoassay, NALFIA). Prior to the detection with NALFIA, a rapid amplification method with tagged primers was applied. In the evaluation of the optimised NALFIA strips, no cross-reactivity was found for any of the antibodies used. The limit of detection was higher than for quantitative PCR (q-PCR), in most cases between 10^4 and 10^5 colony forming units/mL or 0.1–0.9 ng/ μ L DNA. NALFIA strips were applied to 48 isolates from cattle faeces, and results were compared to those achieved by q-PCR. *E. coli* virulence factors identified by NALFIA were

in very good agreement with those observed in q-PCR, showing in most cases sensitivity and specificity values of 1.0 and an almost perfect agreement between both methods (kappa coefficient larger than 0.9). The results demonstrate that the screening method developed is reliable, cost-effective and user-friendly, and that the procedure is fast as the total time required is <1 h, which includes amplification.

Keywords *E. coli* · Verotoxin · Shiga toxin · Screening method · Nucleic acid lateral flow immunoassay · Carbon nanoparticles

Introduction

Detection of pathogenic bacteria is key to the prevention and identification of problems related to health and safety. In spite of the real need for obtaining analytical results in the shortest time possible, traditional and standard bacterial detection methods, even if of sufficient sensitivity, are often expensive and too slow to be of any use; some may take up to 8 days to yield an answer [1, 2]. For this reason, over the recent years, a lot of effort has gone into the development of rapid biosensors of diverse nature. However, their performance is variable and still needs improvement [2]. Most of the reported methods for the detection of pathogenic bacteria are applied to *Escherichia coli* [1, 2].

There are many *E. coli* strains of which quite a number are part of the normal microbiota in the gut. Although most of them are harmless, some of them can cause severe illnesses; one of those is the Shiga toxin-producing *E. coli* (STEC, also called verotoxigenic *E. coli* or enterohemorrhagic *E. coli*) [3]. STEC has emerged as a food poisoning

Electronic supplementary material The online version of this article (doi:10.1007/s00216-010-4334-z) contains supplementary material, which is available to authorized users.

P. Noguera · G. A. Posthuma-Trumpie · M. van Tuil ·
A. P. H. A. Moers · A. van Amerongen (✉)
Wageningen UR Food & Biobased Research,
Biomolecular Sensing & Diagnostics,
P.O. Box 17, 6700 AA Wageningen, the Netherlands
e-mail: aart.vanamerongen@wur.nl

P. Noguera
Instituto de Reconocimiento Molecular y Desarrollo Tecnológico,
Departamento de Química, Universidad Politécnica de Valencia,
Camino de Vera, s/n,
46020 Valencia, Spain

F. J. van der Wal · A. de Boer
Wageningen UR Central Veterinary Institute,
Division Bacteriology and TSEs,
P.O. Box 65, 8200 AB Lelystad, the Netherlands

pathogen that can cause severe and potentially fatal illness like diarrhoea, hemorrhagic colitis or haemolytic uraemic syndrome. The most important reservoirs of STEC, and, consequently, causes of human infections, are domestic ruminants, especially cattle and sheep. Since STEC strains are found as part of the normal intestinal flora of these animals, transmission occurs through, e.g., consumption of undercooked meat, unpasteurised dairy products and vegetables or water contaminated by faeces of carriers [4, 5]. Even though many STEC serotypes have been associated with human illness, the vast majority of reported outbreaks and sporadic cases of STEC infection in humans have been associated with serotype O157:H7 [5]. Some of the STEC virulence factors are: cytotoxins (verotoxins) encoded by the genes *vt1* or *vt2*; the outer membrane protein intimin encoded by the gene *eae* and involved in the attachment to intestine epithelial cells; and the enterohaemolysin encoded by the gene *ehxA*.

Monitoring and evaluation of *E. coli* virulence factors can be done by various methods that include laboratory culture, typing methods, serological methods, detection of DNA sequences and immunochemical methods [2, 5, 6]. The need for timely results has increased the interest in rapid screening methods. This interest is also growing due to the shift of pathogen testing from laboratories (governmental, reference, etc.) to on-site settings in, e.g. food-processing plants [1]. Most *E. coli*-specific methods have a detection limit between 10^3 and 10^5 cells/mL and some rely on the amplification of specific genes of the *E. coli* genome for specific identification [7, 8]. There are biosensors in the literature that can detect a lower amount of cells [2, 6], but in general, these systems have a moderate sensor-to-sensor reproducibility and poor batch-to-batch reproducibility [9], whilst others do not comply with the basic requirements for application in this field, i.e. a low price and ease of use [7]. Moreover, many methods do not distinguish between *E. coli* virulence factors/strains.

The use of nucleic acid lateral flow immunoassays (NALFIA) as biosensors for the detection of specific amplified nucleic acid genes is not new [10]. In NALFIA, nucleic acids can be captured on the lateral flow test strips either in an antibody-independent or antibody-dependent manner [11, 12]. In the first one, a hybridization step is required upon running the sample through the membrane [11–15]. The second and simpler approach uses immobilised antibodies to directly recognise specific tag-labelled amplicons [11, 12, 16, 17]. Some of the advantages of using lateral flow strips are: one-step assay, simple, fast, low cost, versatile and a prolonged shelf life (with no need for refrigeration) [11, 12, 18–20]. Moreover, compared with traditional electrophoresis detection, NALFIA has other advantages like shorter response time and no need for hazardous reagents [16, 17]. NALFIA strips comply with

most of the requirements for biosensors for bacterial detection: low detection limit, species selectivity, strain selectivity, assay time for a single test between 5 and 10 min, no reagent addition needed (except buffer), direct measurement, highly automated format, no operator skills needed to carry out the assay, and also compact size, portable, handheld, and designed for field application [7]. In addition, they comply with two other characteristics: low price and easy manufacture. Another interesting feature is that they are disposable; hence, regeneration time is eliminated. Finally, if NALFIA strips are used with coloured nanoparticles, a visual detection can be easily performed; in this sense, this combination overcomes one of the major shortcomings in the application of immunosensor techniques, which is the cost of the required equipment [8].

The aim of this work was to present the use of carbon nanoparticles in NALFIAs. Carbon nanoparticles were chosen for their high signal-to-noise ratio (black to a white background) [17, 21] and their excellent sensitivity, i.e. low picomolar by visual inspection [18]. NALFIA was applied to the rapid detection and identification of genes encoding various STEC virulence factors (*vt1*, *vt2*, *eae* and *ehxA*) and of the gene coding for *E. coli* 16S rRNA as a control. Genes were amplified with specific tag-labelled primers, and amplicons were sandwiched between neutravidin coated onto carbon nanoparticles and specific anti-tag antibodies immobilised on the NALFIA nitrocellulose membrane. The limit of detection of the NALFIA strips was determined, and the strips were tested with DNA from 48 field strains originating from cattle faeces. Performance of the NALFIA strips was evaluated by comparing the results with those achieved by quantitative PCR (q-PCR).

Material and methods

Reagents

Borate buffer (BB) 100 mM, pH 8.8, was prepared by mixing 100 mM solutions of H_3BO_3 (Merck, Darmstadt, Germany) and $Na_2B_4O_7 \cdot 10 H_2O$ (Sigma-Aldrich Chemie BV, Zwijndrecht, the Netherlands). BB (5 mM) was obtained diluting 100 mM BB appropriately. With these two buffers, bovine serum albumin (BSA, Sigma-Aldrich Chemie BV) and Tween 20 (Merck, Amsterdam, the Netherlands), several buffers were prepared: washing buffer (WB, 5 mM BB, 1% (w/v) BSA), storage buffer (SB, 100 mM BB, 1% (w/v) BSA) and running buffer (RB, 100 mM BB, 1% (w/v) BSA, 0.05% (v/v) Tween 20). NaN_3 was added to all buffers to a final concentration of 0.02% (w/v).

Polyclonal antibodies against digoxigenin (DIG, Roche, Penzberg, Germany), 2,4-dinitrophenol (DNP, Invitrogen Corporation, Camarillo, CA, USA), fluorescein isothiocya-

nate (FITC, AbD-Serotec, Oxford, UK) and Texas Red (TxR, Invitrogen Corporation) were used. A monoclonal antibody against Cy5 (Sigma-Aldrich Chemie BV) was used for the 16S primer anti-tag antibody. As control line, biotinylated anti-rabbit antibody (Pierce Biotechnology, Rockford, IL, USA) was used.

E. coli strains and DNA isolation

E. coli strains used as controls are listed in Table 1. These strains were selected for having different sets of genes coding for virulence factors. The strains were grown overnight aerobically at 37 °C on heart infusion agar (Difco Laboratories, Detroit, MI, USA) supplemented with 5% sheep blood (HIS plates). DNA was isolated by harvesting bacteria from a plate with one streak of an inoculation loop and subsequent resuspension in 300 µL 6% Chelex 100 (Bio-Rad Laboratories, Hercules, CA, USA). The suspension was heated at 56 °C for 15 min, then at 100 °C for 8 min, and centrifuged at 14,000×g for 5 min. Supernatants containing genomic DNA were transferred to a new vessel and stored at -20 °C until use.

To isolate DNA for quantitative purposes, EDL 933 was grown aerobically at 37 °C in 10 mL Bacto Heart Infusion Broth (Becton Dickinson, Sparks, MD, USA). A quantity of 1 mL containing 8.4×10^7 colony forming units (cfu) was taken from a culture in the exponential growth phase for DNA isolation using Chelex, resulting in a DNA preparation with 775 ng/µL.

Polymerase chain reaction

DNA was amplified prior to detection with NALFIA strips. Primers used in the amplification of the genes coding for four STEC virulence factors (*vt1*, *vt2*, *eae* and *ehxA*) and a 16S control specific for *E. coli* are indicated in Table 2. Forward primers were labelled with specific tags TxR (*vt1*), FITC (*vt2*), DIG (*eae*), DNP (*ehxA*) or Cy5 (16S), whereas all reverse primers were labelled with biotin (Eurogentec, Seraing, Belgium). To achieve an amplification reaction in

the shortest time possible with economical equipment, PCR reactions were carried out with Phire Hot Start DNA Polymerase (Finnzymes) and a PIKO Thermal cycler (Finnzymes, Espoo, Finland) using extra thin vessels that allow an increase in heat transfer speed. Since annealing temperature may be affected by the use of tags coupled to primers [22], amplification reactions were optimised with the final aim of using identical conditions for all genes.

The protocol applied had a total length of 30 min and consisted of an initial denaturation at 98 °C (30 s), followed by 30 cycles with a denaturation step at 98 °C (5 s), an annealing step at 61 °C (5 s) and an extension step at 72 °C (5 s), with a final extension step at 72 °C (1 min) followed by a lowering of temperature to 4 °C. The reaction tube contained 2 µL template and 0.4 µL polymerase (as provided in the kit) in a total volume of 20 µL, with final concentrations of 1.5 mM MgCl₂, 0.25 µM dNTPs and 0.9 µM of each primer (forward and reverse). Positive and negative controls were included in all runs. Amplified DNA was directly analysed with a NALFIA strip or stored at -20 °C until use.

Carbon conjugate

Carbon nanoparticles bind proteins non-covalently and, if optimal conditions have been applied, without changing their bioactivity [17, 21, 25, 26]. Therefore, carbon nanoparticles were labelled with neutravidin biotin-binding protein (Pierce Biotechnology) following O'Keefe et al. [27]. Briefly, the procedure to obtain the carbon conjugate was: 10 mg of carbon (Spezial Schwartz 4, Degussa AG, Frankfurt, Germany) was suspended in 1 mL of MilliQ water and sonicated for 5 min (Branson model 250 Sonifier, Danbury, CT, USA). The resulting 1% (w/v) carbon suspension was fivefold diluted in 5 mM BB and sonicated for 5 min. Next, 350 µg neutravidin was added to 1 mL of the diluted carbon suspension and stirred for 3 h at room temperature. This suspension was first centrifuged at 13,636×g for 15 min; the supernatant was removed and the pellet washed with WB to remove unbound protein. To achieve this, 1 mL of WB was added to the pellet and the mixture centrifuged at 13,636×g for 15 min. Subsequently, the supernatant was removed and the pellet was resuspended in 1 mL WB. This process was repeated twice. After the final wash, the pellet was resuspended in 1 mL SB and stored at 4 °C until use. The resulting homogeneous suspension contained 0.2% (w/v) carbon conjugate.

Nucleic acid lateral flow immunoassay

NALFIA strips were manufactured using nitrocellulose (NC) membranes cut to a width of 2.5 cm (HiFlow Plus HFB135, Millipore, Billerica, MA, USA). A Linomat IV

Table 1 Serotypes, sources and virulence factors of selected *E. coli* strains

Strain	Serotype	Source	<i>vt1</i>	<i>vt2</i>	<i>eae</i>	<i>ehxA</i>
EDL 933	O157:H7	Human	+	+	+	+
E32511	O157:H	Human	-	+	+	+
DVI 828	O49:Hnd	Calf	-	-	+	+
TB 154A	O103:H6	Human	+	-	+	+
B2F1	O91:H21	Human	-	+	-	+
Rdec-1	O15:NM	Rabbit	-	-	+	-

+/-: presence/absence

Table 2 Primer and probe sequences used for the amplification of different *E. coli* virulence factors

Primer or probe	Sequence	Amplicon size (bp)	Reference
vt1-F	5'-GGATAATTTGTTTGCAGTTGATGTC-3'	107	[23]
vt1-R	5'-CAAATCCTGTCACATATAAATTATTCGT-3'		
vt1-P	5'-CCGTAGATTATTAACCGCCCTTCCTCTGGA-3'	130	[23]
vt2-F	5'-GGGCAGTTATTTTGCTGTGGA-3'		
vt2-R	5'-GAAAGTATTTGTTGCCGTATTAACGA-3'		
vt2-P	5'-ATGTCTATCAGGCGCTTTTGACCATCTT-3'		
eae-F	5'-CATTGATCAGGATTTTCTGGTGATA-3'	102	[23]
eae-R	5'-CTCATGCGGAAATAGCCGTTA-3'		
eae-P	5'-ATAGTCTCGCCAGTATTCGCCACCAATACC-3'	142	[23]
ehxA-F	5'-CGTTAAGGAACAGGAGGTGTCAGTA-3'		
ehxA-R	5'-ATCATGTTTTCCGCCAATGAG-3'		
ehxA-P	5'-TCATAAGGAATTCCACCGTTTCTGAATTCA-3'	96	[24]
16S-F	5'-CATGCCGCGTGTATGAAGAA-3'		
16S-R	5'-CGGGTAACGTCAATGAGCAAA-3'		
16S-P	5'-TATTAACCTTACTCCCTTCCTCCCGCTGAA-3'		

TLC dispenser (Camag, Berlin, Germany) was used to dispense the specific antibody (test line) and the biotinylated antibody (control line) on the NC membrane at a dose of 1 $\mu\text{L}/5$ mm (diluted in 5 mM BB). Distance between lines was 5 mm. The NC membranes were dried overnight at 37 °C. Then, the NC membranes were fixed on a plastic backing along with a cellulose absorbent pad (Schleicher and Schuell, 's-Hertogenbosch, the Netherlands). Finally, strips were cut at a width of 5 mm using a Bio-Dot Cutter CM4000 (Irvine, CA, USA), used immediately, or packaged in aluminium pouches (with a silica desiccation pad), sealed and stored at room temperature until use.

Following PCR, amplicons were detected by means of NALFIA: The double-tagged amplicons were sandwiched between neutravidin (bound to carbon nanoparticles) and an antibody against the discriminating tag (immobilised on the NC membrane, capture ligand) (see Electronic supplementary material (ESM) Fig. S1). To achieve this, NALFIA assays were performed in a low-binding 96-well microplate. Once reagents were introduced in a well and mixed thoroughly, a strip was dipped vertically and reagents moved upwards through the strip by capillary force. Strips used for qualitative results (presence/absence of virulence factors) were visually examined after 10 min; a positive result (+) was recorded when a line was distinguishable from the background, and a negative result (–) when no line was seen. For quantitative results, strips were dried at 37 °C, and the pixel grey intensity of the test lines was evaluated following conventional flatbed scanning [28].

Immunoassays need to be optimised for each antibody with respect to performance and for the set of antibodies used. To accomplish this, several experiments were per-

formed to find the optimum amount of antibody sprayed on the NC membranes and the amount of PCR product to be used. For this, strips were manufactured with serial dilutions of each of the antibodies and tested with different amounts of PCR product. For one of the antibodies (specific to Cy5), the addition of 2% sucrose (Merck, Darmstadt, Germany) remarkably improved the line quality [29].

Cross-reactivities were studied by testing all amplified genes (using EDL 933 diluted 1/100 as template) on each of the specific NALFIA strips. With the aim of identifying several STEC genes simultaneously, a series of strips was prepared onto which all antibodies were printed. In this case, the width of the NC membrane used was 5 cm and the distance between lines 3 mm.

To evaluate the analytical performance of the NALFIA strips developed, DNA of *E. coli* strain EDL 933 (775 ng/ μL , corresponding with 8.4×10^7 cfu/mL) was subjected to tenfold serial dilutions and divided into six aliquots. Three aliquots were used in the optimised PCR amplification procedure, and for each aliquot, two NALFIA strips were used. For comparison, the remaining three aliquots were analysed using the reference method, q-PCR. For each factor, dried strips were scanned, data (in pixel grey volumes) were transformed to percentage of the maximum, and the resulting percentages were adjusted to a four-parameter logistic curve (Sigmaplot 11, Systat Software, Inc., San Jose, CA, USA). The limit of detection (LOD) was calculated as the signal of the blank plus three times the standard deviation of the blank (SD). For simplicity of interpretation, all quantitative results will be given in colony forming units per millilitre.

Quantitative PCR

To identify whether an isolate is an STEC, q-PCR can be used to detect the various virulence factors and was used as the reference method in this work. Reaction mixtures for q-PCR were assembled in 20- μ L quantities using the TaqMan Fast Universal PCR Master mix according to the manufacturer's instructions (Applied Biosystems, Nieuwerkerk a/d IJssel, the Netherlands) with 2- μ L template DNA. PCRs were performed using the same primers as used for PCR preceding NALFIA (see above; Table 2) and TaqMan probes labelled with the fluorescent dyes 6-FAM or VIC at the 5' end and a quencher at the 3' end, i.e. TAMRA for the *vt1*, *vt2*, *eae* and *ehxA* probes (Applied Biosystems) and BHQ for the 16S probe (Eurogentec, Table 2). The PCRs were performed on an Applied Biosystems 7500 Fast Real-Time PCR System in Fast 7500 run mode using the standard thermal cycler protocol. Positive and negative controls were included in all runs; the cutoff was set at a Ct of 32 based on results with negative controls. All PCRs were performed in duplex reactions (*vt1/vt2*, *eae/ehxA*) or in a single reaction (16S).

Field samples

To test field samples, DNA was obtained from 48 *E. coli* isolates originating from cattle faeces [30]. The isolated DNA was subjected (in duplicate) to the 30-min PCR amplification and subsequently to a NALFIA test strip. The result achieved for each single sample (i.e. the presence or absence of a virulence factor) was compared with the result achieved by q-PCR. When results were identical, it was considered as a true positive (or negative), and when they differed it was considered a false positive (or negative). With these values, several reliability parameters were calculated to evaluate the performance of the NALFIA strips [31–33]: sensitivity, specificity, positive (and negative) predictive values, and efficiency. At present, there is no agreement which of these previous parameters is the most useful one, and there are no guidelines to evaluate results achieved. For this reason, the kappa coefficient (*k*) was also calculated. This statistic coefficient takes into account the possibility of chance agreement, and Landis and Koch [34] provided a starting point to evaluate the strength of the agreement. The use of *k* does not exclude the use of other parameters since predictive values are interpretable as indices of test performance (how well a sample is classified), whereas *k* is interpretable as an index of test quality (how well a test makes use of evidence in classifying samples) [33].

The mathematical equations to calculate all these reliability parameters are given in ESM Table S1, and an Excel spreadsheet specifically developed for evaluating diagnostic tests was used [35].

Results and discussion

Nucleic acid lateral flow immunoassay

NALFIA strips were manufactured with serial dilutions of each of the antibodies and were used with different amounts of PCR product. Line intensity increased with increasing amounts of antibody and PCR product (until a maximum plateau value was reached). The selected final assay conditions were those that achieved good line intensity using the lowest amount of antibody (to make the strips as cost-effective as possible) and the lowest volume of PCR product. For convenience, the final PCR volume used was the same for all factors tested. Examples regarding the selection of optimal conditions are given in the ESM attached.

The selected optimal amount of antibody sprayed on the NC membrane was: 100 μ g/mL for TxR (*vt1*), DIG (*eae*) and biotinylated IgG (control); 125 μ g/mL for DNP (*ehxA*); 250 μ g/mL for FITC (*vt2*); and 300 μ g/mL (in 2% (w/v) sucrose) for Cy5 (16S). Strips were finally used in microplate wells that contained a mixture of 1 μ L of PCR product (sample), 1 μ L carbon–neutravidin conjugate and 98 μ L RB.

To verify that there was no cross-reactivity, optimised strips for the identification of each of the genes were used with all other amplicons, and no cross-reactivity was found. NALFIA strips were also tested with different *E. coli* strains (Table 1) and a blank (water). Once samples had been randomly identified by a number, they were submitted to PCR and their products applied to the optimised NALFIA strips. All genes coding for the virulence factors were correctly identified, allowing the identification of the six *E. coli* strains used (see ESM Table S2). These results were very promising with respect to testing real field samples.

When NALFIA strips were used with different *E. coli* dilutions (submitted to PCR amplification), maximum intensity values was around 10^5 pixel grey volumes. When data were transformed to percentage of the maximum signal, differences in the performance of NALFIA strips having different antibodies were noticeable (Fig. 1). For all systems, the regression values for the adjustment of these data to a four-parameter logistic curve were larger than 0.97, and the LOD increased in the following order: *vt2* < *ehxA* < 16S < *eae* < *vt1* (Table 3), being around 10^5 cfu/mL (0.9 ng/ μ L), which would correspond to 200 cfu in the PCR tube. The lowest LOD was achieved for *vt2* with a value of $6.7 \cdot 10^4$ cfu/mL (0.6 ng/ μ L). LODs of the NALFIA strips developed are similar to those achieved by other systems for the detection of *E. coli* [8], but in our study, virulence factors were specifically identified.

When comparing the LODs of NALFIA strips with those of q-PCR, the LOD achieved by q-PCR in most cases was

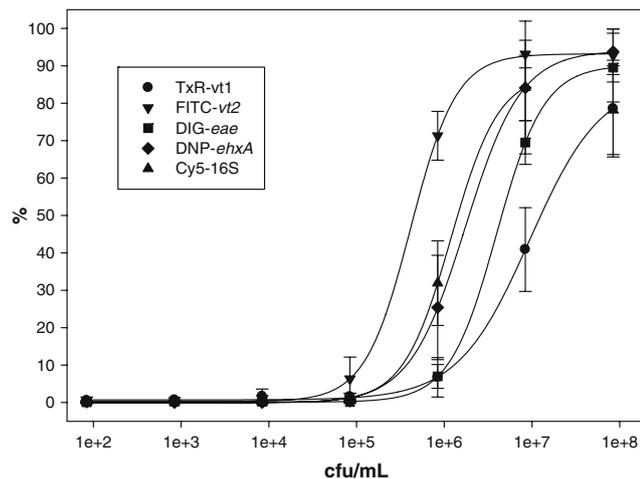


Fig. 1 Sensitivity of the various NALFIAs for each of the *E. coli* virulence factors tested. Average \pm SD ($n=6$)

(at least) two orders of magnitude lower than for NALFIA (Table 3). The sole exception is *vt2*; the LOD achieved in q-PCR (5.9×10^5 cfu/mL or 5.4 ng/ μ L) was higher than the one for NALFIA (6.7×10^4 cfu/mL or 0.6 ng/ μ L). As can be noted, the order of sensitivity achieved by q-PCR is almost contrary to the one achieved by NALFIA strips. These differences in LOD will explain the performance of the NALFIA strips with real samples (presence of false results; see “Field samples”).

Field samples

DNA of each of the 48 cattle faeces isolates was amplified by PCR (30 min) and the resulting amplicons applied to NALFIA tests. The qualitative result achieved for each sample (presence/absence) was compared to the result achieved by q-PCR (considered in this work as the reference method). The results achieved for each virulence factor tested are summarised in Table 4 (see ESM Table S1 for clarification). The presence of FP and FN results is justified by the different performance (LOD) of NALFIA and q-PCR for each of the amplified virulence factors. The three FN found for *vt1* and the one found for *ehxA* are due to the higher LOD of the NALFIA strips (compared to q-PCR); *E. coli* was present, but in a concentration that was not detected [36, 37]. On the other hand, the two FP achieved for *vt2* can be explained if the lower LOD for NALFIA is taken into account (one order of magnitude lower than for q-PCR); that is, in this particular sample, this factor was not detected by q-PCR.

Reliability parameters of the NALFIA strips developed are indicated in Table 5 (standard error values are given in ESM Table S3). For all factors tested, sensitivity and specificity values were very high (larger than 0.85, and in most cases equal to 1.00). In all cases, positive and negative

predictive values (i.e. how well a sample is classified) were larger than 0.90. Moreover, efficiency values (i.e. correct answers) were 0.92 or greater, meaning that more than 90% of the answers were the same as the reference procedure. The kappa coefficient was also calculated, giving values larger than 0.80 and in some cases equal to 1.00. According to Landis and Koch [34], this indicates that there is an almost perfect agreement between NALFIA and the standard used (q-PCR) for *vt1*, *vt2* and *ehxA*, and a perfect agreement for *eae* and 16S. This excellent agreement between the NALFIA strips developed and the reference system used demonstrates that the proposed analytical procedure has been proven to be fit for purpose, i.e. it is valuable for fast screening of *E. coli* virulence factors.

A preliminary multi-analyte NALFIA was prepared with five test lines (one for each of the specific antibodies: TxR, FITC, DIG, DNP and Cy5) and one control line (biotinylated anti-rabbit IgG) with the conditions indicated previously. These NALFIA strips were tested for cross-reactivities (Fig. 2) by running a blank sample (first strip), each of the STEC factors (second to sixth strip), and a sample with all factors (seventh strip). As previously noted, no cross-reactivity was found and all specific amplicons were correctly identified, showing the potential of NALFIA strips for the simultaneous detection of multiple virulence factors.

Conclusions

The performance of the NALFIA strips developed in this work (with a preceding 30-min PCR reaction step) has shown that this system is feasible and reliable and that it allows the specific identification of the four *E. coli* virulence genes (*vt1*, *vt2*, *eae* and *ehxA*) and a marker specific for *E. coli* in approximately 40 min (once DNA is isolated). Results achieved for the detection of *E. coli* virulence factors in real samples by NALFIA (following DNA isolation and PCR amplification) were in very good agreement with those achieved by q-PCR. Most discrepancies can be explained by the different LODs for the various genes in both methods. Nevertheless, NALFIA strips developed have very good sensitivity and specificity,

Table 3 Detection limit achieved by NALFIA and q-PCR for different *E. coli* virulence factors

	Detection limit (cfu/mL)				
	<i>vt1</i>	<i>vt2</i>	<i>eae</i>	<i>ehxA</i>	16S
NALFIA	4.1×10^5	6.7×10^4	3.7×10^5	1.4×10^5	1.8×10^5
q-PCR	8.4×10^2	5.9×10^5	8.4×10^3	8.4×10^3	8.4×10^1

Table 4 Comparison of qualitative results achieved in isolates from cattle faeces for different *E. coli* virulence factors

	Methods	Reference method (q-PCR)										
		<i>vt1</i>		<i>vt2</i>		<i>eae</i>		<i>ehxA</i>		16S		
		pos.	neg.	pos.	neg.	pos.	neg.	pos.	neg.	pos.	neg.	
<i>pos.</i> number of positive samples, <i>neg.</i> number of negative samples	NALFIA	pos.	17	1	30	2	23	0	31	0	47	0
		neg.	3	27	0	16	0	25	1	16	0	1

and results achieved by the strips are in very good agreement with the reference method used (q-PCR). When comparing both systems, q-PCR could have the advantage that (in most cases) the LOD is lower than NALFIA and that the identification is done in one step. However, it has several drawbacks: The first one is the price of the equipment, the second one is that specific knowledge is needed to understand and interpret the results, and the third one is the need of a laboratory setting. When comparing the number of factors that can be analysed, for q-PCR, only a limited number of fluorescent dyes are available, and multiplexing is complicated by the spectral overlap of the various dyes, allowing only a limited combination of dyes in one run (generally up to six). However, as verified in this work, NALFIA strips can also be manufactured with six lines (five test lines and one control line), having several advantages over more sophisticated instrumentation as listed above. Moreover, the same carbon–neutravidin nanoparticle conjugate is presently used to sandwich tag-labelled amplicons in antibody microarrays (results from Wageningen UR Food & Biobased Research, manuscript in preparation). In this format, the number of discrete targets can be easily increased by introducing other tag–antibody combinations. The results of this approach, a 30-min amplification followed by a one-step incubation for 1 h on the microarray, can be obtained by automated data processing following the digitisation of the spots by flatbed scanning or digital photography (10-min total processing time). Finally, if the price per test would be compared, NALFIA is less costly than q-PCR. The price for a duplex q-PCR test (e.g. virulence gene and 16S control) was calculated to be €1.16 (provided multiple samples are tested in the 96-well microtiter plate format). On the other hand,

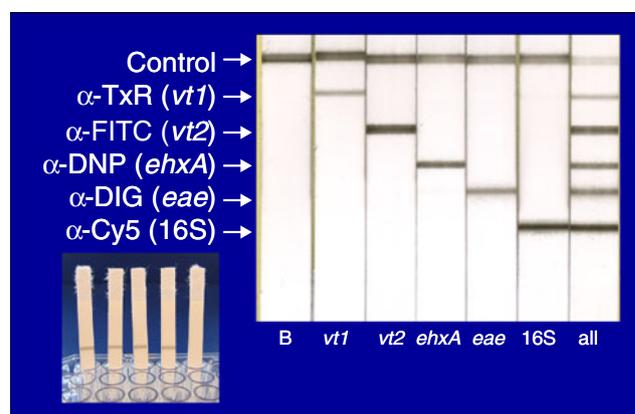
Table 5 Reliability parameters for different *E. coli* virulence factors

	<i>vt1</i>	<i>vt2</i>	<i>eae</i>	<i>ehxA</i>	16S
Sensitivity	0.85	1.00	1.00	0.97	1.00
Specificity	0.96	0.89	1.00	1.00	1.00
Positive predictive value	0.94	0.94	1.00	1.00	1.00
Negative predictive value	0.90	1.00	1.00	0.94	1.00
Efficiency	0.92	0.96	1.00	0.98	1.00
Kappa coefficient (<i>k</i>)	0.83	0.91	1.00	0.95	1.00

cost calculation for a two-line NALFIA (duplex PCR) resulted in €0.41. In conclusion, the NALFIA technology enables a larger multiplexing than q-PCR, is substantially less demanding with respect to laboratory facilities and has a lower price per test.

In this work, it has been demonstrated that the use of NALFIA strips in combination with a quick PCR amplification step comprises most of the requirements for alternative rapid methods [6]: accurate, with very good sensitivity and specificity, reliable, and with a detection limit that enables the correct discrimination of the different *E. coli* virulence factors. The test has also been validated against a standard test (q-PCR), its speed is high (<40 min, once the enrichment step is performed), and the cost of the whole system (strips, reagents) is low, requiring only one instrument (the thermocycler). Other characteristics that this system meets are that NALFIA strips are easily manufactured, have a long shelf life, can be stored under ambient conditions, and are very easy to use. In addition, whilst for q-PCR often small fragments are chosen (100–150 bp), the size of amplicon targets is not critical in NALFIA. This would allow using larger fragments if the variation in a particular region would require so.

Potentially, the developed NALFIA system can be used in the field, near the animal that has to be tested on (zoonotic) pathogens. In that case, transportation of infected samples to testing laboratories would not be necessary,

**Fig. 2** Results achieved with multi-analyte NALFIA for *E. coli* virulence factors. Samples: blank (first strip), each of the STEC factors (second to sixth strip), and all factors (seventh strip)

thereby reducing the risk of contamination outside the place of identification. The user friendliness of the procedure can be further improved by applying ready-to-use amplification reagents combined with isothermal amplification protocols and on-tissue amplification protocols that would make DNA isolation superfluous. No expensive apparatus is needed to perform the test procedure, and the number of specific amplicons that can be tested simultaneously is only dependent on the availability of tag–antibody combinations and on the possibility to develop a multiplex PCR protocol. Moreover, the detection system is broadly applicable since any molecular biological diagnostic question can be translated in primer sets having biotin on one of the primers and a discriminating tag on the other primer. A carbon conjugate with neutravidin (or streptavidin) will always recognise double-labelled amplicons having a biotin on one end of the double-strand chain, and specific anti-tag antibodies on the membrane will bind to their respective tags, thereby revealing the identity of the specific amplicons. Integration of the NALFIA technology in a field-applicable device with which amplification can be done, would further decrease hands-on time and simplify the overall detection procedure.

In conclusion, we have demonstrated that the use of carbon nanoparticles to detect amplified double-strand genetic material by NALFIA results in a screening method that is reliable, simple to use, cost-effective and fast; once DNA is isolated, results are achieved within 40 min, including the amplification step.

Acknowledgments This work was partially supported by the Generalitat Valenciana (BEST/2009/026), the Universidad Politécnica de Valencia (PAID-00-09-2837), and by the Dutch Ministry of Agriculture, Nature and Food Quality (KenniBasis 6 programme). The authors would like to thank Dr. Eva Møller Nielsen at the Danish Veterinary Institute (Copenhagen, Denmark) for providing *E. coli* control strains and Dr. Lutz Geue (Friedrich-Loeffler-Institut, Wusterhausen, Germany) and Dr. Dörte Döpfer (School of Veterinary Medicine, University of Wisconsin, Madison, WI, USA) for field isolates.

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