

Quantitative analysis of penicillins in porcine tissues, milk and animal feed using derivatisation with piperidine and stable isotope dilution liquid chromatography tandem mass spectrometry

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Abstract Penicillins are used universally in both human and veterinary medicine. The European Union (EU) has established maximum residue levels (MRLs) for most β -lactam antibiotics in milk and animal tissues and included them in the National Residue Monitoring Programs. In this study, a novel method is described for the determination and confirmation of eight penicillins in porcine tissues, milk and animal feed by liquid chromatography–tandem mass spectrometry (LC–MS/MS). To prevent degradation of penicillin residues during workup, a derivatisation procedure was developed, by which penicillins were converted to stable piperidine derivatives. Deuterated piperidine derivatives were synthesised for all relevant penicillins, enabling the use of isotope dilution for accurate quantification. Penicillin residues were derivatised in the crude extract with piperidine and isolated using solid-phase extraction. The penicillin piperidine derivatives were determined by LC–MS/MS. The method was validated at the current MRLs, which range from 25–300 $\mu\text{g kg}^{-1}$ in muscle and kidney to 4–30 $\mu\text{g kg}^{-1}$ in milk as well as at the target value of 100 $\mu\text{g kg}^{-1}$ chosen for animal feed, according to the EU requirements for a quantitative confirmatory

method. Accuracy ranged from 94–113% (muscle), 83–111% (kidney) and 87–103% (milk) to 88–116% (animal feed). Intra-day precision (relative standard deviation (RSD)_r) ranged from 5–13% (muscle, $n=18$), 4–17% (kidney, $n=7$) and 5–18% (milk, $n=7$) to 11–32% (animal feed, $n=18$). Inter-day precision (RSD_{RL}, $n=18$) ranged from 6–23% (muscle) to 11–36% (animal feed). From the results, it was concluded that the method was fit for purpose at the target MRLs in animal tissue and target levels for animal feed.

Keywords Penicillins · Piperidine · Derivatisation · LC–MS/MS · Isotope dilution · Validation

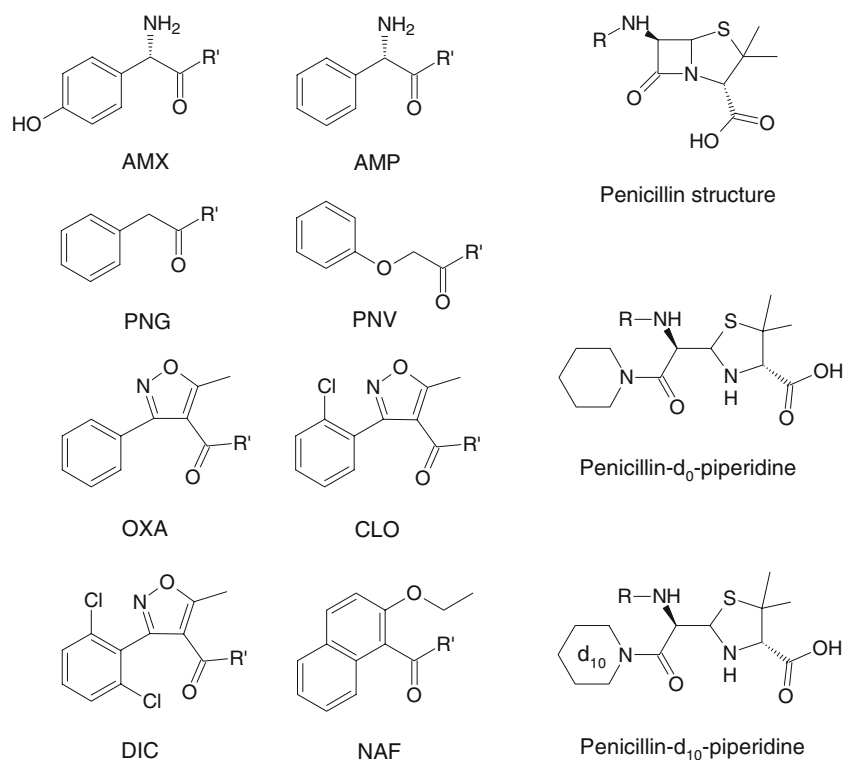
Introduction

The introduction of penicillin to treat bacterial infections has been one of the most important lifesaving medical discoveries of the twentieth century. Today, penicillins are still universally used both in human and veterinary medicine. Unfortunately, indiscriminate use of penicillins and other β -lactam antibiotics has been blamed for the increase of bacterial resistance to antibiotics. Furthermore, the consumption of food products containing penicillin drug residues may be harmful to allergic consumers [1–3]. Therefore, the European Union (EU) has established maximum residue levels (MRLs) for most β -lactam antibiotics (Fig. 1) in milk and animal tissues (see Table 1) and included them in National Residue Monitoring Programs [4]. Penicillins can be administered to livestock in a number of ways (e.g. in feed, water), but only amoxicillin (AMX) has actually been registered both as a feed and water

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Fig. 1 Structures of penicillins, derivatised penicillins and derivatised internal standards



additive (premix 250–500 mg kg⁻¹). Medication in feed is often preferred due to the low stability of penicillins in aqueous solutions [5]. Analysis and control of antibiotics in feedstuffs have become an important issue as feedstuffs can only be prepared under specific conditions as stated in Council Directive 90/167/EEC [6].

Over the years, many methods have been described for the analysis of penicillins in various matrices including animal feeds. Unfortunately, β -lactams are known for their rapid degradation during sample preparation. The use of organic

solvents and acids in extraction and chromatography has been implicated in the degradation of penicillins [7–9]. Losses may also occur during storage (milk [10], meat [11] and feed [5]). To stabilise penicillins during sample processing, several analytical methods apply derivatisation (benzoic anhydride with 1,2,4-triazole mercury (II) reagent [12–14] or formaldehyde with trichloroacetic acid [5]). An additional aim of derivatisation is the introduction of chemical moieties that improve detection by liquid chromatography–ultraviolet or fluorescence [5, 12].

Table 1 MRLs and target values for penicillin residues in animal tissue, milk and feed

Penicillin	Abbreviation	MRL		Target value
		Tissue ^a (μgkg^{-1})	Milk (μgkg^{-1})	Animal feed (μgkg^{-1})
Amoxicillin	AMX	50	4	100
Ampicillin	AMP	50	4	100
Penicillin G (benzylpenicillin)	PNG	50	4	100
Penicillin V (phenoxymethylpenicillin)	PNV	25 ^{bc}	4 ^d	100
Oxacillin	OXA	300	30	100
Cloxacillin	CLO	300	30	100
Dicloxacillin	DIC	300	30	100
Nafcillin	NAF	300	30	100

^a Muscle, liver, kidney

^b Poultry

^c Porcine

^d Target value

Nowadays, liquid chromatography–tandem mass spectrometry (LC–MS/MS)-based methods for the determination of penicillins are widely available [15]. The reproducibility of quantitative LC–MS/MS methods is often improved when isotopically labelled internal standards can be incorporated. However, the introduction of isotopically labelled analogues of the penicillins is hampered by their complex structure. The only suitable penicillin internal standards described thus far are benzylpenicillin- $^{13}\text{C}_2$ [16], benzylpenicillin- d_7 [7, 17], benzylpenicillin- d_5 , ampicillin (AMP)- d_6 and nafcillin (NAF)- d_6 [18]. Alternatively, several methods describe the use of one of the other penicillins as IS [5, 19–23]. The use of other (non-deuterated) penicillins as internal standards is not advisable as these also have veterinary uses and they may exhibit different behaviours during extraction and chromatography as well. So far, no methods have been described that combine derivatisation of penicillins with LC–MS/MS. In this study, the use of piperidine (PIP) to stabilise penicillin residues in matrix by the formation of penicillin–piperidine derivatives is presented. Deuterated piperidine derivatives have been synthesised for all relevant penicillins, enabling the use of isotope dilution for the analysis of penicillins. The development and validation, according to EU guidelines of Commission Decision 2002/657/EC [24], in porcine muscle and kidney tissue as well as in milk and at low carryover levels in feedstuffs is described.

At RIKILT—Institute of Food Safety, feed and porcine tissue samples are screened with an in-house developed microbiological method [25]. Samples suspected to contain β -lactams are subsequently analysed using the confirmatory and quantitative method described in this paper. The method for animal tissues and milk has been in use for over 5 years and the method for feed for over 3 years.

Experimental

Chemicals and reagents

All chemicals and solvents were at least of analytical reagent grade and were purchased from local commercial suppliers. Acetic acid, ammonium acetate (NH_4Ac), phosphoric acid (85%), disodium hydrogen phosphate dibasic salt ($\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$) and formic acid (FA, 98–100%) were obtained from Merck (Darmstadt, Germany). Methanol (MeOH) and acetonitrile (MeCN) were supplied by Biosolve (Valkenswaard, the Netherlands), and piperidine was purchased from Fluka (Zwijndrecht, the Netherlands). Piperidine- d_{11} was obtained from CDN Isotopes (Pointe-Claire, Quebec, Canada). Amoxicillin, ampicillin, penicillin G (PNG), penicillin V (PNV), oxacillin (OXA), cloxacillin (CLO), dicloxacillin (DIC) and nafcillin were supplied by

Sigma-Aldrich (Zwijndrecht, the Netherlands). LC quality water (conductivity at least $10 \text{ M}\Omega \text{ cm}^{-1}$) was prepared with a Milli-Q water purification system (Millipore, St. Louis, MO, USA).

Synthesis of penicillin–piperidine derivatives

Derivatised penicillins were prepared by adding a piperidine solution in water (1.00 ml, 200 mM) to the solid penicillin base (0.020 mmol). The mixture was shaken gently until the penicillin was completely dissolved. The reaction was allowed to proceed for 30 min at room temperature, whereupon acetic acid (0.18 ml, 1 M) was added. Finally, NH_4Ac buffer (100 mM, pH 6.0) was added to obtain a stock solution with a concentration of 1 mg ml^{-1} of penicillin base equivalent. The corresponding deuterated analogues were obtained by using a piperidine- d_{10} solution in water. The derivatised penicillins were stable for 1 year when kept at $-20 \text{ }^\circ\text{C}$. The purity of the piperidine derivatives was assessed with LC–MS/MS and ^1H -nuclear magnetic resonance (NMR) and found to be higher than 90%. No non-derivatised starting material could be detected in the synthesised standards (<1%).

Standard solutions

Stock standard solutions (1 mg ml^{-1}) of penicillin references were prepared in MeOH, taking into account the purity of the reference standard material. The stock solutions were stable for 1 week when stored at $4\text{--}8 \text{ }^\circ\text{C}$.

From the stocks mixed, penicillin working standard solutions (PWS) were prepared. The concentrations of the individual penicillins were chosen to match the specific MRL in muscle and kidney tissue and in milk. For the analysis in tissue samples, a mixed penicillin working solution (PWS-tissue) was prepared by adding $100 \mu\text{l}$ of stock solution of AMX, ampicillin (AMP) and PNG; $50 \mu\text{l}$ of PNV stock; and $600 \mu\text{l}$ of OXA, CLO, DIC and NAF stock solution to a 100-ml volumetric flask and making up to the mark with water. For milk samples, a mixed working solution (PWS-milk) was prepared by adding $16 \mu\text{l}$ of AMX, AMP, PNG and PNV stock solution and $120 \mu\text{l}$ of OXA, CLO, DIC and NAF stock solution to a 100-ml volumetric flask and making up to the mark with water. For feed samples, a mixed working standard (PWS-feed) was prepared by adding $200 \mu\text{l}$ of all penicillin stock solutions to a 100-ml volumetric flask and making up to the mark with water. PWS were stable for 1 week when stored at $4\text{--}8 \text{ }^\circ\text{C}$.

Mixed working standard solutions of penicillin- d_0 -piperidine derivatives (PPWS) and mixed internal standard solutions of the penicillin- d_{10} -piperidine derivatives (PPIS) were prepared from the synthesised standards, matching the specific MRL levels in muscle and kidney tissue and in

milk and were mixed in the same proportions as described for the penicillin working solutions. The PPWS and PPIS were stable for 1 month when stored at 4–8 °C.

Working standards for tissue, milk and feed containing mixtures of the penicillin-d₀-piperidines and penicillin-d₁₀-piperidines were prepared by adding 250 µl of the specific PPWS and 250 µl of the corresponding PPIS in a 10-ml volumetric flask and making up to the mark with water. These working standards were used to check the performance of the LC–MS/MS system.

Instrumentation

An Agilent 1100 Series high-performance liquid chromatography (HPLC) system (Agilent Technologies, USA) coupled to a Quattro Ultima® tandem mass detector (Waters-Micromass, Manchester, UK), both operating under MassLynx® software, was used for sample analysis. The mass spectrometer was operated in electrospray negative mode, and data acquisition was in multiple reaction monitoring mode (MRM). The precursor/product ions monitored are listed in Table 2. The source settings were as follows: capillary voltage 2.6 kV, source temperature 120 °C, desolvation temperature 300 °C, cone voltage 30 V, cone nitrogen gas flow 180 l h⁻¹, desolvation gas flow 580 l h⁻¹. Argon (2.2 × 10⁻³ mbar) was used as the collision gas, and the multiplier was operated at 750 V. The collision energy (17–23 eV) was optimised for each analyte and each MRM (Table 2). The HPLC system was equipped with a 3.0 × 150 mm, 5 µm Symmetry C₁₈ column (Waters, Milford, PA, USA) kept at 40 °C. A binary gradient mobile

phase was used at a flow rate of 0.4 ml min⁻¹ with solvent A (0.2% FA in water) and solvent B (0.2% FA in MeCN/water 9/1 (v/v)). A splitter was used to reduce the solvent flow to 0.15 ml min⁻¹ entering the mass spectrometer. The gradient started isocratically for 1 min at 15% B, followed by a linear increase to 50% B in 7 min, followed by a linear increase to 70% B in 5 min. After an isocratic hold at 70% B for 1 min, the gradient returned to 15% B in 1 min for equilibration of the column. Total run time was 20 min. Sample injection volume was 50 µl.

¹H-NMR spectra were recorded on a Bruker Avance 400 MHz spectrometer. Presaturation was performed on the residual HDO resonance. A 90° pulse was used; the total relaxation delay was 3.7 s; the spectral width was 5,000 Hz. The data were acquired in 16 K data points. Before Fourier transformation and phasing, a one-third-shifted sine squared window multiplication was applied and a zero filling to 128 K data points was applied. Calibration of spectra was achieved by setting the HCD₂ resonance of deuterated methanol to 3.27 ppm.

Sample preparation: porcine muscle and kidney tissue

Porcine kidney and muscle tissue samples (2 g) were minced and homogenised with phosphate buffer (2 ml, 200 mM, pH=6.0). Internal standard mixture (PPIS-tissue, 100 µL) was added to all samples. After an incubation time of 30 min at room temperature, phosphate buffer (100 mM, pH=8.0, 40 ml) was added. The tubes were vortexed for 30 s, followed by addition of piperidine (300 µl). The tubes were vortexed for 30 s, and the pH was checked with a pH

Table 2 LC–MS/MS precursor/product ion combinations (quantifier italicised) monitored in ESI negative mode

Penicillin	Derivatised penicillin	Internal standard	Precursor ion (<i>m/z</i>)	Product ions (<i>m/z</i>)	Collision energy (eV)
AMX	AMX-d ₀ -PIP		449.2	277.2, 371.2	20
		AMX-d ₁₀ -PIP	459.3	381.3	18
AMP	AMP-d ₀ -PIP		433.2	270.2, 355.2	20
		AMP-d ₁₀ -PIP	443.3	365.3	20
PNG	PNG-d ₀ -PIP		418.2	222.2, 340.2	20
		PNG-d ₁₀ -PIP	428.3	350.3	20
PNV	PNV-d ₀ -PIP		434.2	262.2, 356.2	22
		PNV-d ₁₀ -PIP	444.3	366.3	22
OXA	OXA-d ₀ -PIP		485.2	322.2, 407.2	23
		OXA-d ₁₀ -PIP	495.3	417.3	20
CLO	CLO-d ₀ -PIP		519.2	405.2, 441.2	20
		CLO-d ₁₀ -PIP	529.3	451.3	18
DIC	DIC-d ₀ -PIP		553.2	439.2, 475.2	17
		DIC-d ₁₀ -PIP	563.3	485.3	17
NAF	NAF-d ₀ -PIP		498.2	263.2, 420.2	18
		NAF-d ₁₀ -PIP	508.3	430.3	18

stick and if necessary adjusted to 11–12 with piperidine. After agitation for 5 min on a rotary tumbler, phosphoric acid (100 μ l) was added and the samples were vortexed; the pH was adjusted to 8.0 ± 0.5 with phosphoric acid if necessary. Next, the samples were heated in a water bath at 85 °C. After 5 min, the samples were quickly cooled to room temperature by using an ice bath. The samples were centrifuged at $2,700 \times g$ for 10 min, and the supernatants were filtered through a wad of cotton wool into polypropylene tubes. Cloudy extracts were centrifuged at $15,000 \times g$ for 15 min. Oasis[®] HLB (60-mg, 3-cm³ solid-phase extraction (SPE) cartridges, Waters, Manchester, UK) was conditioned using 2 ml of MeOH, 2 ml of water and 2 ml phosphate buffer (100 mM, pH=8.0). The cartridges were loaded with extract (10 ml) and subsequently washed with 2×2 ml of water. After drying under reduced pressure, cartridges were eluted with 3 ml MeOH/water (80/20, v/v). The eluate was evaporated to dryness at 60 °C under a stream of nitrogen and redissolved in 500 μ l 2% MeCN.

Sample preparation: milk

For the analysis of raw bovine milk, the porcine tissue method was used with a few adjustments. To the milk samples (4 g), internal standard mixture (PPIS-milk, 100 μ l) was added. Phosphate buffer (100 mM, pH=8.0, 40 ml) was added, and the tubes were vortexed for 30 s. Piperidine (300 μ l) was added, and after vortex agitation for 30 s, the pH was checked with a pH stick and if necessary adjusted to 11–12 with piperidine. After agitation for 5 min on a rotary tumbler, phosphoric acid (100 μ l) was added and the samples were vortexed; the pH was adjusted to 8.0 ± 0.5 with phosphoric acid if necessary. The samples were centrifuged at $2,700 \times g$ for 10 min, and the supernatants were filtered through a wad of cotton wool into polypropylene tubes. SPE cartridges were loaded with 20 ml of extract, and the final residue was reconstituted in 200 μ l of 2% MeCN.

Sample preparation: animal feed

Feeding stuffs were ground to a particle size of 0.5 mm. To the feed samples (2 g), internal standard mixture (PPIS-feed, 200 μ l) was added. After an incubation time of 30 min at room temperature, phosphate buffer was added (100 mM, pH=8.0, 40 ml). The tubes were agitated for 15 min on a rotary tumbler. The samples were centrifuged for 10 min at $2,700 \times g$, and with a pipette 10 ml of the supernatant was transferred to a 14 ml polypropylene tube. Piperidine (300 μ l) was added, and the samples were vortexed immediately. After agitation, the pH was checked and if necessary adjusted with piperidine to 11–12. After 5 min, extracts were adjusted to pH 8.0 ± 0.5 with phosphoric acid

(100–200 μ l). Subsequent purification with Oasis[®] HLB followed the procedure described for porcine tissue. After evaporation, the residue was reconstituted in 500 μ l of water/MeCN/FA (98/2/0.2, v/v/v).

For accurate quantification, feed samples were analysed using a multi-level standard addition approach. The chosen addition levels depend on the concentrations expected in the feed. In case suspect samples (based on the in-house microbiological screening assay) were analysed, the indicative values derived from the screening assay were used to determine standard addition levels. Depending on the expected concentration, six standard addition levels were chosen and spiked accordingly. To samples suspected to contain more than 5 mg kg⁻¹, a larger volume of PPIS mixture (2 ml) was added. Depending on the results from the microbiological screening, an aliquot of the crude extract was taken and diluted with phosphate buffer (100 mM, pH=8.0). Samples containing less than 1 mg kg⁻¹ were not diluted, whereas samples suspected to contain between 1 and 5 mg kg⁻¹ were diluted five times with phosphate buffer and samples containing between 5 and 50 mg kg⁻¹ were diluted 50 times before derivatisation.

Quality control samples and matrix-matched standard preparation

For tissue and milk samples, calibration was performed using matrix-matched standards (MMS) prepared by adding the appropriate amounts of the penicillin mixed standard solution to blank matrix. For porcine tissue, MMS were prepared corresponding to 0 MRL, 0.25 MRL (25 μ l of PWS-tissue), 0.5 MRL (50 μ l), one MRL (100 μ l), two MRL (200 μ l) and five MRL (500 μ l). Internal standard mixture (PPIS-tissue, 100 μ l) was added to all samples. For milk, MMS were prepared corresponding to 0 MRL, 0.25 MRL (25 μ l of PWS-milk), 0.5 MRL (50 μ l), one MRL (100 μ l), two MRL (200 μ l) and five MRL (500 μ l). Internal standard mixture (PPIS-milk, 100 μ l) was added to all samples. Two blank samples spiked with IS were included in each series for the preparation of matrix-matched recovery standards. Penicillin-d₀-piperidine mixed standard solution (PPWS-tissue) was added to the dried extracts to obtain concentrations corresponding to 0.5 MRL (12.5 μ L) and two MRL (50 μ L) in a total volume of 500 μ l 2% MeCN. For milk, PPWS-milk was added to the dried extracts to obtain concentrations corresponding to 0.5 MRL (25 μ L) and two MRL (100 μ l) in a total volume of 200 μ l 2% MeCN.

At the beginning of each series, a working standard containing the penicillin-d₀-piperidines and the penicillin-d₁₀-piperidines was injected to check the performance of the instrument.

Validation

The method for porcine muscle tissue was validated in a 5-day experimental setup. On three separate occasions, using three different blank porcine muscle materials, samples were fortified with a mixture of penicillins (PWS-tissue) at three levels (0.5 MRL, one MRL and 1.5 MRL). Six replicates, at each of the three fortification levels, were analysed on each day. Accuracy, intra-day precision (repeatability), inter-day precision (within-lab reproducibility, muscle tissue only), recovery, linearity, CC_{α} and CC_{β} were determined. Additionally (day 4), 20 blank samples were analysed to determine selectivity, specificity and limit of detection (LoD; the concentration at which the quantifier product ion has a signal to noise ratio of 3). The method for muscle tissue was tested (day 5) for ruggedness according to Youden [24]. Variables that were tested included the pH of the phosphate buffer, the piperidine derivatisation time, the heating temperature of the derivatised extract, the drying time of the SPE cartridge before elution of the purified extract and the composition of the elution solvent and the time lag before the dried sample was reconstituted. Porcine kidney tissue was additionally validated using six replicates, at each of the three fortification levels on one occasion.

The validation for milk was limited to seven replicate measurements at MRL on 1 day. Intra-day precision and accuracy at MRL level, linearity, specificity and selectivity were determined.

The validation for feedstuffs was carried out on seven separate occasions using multi-level standard addition. Of each of the 20 blank feed samples (representing feeding stuffs intended for poultry, porcine, bovine, ovine), 11 subsamples were processed. Two samples were not spiked with PWS-feed (blanks), six samples were fortified at one of the validation levels (0.1, 0.2, 0.5 mg kg⁻¹) in duplicate and the remaining three samples were fortified at 1, 2 and 5 mg kg⁻¹, respectively. Each validation level was quantified by using the first three addition levels and applying linear regression of the least squares. The sum of the peak areas of the two product ions was used to calculate the response factor relative to the IS. Accuracy, intra-day precision, inter-day precision, linearity, CC_{α} , CC_{β} , specificity, selectivity and stability of the extracts were determined.

Results and discussion

Synthesis of penicillin–piperidine derivatives

For the antibacterial activity of penicillins, an intact β -lactam ring is essential. Acid- or base-catalysed hydrolysis of the β -lactam ring will result in inactive penicilloic acids

and degradation products thereof [26–28]. Penicillins containing a primary amino group may also isomerise to (inactive) diketopiperazine derivatives [27, 28]. Other nucleophiles, such as alcohols and amines, can also react with the β -lactam ring, resulting in the formation of penicilloic esters and (substituted) amides [29, 30]. In case ammonia or simple primary amines are used, the reaction is also called aminolysis. To our knowledge, the reaction of penicillins with secondary amines has not been studied in detail. We found that the reaction of a variety of penicillins with a concentrated solution of piperidine in water proceeded within minutes at room temperature. The synthesis of the derivatised penicillins does not require any special equipment. For the penicillins studied, a single compound was produced in high yield. In all cases, the corresponding piperidine derivative was formed by addition to the β -lactam carbonyl group, followed by ring opening. In Fig. 2, the ¹H-NMR spectrum of amoxicillin-d₀-piperidine (AMX-d₀-PIP) is shown as an example. In AMX β -lactam protons, H₅ and H₆ resonate at 5.40 and 5.58 ppm, while in the ring-opened AMX-d₀-PIP they have shifted to 4.85 and 4.92 ppm. The piperidine protons are found at 1.65 ppm (6H) and 2.96 ppm (4H).

It is possible to isolate the penicillin–piperidine derivatives by crystallisation, but in practise it is more convenient to prepare the standards in situ and store them at –20 °C or lower, without further purification. It was verified by ¹H-NMR that during the reaction no or little epimerisation at the 6 position occurs. However, during long-term storage, some epimerisation may occur in solution, even when the standards are kept at –20 °C. After 12 months at –20 °C, a stock solution of AMX-d₀-PIP contained 15% of the 6- α epimer. Therefore, it was decided to synthesise new standards every 12 months.

The MS/MS spectra of AMX-d₀-PIP and AMX-d₁₀-PIP are presented in Fig. 3. The addition of piperidine is evident from the mass increment of 85 Da in the deprotonated molecular ion (M-H)⁻ (AMX: *m/z* 364). Two major fragments are formed upon collision-induced dissociation: *m/z* 371 (loss of *m/z* 78 or H₂S + CO₂) and *m/z* 277 (loss of *m/z* 172, loss of H₂S + CO₂ + loss of phenol (*m/z* 94)). Both fragments still contain the piperidine ring. In the MS/MS spectrum of the corresponding d₁₀-analogue, the expected mass shift of 10 Da is seen for the (M-H)⁻ and both fragments.

Method development

Penicillins do not only react quickly with piperidine but also with other primary (e.g., methylamine, aminoethanol) and secondary amines (diethyl amine, morpholine, piperazine). Piperidine was chosen as the derivatisation reagent of choice because a suitable deuterated analogue is

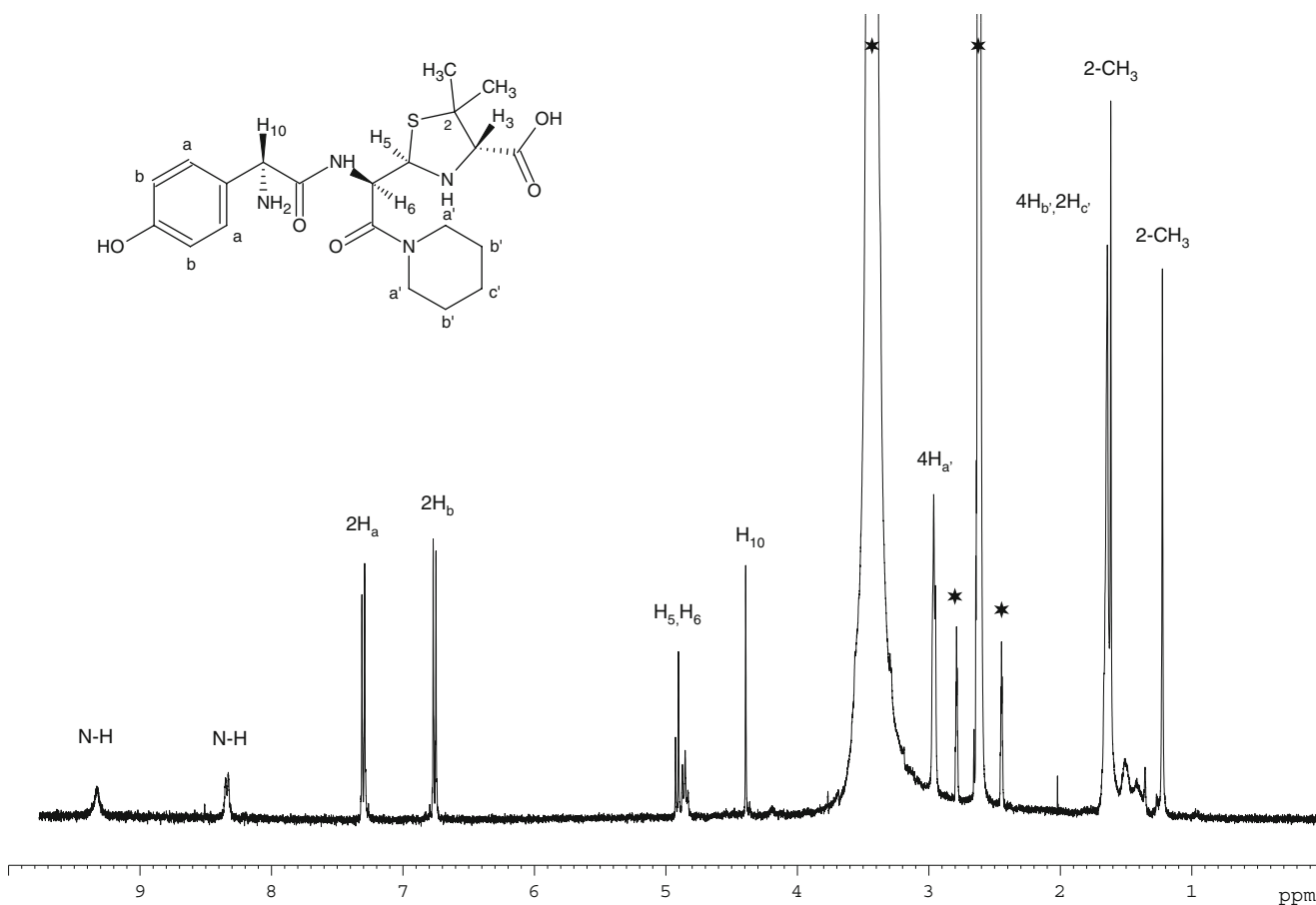


Fig. 2 $^1\text{H-NMR}$ of amoxicillin- d_0 -piperidine, recorded in CD_3OD . Stars indicate solvent and residual water resonances

available and the derivatives produced exhibit favourable chromatographic behaviour. Amoxicillin is the most polar penicillin, eluting shortly after the void volume on a C18 HPLC column. AMX- d_0 -PIP exhibits increased retention, eluting at 4.4 min with the gradient used in this study (Fig. 4). The derivatised penicillins are most conveniently recorded in electrospray negative mode. For all compounds, loss of the m/z 78 fragment is the first or second most prominent fragmentation pathway. Next to the loss of the m/z 78 fragment, for each penicillin derivative, one structure-specific fragment has been included in the MRM method as well (Table 2).

It was verified that the derivatisation is specific for penicillins containing an intact β -lactam and that no reaction occurs with hydrolysed or degraded penicillins that potentially could interfere with the mass spectrometric analysis. Penicillins were degraded in solution by reaction with 0.1 N sodium hydroxide and with 0.1 N hydrochloric acid, until LC-MS/MS analysis indicated the complete conversion to hydrolysed or degraded products [26, 28]. Piperidine was added to the samples and subsequent LC-MS/MS indicated no change in the composition of the samples, nor any formation of penicillin-piperidine deriv-

atives from the hydrolysed or degraded penicillins. Hence, it can be concluded that the derivatisation is very specific for penicillin residues containing an intact β -lactam ring and that, in line with EU regulations, only the pharmacologically active residues are determined [4].

The pH of the extract during derivatisation was identified as a critical factor. In Fig. 5, the effect of pH on the derivatisation efficiency is shown. Piperidine was added in three different amounts to a solution of AMX in phosphate buffer (100 mM) with a pH ranging from 3 to 9. By addition of piperidine, which acts as a base, the pH of the buffer solution is raised. Figure 5 shows that derivatisation is catalysed by high pH (11 or higher); at neutral or acidic pH, the reaction proceeds very slowly and by-products (due to hydrolysis) are produced. Therefore, an excess of piperidine is used to insure an alkaline pH. However, prolonged exposure of the extracts to high pH may in turn lead to degradation of the matrix and this may affect sample cleanup. By adding phosphoric acid after 5 min of reaction, degradation of the matrix is minimised.

Although the use of deuterated internal standards facilitates the accurate quantification of penicillin residues in animal tissues and milk, the results for animal feed

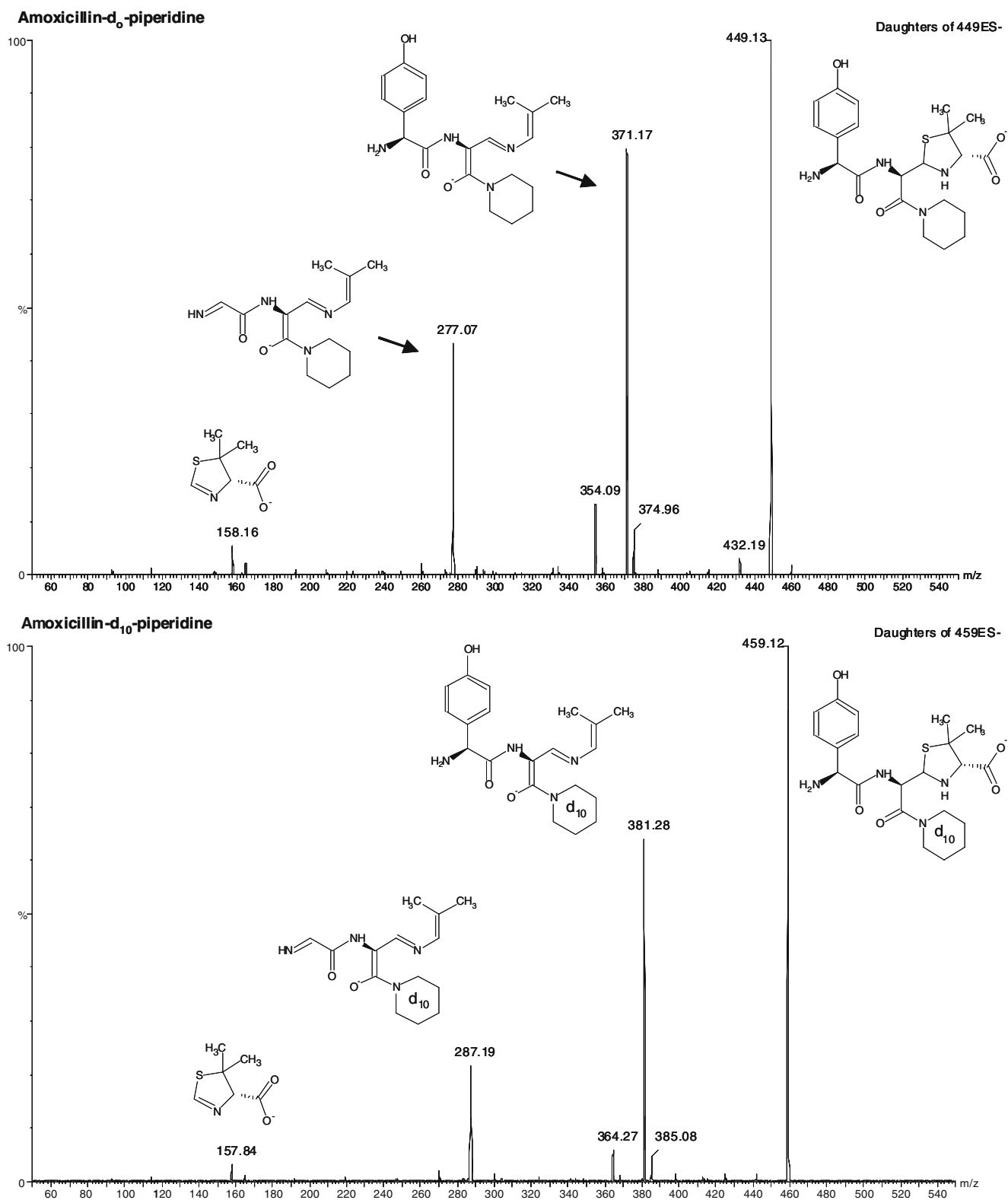


Fig. 3 MS/MS fragmentation spectra of amoxicillin-d₀-piperidine (*top*) and amoxicillin-d₁₀-piperidine (*below*), recorded in electrospray negative mode at a collision energy of 15 eV

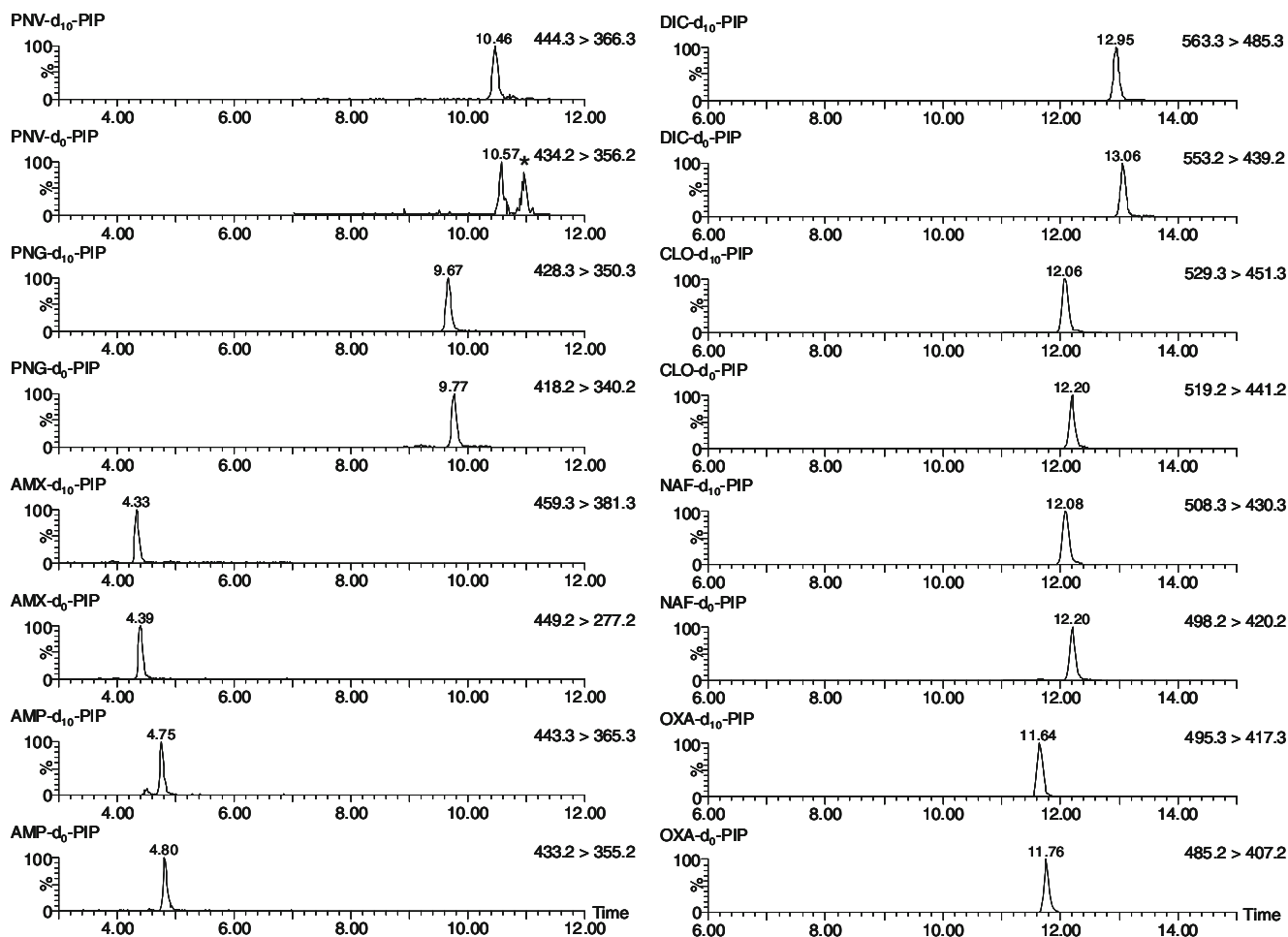


Fig. 4 LC-MS/MS chromatograms obtained for a blank porcine muscle sample spiked with a penicillin mix at MRL. For each penicillin-piperidine derivative, the quantifier trace is shown and the chromatogram for the corresponding IS transition. Stars indicate matrix interference

Fig. 5 Derivatisation of amoxicillin ($5 \mu\text{g ml}^{-1}$) with piperidine as function of the pH of the solution (100 mM phosphate buffer, pH 3 to 9) and the amount of piperidine added ($n=2 \pm \text{SD}$). The effect of piperidine on the final pH of the solution is indicated

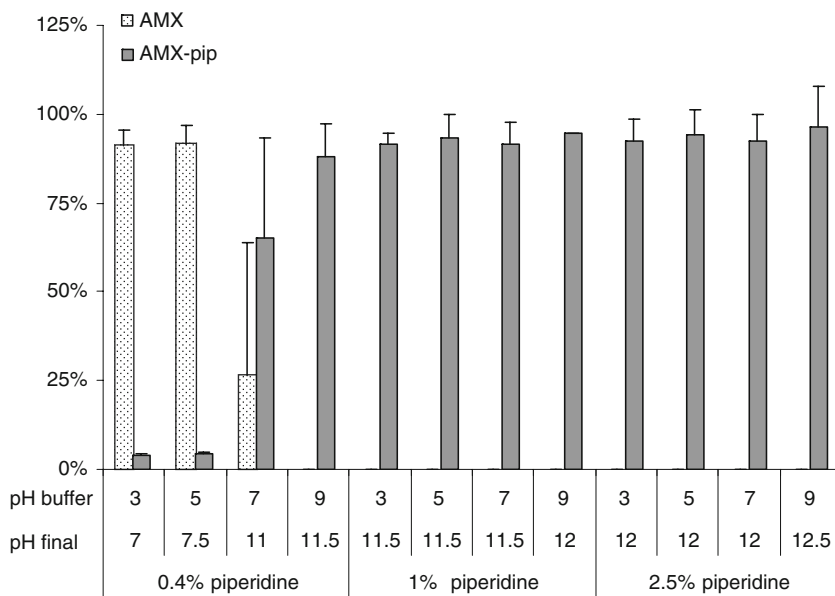


Table 3 Validation results obtained for the determination of penicillins in porcine muscle and kidney tissue

Penicillin	Fortification level (μgkg^{-1})	Muscle ($n=18$)						Kidney ($n=6$)					
		Accuracy (%)	Recovery ^a (%)	RSD _r (%)	RSD _{RL} (%)	CC _{α} (μgkg^{-1})	CC _{β} (μgkg^{-1})	LoD ^b (μgkg^{-1})	Accuracy (%)	Recovery ^a (%)	RSD _r (%)	LoD (μgkg^{-1})	
AMX	25	97	72	7.1	8.6			3.4	97	32	9.4	2.9	
	50	94		6.3	12.1	57	67		95		11.3		
	75	100		10.7	11.8				84		12.3		
AMP	25	105	111	7.1	14.4			5.4	91	120	12.3	2.0	
	50	101		6.0	6.8	56	62		90		4.0		
	75	103		9.2	9.5				83		9.2		
PNG	25	96	89	7.4	9.3			3.8	90	97	4.6	0.7	
	50	95		8.1	8.9	54	62		93		6.0		
	75	98		7.2	8.1				93		11.0		
PNV	12.5	98	103	11.1	23.4			4.7	96	96	17.4	2.3	
	25	98		10.3	11.1	29	33		106		8.7		
	37.5	102		13.4	15.0				100		12.6		
OXA	150	100	89	8.5	12.0			14	103	84	5.4	4.0	
	300	102		5.6	9.4	351	398		104		4.4		
	450	103		5.7	6.5				103		7.3		
CLO	150	96	92	6.8	12.2			14	85	81	4.6	6.3	
	300	98		5.8	6.0	324	353		101		6.2		
	450	113		5.2	5.5				100		7.6		
DIC	150	101	84	5.8	16.3			7.0	93	79	7.0	3.4	
	300	105		6.0	11.2	370	425		106		4.8		
	450	112		6.3	10.4				111		3.9		
NAF	150	96	81	8.7	15.9			2.7	85	79	9.3	1.2	
	300	96		8.5	10.0	338	387		94		4.9		
	450	101		8.3	9.2				91		8.2		

^a Determined at one-half MRL^b Poorest LoD obtained in 5 validation series

initially were less satisfactory, even with internal standard correction. This may be attributed to the large variability in composition, which can be found in feedstuffs. Raw materials and additives of feeds introduce many interfering components, such as oils, fats, proteins and salts, which can occur at very high levels. These constituents may influence the efficiency of sample preparation by causing differences in derivatisation and extraction efficiency and they may induce ion suppression effects during LC–MS/MS analysis. The internal standards are added to samples as derivatives and therefore only correct for ion suppression and variability in cleanup efficiency after derivatisation, but not for differences in derivatisation efficiency. Multi-level standard addition (MLSA) is therefore required for the analysis of animal feeds. MLSA eliminates differences in derivatisation efficiency, thereby improving accuracy and precision of the determination of penicillins in animal feed [31, 32]. Another possibility to reduce ion suppression effects may be dilution of the extract, as described by de Baere et al. [5], but this method was developed for medicated levels (200–500 mg kg⁻¹). This is not an option when penicillins at a carryover level as low as 0.1 mg kg⁻¹ have to be determined. Feeds with concentrations higher than 5 mg kg⁻¹ need to be diluted to comply with the linear dynamic range of the MS detector, but it was not investigated if MLSA could be skipped for these diluted extracts.

Validation

The validation data for porcine muscle tissue were performed according to CD 2002/657/EC [24], and the results obtained are presented in Table 3. In Fig. 4, LC–MS/MS chromatograms are shown for a processed blank muscle tissue sample spiked with penicillins at MRL. Accuracy in muscle tissue varied from 94% to 113% with an intra-day precision (relative standard deviation (RSD)_D, *n*=18) between 5.2% and 13.4% and an inter-day precision (RSD_{RL}, *n*=18) between 5.5% and 23%. Requirements for accuracy (80–110%), repeatability and within-laboratory reproducibility were met for all compounds at all levels with the exception of the accuracy for CLO (113%, 1.5 MRL). Only a few multi-residue HPLC [13, 14] and LC–MS methods [31] have been reported that include all penicillins described in this paper. These studies reported comparable results. Slightly higher variability was observed for PNV, which at least in part, may be caused by the lower fortification levels, compared to the other penicillins. The CC_α and CC_β values that were obtained for all penicillins at MRL comply with the EU criteria and were comparable to those reported for bovine muscle tissue, even though only four replicates per level per day (*n*=12) were used in that study [31]. The average overall recovery determined at 0.5 and two MRL was 70% for AMX and higher than 80% for

the other penicillins. No significant differences in recovery were observed between both fortification levels. Linearity of the MMS curve (0.25–5 MRL) was 0.995 or higher on each of the five validation days. Only in two instances (PNV 0.992 and NAF 0.986, both on day 4) was (slightly) lower linearity observed. For each penicillin, the estimated LoD is five times or more below the specific MRL.

The method was tested for ruggedness in tissue according to Youden. From the experiment, it was concluded that samples must be reconstituted immediately after drying. By analysing 20 different blank porcine tissues, the method was proven to be specific and selective. No interfering signals were observed in any of the blanks at the relevant retention times.

Kidney tissue was validated using a single-day experimental setup because of similarity in matrix (porcine tissue) and fortification levels. The validity of this approach was checked by preliminary experiments during method development. These tests indicated that the performance characteristics were likely to be in the same range as the determined parameters for muscle tissue. Accuracy in kidney tissue varied from 83% to 111% with an intra-day precision (RSD_D, *n*=6) between 4.0% and 17.4% (Table 3). Average recovery in kidney tissue was higher than 80% with the exception of AMX (36%). Lower-than-average recoveries for amoxicillin in kidney tissue were also observed by Becker et al. [31]. They attributed this to loss of retention on SPE by overload of matrix compounds from the extract. Lower recoveries for AMX have also been reported for other matrices such as wastewater and honey [17, 33]. Although recovery for AMX in kidney tissue is significantly lower than in muscle tissue, the accuracies in combination with RSDs obtained indicate that the internal standard corrects adequately for the loss in recovery. No other significant differences were observed between kidney and muscle samples during this study. This is in accordance with the report of Verdon et al. [14], but Becker et al. deemed kidney to be a more difficult matrix [31].

Table 4 Single-day validation results obtained for penicillins in milk (*n*=7) at MRL

Penicillin	Fortification (μgkg ⁻¹)	Accuracy (%)	RSD _r (%)
AMX	4	90	8.4
AMP	4	87	4.9
PNG	4	102	8.8
PNV	4	99	17.5
OXA	30	96	6.8
CLO	30	103	8.0
DIC	30	94	8.8
NAF	30	98	6.6

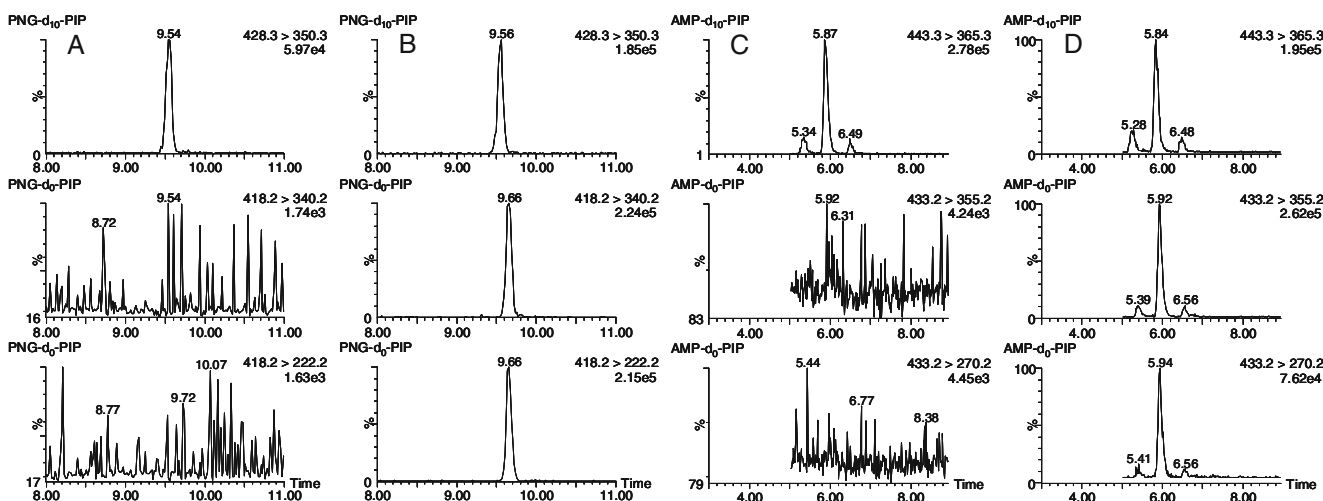


Fig. 6 Selected ion chromatograms for: **a** blank milk sample spiked with $4 \mu\text{g kg}^{-1}$ PNG- d_{10} -PIP IS; **b** a (non-compliant) milk sample obtained from the National Residue Monitoring Program, containing

$5.2 \mu\text{g l}^{-1}$ PNG; **c** blank feed sample spiked with 10 mg kg^{-1} AMP- d_{10} -PIP IS; **d** A feed sample containing 45 mg kg^{-1} AMP (non-compliant). Of the feed extracts, $200 \mu\text{l}$ was used for further SPE cleanup

The method for muscle and kidney tissue was adapted for bovine milk. To accommodate for the (approximately tenfold) lower MRLs in milk, the protocol for porcine tissue was adjusted. A larger portion was processed, the

final sample extract volume was reduced and the injection volume increased. In this way, the required detection limits could be achieved and confirmation of identity could be established for the penicillins (AMX, AMP, PNV, PNG)

Table 5 Validation results obtained for the analysis of penicillins in animal feed

Penicillin	Fortification (mg kg^{-1})	Animal feed ($n=18$)				
		Accuracy (%)	RSD _r (%)	RSD _{RL} (%)	CC _α (mg kg^{-1})	CC _β (mg kg^{-1})
AMX	0.1	107	29	32	0.15	0.20
	0.2	88	23	25		
	0.5	92	15	16		
AMP	0.1	96	18	25	0.14	0.18
	0.2	100	20	21		
	0.5	102	16	17		
PNG	0.1	102	11	14	0.12	0.14
	0.2	106	13	13		
	0.5	112	24	25		
PNV	0.1	111	32	36	0.26	0.33
	0.2	103	20	20		
	0.5	108	24	24		
OXA	0.1	100	11	11	0.12	0.13
	0.2	99	14	18		
	0.5	112	24	24		
CLO	0.1	110	11	14	0.12	0.15
	0.2	103	14	21		
	0.5	118	19	26		
DIC	0.1	106	14	18	0.13	0.16
	0.2	102	13	13		
	0.5	116	21	21		
NAF	0.1	239	16	64	0.97	1.43
	0.2	162	13	44		
	0.5	152	26	37		

with an MRL at $4 \mu\text{g kg}^{-1}$. To check the validity of the adjustments of the method made for bovine milk, the method was additionally validated at MRL using a single-day experimental setup. Accuracy in milk varied from 87% to 103% with an intra-day precision (RSD_r , $n=7$) between 4.9% and 17.5% (Table 4). The accuracies obtained in milk were better than those reported by Kantiani [23]. On the other hand, intra-day precisions were slightly higher than those reported by Sørensen et al. [22] and Becker et al. [31]. Because the method for milk performed well at MRL, no further validation at other concentrations was carried out. LC–MS/MS traces of a compliant and a non-compliant milk sample (containing PNG above the MRL of $4 \mu\text{g kg}^{-1}$) from the National Residue Monitoring Program are shown in Fig. 6.

As there are no EU criteria established for the validation of veterinary drugs in animal feeds, the method for animal feed was validated according to CD 2002/657/EC for practical purposes. The validation was aimed at low carryover levels (100 to $500 \mu\text{g kg}^{-1}$). With the exception of NAF, accuracy in animal feed varied from 88% to 116%, the intra-day precision (RSD_r , $n=18$) between 11% and 32% and the inter-day precision (RSD_{RL} , $n=18$) between 11% and 36% (Table 5). The accuracy for NAF was rather poor, resulting in an overestimation of the concentration (especially at the lowest fortification level) and consequently in a very high CC_α and CC_β . Inspection of the LC–MS/MS data revealed that in some of the samples a matrix interference was co-eluting with NAF- d_0 -PIP. By analysing 20 different animal feeds, it was verified that for the other penicillins the method was specific and selective. No interfering signals were observed in any of the blanks at the relevant retention times. The linearity of the MLSA quantification (based on three additional fortification levels) ranged from 0.996 to 1.000. LC–MS/MS traces of a blank animal feed sample and a non-compliant sample containing 45 mg kg^{-1} AMP are shown in Fig. 6. To our knowledge, a multi-residue method for penicillins at carryover level (0.1 – 2 mg kg^{-1}) in feed has not been reported before. At medicated levels (200 – 500 mg kg^{-1}), mostly single-residue methods have been described for AMX (which is the only penicillin registered for use in feed). The RSD_r and RSD_{RL} obtained by these methods are invariably lower than the ones reported in this study, but this is to be expected considering the much lower target levels in this study [5, 34, 35]. LoDs are much lower for the method reported in this paper.

The derivatisation with piperidine was also tested on the other important group of β -lactam antibiotics, the cephalosporins. Ideally, these compounds together with the penicillins and clavulanate should be incorporated into a general multi-residue β -lactam method. Derivatisation experiments with clavulanate and several cephalosporins have thus far proved unsuccessful or produced the desired

derivatives only with a low yield (results not shown). Apparently, cephalosporins do not react with piperidine in the same straightforward way as the penicillins.

Conclusion

The derivatisation of penicillins with piperidine and the incorporation of the corresponding deuterated penicillin piperidine internal standards not only improved the stability of the penicillins during sample workup but also improved accuracy, repeatability and within-lab reproducibility of the analytical method significantly. The developed multi-residue LC–MS/MS method for the analysis of penicillins in porcine tissue, milk and animal feed has been validated according to Commission Decision 2002/657/EC. Except for the quantitative determination of NAF in animal feed, the obtained validation parameters were all in accordance with the criteria mentioned in the decision. The in-house validation data collected showed that the method is robust and fit for purpose. This was also confirmed by the results of two recent proficiency tests on penicillin residues in milk and in porcine tissue. Excellent results were obtained for penicillin G (Z score -0.26) and cloxacillin (Z score 0.21) in milk [36] and for cloxacillin in muscle tissue (Z score -0.06) [37].

The method has been successfully implemented in the Dutch National Residue Control Program for the analysis of penicillins in kidney, muscle, milk and animal feed samples.

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References

1. Dayan AD (1993) *Vet Microbiol* 35:213–226
2. Dewdney JM, Maes L, Raynaud JP, Blanc F, Scheid JP, Jackson T, Lens S, Verschueren C (1991) *Food Chem Toxicol* 29:477–483
3. Doyle ME (2006) Veterinary drug residues in processed meats—potential health risk, FRI briefings. http://www.wisc.edu/fri/briefs/FRIBrief_VetDrgRes.pdf, March 2006
4. EC (2010) Council Regulation (EC) 37/2010/EC of 22 December 2009 on pharmacologically active substances and their classification regarding maximum residue limits in foodstuffs of animal origin (2010). *Offic J Eur Comm* L15:1–72
5. De Baere S, De Backer P (2007) *Anal Chim Acta* 586:319–325
6. EC (1990) Council Directive 90/167/EEC of 26 March 1990 laying down the conditions governing the preparation, placing on the market and use of medicated feeding stuffs in the Community, Council Directive 90/167/EEC (1990). *Offic J Eur Comm* L92:42–48
7. Riediker S, Stadler RH (2001) *Anal Chem* 73:1614–1621
8. Mastovska K, Lightfield AR (2008) *J Chromatogr A* 1202:118–123

9. Tyczkowska KL, Voyksner RD, Aronson AL (1992) *J Chromatogr A* 594:195–201
10. Riediker S, Rytz A, Stadler RH (2004) *J Chromatogr A* 1054:359–363
11. Verdon E, Fuselier R, Hurtaud-Pessel D, Couëdor P, Cadieu N, Laurentie M (2000) *J Chromatogr A* 882:135–143
12. Boison JO, Salisbury CDC, Chan W, MacNeil JD (1991) *J AOAC Int* 74:497–501
13. Sørensen LK, Snor LK, Elkær T, Hansen H (1999) *J Chromatogr B Biomed Appl* 734:307–318
14. Verdon E, Couëdor P (1999) *J AOAC Int* 82:1083–1095
15. Kantiani L, Farré M, Barceló D (2009) *Trends Anal Chem* 28:729–744
16. Schlösser J, Mehlich A, Ballwanz F, Petz M (1998) *Fresenius' J Anal Chem* 360:498–501
17. Wang J (2004) *J AOAC Int* 87:45–55
18. Goto T, Ito Y, Yamada S, Matsumoto H, Oka H (2005) *J Chromatogr A* 1100:193–199
19. Blanchflower WJ, Hewitt SA, Kennedy DG (1994) *Analyst* 1994:2595–2601
20. Daeseleire E, De Ruyck H, Van Renterghem R (2000) *Rapid Commun Mass Spectrom* 14:1404–1409
21. Fagerquist CK, Lightfield AR, Lehotay S (2005) *Anal Chem* 77:1473–1482
22. Sørensen LK, Rasmussen BM, Boison JO, Keng L (1997) *J Chromatogr B* 694:383–391
23. Kantiani L, Farré M, Sibum M, Postigo C, López de Alda MJ, Barceló D (2009) *Anal Chem* 81:4285–4295
24. EC (2002) Commission Decision 2002/657/EC of 12 August 2002 implementing Council Directive 96/23/EC concerning the performance of analytical methods and the interpretation of results. *Offic J Eur Comm* L221:8–36
25. Pikkemaat MG, Oostra-van Dijk S, Schouten J, Rapallini M, van Egmond HJ (2008) *Food Control* 19:781–789
26. Deshpande AD, Baheti KG, Chatterjee NR (2004) *Curr Sci* 87:1684–1695
27. Olsen MA, Cummings PG, Kennedy-Gabb S, Wagner BM, Nicol GR, Munson B (2000) *Anal Chem* 72:5070–5078
28. Nägele E, Moritz R (2005) *J Am Soc Mass Spectrom* 16:1670–1676
29. Llinás A, Donoso J, Vilanova B, Frau J, Muñoz F, Page MI (2000) *J Chem Soc. Perkin Trans* 2:1521–1525
30. Llinás A, Page MI (2004) *Org Biomol Chem* 2:651–654
31. Becker M, Zittlau E, Petz M (2004) *Anal Chim Acta* 520:19–32
32. Vincent U, Chedin M, Yasar S, von Holst C (2008) *J Pharm Biomed Anal* 47:750–757
33. Benito-Peña E, Partal-Rodera AI, León-González ME, Moreno-Bondi MC (2006) *Anal Chim Acta* 556:415–422
34. Benito-Peña E, Urraca JL, Moreno-Bondi MC (2009) *J Pharm Biomed Anal* 49:289–294
35. Gamba V, Dusi G (2003) *Anal Chim Acta* 483:69–72
36. Fuselier R, Juhel-Gaugain M, Gaudin V, Sanders P (2004) Proficiency testing for control of antibiotic residues in milk. AFSSA 2004/UQ
37. Berendsen BJA, Zuidema T (2007) Proficiency study for penicillins in porcine tissues, RIKILT report 2007.007. <http://library.wur.nl/way/bestanden/clc/1844975.pdf>