



Determination of aflatoxin M₁ and deoxynivalenol biomarkers in infants and children urines from Bangladesh

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Abstract

The mycotoxins aflatoxin B₁ (AFB₁) and deoxynivalenol (DON) are found worldwide in crops and dietary staples. The prevalence and levels of these contaminants can vary greatly, and data in Bangladeshi food commodities are scarce. To characterize human exposure, we have conducted biomonitoring, analyzing AFM₁ (a metabolite of AFB₁) and DON levels in urines of adult cohorts in Bangladesh. Yet, AFM₁ and DON occurrence has not been studied in the very young population of this country. Thus, the same methods, HPLC-FD for AFM₁ and LC-MS/MS for DON analysis, were now applied to determine these biomarkers in urines of infants ($n=49$) and young children ($n=105$) in Rajshahi and Dhaka district. Overall, AFM₁ and DON detection frequency was 43.5% and 33.4%, with 34.7% and 11.5% in infant and 47.6% and 39.4% in children urines, respectively. The mean AFM₁ levels in all infants (9.1 ± 14.3 , max 55.6 pg/mL) and children (8.8 ± 12.9 , max 75.3 pg/mL) were not significantly different. The AFM₁ mean level was slightly higher in Dhaka (9.4 ± 12.4) compared to Rajshahi (8.5 ± 13.9 pg/mL) district. The average DON level was about 2-fold higher in infant (3.8 ± 2.9 , max 6.8 ng/mL) than children urines (1.6 ± 1.8 , max 8.6 ng/mL), and higher in Rajshahi (2.1 ± 2.3 ng/mL) than Dhaka (1.4 ± 1.6 ng/mL) district. The biomarker-based estimated average daily DON intake (29.6 ± 108.3 ng/kg bw in infants and 36.4 ± 81.8 ng/kg bw in children) or the maximum exposure (560 ng/kg bw) do not exceed the current maximum provisional tolerable daily intake value of 1 µg/kg bw for DON, although DON exposure in infants and children is higher than that of Bangladeshi adults. The AFM₁ urine levels in young children are somewhat lower than those found previously in adult cohorts in Bangladesh, but the frequent detection of this biomarker for AFB₁ exposure raises further concerns, also for this vulnerable part of the population. Therefore, continuous surveillance for aflatoxins in Bangladeshi food commodities is clearly required, first to identify major sources of intake and then to reduce exposure.

Keywords Aflatoxins; deoxynivalenol; biomarkers · Infant and children · Bangladesh

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Introduction

Aflatoxins and deoxynivalenol, secondary fungal metabolites produced by various *Aspergillus* and *Fusarium* species, are important contaminants of food commodities including dietary staples (EFSA 2017, 2020; Rushing and Selim 2019; Mishra et al. 2020). Mycotoxin exposure cannot be completely avoided, but it is essential to protect the population against acute and chronic effects. To limit exposures appropriate regulatory standards¹ for these food contaminants are set that consider the hazardous properties of a given mycotoxin and its occurrence.

Aflatoxin B₁ (AFB₁), the most potent mycotoxin, exerts strong hepatotoxic and carcinogenic activity in several animal species. Exposure to aflatoxins, mainly AFB₁, has been implicated in severe diseases in some parts of Africa and in Southeast Asia (Wild and Turner 2002; Williams et al. 2004; Groopman et al. 2008). Epidemiological studies have demonstrated a strong correlation between chronic AFB₁ exposure and risk of developing hepatocellular carcinoma, alone or in tandem with hepatitis B virus infection (IARC 2012; Liu and Wu 2010; Sun et al. 2013). In addition, chronic exposure to aflatoxins has been linked to growth impairment (Gong et al. 2004; Turner 2013) and immune suppression in children (Turner et al. 2003). For aflatoxins known to act as mutagenic carcinogens, maximal levels are set, most strictly for infant food, and enforced in developed countries to minimize exposure of the population (van Egmond et al. 2007; Escola et al. 2019). But, in many developing countries, even when such regulation exists on paper, there are problems to achieve this goal, as described elsewhere in detail (IARC 2015).

Deoxynivalenol (DON) exposure of animals results in a number of adverse effects, including gastroenteritis, growth inhibition, and immunologic dysregulation (Pestka 2010; Alizadeh et al. 2015). Although evidence for health effects in humans related to chronic DON exposure is lacking, given the adverse effects in animals and its frequent occurrence as food contaminant, human exposure to DON is considered as a significant food safety issue (Sudakin 2003; Mishra et al. 2020). Moreover, recent food and/or biomarker-based assessments found that the mean attributed dietary exposure of children and adolescents often exceeds the tolerable daily intake of 1 µg/kg bw. set for DON and its modified forms (JECFA 2011; EFSA 2017).

The prevalence and levels of mycotoxin contamination are known to vary greatly between types of crops, regions, and season. This and variable dietary habits in different regions of the world make exposure assessments for aflatoxins and DON a rather complex task (FAO/WHO 2018; JECFA 2011). In the last decades, biological monitoring has been established as complementary approach to characterize human exposure to mycotoxins, early on for aflatoxins, then for other mycotoxins. The analysis of suitable biomarkers (parent compounds and/or metabolites) has a key role in investigating health concerns related to mycotoxin exposure (Turner et al. 2012), and biomarker analysis in human body fluids covers mycotoxin intake from all dietary sources and exposure by other routes (Degen 2011).

A recent review on biomarker results in human samples (blood, urine, breast milk) documents the variable patterns of mycotoxin exposure in different parts of the world (Al-Jaal et al. 2019). For the developing country Bangladesh data on contaminant levels in food commodities are scarce (Dawlatana et al. 2002; Bhuyian et al. 2013; Roy et al. 2013), and regulatory standards for aflatoxins were only recently established (BFSA 2017). In this context, biomonitoring can provide useful insights into mycotoxin exposure of the Bangladeshi population. Thus, we have conducted biomarker analysis in urines from adult residents of urban and rural areas in Raishahi and in Dhaka district: the results indicate low exposure to the *Fusarium* toxin DON (Ali et al. 2015a, b, 2016a). But AFM₁, a biomarker of exposure to the *Aspergillus* toxin AFB₁, has been detected in many of the urines in all adult cohorts, and at significant levels which raise health concerns (Ali et al. 2016b, 2017).

However, little is known so far about mycotoxin exposure in Bangladeshi children: two studies investigated AFB₁ exposure in young children, one in a rural site in the North of the country, the other in an urban slum in Dhaka (Groopman et al. 2014; Mahfuz et al. 2019). Both have analyzed AFB₁-lysine albumin in blood, a biomarker which integrates exposure during the course of several weeks. Yet, blood sampling requires medical personal whilst sampling of urine is noninvasive, easier to perform in field studies. Analysis of urinary AFM₁ has been used in many studies as biomarker of recent AFB₁ exposure (e.g. Polychronaki et al. 2008; Mitchell et al. 2013; Ayelign et al. 2017; Chen et al. 2018; Ezekiel et al. 2018). Urine is widely used for biomonitoring of mycotoxins which are readily excreted, for example DON: it is found in human urine as parent compound (free DON), but a far larger part are DON-glucuronides (DON-GlcA), mostly DON-15-GlcA and DON-3-GlcA (Turner et al. 2008, 2011; Brera et al. 2015). Many studies therefore apply enzymatic hydrolysis of the conjugated forms and determine 'total DON' (sum of free DON and DON-GlcA) as biomarker of exposure (e.g. Turner et al. 2008, 2010, 2011; Ali et al. 2015b, 2016a; Brera et al. 2015; Papageorgiou et al. 2018;

¹ Scientific bodies evaluate dose–response relationships for the most critical effect observed in animal studies. Then considering also mode of action they derive tolerable daily intake values by a margin of safety approach (e.g. for DON) whereas a margin of exposure approach is used for mutagenic carcinogens like aflatoxins (details in FAO/WHO 2018; JECFA 2011; EFSA 2017; EFSA 2020).

Sarkanj et al. 2018; Wang et al. 2019). The metabolite de-epoxy-DON or DOM-1 is also found in human samples, yet less frequently, and at far lower levels than DON. Although DOM-1 is not a biomarker of exposure, an analysis of this metabolite is of some interest as indicator for detoxication of DON by the gut microbiome (Ali et al. 2016a; Wang et al. 2019).

The present study aimed to assess AFB₁ and DON exposure in infants and children, vulnerable groups in the population, using the same methods for biomarker analysis as applied previously for adult cohorts in two districts of Bangladesh. As young children are known to ingest more food than adults on a kg body weight basis, the intake of contaminants may be higher. Moreover, as both AFB₁ and DON may exert adverse effects on growth and immune function, co-exposure to these mycotoxins is also of interest.

Methods

Chemicals and reagents

Methanol (LC–MS gradient grade) was from Merck (Darmstadt, Germany). HPLC grade methanol and acetonitrile were purchased from Promochem (Wesel, Germany). Standard for AFM₁ solution was from Sigma-Aldrich (Taufkirchen, Germany). Isotope labeled standard ([¹³C₁₅] DON), deoxynivalenol (DON) and de-epoxy DON (DOM-1) were obtained from Romer Labs Diagnostics GmbH (Tulln, Austria). The β -glucuronidase/arylsulfatase from *Helix pomatia* (with specific activity 5.5 U/mL β -glucuronidase, 2.6 U/mL arylsulfatase at 37 °C) was purchased from Roche Diagnostics (Mannheim, Germany) and used with 10-fold hydrolysis buffer (13.6 g sodium acetate hydrate, 1.0 g ascorbic acid, 0.1 g EDTA in 100 mL deionised water, adjusted to pH 5.0 with acetic acid 96%) for enzymatic pretreatment of urines. Immunoaffinity columns AflaTest[®] WB^{SR} and DONTTest[™] (Vicom[®], from Ruttmann, Hamburg, Germany) were used for sample clean-up and enrichment of the target analytes.

Study areas and study subjects

In total, 154 urine samples were collected from Bangladeshi infants and children in Rajshahi (33 infants and 55 children) and Dhaka (16 infants and 50 children) district. Urine samples were collected between January and February 2014, a winter period in Bangladesh. In Rajshahi district, urines were obtained in rural areas (Mongol Para, Bhatpara, Habibpur and Jahubona) under Puthia Upazila. In Dhaka district, urines were obtained in rural and suburban areas (Nalam and Dhamsona) under Savar Upazila. Infants aged 1–12 months and children aged 1–6 years

were included in the study if they were in good health. Demographic (age, sex) and anthropometric (height, weight) data were recorded in a brief questionnaire form. Parents or guardians were informed about the study aims and signed the consent form on behalf the participants. Urine collection containers (pots of 30 mL) and written instructions were given to the participants (parents/guardians) before the day of sample collection. On the next day, morning urine samples were collected from the participant's house. Some urine samples for which information was incomplete were excluded from the study. The collected urine samples were first stored at –20 °C at the laboratory of Biochemistry Department of Gonoshasthaya Samaj Vittik Medical College, Dhaka and sent on dry ice to IfADo, Dortmund for subsequent analysis. The Institute of Biological Sciences of Rajshahi University, Bangladesh and the institutional Internal Review Board of IfADo approved the study.

Sample preparation

Sample preparation of all urines for AFM₁ biomarker analysis was done as described earlier (Ali et al. 2017). In brief, after centrifugation, 5 mL urine aliquots were adjusted to a pH between 5.5 and 7.0 with 1 N hydrochloric acid or 1 M sodium hydroxide. Then the urine was loaded on a AflaTest[®] WB^{SR} column at a flow rate of 1 drop/s. The columns were washed twice with 5 mL of distilled H₂O, then AFM₁ was eluted (flow rate 1 drop/s) with 2 mL of methanol. Then eluates were evaporated to dryness under a stream of nitrogen at 45 °C, and the residue was reconstituted in 250 μ L of acetonitrile/water (25:75). Thus, the analyte enrichment factor was 20.

Due to limited volumes available, the remaining 120 urines (of 26 infants and 94 children) were prepared for DON biomarker analysis. Urine clean-up and enrichment of analytes were done by a slight modification of the procedure used previously (Ali et al. 2015a, b, 2016a). Briefly, 1.5 mL of each urine aliquot was hydrolyzed to cleave DON and DOM-1 conjugates by adding 125 μ L of hydrolysis buffer and 20 μ L of β -Gluc/ArylS enzyme and incubated overnight at 37 °C before sample extraction by immunoaffinity columns. Each column was rinsed with 1 mL of water and the hydrolyzed urine sample was loaded onto a DONTTest[™] column at a flow rate of 1 drop/sec. Then, the column was washed with 3 mL of distilled water and aglycone analytes were eluted (flow rate 1 drop/sec) from the column with 2 mL methanol. Elutes were evaporated to dryness under a stream of nitrogen at 45 °C; the residues were dissolved in 250 μ L water/methanol (90:10), vortexed and filtered through 0.45 μ m pore size PTFE syringe filters before LC–MS/MS analysis. Thus, the enrichment factor was 6.

Biomarker analysis

AFM₁ was determined in urine by HPLC-FD following our previously established method (Ali et al. 2017) on an HPLC Shimadzu system consisting of two LC-10AS pumps, RF-10Ax1 fluorescence detector, SIL-10AD, Vp auto injector, CBM-20A communication module, and Shimadzu LC solution software. A C₁₈ Microsorb-MV100 column (150 × 4.6 mm, 5 μm, from Agilent Technologies, Waldbronn, Germany) fitted with a C₁₈ Metaguard column (10 × 4.6 mm, Microsorb A104MG) was used. The injection volume was 80 μL, and chromatographic separation was achieved by isocratic elution with mobile phase 25% acetonitrile and 75% water at a column temperature of 25 °C and a flow rate of 1 mL/min. The fluorescence detector was set at 360 nm excitation and 440 nm emission wavelengths; the retention time of AFM₁ was 7.6 min. The limit of detection (LOD) was 1.7 pg/mL and limit of quantification (LOQ) was 5 pg/mL for AFM₁. Recovery of the analyte from urine was about 90%.

Urinary levels of DON and its metabolite DOM-1 were determined by liquid chromatography with tandem mass spectrometry with a previously in-house validated method (Ali et al. 2015b, 2016a). In brief, chromatographic separation was carried out at 25 °C on a Nucleosil® C₁₈ column (100–5 material, 125 × 3 mm) with water (mobile phase A) and methanol (mobile phase B) as eluents. LC–MS/MS analysis was done on a Varian 1200-L Quadrupole MS/MS equipped with an electrospray ionization (ESI) source and a Prostar® Varian HPLC system and Varian MS Workstation. DON was monitored by the transitions of *m/z* 295.1 → 265.1 and 295.1 → 138.1 and DOM-1 of *m/z* 279.1 → 248.9 and 279.1 → 231.1. The isotope labeled internal standard ([¹³C₁₅] DON) was used in all quantification steps. The LODs were 0.16 ng/mL and 0.10 ng/mL for DON and DOM-1, and LOQs were 0.30 ng/mL and 0.20 ng/mL for DON and DOM-1. Recoveries for DON and DOM-1 from urine were about 92% and 85%, respectively.

Creatinine analysis

Urinary creatinine levels were measured by a modified Jaffe method on a 96 well plate reader from TecanGenios (Blaszkevicz and Liesenhoff-Henze 2012) to account for variability in urine dilution between individual samples. Biomarker levels determined in pg/mL (for AFM₁) or ng/mL (for DON) were adjusted for creatinine in urines and their concentrations expressed as pg/mg creatinine or ng/mg creatinine, respectively, to facilitate the comparison with some biomarker data published previously.

Exposure assessment

The dietary DON intake was estimated based on results of the urinary DON analysis. The following equation was used to assess the probable daily intake (PDI) of DON among the participants

$$\text{PDI} \left(\frac{\mu\text{g}}{\text{kg}} \text{Body weight} \right) = \frac{C \times V \times 100}{W \times E}$$

With *C* = biomarker concentration (DON μg/L), *V* = daily urine excretion (L), *W* = body weight (kg) and *E* = excretion rate (%). In the calculation, 24 h urinary output was assumed to be 0.5 L for children aged up to 6 years (Gong et al. 2015; Wang et al. 2019). The daily urinary DON excretion rate of 68% (Warth et al. 2013) was used, a value slightly lower than that used by others (Turner et al. 2010). The DON intake estimates (PDI) were then compared to the provisional tolerable daily intake (PMTDI) value of 1 μg/kg bw set by scientific advisory committees (JECFA 2011; EFSA 2017) to assess the risk of DON exposure.

Statistical analysis

The software IBM SPSS version 23 was used to analyses the data. Descriptive analysis was done to determine mean, median and interquartile ranges of the analytes. Urines containing the analyte levels ≥ LOD were used in determining the mean and median values. Differences in biomarker concentrations between the infant and children cohorts, or regions were analyzed by independent sample t-test. Pearson's correlation coefficient test (two-tailed) was applied to evaluate the correlation between biomarker levels with age and anthropometric variables. The box plot represents the distribution of central data where upper and lower limits of the box indicate 25th and 75th percentiles, respectively, and the line inside the box indicate the median value. The *p*-value lower than 0.05 is considered statistically significant.

Results

Characteristics of the study subjects

The basic characteristics of the study subjects are shown in Table 1. In total, 49 infants (35 males and 14 females) and 105 children (59 males and 46 females) were enrolled in the present study. The mean ages were 7.1 ± 3.7 and 37.5 ± 16.5 months for infants and children, respectively. The average height and weight were 62.4 ± 8.1 cm and 7.4 ± 2.2 kg, respectively for infants. In children, the average height and weight were 87.5 ± 13.9 cm and 12.8 ± 3.5 kg,

Table 1 Baseline characteristics of the urine donors

| | All | Rajshahi | Dhaka |
|--------------------|--------------|-------------|-------------|
| <i>N</i> | 154 | 88 | 66 |
| Gender of infants | | | |
| Male | 35 | 26 | 9 |
| Female | 14 | 7 | 7 |
| Gender of children | | | |
| Male | 59 | 31 | 28 |
| Female | 46 | 24 | 22 |
| Age (months) | | | |
| Infants | 7.1 ± 3.7 | 7.1 ± 4.2 | 7.2 ± 2.9 |
| Children | 37.5 ± 16.5 | 39.3 ± 19.8 | 35.5 ± 11.8 |
| Height (cm) | | | |
| Infants | 62.4 ± 8.1 | 62.0 ± 9.0 | 63.1 ± 5.9 |
| Children | 87.5 ± 13.9 | 88.5 ± 15.3 | 86.3 ± 12.2 |
| Weight (kg) | | | |
| Infants | 7.4 ± 2.2 | 6.9 ± 2.3 | 8.4 ± 1.6 |
| Children | 12.8 ± 3.5 | 12.9 ± 4.0 | 12.6 ± 2.8 |
| Creatinine (g/L) | | | |
| Infants | 0.13 ± 0.12 | 0.14 ± 0.14 | 0.12 ± 0.06 |
| Children | 0.43 ± 0.29* | 0.46 ± 0.30 | 0.39 ± 0.27 |

Value given as Mean ± SD

* $p < 0.01$ when compared to infant cohort. p -value obtained from independent sample t test

respectively. The mean creatinine concentration in children urines (0.43 ± 0.29 g/L) was significantly ($p < 0.01$) higher than in the infant (0.13 ± 0.12 g/L) urines.

AFM₁ and DON biomarker levels in urine samples

Results of our biomarker analysis in infants and children are given in Table 2 as non-adjusted and creatinine-adjusted urinary concentrations of AFM₁ and DON. The overall AFM₁ detection frequency was 43.5%, with 34.7% in infants and 47.6% in children urines from two regions. There was no significant difference in the mean level of AFM₁ between all infants (9.1 ± 14.3 , max 55.6 pg/mL) and children (8.8 ± 12.9 , max 75.3 pg/mL) urines. When comparing region, the mean concentration of urinary AFM₁ in all (infant and children) samples was slightly higher in Dhaka (15.4 ± 19.5 pg/mL) than in Rajshahi (3.6 ± 1.4 pg/mL) district ($p < 0.05$), but not significant for children (mean 10.1 ± 15.8 in Dhaka and 7.4 ± 8.4 pg/mL in Rajshahi). Yet, the inter-individual variability of AFM₁ in all infant and children groups is high, as depicted in Fig. 1 (left panel).

The prevalence of the other mycotoxin biomarker was rather low: DON was detected in 11.5% of all infants and in

Table 2 Urinary levels of and AFM₁ and DON in infants and children in Rajshahi and Dhaka district

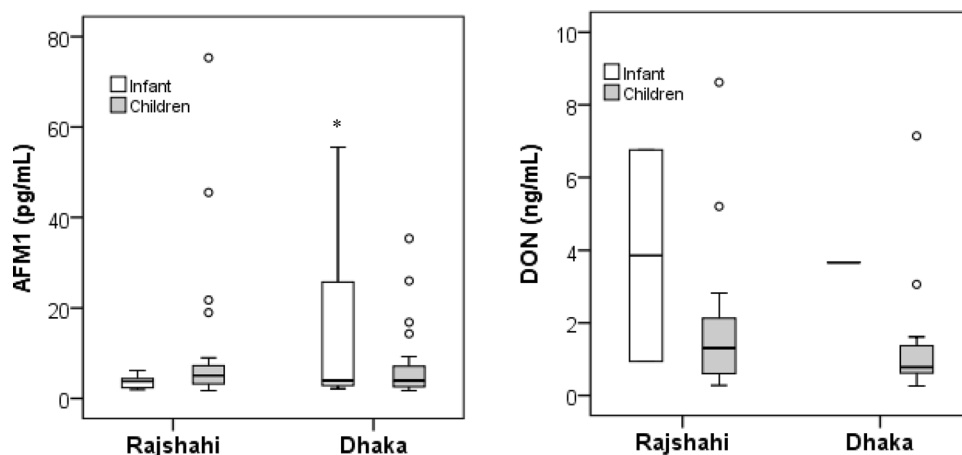
| Region | Category | AFM ₁ | | DON [#] | | | | |
|--------------|----------|------------------|-------------------|------------------------|-------------------------|-------------------|------------------------|------------------------|
| | | <i>N</i> | Mean ± SD (pg/mL) | Median (range) (pg/mL) | Positive <i>n</i> , (%) | Mean ± SD (ng/mL) | Median (range) (ng/mL) | Mean ± SD (ng/mg crea) |
| Rajshahi | Infants | 33 | 3.6 ± 1.4 | 3.8 (1.9–6.1) | 9 (27.3) | 43.1 ± 40.7 | 3.8 (0.9–6.8) | 16.6 ± 5.9 |
| | Children | 55 | 10.1 ± 15.8 | 5.1 (1.8–75.3) | 27 (49.1) | 25.3 ± 29.4 | 1.3 (0.3–8.6) | 4.6 ± 3.9 |
| | All | 88 | 8.5 ± 13.9 | 4.5 (1.8–75.3) | 36 (40.9) | 29.7 ± 32.9 | 1.3 (0.3–8.6) | 5.8 ± 6.51 |
| Dhaka | Infants | 16 | 15.4 ± 19.5* | 4.0 (2.2–55.6) | 8 (50.0) | 97.8 ± 131.4 | 3.67 | 27.2 |
| | Children | 50 | 7.4 ± 8.4 | 3.9 (1.7–35.4) | 23 (46.0) | 22.1 ± 28.8 | 0.8 (0.3–7.1) | 3.4 ± 2.9 |
| | All | 66 | 9.4 ± 12.4 | 3.9 (1.7–55.6) | 31 (47.0) | 41.6 ± 75.9 | 0.8 (0.3–7.1) | 4.6 ± 5.9 |
| Both-regions | Infants | 49 | 9.1 ± 14.3 | 3.8 (1.9–55.6) | 17 (34.7) | 68.8 ± 95.8 | 3.67 (0.9–6.8) | 20.1 ± 12.8 |
| | Children | 105 | 8.8 ± 12.9 | 4.8 (1.7–75.3) | 50 (47.6) | 23.8 ± 28.9 | 1.0 (0.3–8.6) | 3.9 ± 2.5 |
| | All | 154 | 8.9 ± 13.1 | 4.4 (1.7–75.3) | 67 (43.5) | 35.2 ± 56.9 | 1.0 (0.3–8.6) | 5.2 ± 6.2 |

Positive sample refer to urines containing the analyte \geq LOD (LOD: 1.7 pg/mL for AFM₁ and 0.16 ng/mL for DON). Only positive samples were considered during calculation of mean and median values

* $p < 0.05$ when compared to infant cohort in Rajshahi district. p -value obtained from independent sample t -test

[#]Due to limited urine volumes, 'total DON' was analyzed in 120 urines (infants = 26 and children 94) whilst AFM₁ analysis included all 154 samples

Fig. 1 Box plots for urine levels of AFM₁ and DON in infants and children from two regions in Bangladesh. Only positive samples (analyte \geq LOD) are included in the graph. * $p < 0.05$ when AFM₁ level in infant cohort of Dhaka district is compared to Rajshahi district. p -value is obtained from independent sample t -test



39.4% of all children urines, whilst DON-1 was not detectable in any of the samples. The average DON concentration was about 2-fold higher in infants (3.8 ± 2.9 , max 6.8 ng/mL) than in children (1.6 ± 1.8 , max 8.6 ng/mL) urines. As for region comparison, the mean urinary DON level was higher in Rajshahi (2.1 ± 2.3 ng/mL) than in Dhaka (1.4 ± 1.6 ng/mL) district samples. But, differences among groups were not statistically significant, also due to considerable inter-individual variability in biomarker levels (see Fig. 1, right panel).

Estimated dietary DON intake based on urinary analysis

The probable daily DON intake was calculated for the study subjects based on individual data of urine biomarker analysis and some additional parameters (see Methods section). In the entire study cohort, the mean daily DON intake was 34.8 ± 88.1 ng/kg bw, with 29.6 ± 108.3 ng/kg bw in infants and 36.4 ± 81.8 ng/kg bw in children in both regions (Table 3). Participants in Rajshahi district had a slightly higher calculated daily DON intake (36.5 ± 102.8 ng/kg bw) than those in Dhaka district (32.5 ± 63.0 ng/kg bw), and the highest DON intake reached 560 ng/kg bw. But, none of the subjects had an estimated DON intake that exceeds the provisional maximal tolerable daily intake (PMTDI) of 1 μ g/kg bw set by scientific committees (JECFA 2011; EFSA 2017) for DON and its modified forms.

Discussion

Data on the contamination of food commodities with *Aspergillus*, *Penicillium* and *Fusarium* mycotoxins are rather scarce for Bangladesh. To gain some insight into potential risks related to these dietary contaminants, we have previously investigated the occurrence of biomarkers of exposure

to major mycotoxins in the adult population of this country (Ali et al. 2014, 2015a, 2016a, 2016b, 2019; b; Gerding et al. 2015). The present study is aimed to explore the exposure of infants and young children to AFB₁ and DON in Bangladesh. Children are considered as vulnerable group with increased susceptibility to chemicals, including mycotoxins (Makri et al. 2004; Sherif et al. 2009; Lombard et al. 2014). As outlined in the Introduction, the toxic properties of AFB₁ and DON are quite different, the first being primarily known as potent mutagenic carcinogen, and DON (vomitoxin) for adverse effects in the gastrointestinal tract, reduced weight gain and impaired immune function. Child stunting is an emerging topic in the field of aflatoxin-related health outcomes (IARC 2015; FAO/WHO 2018; EFSA 2020); DON exposure may exacerbate this condition, independent of and along with other risk factors (Lombard et al. 2014).

The results of our biomarker analysis, the first for DON in Bangladeshi infants and children, now indicate moderate prevalence of exposure to the trichothecene mycotoxin,

Table 3 Provisional daily intake (PDI) of DON (ng/kg bw)* among the cohort

| Region | Category | N | Mean \pm SD | Maximum |
|--------------|----------|-----|--------------------|---------|
| Rajshahi | Infants | 22 | 27.78 \pm 114.73 | 536.51 |
| | Children | 49 | 40.42 \pm 98.10 | 559.74 |
| | All | 71 | 36.50 \pm 102.87 | 559.74 |
| Dhaka | Infants | 4 | 36.41 \pm 89.18 | 218.45 |
| | Children | 45 | 31.99 \pm 60.08 | 340.00 |
| | All | 49 | 32.51 \pm 63.03 | 340.00 |
| Both regions | Infants | 26 | 29.62 \pm 108.28 | 536.51 |
| | Children | 94 | 36.38 \pm 81.81 | 559.74 |
| | All | 120 | 34.83 \pm 88.14 | 559.74 |

*Dietary DON intake was calculated based on urinary DON levels, adjusted for 24 h urine volume, assuming an 68% excretion rate and individual body weight (see methods section for details). Only positive samples were considered in PDI calculation

whilst the prevalence of AFM₁ in their urines indicates quite frequent intake of AFB₁ in this vulnerable part of the population (Table 2). Before discussing implications of the new results and possible sources of dietary intake, a remark is appropriate on the approach used to 'translate' biomarker data to exposure-related risks: A high renal excretion rate (68–70% within a day; Warth et al. 2013, Turner et al. 2010) enables biomarker-based estimates for DON exposure, and these values are then compared to the tolerable daily intake value set for this mycotoxin (see Table 3). However, only a small fraction of AFB₁ is excreted as AFM₁ in urine (1.5–2%; Zhu et al. 1987) which hampers a reliable back-calculation of biomarker levels to dietary AFB₁ intake. Yet, one can compare new data on prevalence and urinary levels of AFM₁ to results reported from other regions of the world where it served to investigate aflatoxin exposure of children.

Regarding DON biomarker analysis in urine, the detection frequency in the present study (33.3%) is close to that determined in the adult cohort in Rajshahi district (27% in summer and 31% in winter) and a pregnant women cohort in Dhaka (52%) district (Ali et al. 2015b, 2016a). But, the DON concentration in infants and children (mean 1.7, max 8.6) is higher than that found in the adults (mean 0.17, max 1.78 ng/mL in summer and mean 0.16, max 1.21 in winter) and in pregnant women (mean 0.86, max 7.16). The higher DON exposure of young children can be related to higher food consumption per kg body weight than adults and/or children preferring foods such as breads and cookies made from wheat which are more likely contaminated with DON than the typical staple food rice. Yet, no individual of the Bangladesh low age groups exceeds the tolerable daily intake value set for DON (Table 3).

It is also of interest to compare DON biomarker levels in this study to data reported in children from some other countries (Table 4). Exposure to DON in Bangladesh is clearly lower than in two regions of China where 10–73% of the cohorts exceed the TDI (Wang et al. 2019), or in European cohorts: Belgium with 69% above TDI (Heyndrickx et al. 2015), Italy with 25–27.5% above TDI (De Santis et al. 2019), Norway with 20% above TDI (Brera et al. 2015), and the UK, with 33–63% above the TDI (Papageorgiou et al. 2018). In Africa, children in Cameroon have apparently lower DON exposure (Ediage et al. 2013) than those in Tanzania geometric mean 2.5 ng/mL, with 21–54% above TDI in one (Srey et al. 2014) or more in another study (Gong et al. 2015). DON has been detected now in urines of breastfed and non-exclusively breastfed infants from Nigeria (Ezekiel et al. 2020). DON was also found in urines from children and adults in Haiti (Gerding et al. 2015), at levels similar to those found in Swedish children (Mitropoulou et al. 2018). DOM-1, a detoxication product of DON, was not detected in our participants, and also not found in children samples from

Italy or the UK, but in some urines of Norwegian children (Brera et al. 2015).

These biomonitoring studies (Table 4) show a wide range of DON exposures in the pediatric population of different countries, but also differences between regions of a country, as in China (Wang et al. 2019). The extent of DON exposure can be explained in part by differences in dietary habits: food items such as wheat bread, pasta, breakfast cereals, bran rolled flakes and baked goods are major sources of DON exposure in European populations (Brera et al. 2015; EFSA 2017), but these foods are far less often consumed in Bangladesh. Biomarker levels not only differ between countries, but also between years and season (Gratz et al. 2014; Ali et al. 2016a). This reflects the quite variable DON contamination of many crops worldwide (Mishra et al. 2020). Thus, one should keep in mind that biomonitoring sheds a light on the DON exposure in a given cohort and sampling season, and should be followed up, in particular when data indicate exceedance of TDI-values in vulnerable groups (Papageorgiou et al. 2018).

Regarding AFM₁ analysis, the biomarker has been frequently detected (43.5% in the entire cohort), with 34.7% in infants and 47.6% in children urines from two regions (Table 2), and with a high inter-individual variability in AFM₁ levels of all groups (Fig. 1). The new results are first compared with data in Bangladeshi adults and then with findings for children of other countries. The AFM₁ urine concentrations in infants (mean 9.1, max 55.6 pg/mL) and children (mean 8.8, max 75.3 pg/mL) are somewhat lower than those measured in the adult (13.5 pg/mL, max 104 pg/mL in summer and 27.7 pg/mL, max 189.9 pg/mL in winter) and pregnant women (13.9 pg/mL, max 141.5 pg/mL) cohorts of this country (Ali et al. 2017). These biomarker data suggest that foods consumed by adults and by children contain notable levels of aflatoxin B₁, whilst the sources of mycotoxin intake may differ between both groups.

Urine AFM₁ levels in young Bangladeshi children are far lower than levels found in children of some African countries (Table 5), i.e. in Cameroon (Ediage et al. 2013), Guinea (Polychronaki et al. 2008), Nigeria (Ezekiel et al. 2020, 2018; Sarkanj et al. 2018), Sierra Leone (Jonsyn-Ellis 2001) and Tanzania (Chen et al. 2018). However, the AFM₁ mean level in our cohort is higher than that reported in children urines in Egypt (Polychronaki et al. 2008), and it approaches levels found in Ethiopia (Ayelign et al. 2017). Also, the average Bangladeshi values are in a similar range as those found in children in Colombia, South America (Sanchez and Diaz 2019). The children data (Table 5), and a previous overview of AFM₁ biomarker data for adult cohorts of different continents and countries (Ali et al. 2017) illustrate the wide range of exposure to AFB₁. Again, exposure reflects different dietary habits in these populations and also different degrees of aflatoxin contamination in the crops and foods

Table 4 DON biomarker levels in urines from some children cohorts in different countries

| Country, cohort | Positive <i>n</i> (%) | Mean (range) ng/mL | ng/mg creatinine | % exceeding TDI | Method | LOD/LOQ (ng/mL) | Reference |
|-----------------------------|--------------------------|-----------------------|------------------|-----------------|-------------------------|--------------------|---------------------------|
| Bangladesh | | | | | | | |
| Infants | 3/26 (11.5) | 3.8 (0.9–6.8) | 20.1 | 0 | LC–MS/MS ^a | 0.16/0.30 | Ali et al., present study |
| young children | 37/94 (39.3) | 1.6 (0.3–8.6) | 3.9 | 0 | LC–MS/MS ^a | 0.16/0.30 | |
| Belgium, children | 109/155 (70) | 5.2 (0.5–32.5) | 5.5 | 69 | LC–MS/MS ^b | 0.2/0.5 | Heyndrickx et al. 2015 |
| Cameroon, children | 160/220 (73) | 2.22 ^{gm} | na | Na | LC–MS/MS ^b | 0.04/NS | Ediage et al. 2013 |
| China, young children* | | | | | | | |
| Henan | 35/35 (100) | 55.7 (max 224.1) | na | 74.3 | LC–MS/MS ^a | 0.5/1.0 | Wang et al. 2019 |
| Sichuan | 28/30 (93) | 10.1 (max 56.3) | na | 10.0 | | | |
| Haiti, adults and children | 24/142 (17) | 3.2 (<LOQ–16.9) | 3.6 ± na | Na | LC–MS/MS ^b | 0.4/4.0 | Gerding et al., 2015 |
| Italy, children (3–9 years) | | | | | | | |
| Day 1 | 37/40 (93) | 10.8 (1.2–138) | 12.9 | 25 | LC–MS/MS ^a | 0.25/0.50 | De Santis et al. 2019 |
| Day 2 | 37/40 (93) | 11.3 (1.4–140.9) | 14.9 | 27.5 | | | |
| Nigeria, infants | | | | | | | |
| Exclusively breastfed | 7/23 (30) | 3.19 (0.22–19.78) | na | Na | UPLC–MS/MS ^a | 0.05/0.15 | Ezekiel et al. 2020 |
| Non-exclusively breastfed | 23/42 (55) | 5.28 (0.23–21.34) | na | Na | | | |
| Norway children (3–9 years) | 39/40 (98) | 13.2 (1.6–86.9) | 8.2 (0–76.1) | 20 | LC.MS/MS ^a | 0.005/0.015 | Brera et al., 2015 |
| Sweden, children | 47/50 (94) | 3.9 (0.9–12.6) | na | 0 | LC–MS/MS ^b | NS/1.5 | Mitropoulou et al. 2018 |
| Tanzania, young children | 85/166 (51) | 2.5 ^{gm} | Na | 21–54 | LC–MS/MS ^a | 0.25/0.5 | Srey et al. 2014 |
| Tanzania, children | 48/50 (96) | 15.4 ^{gm} | 47.7 | Na | LC–MS/MS ^a | 0.25/0.5 | Gong et al. 2015 |
| UK, children (3–9 years) | 40/40 (100) | 29.2 (1.2–141) | 41.6 (5.3–219.0) | 33–63 | LC–MS(MS) ^a | 0.12/0.25 | Papageorgiou et al. 2018 |

na not available, NS not stated, gm geometric mean

*age group 1–6 years, 3 urines per child collected on 3 consecutive days

^aImmunoaffinity column clean up and tailored method

^bMulti-biomarker method

locally produced and consumed. For instance, in Africa maize and groundnuts continue to be the main sources of aflatoxin exposure (Gong et al. 2018; Xu et al. 2018).

So far there are only two studies on aflatoxin occurrence in food commodities in Bangladesh. Roy et al. (2013) analyzed AFB₁ in rice, lentils, wheat flour, dates, betelnut, red chilli powder, ginger and groundnuts: mean levels in 5 of these 8 commodities were above EU regulatory limits, and the highest levels were found in dates and groundnuts. Bhuiyan et al. (2013) analyzed total aflatoxins in maize, rice and wheat samples collected in all districts of Bangladesh at six times during a year: the highest incidence and level of contamination was found in maize; the incidence and contaminant level were lower, yet still significant in rice and in

wheat, all showing considerable seasonal variability. Both datasets have been used by others to calculate total aflatoxin levels in various food commodities in Bangladesh (Table 1 in Saha Turna and Wu 2019). Then, considering also average dietary consumption data for each of these items, they assessed aflatoxin exposure by dates, groundnuts, lentils, chili/spices, wheat, maize and rice (Table 2 in Saha Turna and Wu 2019). The highest contribution to total aflatoxin exposure was from rice. Rice is the main dietary staple in Bangladesh, and often consumed with curries prepared with several spices. Previously, we noted higher urine AFM₁ levels in adult people who consume more rice per day (Ali et al. 2017) which points also to rice as one important source of AFB₁ intake (Ali 2019). In their paper Saha Turna and

Table 5 AFM₁ level in urines from some children cohorts in different countries

| Country, cohort | Positive <i>n</i> (%) | Mean (range) pg/mL | pg/mg creatinine | Method | LOD/LOQ (pg/mL) | Reference |
|--|--------------------------|-------------------------------|------------------|-------------------------|--------------------|-----------------------------|
| Bangladesh, children | | | | | | |
| Infants | 17/49 (35) | 9.1 (1.9–55.6) | 68.8 | HPLC-FD ^a | 1.7/5.0 | Ali et al., present study |
| young children | 50/107 (48) | 8.8 (1.7–75.3) | 23.8 | | | |
| Cameroon, children | 31/220 (14) | 330 ^{gm} (< 10–4700) | na | LC–MS/MS ^b | 10/20 | Ediage et al. (2013) |
| Colombia, children | 40/96 (42) | 16 (LOD–48.5) | na | HPLC-FD ^a | 2/6 | Sanchez & Diaz (2019) |
| Egypt, children | 4/50 (8) | 5.5 (5.0–6.2) | na | HPLC-FD ^a | 5/- | Polychronaki et al. (2008) |
| Ethiopia, children | 14/200 (7) | 64 (63–70) | na | LC–MS/MS | 25/50 | Ayelign et al. (2017) |
| Guinea, children | 32/50 (64) | 97 (8.0–801) | na | HPLC-FD ^a | 5/- | Polychronaki et al. (2008) |
| Haiti, adults and children | | | | | | |
| Port-au-prince | 20/147 (14) | Na | 43.7 (3.97–202) | HPLC-FD ^a | 4/10 | Schwartzbord et al., (2016) |
| Quartier morin | 48/219 (22) | Na | 116 (2.44–775) | | | |
| Italy, adults and children | 3/52 (6) | 68 (20–146) | na | LC–MS/MS ^b | NS/20 | Solfrizzo et al. (2013) |
| Nigeria, children (2–7 years) | 13/13 (100) | 280 (110–510) | na | ELISA | 6/- | Ezekiel et al. (2018) |
| Nigeria, adults, adolescents and children | 17/120 (14) | 300 (LOD–1500) | na | LC–MS/MS ^b | 50/150 | Ezekiel et al. (2014) |
| Nigeria urines (reanalysis) | 87/120 (72.5) | 40 (1–620) | na | UPLC-MS/MS ^c | –/1 | Sarkanj et al. (2018) |
| Nigeria, infants | | | | | | |
| Exclusively breastfed | 11/23 (4) | 23 (23) | na | UPLC-MS/MS ^a | 0.0003/0.001 | Ezekiel et al. (2020) |
| Non-exclusively breastfed | 55/42 (12) | 166 (32–504) | na | | | |
| Sierra Leone, children | | | | | | |
| Dry season | 104/244 (43) | na (500–374,000) | na | HPLC | 5–50 | Jonsyn-Ellis (2001) |
| Rainy season | 97/190 (51) | na (100–124,000) | na | | | |
| Tanzania (6–14 months) in 3 villages; repeated visits | 72/84 (86%) | 36.5 ^{gm} (15–2840) | na | ELISA | 10–15 | Chen et al. (2018) |

na not available, NS not stated, gmgeometric mean

^aImmunoaffinity column clean up and tailored method

^bMulti-biomarker method ^cmulti-biomarker method with enzymatic hydrolysis and sample clean-up by SPE

Wu (2019) comment also on the high AFB₁ levels in dates, which are consumed mainly during Ramadan in Bangladesh and other Muslim countries. We suggest that along with rice, wheat-based bakery products, also dates may be a relevant source of AFB₁ intake as young children prefer sweet types of food. Furthermore, a recent screening of cow milk and milk products by ELISA reveals frequent occurrence of AFM₁ (Ali et al. unpublished results). As young children are fond of milk and milk based products, this may have also contributed to AFM₁ exposure in our cohort.

AFB₁ exposure has been investigated before in another region of Bangladesh (a rural area in Rangpur) by means of the AFB₁-lysine albumin adduct analysis in blood samples of pregnant women and later on in their 2-year-old children (Groopman et al. 2014). Median levels of this biomarker were 25.35 and 18.08 pg AFB₁-Lys/mg albumin in the first and third trimester, respectively, and 13.79 pg AFB₁-Lys/mg albumin in the children. These results, discussed by the authors in the context of biomarker data for cohorts in other countries, document rather high AFB₁ exposures of their

cohort in the Northwest of Bangladesh between 2008 and 2012. A recent longitudinal study in an urban slum in Dhaka city assessed AFB₁ exposure of children at the age of 7, 15, 24 and 36 months and reported a geometric mean of 1.07 pg AFB₁-Lys/mg albumin and a range of 0.04–123.5 pg AFB₁-Lys/mg albumin (Mahfuz et al. 2019). In this study, a reduction in breastfeeding prevalence, with concomitant introduction of family food, corresponded with an increase in AFB₁-lysine adduct detection at 36 months, and 62% of the children were exposed at the end of their 3rd year of life. Of interest is also the seasonal variation in AFB₁ biomarker prevalence, with the highest detection observed during and after the monsoon period which provides optimal conditions for fungal growth and aflatoxin contamination (Mahfuz et al. 2019).

Overall, the results of our study with determination of AFM₁ metabolite in urine, and the two studies (Groopman et al. 2014; Mahfuz et al. 2019) that measured AFB₁-lysine albumin adduct in blood plasma, document widespread exposure of young children in several parts of Bangladesh. This and

previous data on frequent AFB₁ exposure of adult and pregnant women cohorts (Ali et al. 2017; Groopman et al. 2014) raise concern with regard to dietary intake of this carcinogenic mycotoxin in the Bangladeshi population. Further efforts to analyse aflatoxin contamination are clearly needed to identify major sources of aflatoxin intake, and establish surveillance in food commodities with the aim to protect the population against long-term adverse health effects. Bangladesh has issued in mid 2017 regulation for aflatoxin contamination of certain food items, namely groundnuts, almonds, Brazil nuts, hazelnuts, pistachios and AFM₁ in milk (BFSA 2017). Yet, a recent risk assessment of aflatoxin-related liver cancer in Bangladesh concluded that the new regulations are unlikely to significantly reduce the risk of this cancer in the country (Saha Turna and Wu 2019). Indeed, considering food contaminant data available (vide supra), cereal-based commodities including rice, as well as pulses, spices and other items are likely to contribute far more to overall AFB₁ exposure than various types of nuts. Thus, we recommend to conduct regular surveys on aflatoxin contamination, at least in major staples (Ali 2019), and consider also further biomonitoring as this integrates human exposure from all sources.

Conclusion

This study applied sensitive biomonitoring methods to assess for the first time aflatoxin and DON exposure among infant and children cohorts in Bangladesh. DON exposure appears to be of low concern, with intake estimates below tolerable levels. But, the prevalence and levels of AFM₁ in infant and children urines indicate widespread contamination of the children's diets with the carcinogenic mycotoxin AFB₁, a finding which raises serious health concerns for this vulnerable population. Continuous surveillance of aflatoxins in Bangladeshi food commodities are urgently needed, in order to identify major sources of intake, and then take appropriate steps to further reduce risks from exposure-related adverse health effects.

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Compliance with ethical standards

Conflict of interest The authors have no conflict of interest to declare.

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