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# Effects of exogenous hydrogen sulfide on brain metabolism and early neurological function in rabbits after cardiac arrest

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CA. Inhalation of 80 ppm H<sub>2</sub>S significantly increased CEO<sub>2</sub>  $(25.04 \pm 7.11 \text{ vs. } 16.72 \pm 6.12 \%)$ and decreased AJVD(glu)  $(0.77 \pm 0.29 \text{ vs. } 1.18 \pm 0.38 \text{ mmol}/$ L) and lactate (5.11  $\pm$  0.43 vs.  $6.01 \pm 0.64$  mmol/L) at 30 min after resuscitation when compared with the CA group (all P < 0.05). In addition, neurologic deficit scores, viable neuron counts, and survival rate were significantly better whereas S100B was decreased after H<sub>2</sub>S inhalation. Conclusions: The present study reveals that inhalation of 80 ppm H<sub>2</sub>S reduced neurohistopathological damage and improves early neurological function after CA and resuscitation in rabbits. The increased CEO<sub>2</sub> and decreased AJVD(glu) and enhanced lactate clearance may be involved in the protective effects.

**Keywords** Hydrogen sulfide · Brain metabolism · Neurologic manifestations · Heart arrest · Resuscitation

# Introduction

Over the last decade, hydrogen sulfide (H<sub>2</sub>S) has been recognized as a gasotransmitter, playing an important role in brain function [1–3]. Endogenous H<sub>2</sub>S can be produced from cysteine by cystathionine  $\beta$ -synthase (CBS) in the central

nervous system and cystathionine  $\gamma$ -lyase (CSE) in the vessels and other tissues. Recently, higher levels of endogenous  $H_2S$  have been detected in the brain than in other organs. This suggests that  $H_2S$  may have a neuromodulatory role [3].

Some of the neuroprotective effects of  $H_2S$  can be attributed to hypometabolism and hypothermia. For

example, Blackstone et al. [1] observed that inhalation of H<sub>2</sub>S decreased the brain metabolic rate in mice by more than 50 % within 5 min. The metabolic rate is also associated with reductions in body temperature. However, the applicability of murine models of metabolism to larger animals has been questioned [4–6]. Furthermore, all the aforementioned studies focused on either whole-body metabolism or core temperature. However, changes in systemic metabolism, including oxygen extraction after cardiac arrest (CA), may vary more dramatically than brain metabolism [7]. To predict neurological outcomes, brain metabolism, especially cerebral extraction of oxygen (CEO<sub>2</sub>), must be taken into account to a greater extent than systemic metabolism [8, 9]. However, studies of brain metabolism and the effects of H<sub>2</sub>S on brain metabolism are rare. The effects of H<sub>2</sub>S on neurological function after CA and resuscitation remain poorly understood.

The aim of this study was to investigate the effects of inhaled exogenous  $H_2S$  on brain metabolism and neurological function in rabbits after CA and resuscitation.

# **Materials and methods**

For detailed materials and methods, see the electronic supplementary material (ESM).

# Animal surgical procedures

This study was approved by the Institutional Animal Care and Use Committee of Harbin Medical University in Harbin, Heilongjiang, China, and followed national guidelines for the treatment of animals. Adult male New Zealand white rabbits, weighing 2.5–3.0 kg, were used in this study. Food and water were available ad libitum until the morning of the experiment.

Anesthesia was performed with 1-2 % halothane and 30 % oxygen. Ventilation was controlled with a small animal ventilator (Kent Scientific, Litchfield, CT, USA) to maintain arterial pH at 7.35–7.45, PaCO<sub>2</sub> at 35–45 mmHg, and PaO<sub>2</sub> over 90 mmHg. All animals were mechanically ventilated except during the asphyxial CA procedures. Rectal temperatures were monitored continuously with thermal probes (BIOPAC Systems Inc., Santa Barbara, CA, USA) and the ambient temperature of 25 °C was maintained throughout the experiment. A standard lead II ECG was recorded continuously using subdermal needle-type electrodes placed in the limbs. All of the physiology data were monitored and recorded using the BIOPAC MP150 physiometer (BIOPAC Systems Inc., Santa Barbara, CA, USA).

Two saline-filled 18-gauge polyethylene catheters were inserted into the carotid artery for blood pressure

measurement and into the left femoral vein for infusion and drug administration. Another 18-gauge catheter was placed in the right internal jugular vein and advanced in the retrograde direction until increased resistance was met at the base of the skull. The catheter was then withdrawn a few millimeters and secured, after which the blood flowed readily. The position of the catheter tip was at the right internal jugular bulb and confirmed by autopsy in animals that failed resuscitative efforts.

After preparation and subsequent stabilization, the animals were randomly assigned to four groups: sham group, sham/H<sub>2</sub>S group, CA group, and CA/H<sub>2</sub>S group (details in ESM).

Cardiac arrest and resuscitation

Cardiac arrest was induced in CA and CA/H<sub>2</sub>S groups by asphyxia as previously described [10, 11]. Asphyxia was induced by administering an additional dose of vecuronium and clamping the endotracheal tube. The cardiac arrest period was considered to begin when MAP decreased to less than 10 mmHg and left untreated for 3 min. Total asphyxia time was approximately 8 min. A detailed description of this model including the exact resuscitative procedures is provided in the ESM.

Measurement of brain temperature

Two digital electronic thermometers with thermocouple probes were inserted for continuous monitoring of the cortex and hippocampus temperature (details in ESM).

# Determination of S100B level

Blood samples were draw from the internal jugular venous bulb and centrifuged for measuring the concentrations of S100B with a sandwich ELISA kit (details in ESM).

Neurological assessment and survival rate

All animals were evaluated before the experiment to ensure normal neurological function. Neurological dysfunction was evaluated daily in surviving animals with a clinical score previously validated in rabbits (0-10 %normal, 100 % brain death) [12]. Neurological dysfunction score (NDS) and the survival rates were assessed by the same investigator who was unaware of the group assignment during the 7 days after return of spontaneous circulation (ROSC).

#### Neuronal counts

On the 7th day after ROSC, the rabbits were deeply anesthetized with halothane. Hippocampi were removed and sectioned to 3  $\mu$ m for hematoxylin and eosin staining. The numbers of viable neurons were counted in the CA1 region per high-power field (×400) (details in ESM).

#### Concentration of hydrogen sulfide

The concentrations of  $H_2S$  in the plasma at baseline (t0), 30 min after ROSC (t1), and 60 min after ROSC (t2) were measured according to the method described previously (details in ESM) [13].

# Measurement of brain metabolism

Cerebral extraction of oxygen (CEO<sub>2</sub>), arterio-jugular venous differences of glucose [AJVD(glu)], and lactate clearance were measured at t0, t1, and t2. A detailed description of brain metabolism is provided in the ESM.

#### Statistical analysis

Data are presented as mean  $\pm$  SD. Intergroup differences in temperature, H<sub>2</sub>S concentration, CEO<sub>2</sub>, AJVD(glu), lactate, and S100B were assessed by two-way repeatedmeasures analysis of variance (ANOVA) with Bonferroni post hoc test. Differences in lactate clearance and neuron numbers were analyzed by one-way ANOVA with Bonferroni correction for post hoc comparison between multiple experimental groups. NDSs were compared with those of the corresponding CA group by use of a Mann– Whitney nonparametric test. Kaplan–Meier survival curves were compared using log-rank test. Significance was considered at the level of P < 0.05. Statistical analysis was performed using SPSS<sup>®</sup> software (SPSS Inc., Chicago, IL, USA).

# Results

Forty-five rabbits were included in the present study. Twelve of 15 rabbits were successfully resuscitated in the CA and CA/H<sub>2</sub>S groups. The characteristics of CA and resuscitation showed no statistical differences between the CA and CA/H<sub>2</sub>S groups (ESM Table 1). There were no significant differences in physiological variables among all groups at the baseline period (ESM Table 2).

#### Rectal and cerebral temperature

Although the temperature was slightly lower in the  $H_2S$  treated animals, no statistical differences were observed in rectal, cortex, and hippocampus temperatures between CA and CA/H<sub>2</sub>S groups during the 60 min after ROSC. There were also no significant differences in rectal, cortex, and hippocampus temperature between sham and sham/H<sub>2</sub>S groups (Fig. 1).

# S100B

There was no significant difference in S100B between sham and sham/H<sub>2</sub>S groups. The level of serum S100B was significantly higher in the CA group than in the sham group at t1 (9.69 ± 1.50 vs. 7.00 ± 1.06 ng/ml, P < 0.05) and t2 (14.16 ± 1.99 vs. 6.99 ± 0.97 ng/ml, P < 0.05). However, this increase was significantly less pronounced in the CA/H<sub>2</sub>S group at t1 (8.63 ± 1.19 ng/ ml) and t2 (10.85 ± 1.44 ng/ml) than in the CA group (both P < 0.05) (Fig. 2).

NDS and overall survival rate after ROSC

A significant difference in survival rate was shown between the CA/H<sub>2</sub>S group and CA group (P < 0.05) (Fig. 3a). At the end of the follow-up, five rabbits [42 % (5 of 12)] in the CA group and 10 rabbits [83 % (10 of 12)] in the CA/H<sub>2</sub>S group survived until day 7. It is noteworthy that the deaths occurred primarily between days 2 and 4. This is a time when death due to CA causes is uncommon and neurological causes predominate.

As shown in Fig. 3b, at 24 h after ROSC, the NDS was significantly attenuated in the CA/H<sub>2</sub>S group compared with the CA group (P < 0.05) (Fig. 3b).

#### Neuronal counts

On the 7th day after CA and resuscitation, the neuronal density of the CA group was much lower than that of the sham group (Fig. 4a). The pyramidal neurons appeared swollen and arranged in irregular patterns. Nuclear pyknosis, karyorrhexis, and vacuolization were also observed. However, neuronal density and cell morphology were well preserved in the CA/H<sub>2</sub>S group. For comparison, the numbers of viable neurons per high-power field (×400) in the CA and CA/H<sub>2</sub>S groups were significantly lower than that of the sham group (both P < 0.05). The hippocampal CA1 counts were 24 ± 6 in the CA/H<sub>2</sub>S group, which was significantly more than in the CA group (10 ± 3, P < 0.05) but still less than in the sham group (39 ± 6,

**Fig. 1** Rectal, cortex, and hippocampus temperatures in the different experimental groups. *BL* baseline







Fig. 2 Concentration of S100B as percentage of baseline values at t0, t1, and t2 for each group. *T0*, *T1*, and *T2* represent baseline, 30 and 60 min after restoration of spontaneous circulation. \*P < 0.05 versus sham group.  $^{\#}P < 0.05$  versus CA group

P < 0.05). No significant differences in CA1 neuron counts were observed between the sham and sham/H<sub>2</sub>S groups (Fig. 4b).

# H<sub>2</sub>S concentration

As can be seen in Fig. 5, the concentration of  $H_2S$  was significantly increased after inhalation of exogenous  $H_2S$  in the sham/ $H_2S$  group at t1 and t2 ( $3.26 \pm 0.47$  and  $3.74 \pm 0.29 \mu$ mol/L, respectively) compared with the sham group ( $2.29 \pm 0.07$ ,  $2.38 \pm 0.14 \mu$ mol/L, respectively, both P < 0.05). These changes were also observed between the CA and CA/ $H_2S$  groups at t1 ( $2.63 \pm 0.33$  vs.  $6.43 \pm 0.51 \mu$ mol/L) and t2 ( $2.92 \pm 0.41$  vs.  $8.16 \pm 0.38 \mu$ mol/L) (both P < 0.05). We also observed higher concentrations of  $H_2S$  in the CA group relative to the sham group at t1 and t2. This difference was significant at t2 (P < 0.05). The levels of  $H_2S$  with  $H_2S$  therapy were significantly higher in animals undergoing CA compared to shams (P < 0.05 for both t1 and t2).

Brain metabolic measures

 $CEO_2$ 

As presented in Fig. 6, the levels of CEO<sub>2</sub> at t1 were  $16.72 \pm 6.12$  % in the CA group and  $25.04 \pm 7.11$  % in the CA/H<sub>2</sub>S group. These were both significantly higher than those of the sham group ( $8.46 \pm 4.73$  %, both P < 0.05). This tendency was still observable at t2 ( $19.69 \pm 6.02$ ,  $22.36 \pm 7.70$ , and  $9.31 \pm 3.29$  %, respectively, P < 0.05). The CA/H<sub>2</sub>S group had a higher level of CEO<sub>2</sub> than the CA group at t1 (P < 0.05) and t2. No significant differences were observed between the



Fig. 3 a Kaplan–Meier survival curves in the different experimental groups submitted to cardiac arrest and resuscitation. b Neurological dysfunction scores at day 1 after resuscitation in the different experimental groups. *Open circles* represent individual scores; *thick line* the median value of the corresponding group. #P < 0.05 versus CA group

sham and sham/H<sub>2</sub>S groups throughout the 60 min of the experiment.

# Arterio-jugular venous differences in glucose concentration

The values of AJVD(glu) were significantly higher in the CA group  $(1.18 \pm 0.38 \text{ mmol/L}, P < 0.05)$  but only slightly higher in the CA/H<sub>2</sub>S group  $(0.77 \pm 0.29 \text{ mmol/}$ L) than sham values  $(0.44 \pm 0.14 \text{ mmol/L})$  at t1. There was a significant decrease in AJVD(glu) in the CA/H<sub>2</sub>S group compared with the CA group at t1 (P < 0.05). The increase in AJVD(glu) in the CA group returned to normal values at t2 (Fig. 6). No significant differences in AJVD(glu) were observed between the sham and sham/H<sub>2</sub>S groups at any time.



**Fig. 4 a** Representative histological images of the hippocampus CA1 region from sham, sham/H<sub>2</sub>S, CA, and CA/H<sub>2</sub>S groups at 7 days after restoration of spontaneous circulation. All images were captured at  $\times 400$  magnification. *Scale bar* indicates 100 µm. **b** Number of viable neurons in the CA1 region belonging to different groups. \**P* < 0.05 versus sham group. #*P* < 0.05 versus CA group



Fig. 5 Concentration of plasma H<sub>2</sub>S for each group. *T0*, *T1*, and *T2* represent baseline, 30 and 60 min after restoration of spontaneous circulation. \*P < 0.05 versus sham group.  $^{\#}P < 0.05$  versus CA group,  $^{\$}P < 0.05$  versus sham/H<sub>2</sub>S group

#### Lactate and lactate clearance

A statistically significantly higher lactate level was observed in the CA (6.01  $\pm$  0.64 mmol/L) and CA/H<sub>2</sub>S group (5.11  $\pm$  0.43 mmol/L) at t1 than in the sham group (3.50  $\pm$  0.33 mmol/L, both *P* < 0.05). Differences in lactate levels between the CA and CA/H<sub>2</sub>S groups reached statistical significance at t1 (*P* < 0.05) and widened by t2 (6.11  $\pm$  0.77 vs. 3.83  $\pm$  0.56 mmol/L, *P* < 0.05) (Fig. 6).

After ROSC, lactate clearance of Lactate (t0–t1) and Lactate (t0–t2) were negative, indicating an increase in lactate level. However, in the CA/H<sub>2</sub>S group, Lactate (t1–t2) was positive, indicating lactate clearance after ROSC. As shown in Fig. 6, the values of Lactate (t0–t1) (74.22  $\pm$  12.59 %) and Lactate (t0–t2) (77.04  $\pm$  28.20 %) were significantly higher in the CA group than in the sham group (0.48  $\pm$  5.76, 3.49  $\pm$  6.93 %, both *P* < 0.05). However, Lactate (t0–t1) (45.98  $\pm$  13.43 %) and Lactate (t0–t2) (9.72  $\pm$  19.67 %) were significantly lower in CA/H<sub>2</sub>S group than in the CA group (both *P* < 0.05).

# Discussion

The current study investigated the effects of exogenous  $H_2S$  on cerebral metabolism and neurological function following CA and resuscitation. The key findings were that  $H_2S$  reduced neurohistopathological damage and improved early neurological function after CA and resuscitation. Increased CEO<sub>2</sub>, decreased AJVD(glu), and enhanced lactate clearance were observed at the same time.

In this study, we observed that inhaling exogenous 80 ppm H<sub>2</sub>S gas significantly increased the concentration of  $H_2S$  in plasma. Interestingly, we observed an increase of H<sub>2</sub>S after ROSC even without inhalation of exogenous H<sub>2</sub>S in the CA group. Several researchers have described significant changes of H<sub>2</sub>S in plasma levels in various disease states [14-16]. A recent study with cerebellar slices and intact mouse brains identified that during hypoxia inhibition of heme oxygenase 2 mediated CO production, with a corresponding release of the tonic inhibition of CBS, and allowing CBS to generate H<sub>2</sub>S [17]. On the other hand, the decreased capacity of rhodanese, which is the key component in the H<sub>2</sub>S removal system under low O<sub>2</sub> tensions, may effectively increase the  $H_2S$  concentrations [18]. Eto and colleagues [2] found that the excitatory neurotransmitter L-glutamate greatly enhanced the production of  $H_2S$ . And many studies have confirmed that there is a massive outpouring of extracellular glutamate after CA or cerebral ischemia and reperfusion injury [19, 20]. On the basis of these studies we hypothesize that hypoxia and glutamate excess drive increased post-ischemic H<sub>2</sub>S levels and the synergistic increase with H<sub>2</sub>S therapy.



**Fig. 6** Cerebral extraction of oxygen, arterio-jugular venous differences of glucose, and lactate in sham, sham/H<sub>2</sub>S, CA, and CA/H<sub>2</sub>S groups. Lactate clearance shows the clearance of lactate between t0 and t1 [*Lactate* (t0-t1)], between t0 and t2 [*Lactate* (t0-t1)]

t2)], and between t1 and t2 [Lactate (t1-t2)] for each group. T0, T1, and T2 represent baseline, 30 and 60 min after restoration of spontaneous circulation. \*P < 0.05 versus sham group. \*P < 0.05 versus CA group

Much research has been published on the clinical utility of S100B in predicting neurological outcomes after CPR [21, 22]. In this study, we found that serum S100B levels were significantly increased after ROSC, whereas inhalation of  $H_2S$  significantly reduced the level of S100B, which predicted a better neurological outcome. This was confirmed by the results of NDS. The 24-h NDS data, when survivors were nearly equal between CA and CA/H<sub>2</sub>S groups, clearly showed less neurological injury in H<sub>2</sub>S treated animals. And this protective effect continued throughout the experiment, which was proven by the increase in viable neurons and survival at 7 days after ROSC. Taking all these factors together, the present data provide strong evidence that  $H_2S$  can improve early neurological outcomes after CA and resuscitation.

Although decreased body temperature and metabolism were observed in mice exposed to  $H_2S$  [1, 23], these results had been questioned in piglets [4], sheep [5], and even rats [6]. Similarly in our rabbit model, rectal, cortex, and hippocampus temperatures were not decreased significantly in  $H_2S$  treated animals. Owing to their large surface area/mass ratio, mice can experience rapid drops in core body temperature, whereas these are rare or impossible in larger animals. Another key factor was the low  $O_2$  tension which was indispensable for  $H_2S$ -dependent hypometabolism [6]. Stein and colleagues [6] found that rats were largely insensitive to  $H_2S$ -induced low body temperature unless  $O_2$  tension was decreased to 10.5 %. This finding was supported by our study with temperature results during the short periods of exposure to lowered  $O_2$  tensions.

In the present study, we observed an increased CEO<sub>2</sub> after CA which is compatible with previous studies [24]. However, Lemiale and colleagues [25] found that CEO<sub>2</sub> reached nearly normal values over time in survivors. These different results of CEO<sub>2</sub> could probably be explained in part by the therapeutic hypothermia and midazolam used for sedation in Lemiale's study which can depress the brain metabolism. Another explanation could be the different degree of brain damage. In Lemiale's study, more severe brain damage was found in 10 of 18 patients and nearly half of them had serious seizures [25]. More severe brain damage meant extensive loss of viable neurons with a concomitant decrease in cerebral metabolism [24, 26].

The current findings also demonstrate that less glucose extraction and faster lactate clearance occur after inhalation of H<sub>2</sub>S. Taking into account the higher CEO<sub>2</sub>, one possibility could be a switch from anaerobic glycolysis induced by asphyxia to oxidative phosphorylations which consume more oxygen and less glucose while producing less lactate. In the latest study [27], evidence had shown that H<sub>2</sub>S could correct the pathophysiological switch between oxidative phosphorylation and glycolysis, which gave a powerful support to our study.

Many studies demonstrated that the increased CEO<sub>2</sub> [25, 26] or lower CJVO<sub>2</sub> (content of oxygen in the jugular venous blood) [24, 28] and effective early lactate clearance were associated with improvement of neurological outcome and decreases in both early and late mortality in post-CA patients [29, 30]. Consistent with these findings, a better neurological outcome was confirmed with less S100B, more viable neurons, less NDS, and higher survival rate in H<sub>2</sub>S treated animals which had increased CEO<sub>2</sub>, decreased AJVD(glu), and enhanced lactate clearance.

This study has several limitations. Firstly, we used 80 ppm H<sub>2</sub>S because this dose had been widely used in previous studies and no obvious neurotoxic effects were observed [1, 4, 31]. Although this will make it easier to draw comparisons, the relationship between H<sub>2</sub>S concentration and favorable outcome has not been verified. Furthermore, the lack of measurement of CBF and ATP generation/ox-phos limits our ability to draw a conclusion whether the brain metabolism changes are related to delivery or consumption. Future studies are warranted to further investigate the relationship between H<sub>2</sub>S effects and CBF and ATP generation/ox-phos. Lastly, though our findings demonstrate improvement of early neurological function after inhalation of 80 ppm H<sub>2</sub>S in a rabbit CA model, the translational implication of H<sub>2</sub>S in the human clinical setting is uncertain and needs to be investigated.

In summary, our study revealed that inhalation of 80 ppm  $H_2S$  reduced neurohistopathological damage and improved early neurological function after CA and resuscitation in rabbits. We suggest that improvements in cerebral metabolism evidenced by increasing CEO<sub>2</sub>, decreasing AJVD(glu), and clearing lactate may be involved in the protective effects. A better understanding of the relationship between  $H_2S$  effects and CBF and ATP generation/ox-phos in future studies will be important in clarifying the underlying mechanisms.

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**Conflicts of interest** There was no conflict of interest in this study.

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