

M. P. Fink  
T. W. Evans

## Mechanisms of organ dysfunction in critical illness: report from a Round Table Conference held in Brussels

---

Received: 10 July 2001  
Accepted: 11 December 2001  
Published online: 8 February 2002  
© Springer-Verlag 2002

M.P. Fink (✉)  
Departments of Critical Care Medicine and Surgery,  
University of Pittsburgh Medical School, 616 Scaife Hall,  
3550 Terrace Street, Pittsburgh, PA 15261, USA  
e-mail: finkmp@ccm.upmc.edu

T.W. Evans  
Unit of Critical Care, Imperial College School of Medicine,  
Royal Brompton Hospital, London, UK

---

### Introduction

The most common cause of death in patients with sepsis or other forms of critical illness is deterioration in the function of multiple organs, now termed multiple organ dysfunction syndrome (MODS). Commonly affected organs include the lungs, liver, and kidneys, and the clinical manifestations of arterial hypoxemia, decreased pulmonary compliance, cholestatic jaundice, oliguria, and azotemia are familiar to all intensivists. Manifestations of central nervous system dysfunction, ranging from subtle alterations in mental status to frank coma, are also common in patients with sepsis. Derangements in gastrointestinal function, including loss of normal peristalsis (i.e., ileus) and enterocytic barrier function, are also common, albeit difficult to quantify, manifestations of MODS. Although a fatal outcome in patients with sepsis and septic shock is virtually always accompanied by MODS, the histopathology of fatal sepsis is remarkably bland. For example, histological sections of liver or kidney tissue from patients dying from sepsis or septic shock occasionally show evidence of focal necrosis or apoptosis (“programmed cell death”), but massive loss of parenchymal mass is rarely observed [1]. Thus the physiological basis for organ dysfunction in sepsis remains a puzzling problem.

To provide a “snapshot” of the state of the art of research into this fascinating problem a Roundtable Conference was recently held in Brussels to discuss and review “Mechanisms of Organ Dysfunction in Critical Illness.” The Roundtable Conference was jointly sponsored the European Society of Intensive Care Medicine, the Society of Critical Care Medicine, and the American Thoracic Society. The list of participants included the following clinical and basic scientists from several European countries, Canada, and the United States: A.J. Bauer, T.G. Buchman, W.A. Buurman, G.P. Downey, T.W. Evans, M.P. Fink, A.P. Halestrap, C. Ince, P. Kochanek, X.M. Laverne, M.A. Matthay, J.C. Marshall, B.A. Molitoris, R. Neviere, D. Payen, P. Radermacher, J. Saklatvala, P.T. Schumacker, W. Sibbald, A.S. Slutsky, C. Szabó, K.J. Tracey, R.J. Traystman, T. van der Poll, B. Vallet, J.-L. Vincent, and J.M. Weinberg

---

### Mitochondrial dysfunction and “cytopathic hypoxia”

A derangement in cellular energy metabolism seems a likely proximate cause of cellular – and hence organ – dysfunction in critical illness. Adenosine triphosphate (ATP) is the “energy currency” for cells. The free energy for the hydrolysis reaction,  $ATP \rightarrow$  adenosine diphosphate (ATP) + inorganic phosphate (Pi) + hydrogen ion ( $H^+$ ), is used to power various metabolic processes in cells, such as the activities of membrane-bound transporters and the synthesis and degradation of proteins. In most cells ATP production occurs primarily in the mitochondria through the process called oxidative phosphorylation. In this process, reducing equivalents, namely the reduced forms of nicotinamide adenine dinucleotide (NADH) and flavin adenine dinucleotide ( $FADH_2$ ) are oxidized by molecular oxygen to their oxidized forms (i.e.,  $NAD^+$  and FAD, respectively). The energy released by this process is used to create a gradient of hydrogen ions across the inner mitochondrial membrane. As pro-

tons move down this gradient through a specialized enzyme complex, the  $F_0F_1$ ATPase, the energy stored in the gradient as an electrochemical potential is released and coupled to the formation of ATP from ADP, Pi, and  $H^+$ . The reducing equivalents, NADH and  $FADH_2$ , are produced through the catabolism of fuels produced during anaerobic glycolysis, the  $\beta$ -oxidation of fats, and the oxidation of acetyl-coenzyme A in the tricarboxylic acid cycle.

The normal functioning of this remarkable cellular machinery depends of course on the adequate availability of oxygen. However, the normal functioning of the mitochondrial ATP factory also depends upon many other factors, such as the availability of catalytic quantities of NADH/NAD<sup>+</sup>, integrity of the various enzyme complexes that carry out electron transport, and appropriate maintenance of the protonic gradient across the inner mitochondrial membrane. Accordingly, energy metabolism might be deranged either on the basis of decreased delivery of oxygen to cells or an acquired intrinsic alteration in the functioning of mitochondria.

One potentially important form of mitochondrial dysfunction can be caused by the opening of a nonspecific pore in the mitochondrial inner membrane. This mitochondrial permeability transition pore (MPTP) is normally closed. However, under certain pathological conditions, such as during reperfusion after a period of ischemia or after exposure of the cell to various toxins, the MPTP opens, leading to dissipation of the mitochondrial protonic gradient ("mitochondrial depolarization") and hence disruption of ATP synthesis [2]. Although the MPTP was first described many years ago, major advances in our understanding of its importance were not made until the discovery that submicromolar concentrations of the immunosuppressive agent cyclosporin A inhibit opening of the pore [3]. Opening of the MPTP has been implicated in various forms of organ dysfunction, including hepatocyte damage due to exposure to tumor necrosis factor [4], myocardial reperfusion injury [5], and neuronal damage induced by glucose and oxygen deprivation [6]. Of considerable interest are data indicating that opening of the MPTP can be blocked by analogues of cyclosporin A that lack the immunosuppressive activity of the parent compound [6]. Even some unrelated agents, such as the cardioprotective compound carvedilol [7], may work by blocking opening of the MPTP. Pharmacological blockade of the MPTP may therefore become a viable therapeutic strategy in the management of critically ill patients.

Activation of the enzyme poly(ADP-ribose) polymerase (PARP), also sometimes called poly(ADP-ribose) synthetase, is another potential cause of "cytopathic hypoxia" (intrinsic derangement in cellular respiration) during critical illness [8]. Under normal conditions PARP is relatively inactive in cells. However, PARP is activated by the presence of single-strand breaks in nuclear DNA,

which can be induced by various endogenously generated oxidants, such as peroxynitrite (ONOO<sup>-</sup>) and hydrogen peroxide ( $H_2O_2$ ) [9]. The natural substrate for PARP is NAD<sup>+</sup>. When activated, PARP cleaves NAD<sup>+</sup> into ADP-ribose and nicotinamide. PARP then covalently attaches the ADP-ribose units to various nuclear proteins, forming in the process a branched homopolymer, poly-ADP-ribose. Simultaneously, poly-ADP-ribose is degraded by various nuclear enzymes, especially poly-ADP-ribose glycohydrolase [10]. Thus the coupled actions of PARP plus poly-ADP-ribose glycohydrolase are the biochemical equivalent of an "NADase." As a consequence the activation of PARP can lead to profound depletion of cellular levels of NAD and thereby to an "energy crisis" in cells [11, 12, 13]. Pharmacological inhibition of PARP has been shown to be beneficial in animal models of sepsis and/or multiple organ dysfunction [11, 14]. New, potent PARP inhibitors may represent a novel important therapeutic approach for the management or prevention of organ system dysfunction in critical illness [15].

Ischemic injury to cells can lead to derangements in mitochondrial function even after normal tissue oxygenation is restored. Thus respiration by renal tubules subjected to an episode of hypoxia is inhibited during reoxygenation, predominantly because of dysfunction of the mitochondrial enzyme complex, complex I [16]. ATP depletion within the mitochondria may contribute irreversible damage to the organelle during episodes of tissue ischemia and cellular hypoxia. Accordingly, pharmacological maneuvers to (at least partially) preserve ATP content during hypoxia might promote cellular recovery during reoxygenation. Indeed, during ischemia and other insults that limit mitochondrial oxidative phosphorylation, ATP production by anaerobic glycolysis can prevent generalized cell injury and, via reverse operation of the inner membrane ATP synthase ( $F_0F_1$ -ATPase), maintain mitochondrial energization and prevent opening of MPTP [17, 18]. However, anaerobic glycolysis is eventually suppressed during myocardial ischemia and is inherently limited or absent in other cells such as neurons and kidney proximal tubular cells. Interestingly, anaerobic mitochondrial metabolism can generate ATP and maintain mitochondrial energization via at least two pathways: substrate-level phosphorylation during the conversion of  $\alpha$ -ketoglutarate to succinate by  $\alpha$ -ketoglutarate dehydrogenase, and electron transport in complexes I and II driven by reduction of fumarate to succinate coupled to the oxidation of reduced ubiquinone that is generated via NADH from citric acid cycle reducing equivalents [16, 19]. Anaerobic generation of ATP by mitochondria is promoted by providing cells with a combination of substrates, containing  $\alpha$ -ketoglutarate and apartate, and the use of this substrate cocktail has been shown to support mitochondrial function in renal tubules subjected to a hypoxia/reoxygenation stress [16, 19].

It is becoming increasingly apparent that mitochondria play hugely important roles in cellular physiology in ways that are only indirectly related to the function of these organelles as factories for producing ATP. For example, during hypoxia mitochondria tend to “leak” partially reduced forms of molecular oxygen, so-called “free radicals,” and these potent oxidants can initiate proinflammatory signal transduction pathway, including activation of the transcription factor nuclear factor- $\kappa$ B, important in the regulation of tumor necrosis factor expression [20].

### **Leukocytes, cell signaling mechanisms, and alterations in capillary perfusion**

Leukocyte trafficking and senescence, and cytokine-mediated cell signaling play important roles in mediating the microcirculatory dysfunction that characterizes clinical sepsis. Chemokine-induced neutrophil migration from the pulmonary vasculature into the alveoli involves neutrophils transiting the endothelium, the interstitial matrix, and tight junctions between adjacent alveolar epithelial cells. Neutrophils therefore can damage the normal alveolar fluid-clearance mechanism and thus promote alveolar flooding in conditions association with pulmonary inflammation and acute lung injury [21]. Moreover, the efficiency of this alveolar clearance system is, at least in experimental models, influenced in part by sheer forces. Thus in experimental models the application of lower tidal volumes is associated with improved solute and ion clearance [22]. Clinical studies confirm that the ability of the alveoli to remove ions and solutes holds prognostic significance in patients with established acute respiratory distress syndrome [23]. Inflammatory stimuli induce species-specific phenotypic changes in pulmonary vascular smooth muscle, leading to the expression of enzyme systems producing vasomotor substances including endothelins, which appear to induce smooth muscle cell proliferation. The clinical importance of pulmonary vascular remodeling in acute respiratory distress syndrome has yet to be explored in any detail, but prostacyclin analogues appear to inhibit this endothelin-mediated effect [24].

Apoptosis (programmed cell death) of neutrophils can be triggered via both cell surface receptors and the release of cytochrome *c* by mitochondria [25]. These processes are tightly regulated by inhibitors of apoptosis proteins and caspases [26]. Caspases can have both pro- and antiapoptotic actions, and their therapeutic manipulation to affect inflammatory processes is theoretically highly attractive. Neutrophils are also a potent source of proinflammatory cytokines, principal among which are tumor necrosis factor and interleukin 1. Their effects appear to be regulated in part by the coexpression of anti-inflammatory cytokines, including interleukin 10. Re-

cently published work suggests that interleukin 10 is critically important in determining the lethality of sepsis in mice. For example, when subjected to a septic challenge, mice with a genetic inability to produce interleukin 10 have a markedly higher death rate and incidence of organ failure than wild-type controls [27].

The way in which inflammatory processes lead to various manifestations of endothelial dysfunction, including alterations in cell adhesion, coagulation, and vasomotor regulation, is gradually becoming clearer. Some *in vivo* studies using intravital techniques have suggested that the density of perfused capillaries is altered in sepsis such that some microvascular units are hyperperfused, possibly leading to apparent shunt flow, whereas other units are underperfused (or even “stopped-up” entirely) due to the presence of aggregates of red cells and/or leukocytes [28, 29]. Other studies, however, suggest that sepsis-induced organ dysfunction can occur even in the absence of alterations in microvascular perfusion [30].

The extent to which changes in microvascular perfusion lead directly to organ system failure was the subject of persistent debate, the strands of which permeated discussions following all presentations. Thus, although capillary hyper- and hypoperfusion can now be visualized in a number of microvascular beds clinically and in animal models, it does not appear to be invariably associated with organ system failure and death. Indeed, the extent to which multiorgan system failure can be attributed solely to microcirculatory abnormalities continues to be hotly contested by those who feel that cellular dysfunction, possibly at a mitochondrial level, is more likely responsible. Ongoing clinical studies using powerful techniques such as orthogonal polarization spectral imaging, which permit visualization of the microcirculation in patients, should prove to be very informative in this regard [31, 32].

Inappropriately high tidal volumes applied to the lung can also lead to the generation and dispersal of inflammatory cytokines into the pulmonary and systemic circulations, with important implications for distant organ function [33, 34]. However, inappropriate sheer stresses applied to the lung in this fashion may also lead to neutrophil apoptosis in kidney cells through the effects of transmitted pressures rather than systemic inflammation. Moreover, the rhythmic application of sheer to isolated cells systems in culture can result in mitochondrial activation, activation of nuclear factor- $\kappa$ B, and increased expression of proinflammatory substances that might explain many of the vascular abnormalities described above.

### **The kidney and gastrointestinal tract**

Acute renal failure is a common clinical event affecting 2–5% of hospitalized patients and up to 10–30% of those

in intensive care units [35]. Renal ischemia is one of the most important antecedent factors in acute renal failure, reported to be causative in 50% of cases [35]. Until recently necrosis was believed to be the major form of cell death involved in acute renal failure. This belief gave birth to a commonly used, but nonetheless misleading, synonym for acute renal failure: acute tubular necrosis. When cells undergo necrosis, the cells swell and lose their membrane integrity. As a consequence there is leakage of intracellular compounds into the extracellular milieu. This leakage of intracellular material into surrounding tissue is believed to initiate the ischemia/reperfusion-induced inflammatory response, which is considered to be a major cause of the resultant tissue damage and organ dysfunction. Recently, however, apoptosis has been implicated in the pathophysiology of renal injury secondary to ischemia/reperfusion. Apoptosis is a highly conserved biological process essential in maintaining normal homeostasis. As noted above, apoptosis is characterized by activation of caspases, the intracellular execution enzymes of apoptotic cell death. Activated caspases cleave a wide range of intracellular substrates, ultimately leading to formation of apoptotic bodies that can be ingested by neighboring cells or macrophages without provoking an inflammatory response. Evidence for the occurrence of apoptotic cell death in ischemia/reperfusion injury was originally reported by Schumer et al. [36] in 1992, who demonstrated apoptotic cell death upon reperfusion after renal ischemia in the rat. Since then evidence has accumulated that apoptosis is not just an epiphenomenon of ischemia/reperfusion injury, but that it plays a functional role in the pathogenesis of renal ischemia/reperfusion injury [37, 38]. Accordingly, pharmacological inhibition of caspase activation, and hence apoptosis, someday may prove to be a clinically relevant strategy to prevent the development of acute renal failure in patients with critical illness.

The proper functioning of many different cell types, including renal and intestinal epithelial cells, depends on the maintenance of normal cytoskeletal organization. The protein actin forms the foundation for the cytoskeleton. The actin-based cytoskeleton is a dynamic structure requiring continuous actin assembly and disassembly. This assembly-disassembly process is regulated by a variety of intracellular mechanisms that include regulatory proteins such as the rho family GTPases and actin effector proteins such as actin-depolymerizing factor (depolymerizes F-actin in a pH-dependent fashion and sequesters G-actin), capping protein (caps the barbed end and stabilizes F-actin), and gelsolin (severs F-actin in a  $\text{Ca}^{2+}$  dependent fashion, caps and stabilizes F-actin, and nucleates actin filaments) [39]. Activation of actin depolymerizing factor by dephosphorylation may play a central role in mediating the cytoskeletal alterations induced by ischemia/reperfusion injury or other forms of cellular stress. The actin depolymerizing factor/cofilin (AC) fam-

ily of proteins locate to apical membrane blebs shed from proximal tubule cells into the lumen during ischemia. This family of ubiquitous intracellular proteins plays an essential role in many actin regulated cellular processes such as the establishment of cellular polarity, cytokinesis, phagocytosis, motility, and fluid phase endocytosis. The primary function of AC proteins is their ability to act as "dynamizing factors" by greatly increasing actin filament turnover rates [40, 41, 42, 43]. Although the activity of AC proteins can be affected by several mechanisms, the primary mode of regulation is through phosphorylation of the amino terminal serine [44, 45, 46]. Phosphorylation of AC proteins inhibits their binding to actin and results in inactivation. Dephosphorylation of AC can be induced in response to a variety of cell stimuli, including ATP depletion.

Because patients can be fed parenterally, the gut was until fairly recently viewed as a "quiescent" organ during critical illness [47]. However, there now is good evidence that marked derangements in gut barrier function portend a bad outcome, at least for certain groups of critically ill patients [48, 49]. Moreover, several clinical studies support the view that providing enteral nutrition to critically ill patients improves outcome [50, 51]. Thus understanding the basis for both gut barrier dysfunction and the loss of normal peristalsis (i.e., the development of ileus) in critical illness assumes considerable importance.

Alterations in intestinal barrier function in critical illness appear to be caused by multiple factors, including induction of the inducible isoform of nitric oxide ( $\text{NO}\cdot$ ) synthase and overproduction of  $\text{NO}\cdot$  [52, 53], cytoskeletal alterations, derangements in cellular respiration [54], and, possibly, increased epithelial apoptosis [1, 55]. Recently obtained data strongly suggest that induction of PARP is a key factor. Ileus, signifying the impairment of coordinated propulsive intestinal peristalsis, remains a well documented and virtually inevitable consequence of open abdominal surgery, sepsis, and shock. Although the pathophysiology of ileus in critical illness remains incompletely understood, accumulating data support the view that infiltration of the muscular coats of the bowel by neutrophils and activation of resident macrophages, plays a crucial role [56, 57].

#### Brain and nervous system

Progress in the successful application of novel therapies in the treatment of severe traumatic brain injury in humans has been disappointing. Currently treatment is limited to field stabilization and supportive neurointensive care, the application of clinically accepted strategies to control intracranial hypertension, and surgical resection of mass lesions. A number of novel therapies have shown promise in contemporary experimental models of

severe traumatic brain injury, but none has been successfully translated to clinical use via a randomized controlled clinical trial demonstrating a beneficial effect on outcome. Examples of important failures include the trials of two agents targeting oxidative injury, tirilazad [58] and superoxide dismutase [59], and one targeting excitotoxicity, Selfotel [60]. Similarly dismal results have been obtained in clinical studies of new agents for the management of stroke [61, 62, 63]. Recent preclinical studies, however, suggest that pharmacological inhibition of PARP may prove to be very promising for this indication [64].

A very promising strategy to treat cerebral ischemia has recently emerged, focusing upon blocking the deleterious effects of a small molecule, 3-aminopropanal, which appears to be important in cause of neuronal damage in stroke [65]. 3-Aminopropanal is a metabolic product of the degradation of the polyamines spermine and spermidine by polyamine oxidase. Levels of spermine and spermidine fall during cerebral ischemia, and polyamine oxidase activity increases, resulting in increased production of 3-aminopropanal. 3-Aminopropanal is a potent cytotoxin that causes neuronal necrosis and glial cell apoptosis [65]. 3-Aminopropanal modified proteins can be detected in the cerebrospinal fluid of patients with

stroke or head injury (Connolly and Ivanova, unpublished data). Administration of experimental therapeutics that either inhibit polyamine oxidase activity or react directly with 3-aminopropanal prevent brain damage in animals with cerebral ischemia or brain trauma [66, 67, 68].

It is increasingly apparent that the brain and neuroendocrine systems play hugely important roles in determining the responses of the whole organism to trauma and proinflammatory stimuli. For example, it has recently been shown that efferent signals transmitted through the vagal nerve dramatically down-regulate systemic cardiovascular and immunological responses to an injection of lipopolysaccharide in mice [69]. Moreover, evidence is accumulating that a key feature of critical illness in humans is a decrease in the variability of heart rate, a finding that suggests a loss of the normal neuroendocrine linkages between the heart and other organs [70, 71, 72]. Of course, intensivists recognize that changes in mental status, ranging from subtle abnormalities in cognition to agitation to frank coma, are common findings in critically ill patients. Whether these alterations in mental status and/or a loosening of the neuroendocrine linkages between organ systems is adaptive or maladaptive responses remains to be determined.

## References

- Hotchkiss RS, Swanson PE, Freeman BD, Tinsley KW, Cobb JP, Matuschak GM, Buchman TG, Karl IE (1999) Apoptotic cell death in patients with sepsis, shock, and multiple organ dysfunction. *Crit Care Med* 27:1230–1251
- Granger DL, Taintor JL, Cook JL, Hibbs JB Jr (1980) Injury of neoplastic cells by murine macrophages leads to inhibition of mitochondrial respiration. *J Clin Invest* 65:357–370
- Crompton M, Ellinger H, Costi A (1988) Inhibition by cyclosporin A of a  $Ca^{2+}$ -dependent pore in heart mitochondria activated by inorganic phosphate and oxidative stress. *Biochem J* 255:357–360
- Pastorino JG, Simbula G, Yamamoto K, Glascott PA, Rothman RJ, Farber JL (1996) The cytotoxicity of tumor necrosis factor depends on induction of the mitochondrial permeability transition. *J Biol Chem* 271:29792–29798
- Halestrap AP, Connern CP, Griffiths EJ, Kerr PM (1997) Cyclosporin A binding to mitochondrial cyclophilin inhibits the permeability transition pore and protects hearts from ischaemia/reperfusion injury. *Mol Cell Biochem* 174:167–172
- Khaspeckov L, Friberg H, Halestrap A, Victorov I, Wieloch T (1999) Cyclosporin A and its non-immunosuppressive analogue N-Me-Val-4-cyclosporin A mitigate glucose/oxygen deprivation-induced damage to rat cultured hippocampal neurons. *Eur J Neurosci* 11:3194–3198
- Rolo AP, Oliveira PJ, Moreno AJ, Palmeira CM (2001) Protective effect of carvedilol on chenodeoxycholate induction of the permeability transition pore. *Biochem Pharmacol* 61:1449–1454
- Fink MP (2001) Cytopathic hypoxia: mitochondrial dysfunction as a mechanism contributing to organ dysfunction in sepsis. *Crit Care Clin* 17:219–237
- Szabó C, Dawson VL (1998) Role of poly (ADP-ribose) synthetase in inflammation and ischaemia-reperfusion. *Trends Pharmacol Sci* 19:287–298
- Desnoyers S, Shah GM, Brochu G, Hoflack JC, Verreault A, Poirier GG (1995) Biochemical properties and function of poly (ADP-ribose) glycohydrolase. *Biochimie* 77:433–438
- Szabó C, Zingarelli B, Salzman AL (1996) Role of poly-ADP ribosyltransferase activation in the vascular contractile and energetic failure elicited by exogenous and endogenous nitric oxide and peroxynitrite. *Circ Res* 78:1051–1063
- Szabó C, Zingarelli B, O'Connor M, Salzman AL (1996) DNA strand breakage, activation of poly-ADP ribosyl synthetase, and cellular energy depletion are involved in the cytotoxicity in macrophages and smooth muscle cells exposed to peroxynitrite. *Proc Natl Acad Sci USA* 93:1753–1758
- Szabó C, Saunders C, O'Connor M, Salzman AL (1997) Peroxynitrite causes energy depletion and increases permeability via activation of poly (ADP-ribose) synthetase in pulmonary epithelial cells. *Am J Respir Cell Mol Biol* 16:105–109
- Cuzzocrea S, Zingarelli B, Costantino G, Sottile A, Teti D, Caputi AP (1999) Protective effect of poly (ADP-ribose) synthetase inhibition on multiple organ failure after zymosan-induced peritonitis in the rat. *Crit Care Med* 27:1517–1523
- Garcia Soriano F, Virag L, Jagtap P, Szabo E, Mabley JG, Liaudet L, Marton A, Hoyt DG, Murthy KG, Salzman AL, Southan GJ, Szabo C (2001) Diabetic endothelial dysfunction: the role of poly (ADP-ribose) polymerase activation. *Nat Med* 7:108–113

16. Weinberg JM, Venkatachalam MA, Roeser NF, Nissim I (2000) Mitochondrial dysfunction during hypoxia/reoxygenation and its correction by anaerobic metabolism of citric acid cycle intermediates. *Proc Natl Acad Sci USA* 97:2826–2831
17. Simbula G, Glascott PA Jr, Akita S, Hoek JB, Farber JL (1997) Two mechanisms by which ATP depletion potentiates induction of the mitochondrial permeability transition. *Am J Physiol* 273:C479–C488
18. Nieminen AL, Saylor AK, Herman B, Lemasters JJ (1994) ATP depletion rather than mitochondrial depolarization mediates hepatocyte killing after metabolic inhibition. *Am J Physiol* 267:C67–C74
19. Weinberg JM, Venkatachalam MA, Roeser NF, Saikumar P, Dong Z, Senter RA, Nissim I (2000) Anaerobic and aerobic pathways for salvage of proximal tubules from hypoxia-induced mitochondrial injury. *Am J Physiol* 279:F927–F943
20. Chandel NS, Trzyna WC, McClintock DS, Schumacker PT (2000) Role of oxidants in NF-kappa B activation and TNF-alpha gene transcription induced by hypoxia and endotoxin. *J Immunol* 165:1013–1021
21. Laffon M, Lu LN, Modelska K, Matthay MA, Pittet JF (1999) Alpha-adrenergic blockade restores normal fluid transport capacity of alveolar epithelium after hemorrhagic shock. *Am J Physiol* 277:L760–L768
22. Lecuona E, Saldias F, Comellas A, Ridge K, Guerrero C, Sznajder JI (1999) Ventilator-associated lung injury decreases lung ability to clear edema in rats. *Am J Respir Crit Care Med* 159:603–609
23. Ware LB, Matthay MA (2001) Alveolar fluid clearance is impaired in the majority of patients with acute lung injury and the acute respiratory distress syndrome. *Am J Respir Crit Care Med* 163:1376–1383
24. Wort SJ, Mitchell JA, Woods M, Evans TW, Warner TD (2000) The prostacyclin-mimetic cicaprost inhibits endogenous endothelin-1 release from human pulmonary artery smooth muscle cells. *J Cardiovasc Pharmacol* 36 [5 Suppl 1]:S410–S413
25. Watson RW, O'Neill A, Brannigen AE, Coffey R, Marshall JC, Brady HR, Fitzpatrick JM (2001) Regulation of Fas antibody induced neutrophil apoptosis is both caspase and mitochondrial dependent. *FEBS Lett* 453:67–71
26. Akgul C, Moulding DA, Edwards SW (2001) Molecular control of neutrophil apoptosis. *FEBS Lett* 487:318–322
27. Sewnath ME, Olszyna DP, Birjmohun R, ten Kate FJ, Gouma DJ, van der Poll T (2001) IL-10-deficient mice demonstrate multiple organ failure and increased mortality during escherichia coli peritonitis despite an accelerated bacterial clearance. *J Immunol* 166:6323–6331
28. Hoffmann JN, Vollmar B, Inthorn D, Schildberg FW, Menger MD (2000) The thrombin antagonist hirudin fails to inhibit endotoxin-induced leukocyte/endothelial cell interaction and microvascular perfusion failure. *Shock* 14:528–534
29. Piper RD, Pitt-Hyde ML, Li F, Sibbald WJ, Potter RF (1996) Microcirculatory changes in rat skeletal muscle in sepsis. *Am J Respir Crit Care Med* 154:931–937
30. Neviere RR, Pitt-Hyde ML, Piper RD, Sibbald WJ, Potter RF (1999) Microvascular perfusion deficits are not a prerequisite for mucosal injury in septic rats. *Am J Physiol* 276:G933–G940
31. Mathua KR, Vollebregt KC, Boer K, De Graaff JC, Ubbink DT, Ince C (2001) Comparison of OPS imaging and conventional capillary microscopy to study the human microcirculation. *J Appl Physiol* 91:74–78
32. Groner W, Winkelmann JW, Harris AG, Ince C, Bouma GJ, Messmer K, Nadeau RG (1999) Orthogonal polarization spectral imaging: a new method for study of the microcirculation. *Nat Med* 5:1209–1212
33. Mourageon E, Isowa N, Keshavjee S, Zhang X, Slutsky AS, Liu M (2000) Mechanical stretch stimulates macrophage inflammatory protein-2 secretion from fetal rat lung cells. *Am J Physiol* 279:L699–L706
34. Ranieri VM, Suter PM, Tortorella C, De Tullio R, Dayer JM, Brienza A, Bruno F, Slutsky AS (1999) Effect of mechanical ventilation on inflammatory mediators in patients with acute respiratory distress syndrome: a randomized controlled trial. *JAMA* 282:54–61
35. Thadhani R, Pascual M, Bonventre JV (1996) Acute renal failure. *N Engl J Med* 334:1448–1460
36. Schumer M, Colombel MC, Sawczuk IS, Gobe G, Connor J, O'Toole KM, Olsson CA, Wise GJ, Buttyan R (1992) Morphologic, biochemical, and molecular evidence of apoptosis during the reperfusion phase after brief periods of renal ischemia. *Am J Pathol* 140:831–838
37. Daemen MARC, van't Veer C, Denecker G, Heemskerk VH, Wolfs TGAM, Clauss M, Vandenabeele P, Buurman WA (1999) Inhibition of apoptosis induced by ischemia-reperfusion prevents inflammation. *J Clin Invest* 104:541–549
38. Feldenberg LR, Thevananther S, del Rio M, de Leon M, Devarajan P (1999) Partial ATP depletion induces Fas- and caspase-mediated apoptosis in MDCK cells. *Am J Physiol* 276:F837–F846
39. Molitoris BA (1997) Putting the actin cytoskeleton into perspective: pathophysiology of ischemic alterations. *Am J Physiol* 272:F430–F433
40. Schwartz N, Hosford M, Sandoval RM, Wagner MC, Atkinson SJ, Bamburg J, Molitoris BA (1999) Ischemia activates actin depolymerizing factor: role in proximal tubule microvillar actin alterations. *Am J Physiol* 276:F544–F551
41. Bamburg JR (1999) Proteins of the ADF/cofilin family: essential regulators of actin dynamics. *Annu Rev Cell Dev Biol* 15:185–230
42. Eddleston JM, Vohra A, Scott P, Tooth JA, Pearson RC, McCloy RF, Morton AK, Doran BH (1991) A comparison of the frequency of stress ulceration and secondary pneumonia in sucralfate- or ranitidine-treated intensive care unit patients. *Crit Care Med* 19:1491–1496
43. Thompson PA, Jelinek DF, Lipsky PE (1984) Regulation of human B cell proliferation by prostaglandin E<sub>2</sub>. *J Immunol* 133:2446–2450
44. Lappalainen P, Drubin DG (1997) Cofilin promotes rapid actin filament turnover in vivo. *Nature* 388:78–82
45. Richard C, Lemonnier F, Thibault M, Couturier M, Auzepy P (1990) Vitamin E deficiency and lipoperoxidation during adult respiratory distress syndrome. *Crit Care Med* 18:4–9
46. Huber M, Beutler B, Keppler D (1988) Tumor necrosis factor? stimulates leukotriene production in vivo. *Eur J Immunol* 18:2085–2088
47. Wilmore DW, Smith RJ, O'Dwyer ST, Jacobs DO, Ziegler TR, Wang X-D (1988) The gut: a central organ after surgical stress. *Surgery* 104:917–923
48. Ammori BJ, Leeder PC, King RFGJ, Barclay GR, Martin IG, Larvin M, McMahon MJ (1999) Early increase in intestinal permeability in patients with severe acute pancreatitis: correlation with endotoxemia, organ failure, and mortality. *J Gastrointest Surg* 3:252–262
49. Oudemans-van Straaten HM, Jansen PG, Hoek FJ, van Deventer SJ, Sturk A, Stoutenbeek CP, Tytgat GN, Wildevuur CR, Eysman L (1996) Intestinal permeability, circulating endotoxin, and postoperative systemic responses in cardiac surgery patients. *J Cardiovasc Anesth* 10:187–194

50. Kudsk KA, Croce MA, Fabian TC, Minard G, Tolley EA, Poret HA, Kuhl MR, Brown RO (1992) Enteral versus parenteral feeding: effects on septic morbidity after blunt and penetrating abdominal trauma. *Ann Surg* 215:503–513
51. Moore FA, Moore EE, Jones TN, McCroskey BL, Peterson VM (1989) TEN versus TPN following major abdominal trauma-reduced septic morbidity. *J Trauma* 29:916–923
52. Salzman AL, Menconi MJ, Unno N, Ezzell RM, Casey DM, Gonzalez PK, Fink MP (1995) Nitric oxide dilates tight junctions and depletes ATP in cultured Caco-2BBE intestinal epithelial monolayers. *Am J Physiol* 268:G361–G373
53. Unno N, Wang H, Menconi MJ, Tytgat SHAJ, Larkin V, Smith M, Morin MJ, Chavez A, Hodin RA, Fink MP (1997) Inhibition of inducible nitric oxide synthase ameliorates lipopolysaccharide-induced gut mucosal barrier dysfunction in rats. *Gastroenterology* 113:1246–1257
54. King CJ, Tytgat S, Delude RL, Fink MP (1997) Ileal mucosal oxygen consumption is decreased in endotoxemic rats but is restored toward normal by treatment with aminoguanidine. *Crit Care Med* 27:2518–2524
55. Coopersmith CM, O'Donnell D, Gordon JI (1999) Bcl-2 inhibits ischemia-reperfusion-induced apoptosis in the intestinal epithelium of transgenic mice. *Am J Physiol* 276:G677–G686
56. Hierholzer C, Kalff JC, Chakraborty A, Watkins SC, Billiar TR, Bauer AJ, Tweardy DJ (2001) Impaired gut contractility following hemorrhagic shock is accompanied by IL-6 and G-CSF production and neutrophil infiltration. *Dig Dis Sci* 46:230–241
57. Kalff JC, Buchholz BM, Eskandari MK, Hierholzer C, Schraut WH, Simmons RL, Bauer AJ (1999) Biphasic response to gut manipulation and temporal correlation of cellular infiltrates and muscle dysfunction in rat. *Surgery* 126:498–508
58. Marshall LF, Maas AI, Marshall SB, Bricolo A, Fearnside M, Iannotti F, Klauber MR, Lagarrigue J, Lobato R, Persson L, Pickard JD, Piek J, Servadei F, Wellis GN, Morris GF, Means ED, Musch B (1998) A multicenter trial on the efficacy of using tirilazad mesylate in cases of head injury. *J Neurosurg* 89:519–525
59. Muizelaar JP, Marmarou A, Young HF, Choi SC, Wolf A, Schneider RL, Kontos HA (1993) Improving the outcome of severe head injury with the oxygen radical scavenger polyethylene glycol-conjugated superoxide dismutase: a phase II trial. *J Neurosurg* 78:375–382
60. Morris GF, Bullock R, Marshall SB, Marmarou A, Maas A, Marshall LF (1999) Failure of the competitive N-methyl-D-aspartate antagonist Selfotel (CGS 19755) in the treatment of severe head injury: results of two phase III clinical trials. The Selfotel Investigators. *J Neurosurg* 91:737–743
61. Lees KR, Lavelle JF, Cunha L, Diener HC, Sanders EA, Tack P, Wester P, GAIN Phase II European Study Group (2001) Glycine antagonist (GV150526) in acute stroke: a multicentre, double-blind placebo-controlled phase II trial. *Cerebrovasc Dis* 11:20–29
62. Diener HC, Cortens M, Ford G, Grotta J, Hacke W, Kaste M, Koudstaal PJ, Wessel T (2001) Lubeluzole in acute ischemic stroke treatment: a double-blind study with an 8-hour inclusion window comparing a 10-mg daily dose of lubeluzole with placebo. *Stroke* 31:2543–2551
63. Clark WM, Raps EC, Tong DC, Kelly RE (2000) Cervene (Nalmefene) in acute ischemic stroke: final results of a phase III efficacy study. The Cervene Stroke Study Investigators. *Stroke* 31:1234–1239
64. Takahashi K, Pieper AA, Croul SE, Zhang J, Snyder SH, Greenberg JH (1999) Post-treatment with an inhibitor of poly (ADP-ribose) polymerase attenuates cerebral damage in focal ischemia. *Brain Res* 829:46–54
65. Ivanova S, Botchkina GI, Al-Abed Y, Mestrell M 3rd, Batiwalla F, Dubinsky JM, Iadecola C, Wang H, Gregersen PK, Eaton JW, Tracey KJ (1988) Cerebral ischemia enhances polyamine oxidation: identification of enzymatically formed 3-aminopropanal as an endogenous mediator of neuronal and glial cell death. *J Exp Med* 188:327–340
66. Cockcroft KM, Mestrell M 3rd, Zimmerman GA, Risucci D, Bloom O, Cerami A, Tracey KJ (1996) Cerebroprotective effects of aminoguanidine in a rodent model of stroke. *Stroke* 27:1393–1398
67. Dogan A, Rao AM, Baskaya MK, Hatcher J, Temiz C, Rao VL, Dempsey RJ (1999) Contribution of polyamine oxidase to brain injury after trauma. *J Neurosurg* 90:1078–1082
68. Rao AM, Hatcher JF, Dogan A, Dempsey RJ (2000) Elevated N1-acetylspermidine levels in gerbil and rat brains after CNS injury. *J Neurochem* 74:1106–1111
69. Borovikova LV, Ivanova S, Zhang M, Yang H, Botchkina GI, Watkins LR, Wang H, Abumrad N, Eaton JW, Tracey KJ (2000) Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature* 405:458–462
70. Haji-Michael PG, Vincent J-L, Degaute JP, van de Borne P (2000) Power spectral analysis of cardiovascular variability in critically ill neurosurgical patients. *Crit Care Med* 28:2578–2583
71. Goldstein B, Fiser DH, Kelly MM, Mickelsen D, Ruttimann U, Pollack MM (1998) Decomplexification in critical illness and injury: relationship between heart rate variability, severity of illness, and outcome. *Crit Care Med* 26:352–357
72. Godin PJ, Buchman TG (1996) Uncoupling of biological oscillators: a complementary hypothesis concerning the pathogenesis of multiple organ dysfunction syndrome. *Crit Care Med* 24:1107–1116