

- morphism; allele 814: lack of association with type I diabetes in Basques. *Diabetologia* 41: 1121–1123
6. World Health Organisation Study Group on Diabetes Mellitus (1985) Technical Report Series No 727, WHO Geneva
7. Kristiansen OP, Larsen ZM, Johannesen J, Mandrup-Poulsen T, Nerup J, Pociot F (1997) Linkage analysis of IDDM 13 in

- Danish IDDM multiplex families. *Diabetologia* 40 [Suppl]: A 53 (abstract)
8. Djlali-Saiah I, Larger E, Harfouch-Hammoud E et al. (1998) No major role for the CTLA-4 gene in the association of autoimmune thyroid disease with IDDM. *Diabetes* 47: 125–127

UKPDS 31: Hepatocyte nuclear factor-1alpha (the MODY3 gene) mutations in late onset Type II diabetic patients in the United Kingdom

Dear Sir,

To date five genes of maturity-onset diabetes of the young (MODY) have been cloned [1–5]. Patients with MODY3 (hepatocyte nuclear factor 1 alpha) mutations are usually nonobese and can progress to severe, insulin requiring diabetes masquerading as autoimmune Type I (insulin-dependent) diabetes mellitus. The *HNF1α* gene has been reported to have mutations in 5.5% of Japanese subjects with Type I diabetes [6]. Two studies have indicated that the MODY3 region of chromosome 12 contains a gene which could have a role in late onset Type II (non-insulin-dependent) diabetes mellitus. Evidence of linkage has been shown between MODY3-linked markers and Type II diabetes in a subpopulation with a history of diabetic nephropathy [7]. A genome scan in Finnish families also shows linkage in the MODY3 region with Type II diabetes in a subset of patients having low insulin concentrations [8, 9]. It is thus possible that mutations in *HNF1α* could present as classical Type II diabetes. We hypothesised that *HNF1α* mutations might be identified in newly diagnosed, non-obese patients, BMI less than 27 kg/m², age 25–50 years, with high fasting plasma glucose (fpg) greater than 12 mmol/l and a family history of diabetes ($n = 68$) or in similar patients with fpg less than 8 mmol/l with or without family history ($n = 29$), similar to features of MODY patients with glucokinase mutations [1, 2]. We also studied randomly selected Type II diabetic patients ($n = 71$) and subjects who had gestational diabetes ($n = 38$). Our prior hypothesis was that *HNF1α* mutations might have led to unrecognised, severe beta-cell deficient diabetes in adult patients with Type II diabetes, similar to that which develops in MODY patients with *HNF1α* mutations [1, 10]. Therefore we have systematically analysed the coding, splice site and 474 bases of putative promoter sequences of the hepatocyte nuclear factor 1 alpha gene in 204 white Caucasian Type II diabetes patients in the United Kingdom. Control subjects were non-diabetic partners of diabetic patients matched for age and population in the United Kingdom prospective diabetes study (UKPDS). Sequence was analysed by direct fluorescent sequencing (Perkin-Elmer-ABI, Cheshire, UK, dye primer) in 92 and by single-stranded conformation polymorphism (SSCP) in 112 patients. SSCP variants were confirmed by sequencing. We found five potential variants in the heterozygous state in individual patients, these variants and the characteristics of each patient are summarised in Table 1. We identified two potentially functional variants in the promoter region in two patients. A 2 bp deletion of nucleotides –8 and –9 was identified in one subject of 138 patients and 84 control subjects in a putative C/EBP binding site (sequence GAATTTCCCCAG deleted to GA--TTCCCCAG) close to exon 1. This sequence is conserved in rat, mouse, chicken and frog promoters [11]. A poly-

merase chain reaction (PCR)-restriction fragment polymorphism (RFLP) assay using Apo I confirmed that this deletion was unique to this subject. The second heterozygous variant was G237A, a base conserved in rat and mouse as part of a block of eight residues [12]. This variant was not found in 38 control subjects or 147 other diabetic patients. Three variants from screening of the coding region and flanking exon-intron boundaries, not found in other populations [1, 11, 13, 14], have been identified. We identified a patient who had an exon 7, T492I mutation (C > T) in the transactivation domain that introduced a branch chain amino acid (Threonine → Isoleucine) without a charge change in a nonobese patient who had no family history of diabetes. This variant was not found in 79 control subjects or 197 other diabetic patients. This residue is conserved in several species, e.g. rat, mouse, hamster, xenopus and salmon. The second patient had an exon 7, S498R variant (C > A) with a charge change (Serine → Arginine) in an obese patient with a family history of diabetes treated with insulin and a high fasting glucose (13 mmol/l) at diagnosis. This variant was not found in 79 controls or 197 other diabetic patients. A nonpolar amino acid at this site is conserved across species and the substitution by a basic arginine could be pathogenic. The third exonic variant was found in one patient in exon 4, A301T (G > A) in the transactivation domain. This variant was not found in 105 control subjects or 175 other diabetic patients. This was not a conserved residue and there was no charge change (Alanine → Threonine). We found four of five subjects with novel variants, including the two with promoter variants among the 29 nonobese subjects with fpg less than 8 mmol/l which was more than expected by chance (Fisher's exact test, $p = 0.0019$). We identified three common missense variants described in a previous study [1], I27L, A98 V and S487N. The allele frequencies of these were not significantly different (Chi squared test, data not shown) between diabetic patients and control subjects and it is unlikely that these contribute to diabetes. In addition, the allele frequencies of 12 single nucleotide and noncoding variants (Table 2 in [1], data not shown) showed no difference in prevalence between the diabetic subgroups and the control subjects. This reduces the likelihood of a common functionally important variant contributing to Type II diabetes since it would be expected that some of the 15 single nucleotide polymorphisms would be associated with Type II diabetes because they would be in linkage disequilibrium with a pathogenic mutation. Thus there is no evidence for a common mutation in *HNF1α* or its promoter region that we have failed to identify. These data collected in the United Kingdom do not support the existence of a common mutation, presenting as late onset Type II diabetes, in the *MODY3* gene and is in accord with data in the literature studying Japanese and Danish subjects [13–15]. *HNF1α* mutations probably account for a maximum of 3% of Type II diabetes in the United Kingdom, although we have not tested these mutations functionally. It is possible that this region of chromosome 12 could contain another gene contributing to Type II diabetes, as suggested by genetic mapping studies [7, 8].

Yours sincerely,

R.D. Cox¹, L. Southam¹, Y. Hashim², V. Horton², Z. Mehta², J. Taghavi¹, M. Lathrop¹, R. Turner²

¹ Wellcome Trust Centre for Human Genetics, University of Oxford, Windmill Road, Headington, Oxford. OX3 7BN, UK.

² Address as for corresponding author.

Corresponding author: Dr. Y. Hashim, Diabetes Research Laboratories, Nuffield Department of Clinical Medicine, Oxford University, Radcliffe Infirmary, Woodstock Road, Oxford OX2 6HE, UK

Table 1. Patient characteristics of novel missense or promoter polymorphisms

Site	Codon	bp change (aa change)	Family history of diabetes	Diagnosis			Subsequent therapy at 6 years
				Age (yrs)	Fasting glucose (mmol/l)	BMI (kg/m ²)	
Promoter	-237	G > A	Yes (not insulin)	42	6.2	25	Diet, fasting glucoses 6.4–8.5 mmol/l
Promoter	-8–9	2 bp deletion	No	33	7.2	22	Diet, fasting glucoses 4.9–5.3 mmol/l
Exon 4	301	G > A (Ala/Thr)	No	45	7.2	25	Insulin 24 U (1), fasting glucoses 4.9–8.5 mmol/l
Exon 7	492	C > T (Thr/Ile)	No	37	8	23	Chlorpropamide 100 mg (2), fasting glucoses 5.3–7.2 mmol/l
Exon 7	498	C > A (Ser/Arg)	Yes	46	13	38	Glibenclamide (3) 20 mg & metformin 2550 mg, fasting glucose 12 mmol/l

(1) Insulin requiring when symptomatic on maximum sulphonylurea and metformin therapies

(2) Allocated to chlorpropamide as fasting glucose > 6.0 mmol/l

(3) Allocated to diet and transferred to sulphonylurea therapy when fasting glucose > 15 mmol/l

References

- Yagamata K et al. (1996) Mutations in the hepatocyte nuclear factor-1 alpha gene in maturity onset diabetes of the young (MODY3). *Nature* 384: 455–458
- Vionnet N, Stoffel M, Takeda J et al. (1992) Nonsense mutation in the glucokinase gene causes early-onset non-insulin dependent diabetes mellitus. *Nature* 356: 721–722
- Yagamata K, Furuta H, Oda N et al. (1996) Mutations in the hepatocyte nuclear factor 4 alpha gene in maturity onset diabetes of the young (MODY1). *Nature* 384: 458–460
- Stoffers DA, Ferrer J, Clarke WL, Habener JF (1997) Early-onset Type-II diabetes mellitus (MODY4) linked to IPF1. *Nature Genetics* 17: 138–139
- Horikawa Y, Iwasaki N, Hara M et al. (1997) Mutations in hepatocyte nuclear factor-1 beta (TCF2) associated with MODY. *Nature Genetics* 17: 384–385
- Yamada S, Nishigori H, Onda H et al. (1997) Identification of mutations in the hepatocyte nuclear factor (HNF)-1 alpha gene in Japanese subjects with IDDM. *Diabetes* 46: 1643–1647
- Bowden DW, Sale M, Howard TD (1997) Linkage of genetic markers on human chromosomes 20 and 12 to NIDDM in caucasian sib pairs with a history of diabetic nephropathy. *Diabetes* 46: 882–886
- Mahtani MM et al. (1996) Mapping of a gene for Type II diabetes associated with an insulin secretion defect by a genome scan in Finnish families. *Nature* 14: 90–94
- Lehto M et al. (1997) Characterisation of the MODY3 phenotype. Early-onset diabetes caused by an insulin secretion defect. *J Clin Invest* 99: 582–591
- Hattersley AT (1998) Maturity-onset diabetes of the young: clinical heterogeneity explained by genetic heterogeneity. *Diabetic Medicine* 15: 15–24
- Vaxillaire M, Rouard M, Yagamata K et al. (1997) Identification of nine novel mutations in the hepatocyte nuclear factor 1 alpha gene (HNF-1a) associated with maturity-onset diabetes of the young (MODY3). *Hum Mol Genet* 6: 583–586
- Kaisaki P, Menzel S, Lindner T et al. (1997) Mutations in the hepatocyte nuclear factor-1-alpha gene in MODY and early-onset NIDDM, evidence for a mutational hotspot in exon 4. *Diabetes* 46: 528–535
- Yamada S, Nishigori H, Onda H et al. (1997) Mutations in the Hepatocyte Nuclear Factor -1 alpha Gene (MODY3) Are Not a Major Cause of Late-Onset NIDDM in Japanese Subjects. *Diabetes* 46: 1512–1513
- Iwasaki N, Oda N, Ogata M et al. (1997) Mutations in the hepatocyte nuclear factor-1 alpha/MODY3 gene in Japanese subjects with early- and late-onset NIDDM. *Diabetes* 46: 1504–1508
- Urhammer SA, Rasmussen SK, Kaisaki PJ et al. (1997) Genetic variation in the hepatocyte nuclear factor-1 alpha gene in Danish Caucasian with late-onset NIDDM. *Diabetologia* 40: 473–475

Molecular genetics of MODY in Germany

Dear Sir,

Ledermann has suggested that approximately 2% of all diabetic patients in Germany are likely to have maturity-onset diabetes of the young (MODY) [1]. The contribution of mutations in the known MODY genes to the overall prevalence of this form of

diabetes in Germany is not known. As a first step in addressing this problem, we have studied 11 families of German ancestry with a probable diagnosis of MODY, i.e. diagnosis of diabetes before 25 years of age in at least one family member and other first-degree relatives with MODY or Type II (non-insulin-dependent) diabetes (Fig. 1). Family Dresden-1 was from the state of North Rhine-Westphalia, families Dresden-2, -4, -6, -7, -13, -15, and -17 from Saxony, and families Dresden-3, -5, and -11 were from Thuringia. The average age at diagnosis of diabetes in the probands was 18.2 ± 6.6 (range 12–25 years) (means \pm SEM) and included four men and seven women. Of the eleven probands, six, two and three were currently being treated with insulin, oral hypoglycaemic agents and diet, respectively. The average body mass index (BMI) was 25.7 ± 5.9 kg/m² (range

Corresponding author: T.Lindner, M.D., Ph. D., Howard Hughes Medical Institute, The University of Chicago, 5841 South Maryland Avenue, MC1028, Chicago IL 60637, USA