

## Is omega-3 key to unlocking inflammation in obesity?

P. J. White · A. Marette

Published online: 7 July 2006  
© Springer-Verlag 2006

Low-grade inflammation has been identified as a key player in the development of the metabolic syndrome in obese subjects, leading the way to type 2 diabetes and cardiovascular diseases. Insulin resistance, a primary component of the metabolic syndrome, is characterised by chronically elevated concentrations of proinflammatory cytokines, acute phase proteins, and enhanced activation of proinflammatory signalling in insulin-responsive tissues of obese subjects.

The research effort thus far has established that, in obesity, the expanding adipose tissue makes a substantial contribution to the development of obesity-linked inflammation via the secretion of various proinflammatory cytokines, chemokines and adipokines. Recent investigations into the mechanisms responsible for this phenomenon have outlined a role for the macrophage [1–3]. In fact, macrophage accumulation in adipose tissue is positively correlated with adipocyte size and contributes to the expression of proinflammatory mediators of insulin resistance, such as TNF- $\alpha$ , IL-6 and inducible nitric oxide synthase (iNOS) [1, 2]. Furthermore, both inhibition of macrophage function, via myeloid-specific I $\kappa$ B kinase- $\beta$  deletion [3], and prevention of macrophage accumulation in adipose tissue, via deletion of C–C motif chemokine receptor-2 [4], prevent the development of the inflammatory phenotype and insulin resistance with obesity. Hence, there is growing support for the current notion that

macrophage infiltration into adipose tissue is central to obesity-related metabolic disorders.

Pioneering work by Storlien and colleagues has shown that long-chain omega-3 polyunsaturated fatty acids (LCn-3PUFA) can prevent the development of diet-induced insulin resistance in rats [5]. Furthermore, a growing number of studies support a link between this beneficial effect of LCn-3PUFA and an anti-inflammatory mechanism. LCn-3PUFA have been widely reported to have anti-inflammatory effects in a range of chronic inflammatory conditions, including rheumatoid arthritis and Crohn's disease [6, 7]. Treatment of obese subjects with LCn-3PUFA in a clinical setting has also been shown to reduce circulating levels of both proinflammatory cytokines and acute phase proteins [8, 9]. Whether these anti-inflammatory actions of LCn-3PUFA, and their positive influence on the metabolic syndrome, can be linked to local blunting of adipose tissue inflammation is not yet known.

In this issue of Diabetologia, Todoric et al. [10] present evidence that the inclusion of LCn-3PUFA in a high-fat diet prevents the development of an inflammatory gene expression profile and macrophage infiltration in the adipose tissue of obese diabetic *db/db* mice. Despite significantly enhancing weight gain, LCn-3PUFA treatment completely prevented the diet-induced adipose tissue switch to an inflammatory profile, attenuating the upregulation of an array of genes, notably those encoding the macrophage surface marker CD68, macrophage chemotactic protein-1, and the lipopolysaccharide receptor CD14. Incorporation of LCn-3PUFA into the high-fat diet also prevented the downregulation of genes involved in lipid metabolism, including those for fatty acid synthase and hormone-sensitive lipase, in adipose tissue of the obese mice. The anti-inflammatory effect of LCn-3PUFA was related to greatly diminished macrophage migration into

P. J. White · A. Marette (✉)  
Department of Anatomy—Physiology and Lipid Research Unit,  
Laval University Hospital Research Centre,  
2705 Laurier Blvd,  
G1V 4G2 Ste-Foy, QC, Canada  
e-mail: andre.marette@crchul.ulaval.ca

adipose tissue and substantially enhanced circulating adiponectin levels. *n*-3PUFA as part of the high-fat diet also lowered circulating triglyceride to levels below those seen in chow-fed mice. Although these data suggest that improvement of metabolic status in obesity by *LCn*-3PUFA involves regulation of the adipose tissue gene expression profile and the prevention of macrophage infiltration into fat depots, further research is required to elucidate the exact mechanisms responsible.

Indeed, there are a multitude of potential mechanisms that could account for the anti-inflammatory actions elicited by *LCn*-3PUFA in this study (Fig. 1). For example, the competitive inhibition of arachidonate-derived proinflammatory leukotriene and prostaglandin production by *LCn*-3PUFA [11–13] could limit inflammation in adipose tissue. Negative modulation of monocyte function by *LCn*-3PUFA [14] could also account for the reduced macrophage infiltration. Interestingly, more recent studies have also identified *LCn*-3PUFA as precursors of two novel families of potent anti-inflammatory lipid mediators, the resolvins and protectins [15]. We may conjecture that these eicosanoids could contribute to the *LCn*-3PUFA-mediated inhibition of adipose tissue inflammation reported by Todoric and colleagues by favouring the resolution of the inflammatory process. Alternatively, *LCn*-3PUFA may blunt adipose tissue inflammation by acting as endogenous ligands for the peroxisome proliferator-activator receptors (PPARs)  $\alpha$ ,  $\delta$  and  $\gamma$ , which have recently been recognised for their anti-inflammatory actions in adipocytes and macrophages [16–20].

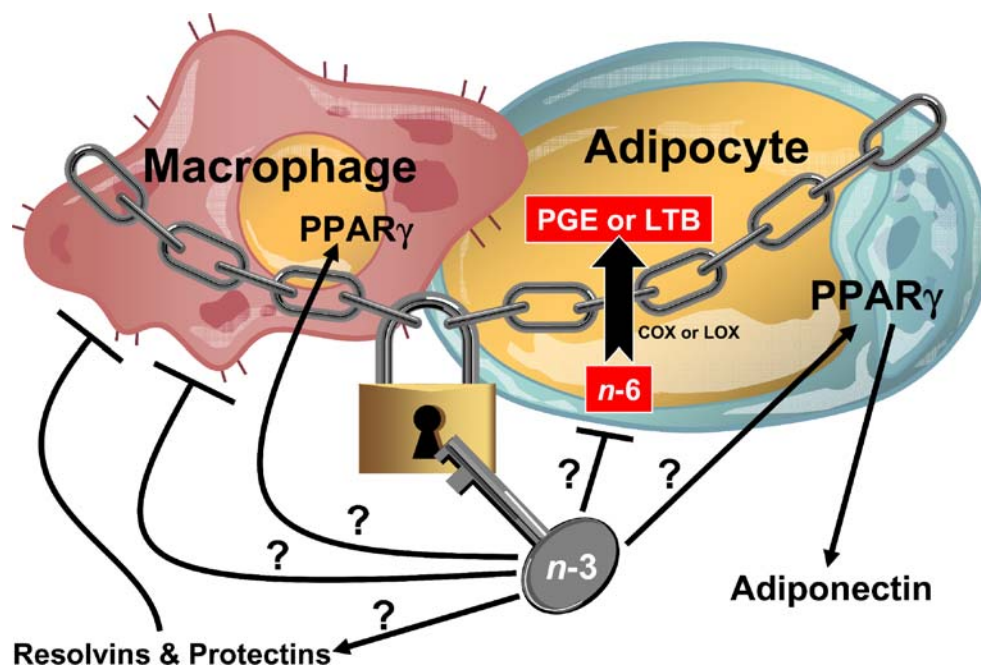
Of the three known mammalian PPAR isoforms, *LCn*-3PUFA appear to bind most efficiently with PPAR $\gamma$  [21].

On the basis of current literature, this latter target of *LCn*-3PUFA appears to be the most plausible mechanistic explanation for the findings of Todoric and colleagues. Accordingly, PPAR $\gamma$  agonists have been reported to inhibit macrophage activation [22] and infiltration into adipose tissue [2]. Moreover, a very recent study using the PPAR $\gamma$  inhibitor bisphenol A diglycidyl ether (BADGE) demonstrated that treatment with *LCn*-3PUFA augments circulating adiponectin levels via a PPAR $\gamma$ -dependent mechanism [17]. Given that adiponectin is known to exert anti-inflammatory effects and enhance insulin sensitivity [23, 24], it is conceivable that *LCn*-3PUFA could impede the adipose tissue switch to an inflammatory gene expression profile in response to obesity via a PPAR $\gamma$ - and adiponectin-dependent mechanism.

In accord with this theory, the use of PPAR $\gamma$  agonists is characterised by enhanced weight gain [25] similar to that seen with *LCn*-3PUFA treatment in this study. However, it should be noted that not all studies using *LCn*-3PUFA reported increased weight gain. Indeed, to the contrary, the prevention of weight gain in insulin resistance prevention studies is often reported following treatment with *LCn*-3PUFA [26]. This heterogeneity as regards the influence of *LCn*-3PUFA on weight gain may stem from differences in the animal models used, the mode of treatment (e.g. use for prevention or reversal of insulin resistance), or might result from differential degrees of *LCn*-3PUFA signalling through the PPAR $\gamma$  and PPAR $\alpha$  receptors in these studies [25].

In conclusion, these mechanisms remain to be explored and it is very likely that the results presented in the article by Todoric and colleagues reflect a combination of the pleiotropic functions of *LCn*-3PUFA at multiple sites within the adipose

**Fig. 1** Proposed mechanisms for the anti-inflammatory actions of *LCn*-3PUFA in adipose tissue. *COX* Cyclooxygenase, *PGE* prostaglandin, *LOX* 5-lipoxygenase, *LTB* leukotriene



tissue. Additional studies are certainly warranted to examine whether such effects of LCN-3PUFA are present in other insulin-sensitive tissues, including liver and skeletal muscle, and to elucidate the underlying mechanisms. It now seems critical to determine whether the adipose-specific blunting of an inflammatory profile is the principal mechanism by which LCN-3PUFA prevent diet-induced insulin resistance.

**Acknowledgement** A. Marette is the recipient of a CIHR Investigator Award and is a National Researcher of the Fonds de la Recherche en Santé du Québec.

## References

- Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW Jr (2003) Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 112:1796–1808
- Xu H, Barnes GT, Yang Q et al (2003) Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. *J Clin Invest* 112:1821–1830
- Arkan MC, Hevener AL, Greten FR et al (2005) IKK-beta links inflammation to obesity-induced insulin resistance. *Nat Med* 11:191–198
- Weisberg SP, Hunter D, Huber R et al (2006) CCR2 modulates inflammatory and metabolic effects of high fat feeding. *J Clin Invest* 116:115–124
- Storlien LH, Kraegen EW, Chisholm DJ, Ford GL, Bruce DG, Pascoe WS (1987) Fish oil prevents insulin resistance induced by high-fat feeding in rats. *Science* 237:885–888
- Geusens P, Wouters C, Nijs J, Jiang Y, Dequeker J (1994) Long term effect of omega-3 fatty acid supplementation in active rheumatoid arthritis. A 12 month, double blind, controlled study. *Arthritis Rheum* 37:824–829
- Belluzzi A, Brignola C, Campieri M, Pera A, Boschi S, Miglioli M (1996) Effect of an enteric-coated fish-oil preparation on relapses in Crohn's disease. *N Engl J Med* 334:1557–1560
- Rallidis LS, Paschos G, Liakos GK, Velissaridou AH, Anastasiadis G, Zampelas A (2003) Dietary  $\alpha$ -linolenic acid decreases C-reactive protein, serum amyloid A and interleukin-6 in dyslipidaemic patients. *Atherosclerosis* 167:237–242
- Trebble T, Arden NK, Stroud MA et al (2003) Inhibition of tumour necrosis factor- $\alpha$  and interleukin 6 production by mononuclear cells following dietary fish-oil supplementation in healthy men and response to antioxidant co-supplementation. *Br J Nutr* 90:405–412
- Todoric J, Löffler M, Huber J et al (2006) Adipose tissue inflammation induced by high-fat diet in obese diabetic mice is prevented by *n*-3 polyunsaturated fatty acids. *Diabetologia* DOI 10.1007/s00125-006-0300-x
- Lee TH, Mencia-Huerta JM, Shih C, Corey EJ, Lewis RA, Austen KF (1984) Effects of exogenous arachidonic, eicosapentaenoic, and docosahexaenoic acids on the generation of 5-lipoxygenase pathway products by ionophore-activated human neutrophils. *J Clin Invest* 74:1922–1933
- Needleman P, Raz A, Minkes MS, Ferrendelli JA, Sprecher H (1979) Triene prostaglandins: prostacyclin and thromboxane biosynthesis and unique biological properties. *Proc Natl Acad Sci U S A* 76:944–948
- Corey EJ, Shih C, Cashman JR (1983) Docosahexaenoic acid is a strong inhibitor of prostaglandin but not leukotriene biosynthesis. *Proc Natl Acad Sci U S A* 80:3581–3584
- Hughes DA (1998) In vitro and in vivo effects of *n*-3 polyunsaturated fatty acids on human monocyte function. *Proc Nutr Soc* 57:521–525
- Bannenberg GL, Chiang N, Ariel A et al (2005) Molecular circuits of resolution: formation and actions of resolvins and protectins. *J Immunol* 174:4345–4355
- Kliwer SA, Sundseth SS, Jones SA et al (1997) Fatty acids and eicosanoids regulate gene expression through direct interactions with peroxisome proliferator-activated receptors  $\alpha$  and  $\gamma$ . *Proc Natl Acad Sci U S A* 94:4318–4323
- Neschen S, Morino K, Rossbacher JC et al (2006) Fish oil regulates adiponectin secretion by a peroxisome proliferator-activated receptor- $\gamma$ -dependent mechanism in mice. *Diabetes* 55:924–928
- Pilon G, Dallaire P, Marette A (2004) Inhibition of inducible nitric-oxide synthase by activators of AMP-activated protein kinase: a new mechanism of action of insulin-sensitizing drugs. *J Biol Chem* 279:20767–20774
- Lee CH, Chawla A, Urbiztondo N et al (2003) Transcriptional repression of atherogenic inflammation: modulation by PPAR $\delta$ . *Science* 302:453–457
- Tsuchida A, Yamauchi T, Takekawa S et al (2005) Peroxisome proliferator-activated receptor (PPAR)  $\alpha$  activation increases adiponectin receptors and reduces obesity-related inflammation in adipose tissue: comparison of activation of PPAR $\alpha$ , PPAR $\gamma$ , and their combination. *Diabetes* 54:3358–3370
- Xu HE, Lambert MH, Montana VG et al (1999) Molecular recognition of fatty acids by peroxisome proliferator-activated receptors. *Mol Cell* 3:397–403
- Ricote M, Li AC, Willson TM, Kelly CJ, Glass CK (1998) The peroxisome proliferator activated receptor- $\gamma$  is a negative regulator of macrophage activation. *Nature* 319:79–82
- Berg AH, Combs TP, Du X, Brownlee M, Scherer PE (2001) The adipocyte-secreted protein Acrp30 enhances hepatic insulin action. *Nat Med* 7:947–953
- Yamauchi T, Kamon J, Waki H et al (2001) The fat derived hormone adiponectin reverses insulin resistance associated with both lipoatrophy and obesity. *Nat Med* 7:941–946
- Carmona MC, Louch K, Nibbelink M et al. (2005) Fenofibrate prevents rosiglitazone-induced body weight gain in *ob/ob* mice. *Int J Obes* 29:864–871
- Wang H, Storlien LH, Huang XF (2002) Effects of dietary fat types on body fatness, leptin, and ARC leptin receptor, NPY, and AgRP mRNA expression. *Am J Physiol Endocrinol Metab* 282: E1352–E1359