

Short Communication

The *PIA1/A2* polymorphism of platelet glycoprotein IIIa is not associated with the risk of type 2 diabetes. The Ludwigshafen Risk and Cardiovascular Health Study

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Abstract

Aims/hypothesis. The *PIA1/A2* polymorphism of platelet glycoprotein IIIa (GPIIIa) has been implicated in the pathogenesis of type 2 diabetes. We studied this polymorphism in a homogenous, extensively phenotyped cohort using the candidate gene approach.

Methods. The *PIA1/A2* polymorphism was determined in 1051 patients with type 2 diabetes and in 2247 individuals without type 2 diabetes.

Results. In patients with type 2 diabetes, genotype frequencies were as follows: *PIA1/A1* 71.4%, *PIA1/A2* 26.0%, and *PIA2/A2* 2.7%. In individuals without type 2 diabetes, genotype frequencies were 71.6%, 25.7% and 2.8%, respectively. The *PIA2* allele was not associated with fasting and postprandial glucose, glycated

haemoglobin, insulin, proinsulin, C-peptide and calculated indices of insulin resistance or pancreatic beta cell function. The *PIA2* allele was also not significantly associated with angiographic CHD (adjusted odds ratio [OR] 1.13; 95% CI, 0.93–1.39) or with a history of previous myocardial infarction (adjusted OR 1.09; 95% CI, 0.87–1.37).

Conclusions/interpretation. The GPIIIa *PIA1/A2* polymorphism is not associated with type 2 diabetes, glucose metabolism, angiographic CHD or myocardial infarction.

Keywords Coronary heart disease · Genetics · Glucose · Human platelet glycoprotein IIIa polymorphism · Myocardial infarction · Type 2 diabetes mellitus

Introduction

Platelet glycoprotein IIb/IIIa is a membrane receptor for fibrinogen and von Willebrand factor. The *PIA1/A2* polymorphism of the glycoprotein IIIa (GPII-

Ia) subunit results from a T to C transition of the *GPII-Ia* gene, replacing leucine by proline at amino acid residue 33 of GPIIIa [1]. This polymorphism has been implicated in vascular disease (review see [2]). A recent case-control study of 215 subjects reported a more than two-fold higher prevalence of *PIA2* in patients with type 2 diabetes than in individuals with normal glucose metabolism [3]. In an attempt to substantiate this observation, we analysed the association between the GPIIIa *PIA1/PIA2* polymorphism and type 2 diabetes and CHD in 3298 individuals.

Subjects and methods

Study design. Our study includes Caucasian patients hospitalised for coronary angiography [4]. Clinically relevant CHD was defined as the presence of a visible luminal narrowing ($\geq 20\%$ stenosis) in at least one of 15 coronary segments [5].

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Abbreviations: GPIIIa, glycoprotein IIIa · OR, odds ratio

Table 1. Characteristics of CHD patients and control subjects^a

	No type 2 diabetes		type 2 diabetes		<i>p</i> type 2 diabetes vs. no type 2 diabetes
	Men	Women	Men	Women	
<i>n</i> =	1571	676	728	323	
Age, years	60.3±11.1	63.4±10.7	65±9.0	67.6±8.6	<0.001 ^b
BMI, kg/m ²	27.2±3.6	26.6±4.3	28.3±4.1	28.8±5.0	<0.001 ^c
Smoker (former and current), <i>n</i> (%)	1182 (75.2)	575 (79.0)	246 (36.4)	110 (34.1)	0.004 ^c
CHD, <i>n</i> (%)	1261 (80.3)	394 (58.3)	660 (90.7)	252 (78.0)	<0.001 ^c
Hypertension ^d , <i>n</i> (%)	1033 (65.8)	484 (71.6)	605 (83.1)	275 (85.1)	<0.001 ^c
Beta blocker, <i>n</i> (%)	1023 (65.1)	392 (58.0)	459 (63.0)	212 (65.6)	NS ^c
ACE inhibitors, <i>n</i> (%)	810 (51.6)	312 (46.2)	448 (61.5)	193 (59.8)	<0.001 ^c
AT1 receptor antagonists, <i>n</i> (%)	61 (3.9)	30 (4.4)	36 (4.9)	20 (6.2)	NS ^c
Calcium channel blockers, <i>n</i> (%)	195 (12.4)	110 (16.3)	142 (19.5)	68 (21.1)	<0.002 ^c
Aspirin and/or other antiplatelet agents, <i>n</i> (%)	1154 (73.5)	439 (64.9)	531 (72.9)	232 (71.8)	NS ^c
Lipid-lowering drugs, <i>n</i> (%)	792 (50.4)	283 (41.9)	374 (51.4)	142 (47.1)	NS ^c
Systolic blood pressure, mm Hg	139±23	139±24	146±23	146±24	<0.001 ^b
Diastolic blood pressure, mm Hg	81±11	79±11	82±11	80±12	NS
Fasting plasma glucose, mmol/l	5.45±0.59	5.36±0.55	7.90±2.47	8.09±2.79	<0.001 ^b
LDL cholesterol, mmol/l	2.98±0.83	3.26±0.97	2.86±0.86	3.07±0.94	<0.001 ^b
HDL cholesterol, mmol/l	0.97±0.26	1.18±0.30	0.90±0.23	1.01±0.27	<0.001 ^b
Triglycerides, mmol/l					
Geometric mean ± standard deviation	1.74±1.44	1.56±0.98	1.98±1.52	2.15±1.38	<0.001 ^{b,e}
Median (25th and 75th percentile)	1.68 (1.26–2.30)	1.52 (1.16–2.05)	1.91 (1.43–2.56)	2.05 (1.52–2.88)	
HbA _{1c} , %	5.82±0.58	5.86±0.54	7.24±1.60	7.51±1.65	<0.001 ^b

^a All values are means ± standard deviations, unless stated otherwise. ^b Adjusted for sex and age by analysis of variance; ^c adjusted for sex and age by logistic regression; ^d history of hypertension and/or blood pressure >140/90 mm Hg; ^e analysis

of variance of logarithmically transformed values adjusted for sex and age. Levene's test of equality of error variances: *p*=0.153 for the null hypothesis of equal variances across the groups. AT1, angiotensin I

Patients with type 1 diabetes were excluded from this analysis. The study was approved by the ethics review committee at the "Landesärztekammer Rheinland-Pfalz" (Medical Council of Rhineland Palatinate, Mainz, Germany) and written informed consent was obtained from the participants.

Laboratory procedures. Fasting blood glucose was determined enzymatically using the hexokinase method (Roche Diagnostics, Mannheim, Germany). Cholesterol and triglycerides were measured using enzymatic reagents from Wako (Neuss, Germany) [4]. LDL and HDL cholesterol were measured by a combined ultracentrifugation–precipitation method [4]. HbA_{1c} was determined by high-performance liquid chromatography (DIAMAT, Chromsystem Instruments & Chemicals, Martinsried, Germany). Enzyme immunoassays were used to determine the following: insulin (AIA-PACK IRI, Eurogenetics, Eschborn, Germany), C-peptide (AIA-PACK C-peptide, Eurogenetics), and pro-insulin (IBL, Hamburg, Germany). The GPIIIa PIA1/A2 polymorphism was determined with a multilocus genotyping assay [6]. The accuracy of this assay has recently been evaluated by direct sequencing or restriction fragment length analysis [7]. We determined the reproducibility of this method in a subset of 165 individuals who were enrolled for a second or third investigation in the Ludwigshafen Risk and Cardiovascular Health Study (*n*=147 twice, *n*=18 three times). A total of 183 (147+18+18) additional DNA samples were genotyped in a blinded fashion. In each of the 183 samples, we obtained consistent results.

Metabolic studies. An OGTT was performed in individuals not previously diagnosed as having type 2 diabetes. Normoglycemia, IGT and type 2 diabetes were diagnosed according to the American Diabetes Association (ADA) criteria [8]. In addition, individuals with a history of diabetes or treatment with hypoglycaemic agents were defined as diabetic. Insulin sensitivity and pancreatic beta cell function were estimated using HOMA-IR and %beta respectively [9]. Subjects with LDL cholesterol levels greater than 4.1 mmol/l or triglyceride levels greater than 2.3 mmol/l or HDL cholesterol lower than 1 mmol/l were considered dyslipidaemic.

Statistical analysis. Continuous variables between CHD patients and control subjects were compared by analysis of variance, adjusting for sex and age (Table 1). Associations between genotypes and clinical and biochemical variables (Table 2) were evaluated by the Mann–Whitney rank sum test. Associations between categorical variables were examined by logistic regression analysis, including covariables as indicated (Table 3). In models assuming a co-dominant (additive) effect of the PIA2-allele, the genotypes PIA1/A1, PIA1/A2 and PIA2/A2 were coded as 0, 1, and 2 respectively. When assuming a dominant effect, genotype PIA1/A1 was coded as 0, and PIA1/A2 and PIA2/A2 combined as 1. When assuming a recessive effect, genotypes PIA1/A1 and PIA1/A2 were coded as 0, and PIA2/A2 as 1. The SPSS statistical package, version 11.0 for Windows (SPSS, Chicago, Ill., USA) was used.

Table 2. Effects of the PIA1/A2 polymorphism on glucose and insulin metabolism

	GPIIIa PIA1/A2 genotype			<i>p</i>	<i>p</i>
	A1/A1	A1/A2	A2/A2		
<i>n</i> (%)	1433 (71.8)	501 (25.1)	63 (3.2)		
Normal glucose tolerance, <i>n</i> (%)	596 (71.8)	211 (25.4)	22 (2.8)		
IGT, <i>n</i> (%)	513 (72.8)	169 (24.0)	23 (3.3)		
Type 2 diabetes, <i>n</i> (%)	324 (70.1)	121 (26.2)	17 (3.7)		
HbA _{1c} , % ± SD	5.95±0.74	5.97±0.82	5.92±0.72	NS	NS
Fasting plasma glucose, mmol/l, ± SD	5.72±1.11	5.76±1.15	5.67±1.07	NS	NS
OGTT 1-h plasma glucose, mmol/l, ± SD	9.49±3.00	9.51±2.95	10.00±2.92	NS	NS
OGTT 2-h plasma glucose, mmol/l, ± SD	7.55±3.21	7.67±3.57	7.85±3.41	NS	NS
Fasting insulin, mU/l ± SEM	11.6±0.3	11.9±0.6	10.2±0.9	NS	NS
Fasting C-peptide, µg/l ± SEM	2.38±0.05	2.36±0.09	2.17±0.17	NS	NS
Fasting pro-insulin, ng/l ± SEM	10.0±0.3	9.4±0.4	9.7±1.3	NS	NS
HOMA-IR, mmol·mU ⁻¹ ·l ⁻² ,	2.16	2.19	2.34	NS	NS
Median (25th and 75th perc.)	(1.35–3.52)	(1.41–3.32)	(1.40–3.63)		
Beta%, mU/mmol,	87	86	90	NS	NS
Median (25th and 75th percentile)	(57–134)	(59–122)	(62–124)		
Fasting pro-insulin/insulin, ng/mU,	0.81	0.80	0.92	NS	NS
Median (25th and 75th percentile)	(0.48–1.41)	(0.50–1.25)	(0.44–1.27)		
Fasting pro-insulin/C-peptide, ng/µg,	3.6	3.6	3.4	NS	NS
Median (25th and 75th percentile)	(1.9–9.5)	(1.9–8.4)	(2.1–9.5)		

HOMA-IR, homeostasis model assessment for insulin resistance

Table 3. Odds ratios for angiographic CHD and myocardial infarction according to the GPIIIa PIA1/A2 polymorphism

GPIIIa PIA1/A2 genotype	Model 1 OR (95% CI)	<i>p</i>	Model 2 OR (95% CI)	<i>p</i>	Model 3 OR (95% CI)	<i>p</i>
Angiographic CHD^a						
A1/A1	1.0 _{reference}		1.0 _{reference}		1.0 _{reference}	
A1/A2	1.07 (0.88–1.30)	NS	1.13 (0.93–1.39)	NS	1.13 (0.92–1.39)	NS
A2/A2	1.09 (0.65–1.82)	NS	1.18 (0.68–2.04)	NS	1.16 (0.66–2.04)	NS
A1/A1 vs A1/A2 or A2/A2	1.08 (0.90–1.29)	NS	1.14 (0.94–1.38)	NS	1.13 (0.93–1.39)	NS
A1/A1 or A1/A2 vs A2/A2	1.07 (0.64–1.78)	NS	1.14 (0.66–1.97)	NS	1.13 (0.64–1.97)	NS
MI^b						
A1/A1	1.0 _{reference}		1.0 _{reference}		1.0 _{reference}	
A1/A2	1.05 (0.86–1.30)	NS	1.13 (0.90–1.41)	NS	1.11 (0.88–1.41)	NS
A2/A2	1.06 (0.60–1.86)	NS	0.95 (0.52–1.73)	NS	0.93 (0.49–1.74)	NS
A1/A1 vs A1/A2 or A2/A2	1.05 (0.86–1.29)	NS	1.11 (0.89–1.38)	NS	1.09 (0.87–1.37)	NS
A1/A1 or A1/A2 vs A2/A2	1.04 (0.60–1.83)	NS	0.92 (0.50–1.67)	NS	0.90 (0.48–1.69)	NS

^a Comparison of patients with angiographic CHD (*n*=2567) and individuals in whom significant (≥20% stenosis) CHD had been ruled out by angiography (*n*=731). ^b Comparison of patients with a clinical history of myocardial infarction (*n*=1357) and individuals without myocardial infarction and the absence

of significant (≥20% stenosis) CHD (*n*=727). Model 1: unadjusted. Model 2: adjusted for age and sex. Model 3: adjusted, in addition, for type 2 diabetes, BMI, smoking, hypertension and dyslipidaemia

Results

Patients with type 2 diabetes (*n*=1051) were significantly older and had a higher mean BMI than individuals without type 2 diabetes (*n*=2247). Current or past smoking was less prevalent in the former; angiographic CHD and hypertension were more frequent. More patients with type 2 diabetes than those without were taking ACE inhibitors and calcium channel blockers,

had higher systolic blood pressure, higher fasting glucose, higher triglycerides and lower HDL cholesterol. Mean LDL cholesterol was significantly lower in patients with type 2 diabetes than in subjects who did not have type 2 diabetes (Table 1).

The genotypes PIA1/A1, PIA1/A2, PIA2/A2 occurred at similar frequencies both in patients with type 2 diabetes (71.4%, 26.0%, and 2.7% respectively), and in non-diabetic individuals (71.6%, 25.7%, and

2.8% respectively). Logistic regression analysis assuming co-dominant (additive), dominant or recessive effects of the PIA2 allele did not reveal any association with type 2 diabetes, regardless of whether or not we included age, sex, BMI or CHD status as co-variables (data not shown).

To investigate whether the GPIIIa PIA1/A2 polymorphism affected glucose metabolism, we analysed 1997 individuals, in whom an OGTT was performed and who were not being treated with oral hypoglycaemic agents or insulin. This subgroup consisted of subjects with normal glucose tolerance, IGT and newly diagnosed type 2 diabetes. The PIA1/A2 polymorphism was not significantly associated with any of the measures of glucose metabolism (Table 2).

The A2 allele was also not significantly associated with the prevalence of angiographic CHD and a history of myocardial infarction. Adjustments for established cardiovascular risk factors including age, sex, BMI, type 2 diabetes, smoking, hypertension and dyslipidaemia slightly increased ORs, but none of the associations reached statistical significance (Table 3).

Discussion

The current study has two key results. First, we did not obtain any evidence for a relationship between the PIA1/A2 polymorphism and glucose metabolism. Second, the PIA1/A2 polymorphism was not associated with CHD or with a history of myocardial infarction, a finding which is completely consistent with previous findings [2].

Type 2 diabetes is influenced by genetic factors. So far, linkage scans and association studies have revealed only few consistent susceptibility loci for type 2 diabetes [10]. As classical linkage may be of limited merit in the genetic analysis of polygenic diseases, association studies might be the strategy of choice to unravel the genetic causes of type 2 diabetes [11]. However, association studies have produced highly discrepant results, especially when numbers of cases and controls have been small. In many instances, the positive associations seen in small studies have been disproven in subsequent studies of large cohorts. A recent study [3], claiming an association between the GPIIIa PIA1/A2 polymorphism and type 2 diabetes, included 112 patients with type 2 diabetes and 103 control subjects, with the two groups originating from different clinical settings. The type 2 diabetes patients enrolled in that study were recruited at a diabetes clinic, the control subjects were people presenting with chest-pain at a cardiology department. This recruitment strategy is clearly different from that used in our study, which exclusively enrolled patients scheduled for angiography. An important reason for the false-positive findings by Tschoepe and co-workers [3] may be that the PIA2 allele occurred at a fre-

quency of 14.6% only in their non-diabetic control subjects, a value substantially lower than the prevalence of 28.5% found in the current study and the prevalence of 26.2% found in a recent meta-analysis of 8829 individuals without previous myocardial infarction [2].

Our study, however, may still have limitations. We exclusively enrolled individuals with clinically indicated coronary angiography, and this may have resulted in a referral bias. As to the definition of the coronary artery status, this is also a possible strength of our study. The prevalence of clinically asymptomatic coronary atherosclerosis has been reported to be high at or above 50 years of age [12]. Hence, angiography-based recruitment of control subjects rules out the possibility of individuals with significant, yet clinically unapparent coronary artery disease being inadvertently allocated to the control group. Surprisingly, our control group appears well representative, since the major cardiovascular risk factors occur at a frequency similar to that found in the general population. The prevalence of hypertension is close to that found in a random probability sample from Germany [13]. Diabetes mellitus appears two to three times more frequently than in the German population [14]. This, however, is probably due to the fact that we did not rely on self-reports. Rather, we measured fasting glucose and performed an oral glucose challenge in individuals not previously known to have diabetes mellitus. Based on clinical history or fasting glucose measurements, the National Health and Nutrition Examination survey 1999–2000 reports prevalences of diabetes mellitus of 9.2% in adults between 40 and 59 years of age and 19.3% in adults older than 60 years [15]. In the current study, 12.1% of the control subjects had diabetes mellitus according to this criterion, while another 5.6% were detected by elevated post-challenge glucose only.

In summary, our results strongly argue against a role of this polymorphism in the development of type 2 diabetes and indicate that sample sizes have to be adequately large in genetic association studies to avoid false positive findings.

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