



# Analysis of the action mechanisms and targets of herbal anticonvulsants highlights opportunities for therapeutic engagement with refractory epilepsy

Sobia Tabassum<sup>1</sup> · Susan Shorter<sup>2</sup> · Saak V. Ovsepian<sup>2,3</sup>

Received: 11 December 2023 / Revised: 26 March 2024 / Accepted: 5 April 2024  
© Crown 2024

## Abstract

Epilepsy is a neurological disorder characterized by spontaneous and recurring seizures. It poses significant therapeutic challenges due to diverse etiology, pathobiology, and pharmacotherapy-resistant variants. The anticonvulsive effects of herbal leads with biocompatibility and toxicity considerations have attracted much interest, inspiring mechanistic analysis with the view of their use for engagement of new targets and combination with antiseizure pharmacotherapies. This article presents a comprehensive overview of the key molecular players and putative action mechanisms of the most common antiepileptic herbals demonstrated in tissue culture and preclinical models. From the review of the literature, it emerges that their effects are mediated via five distinct mechanisms: (1) reduction of membrane excitability through inhibition of cation channels, (2) improvement of mitochondrial functions with antioxidant effects, (3) enhancement in synaptic transmission mediated by GABA<sub>A</sub> receptors, (4) improvement of immune response with anti-inflammatory action, and (5) suppression of protein synthesis and metabolism. While some of the primary targets and action mechanisms of herbal anticonvulsants (1, 3) are shared with antiseizure pharmacotherapies, herbal leads also engage with distinct mechanisms (2, 4, and 5), suggesting new drug targets and opportunities for their integration with antiseizure medications. Addressing outstanding questions through research and in silico modeling should facilitate the future use of herbals as auxiliary therapy in epilepsy and guide the development of treatment of pharmacoresistant seizures through rigorous trials and regulatory approval.

**Keywords** Epilepsy · Medicinal herbs · Biomarkers · Antiseizure medications · Ion channels · mTOR signaling · Neuroprotection

## Abbreviations

ASM	Antiseizure medications	mEC	Medial entorhinal cortex
NMDA	<i>N</i> -Methyl- <i>D</i> -aspartate	PTZ	Pentylentetrazol
SE	Status epilepticus	RIN	Rhynchophylline
SREDS	Spontaneous recurrent epileptiform discharges	$I_{NaP}$	Persistent sodium current
VGCC	Voltage-gated calcium channels	SSA	Saikosaponin A
FS	Febrile seizures	HNC	Hippocampal neuron culture
VGSC	Voltage-gated sodium channels	ROS	Reactive oxygen species
TLE	Temporal lobe epilepsy	TNF	Tumor necrosis factor
		IL	Interleukin
		DPPH	2,2-Diphenyl-1-picrylhydrazyl
		ANTS	2,2'-Azino-bis 3-ethylbenzothiazoline-6-sulfonic acid
		KA	Kainic acid
		E/I	Excitatory–inhibitory ratio
		GABA	$\gamma$ -Amino butyric acid
		PTA	Pinellia total alkaloids
		GAT-1	GABA transporter-1
		GAD65	Glutamate decarboxylase 65
		GABA-T	GABA transaminase

✉ Saak V. Ovsepian  
s.v.ovsepian@gre.ac.uk

<sup>1</sup> Department of Biological Sciences, Faculty of Sciences, International Islamic University, Islamabad, Pakistan

<sup>2</sup> Faculty of Engineering and Science, University of Greenwich London, Chatham Maritime, Kent ME4 4TB, UK

<sup>3</sup> Faculty of Medicine, Tbilisi State University, Tbilisi 0177, Republic of Georgia

PTE	Posttraumatic epilepsy
FAC	Ferric ammonium citrate
12/15-LOX	12/15-Lipoxygenase
HMGB1	High-mobility group box 1
BDNF	Brain-derived neurotrophic factor
p-NF- $\kappa$ B	Phosphorylated transcription factor nuclear factor kappa B
mTOR	Mammalian target of the rapamycin
PCR	Polymeric chain reaction
COX-2	Cyclooxygenase 2
NO	Nitric oxide

## Introduction

Epilepsy is one of the most common and potentially fatal neurological conditions affecting ~70 million individuals worldwide. According to the World Health Organization, the average incidence of epilepsy in industrialized countries is ~50 per 100,000 population, with nearly double of that in developing countries [1, 2]. Despite remarkable progress in research and development of effective pharmacotherapies, in 20–30% of cases, antiepileptic drugs fail to control clinical seizures [3–5]. Affecting one in four of the general population of patients, pharmacoresistant epilepsy is associated with high risks of injuries related to seizures and major psychosocial trauma, occasionally leading to disability and premature death [6, 7]. Moreover, the majority of antiseizure medications (ASM) have multiple adverse neurological effects, including drowsiness, headaches, dizziness, agitation, blurred vision, and uncontrollable shaking, calling for new medicines that are tolerated better and have minimal side effects [8, 9]. Notably, some of the frontline antiseizure pharmacotherapies may also cause endocrinal disturbances, with effects particularly detrimental in children, which may have lasting unfavorable consequences [10, 11]. Analysis of the impact of long-term use of ASM on thyroid functions of children, for instance, revealed a reduction in thyroxine ( $T_4$ ) and triiodothyronine ( $T_3$ ) levels in serum by some, implying the contribution of thyroid dysfunctions [12]. With described untoward effects and lack of response to antiseizure medications in 20–30% subjects, there is a pressing need for mechanistic research to pursue new and affordable therapeutics, aiming not only at better management of seizures but also at more natural and cost-effective interventions with fewer side effects. To this end, anticonvulsant herbals used over millennia have generated renewed interest [13, 14]. Utilized mainly as stand-alone treatment, herbals with anticonvulsant properties have displayed several advantages over pharmacotherapies, demonstrating superior biocompatibility, less toxicity, and milder neurological side effects [13, 15, 16]. As natural products, anticonvulsant herbals are also more affordable and are increasingly used

in developing countries [17, 18]. Remarkably, in addition to suppressing epileptic seizures through a direct stabilizing effect on neuronal excitability by acting on ion channels and neurotransmitter receptors, herbal anticonvulsants have also been reported to alleviate neuroinflammation and immune response in the brain, inhibit autophagy, and improve the homeostatic processes [19, 20]. These additional outcomes have been suggested to contribute to the secondary antiseizure actions of herbal anticonvulsants, with increasing recognition of their relevance to therapy of pharmaco-resistant variants of epilepsy [20–22].

Although, in present days, the practice of antiseizure herbals for the therapy of epilepsy is primarily based on established traditions and anecdotal efficacy reports, some have passed rigorous scientific trials and have been licensed by the US Food and Drug Administration as anti-epilepsy leads [23, 24]. As it emerges from present discussions of *in vitro* and *in vivo* preclinical studies, the antiepileptic effects of herbals are mediated via five distinct mechanisms: (1) attenuation of the neuronal excitability through modulation of ion channel activity, (2) enhancement of mitochondrial functions with antioxidant effects, (3) allosteric enhancement of GABA<sub>A</sub> receptor mediated currents, (4) suppression of inflammatory response, and (5) modulation of protein synthesis and metabolism. While some of these effects (1, 3) are shared with established antiseizure drugs, others (2, 4, and 5) are specific to herbal anticonvulsants, suggestive of novel targets for pharmacotherapies as well as opportunities of their combination with ASM for additive effects.

This article presents a comprehensive overview of the putative action mechanisms and targets of the most widely used antiepileptic herbals in tissue culture and animal models. The emerging data not only advocate the possible uses of herbal leads as adjuvants to antiseizure drugs but also highlight their potential as a guide for drug discovery efforts towards new leads and targets relevant to refractory forms of epilepsy.

## Methods

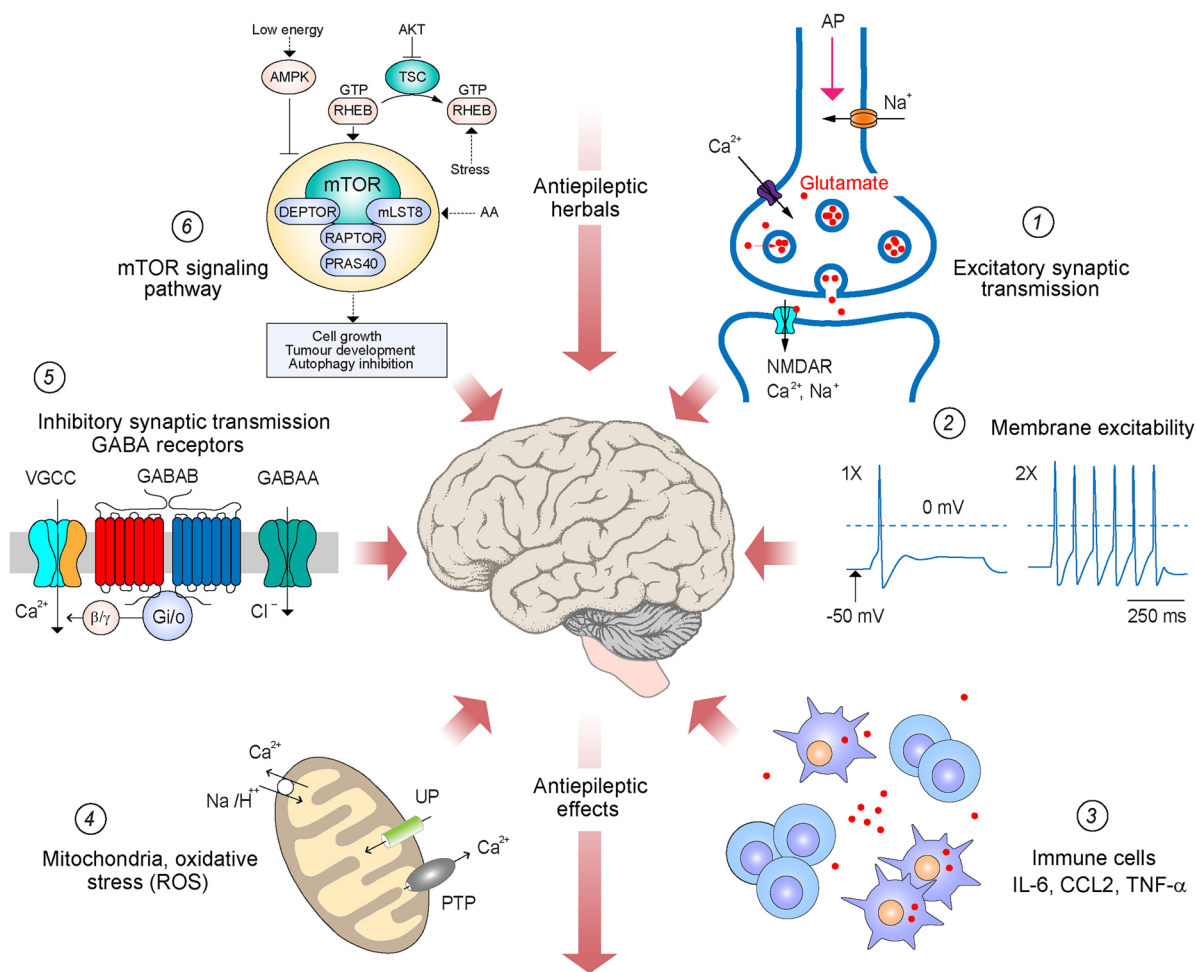
All authors conducted a literature search using scientific databases such as PubMed and ScienceDirect. Where necessary, Google Scholar, Academia, and ResearchGate have been used as additional sources of information. Keywords for search were antiepileptic herbals; antiseizure plant medicine; herbal anticonvulsant; traditional medicine for epilepsy; animal models of epilepsy; pharmaco-resistant epilepsy and herbal therapies; epilepsy biomarkers; ion channels and herbal medicine; mTOR signalling and epilepsy; herbal neuroprotection. The reference list of articles was scanned to identify further information relevant to the

current analysis. A summary of all references was drafted, followed by thematic grouping and manuscript writing. Figures are prepared using Adobe Illustrator Artwork 16.0 of the Adobe Creative Suit version 6.0. The table has been generated using Microsoft Word. EndNote X8.2 was used for citations, with references formatted per Journal Guidelines.

## Anticonvulsant herbs reducing neuronal excitability

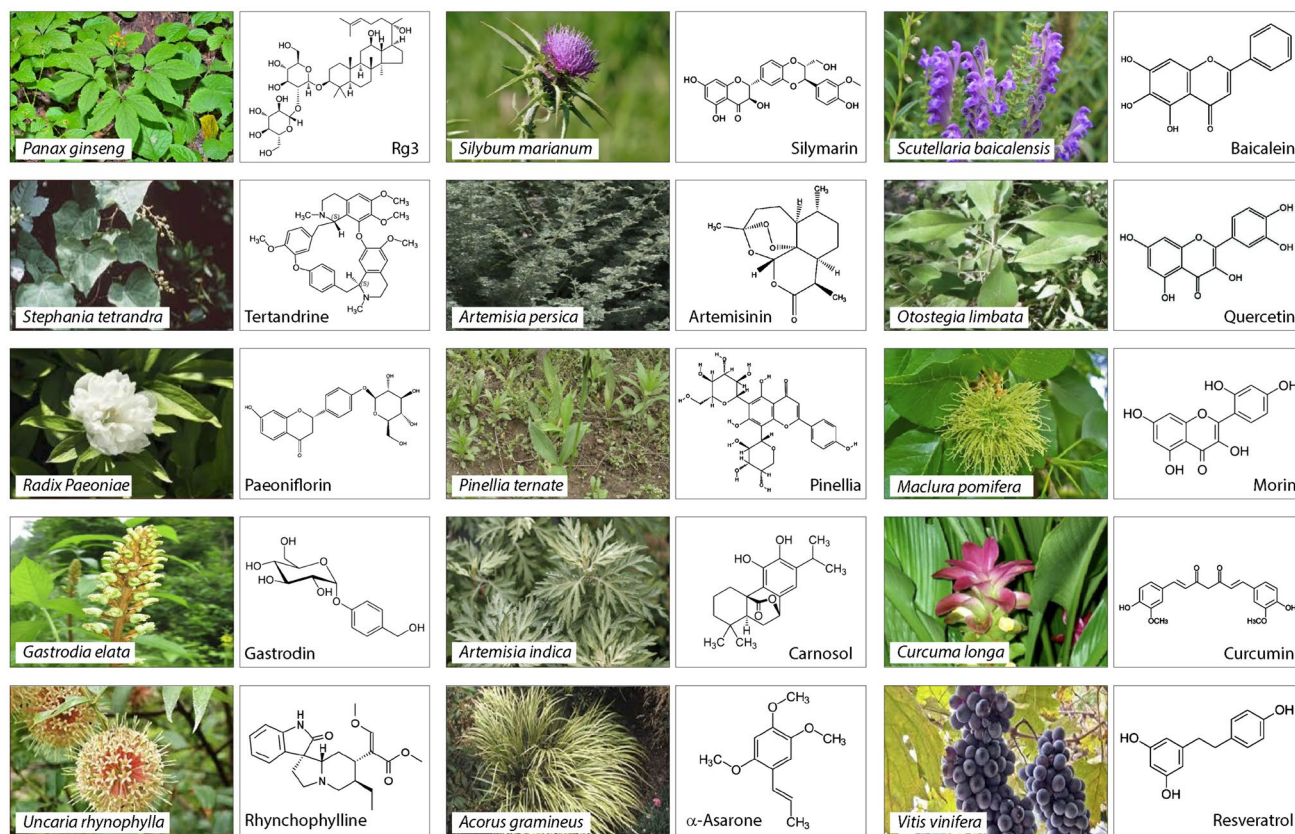
Abnormal activity of ion channels and receptors accounts for a significant portion of epilepsy cases, with membrane hyperexcitability linked to the initiation and propagation of seizures [25–27]. Most antiepileptic effects of medicinal plants acting on ion channels and receptors are attributed

to lowering the excitability of glutamatergic neurons, with attenuation of excitatory transmission [28, 29] (Fig. 1). Several studies demonstrated the antiseizure effects of ginsenoside Rg3 enriched in *Panax ginseng*, capable of restoring  $\text{Ca}^{2+}$  homeostasis by inhibiting the *N*-methyl-*D*-aspartate (NMDA) glutamatergic receptor [30, 31] (Fig. 2). In epilepsy models of cultured hippocampal neurons, Rg3 inhibits intracellular  $\text{Ca}^{2+}$  oscillations and lowers its concentration by suppressing  $\text{Ca}^{2+}$  influx mediated by NMDA receptor [31]. These effects suggest that the anticonvulsive activity of Rg3 could also counter cytotoxicity related to high intracellular  $\text{Ca}^{2+}$  caused by NMDA receptor hyperactivity in status epilepticus (SE) or during spontaneous recurrent epileptiform discharges (SREDS) [31]. Inhibition of  $\text{Ca}^{2+}$  influx with attenuation of the activity



**Fig. 1** Schematic representation of major targets and putative mechanisms implicated in antiepileptic effects of herbs. (1) Synaptic transmission at glutamatergic connections, post-synaptic NMDA receptor, presynaptic voltage-gated  $\text{Ca}^{2+}$ , and voltage-gated  $\text{Na}^{+}$  channels; (2) membrane excitability and action potential firing through modulation of ion channel activity; (3) neuroimmune and inflammatory response in the brain; (4) enhancement of mitochondrial functions and restoring ROS signaling; (5) modulation of inhibitory neu-

rotransmission by altering GABAergic drive; and (6) downregulation of mTOR signaling. TSC: tuberous sclerosis complex; GTP: guanosine 3-phosphate; AA: amino acids; AMPK: AMP-activated protein kinase; AKT: protein kinase B; RHEB: Ras homolog enriched in brain; DEPTOR: DEP domain-containing mTOR-interacting protein; RAPTOR: regulatory-associated protein of mTOR; PRAS40: proline-rich Akt substrate of 40 kDa



**Fig. 2** Illustration of selected medicinal herbals with anticonvulsant effects and chemical structure of the active lead. Individual panels are taken from Britannica plants: <https://www.britannica.com/plant/ginseng>

of glutamatergic neurons also underlies the anticonvulsant effects of *Stephania tetrandra* [32]. Unlike Rg3 acting upon the NMDA receptor, the active lead of *S. tetrandra*, tetrandrine, blocks voltage-gated calcium channels (VGCC), as reported in several excitable cells, including rat neurohypophysial nerve terminals [32] (Fig. 2). Notably, tetrandrine can improve the antiepileptic effects of antiseizure pharmacotherapies such as phenytoin and valproate on milder seizures in refractory epilepsy models in rats induced by doxorubicin. This effect appears to be partly mediated via lowering the expression of multidrug-resistant P-glycoprotein in tested cortical and hippocampal neurons [33]. Inhibition of  $\text{Ca}^{2+}$  influx is also thought to contribute to tempering the hyperexcitability associated with febrile seizures (FS) by *Radix paeoniae*, with paeoniflorin as the active ingredient [34] (Fig. 2). In the hyperthermia-induced FS model of immature rats, paeoniflorin inhibited the glutamate-induced elevation of intracellular  $\text{Ca}^{2+}$  with membrane depolarization and cytotoxicity associated with the hyperactivity of NMDA receptors [34].

Voltage-gated sodium channels (VGSC) are also an important drug target for managing epilepsy [35, 36]. Hyperactivity of VGSC is essential in the generation and propagation

of seizures. Therefore, suppressing  $\text{Na}^{+}$  influx in neurons by herbals is anticipated to have antiepileptic effects [13, 37]. Gastrodin, a phenolic glucoside from *Gastrodia elata* has been shown to exhibit antiepileptic activity by attenuating  $\text{Na}^{+}$  currents, reducing the severity and duration of SE in the pilocarpine rat model of temporal lobe epilepsy (TLE) [38] (Figs. 1 and 2). This effect protected neurons of the layer III medial entorhinal cortex from degeneration in SE and lowered the damage to neurons of the layer II medial entorhinal cortex (mEC). In addition, gastrodin could attenuate the inflammatory responses to pentylenetetrazol (PTZ)-induced seizures in mice [39]. Similar effects but through inhibition of specific subtype of  $\text{Na}^{+}$  channels have been reported for rhynchophylline (RIN), an alkaloid derived from *Uncaria rhynophylla*, which showed a potent anticonvulsant effect in a pilocarpine-induced SE rat model of TLE [40]. The tempering effect of RIN on neuronal hyperexcitability was mediated via inhibition of persistent sodium current ( $I_{\text{NaP}}$ ) and NMDA receptor. Furthermore, RIN has shown protective effects on medial entorhinal cortex layer III neurons and inhibited spontaneous epileptiform discharge of mEC layer II neurons in SE rats [40]. Finally, antiepileptic effects via dual NMDA receptor and  $I_{\text{NaP}}$  targeting have been described

for saikosaponin A (SSA) isolated from *Radix bupleuri* [41]. Such binary action was especially effective in terminating spontaneous recurrent epileptiform discharges in the hippocampal neuron culture (HNC) model of acquired epilepsy and continuous epileptiform high-frequency bursts of SE [42] (Table 1).

### Anticonvulsant herbals with antioxidant effects

Reactive oxygen species (ROS) are the leading cause of oxidative stress, which can contribute to the hyperexcitability of glutamatergic neurons and epileptogenesis [57, 58]. ROS play a critical role in the molecular signaling of neurons, regulating neuronal activity, membrane excitability, and synaptic plasticity mechanisms (Fig. 1). When in excess, ROS become highly toxic, interfering with various physiological processes and functions [55, 59]. In epilepsy, oxidative stress with increased ROS is explored as one of the early

biomarkers contributing to the generation of seizures and cytotoxicity. Substantial data suggests that oxidative stress might be especially prominent in refractory epilepsy, with specific mechanisms remaining elusive [60, 61]. Therefore, restoring the oxidative balance using pharmacotherapies and herbal medicines might temper epileptic seizures and avert neuronal damage with cytotoxicity [45, 46].

The antioxidant effects of *Artemisia* extracts with the prevention of oxidative stress and neuroprotection via modulation of the endogenous antioxidants have been shown by several groups [62, 63] (Fig. 2, Table 1). Analysis of the effects of hydroalcoholic compounds of *A. persica* on PTZ-induced seizures and memory impairments in experimental mice have shown amelioration of convulsions and memory improvements [43]. In the same model, the extracts of *Artemisia* increased the antioxidant capacity of the serum and brain tissue. These favorable outcomes correlate with the reduced frequency of spinning and jumping of epileptic mice

**Table 1** Summary of reviewed medicinal plants demonstrating anticonvulsant activity in preclinical models, with active leads, molecular targets, action mechanisms and corresponding references

Medicinal plants	Compounds	Targets	Action mechanisms	Preclinical models	References
<i>Panax ginseng</i>	Rg3	NMDAR	Inhibition/NMDAR	Neuronal culture	[27]
<i>Stephania tetrandra</i>	Tetrandrine	VGCC	Blockade/Ca <sup>2+</sup>	Rat (nerve terminals) Doxorubicin model	[28, 29]
<i>Radix paeoniae</i>	Paeoniflorin (PF)	NMDAR	Inhibition/NMDAR	Rat (hyperthermia febrile seizures model)	[30]
<i>Gastrodia elata</i>	Gastrodin (GAS)	VGSC; free radicals	Blockade/Na <sup>+</sup> Antioxidant	Rat (pilocarpine TLE model; KA model)	[34, 43]
<i>Uncaria rhynchophylla</i>	Rhynchophylline (RIN)	NMDAR	Inhibition/Na <sup>+</sup>	Rat (pilocarpine SE model)	[36]
<i>Radix bupleuri</i>	Saikosaponin A (SSA)	NMDAR; mTOR	Inhibition/Na <sup>+</sup> ; mTOR	Neuronal culture Rat (PTZ model)	[38, 44]
<i>Artemisia persica</i>	Artemisinin	Free radicals; Inflammation	Anti-oxidation, IL-1 $\beta$ , TNF- $\alpha$ inhibition	Mouse (PTZ model)	[45]
<i>Silybum marianum</i>	Silymarin	Free radicals	Anti-oxidation	Mouse (PTZ model)	[46]
<i>Pinellia ternate</i>	Pinellia	GABAAR	Activation	Rat (pilocarpine model)	[47]
<i>Artemisia indica</i>	Carnosol, ursolic & oleanolic acid	GABAAR	Activation	Mouse (PTZ model)	[48]
<i>Acorus gramineus</i>	$\alpha$ -Asarone	GABAAR NMDAR, VGSC	Activation GABAAR Blockade/Na <sup>+</sup>	Mouse (PTZ model)	[49]
<i>Scutellaria, baicalensis</i>	Baicalein	IGF-1 receptor Inflammation	Attenuation of IL-1 $\beta$ , IL-6, and TNF- $\alpha$ expression	Rat (pilocarpine model)	[50]
<i>Ruta graveolens</i>	Quercetin, Rutin	TLR4/NF-kB Inflammation	IL-1 $\beta$ , IL-6, TNF- $\alpha$ inhibition, HMGB1 expression	Rat (KA model)	[51]
<i>Citrus limon</i>	Hesperidin	Inflammation	BDNF and IL-10 inhibi- tion	Zebrafish larvae (PTZ model)	[52]
<i>Otostegia limbata</i>	Quercetin, Rutin	Inflammation	TNF- $\alpha$ and p-NF-K $\beta$ inhibition	Mouse (PTZ model)	[53]
<i>Maclura pomifera</i>	Morin	mTOR	Inhibition	Mouse (KA model)	[54]
<i>Curcuma longa</i>	Curcumin	mTOR	Inhibition	Neuronal culture	[55]
<i>Vitis vinifera</i>	Resveratrol	AMPK/mTOR	Inhibition	Rat (pilocarpine model)	[56]

and the attenuation of their tonic seizures. Interestingly, the anticonvulsant effects of *A. persica* extracts can be enhanced by diazepam, while flumazenil has the opposite effect. Histological examination of brain tissue showed reduced IL-1 $\beta$  and TNF- $\alpha$  expression by *A. persica* in the brain of the PTZ-treated mouse model [43]. In vitro antioxidant effects of the hydroalcoholic *A. persica* extracts was also verified by using tests for oxidative activity, which demonstrated enhancement in 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radicals scavenging, stabilization of 2,2'-azino-bis 3-ethylbenzothiazoline-6-sulfonic acid (ABTS) activity and hydroxyl free radicals scavenging, as well as restoration of Fe<sup>3+</sup> level with higher ferric ion reducing activity [43].

*Silybum marianum* is another anticonvulsive herbal with antioxidant and antiseizure effects, lowering the frequency and duration of convulsions in PTZ experimental mice and reducing PTZ-induced lethality [64] (Fig. 2). This effect was mediated partly via attenuation of the oxidative stress in the brain with the increase in superoxide dismutase and catalase activity and a reduced level of lipid peroxidation [64]. Finally, antioxidant effects have also been implicated in the ameliorative influence of *Gastrodia elata* on several neurological conditions. As a traditional Chinese medical herb, *G. elata* with active ingredient gastrodin demonstrated promising therapeutic outcomes in preclinical models of epilepsy, Alzheimer's disease, and Parkinson's disease [65, 66]. The anticonvulsive effects of *G. elata* were reported in the kainic acid (KA) rat model, potentially related to its free radical scavenging [67] (Fig. 2). The oral administration of *G. elata* significantly and dose-dependently reduced the level of lipid peroxides in the rat brain, leading to fewer incidents of wet dog shakes, paw tremors, and facial myoclonia [67].

### Anticonvulsant herbals targeting $\gamma$ -aminobutyric acid (GABA) receptors

The disbalance of excitatory/inhibitory (E/I) synaptic activity in cortical and hippocampal glutamatergic neurons plays a crucial role in the pathobiology of several neurological and neurodegenerative conditions [68–71]. In epilepsy, this change leads to the disinhibition of neuronal networks, resulting in paroxysmal hyperactivity, profuse glutamate release and associated cytotoxicity [4, 39]. GABA is the most prevalent inhibitory neurotransmitter in the central nervous system, stabilizing neuronal activity in the brain and spinal cord, with a decrease in GABAergic synaptic drive leading to epileptogenesis [47, 72]. As a transmitter, GABA acts via three groups of specific receptors: GABA<sub>A</sub>, GABA<sub>B</sub>, and GABA<sub>C</sub> [48, 49]. Upon binding to GABA, GABA<sub>A</sub> receptors mediate Cl<sup>-</sup> influx across the membrane, leading to neuronal hyperpolarization and reduction in excitability, preventing spontaneous and induced seizures in animal studies and epilepsy patients [73] (Fig. 1).

Several medicinal plants produce and accumulate large amounts of metabolites known as flavonoids, which share structural similarities with positive modulators of GABA<sub>A</sub> receptors - benzodiazepines [74]. Like benzodiazepines, these flavonoids are potent anticonvulsants, acting primarily as GABA<sub>A</sub> receptor enhancers, potentiating inhibitory synaptic currents [75]. One of the most utilized antiseizure herbs, *Pinellia ternate*, contains high concentrations of *Pinellia* total alkaloids (PTA), which enhances GABA<sub>A</sub> currents, preventing pilocarpine-induced convulsions in a rat model [76] (Fig. 2, Table 1). As a result, the incidence and intensity of seizures are significantly reduced compared to pilocarpine-treated rats without PTA. Notably, prolonged exposure to PTA led to higher expression of GABA<sub>A</sub> receptors and suppressed the levels of GABA transporter-1 (GAT-1). The mRNA analysis of GABA<sub>A</sub> receptor showed upregulation of low-expressed mRNA of GABA<sub>A</sub>R  $\alpha$ 5 subunit,  $\delta$  subunit and  $\gamma$ 2 subunit along with enhancement of glutamate decarboxylase 65 (GAD65), while GAT-1, GABA transaminase (GABA-T) mRNA levels showed opposite trend [76].

An earlier study isolated and characterized three active ingredients, carnosol, ursolic acid, and oleanolic acid, from another antiepileptic plant, *Artemisia indica*, acting on GABA<sub>A</sub> receptors with anticonvulsant effects [77] (Fig. 1). These substances have enhanced the response of GABA<sub>A</sub> receptors to GABA in a concentration-dependent manner, operating as a positive modulator of  $\alpha$ 1 $\beta$ 2 $\gamma$ 2L containing GABA<sub>A</sub> receptors at the benzodiazepine binding site [77]. Analysis of the antiepileptic activity of carnosol, ursolic acid, and oleanolic acid on PTZ-induced convulsions in mice showed that all three have potent antiseizure effects [77]. Finally,  $\alpha$ -asarone is another widely recognized antiepileptic herbal extract targeting GABA<sub>A</sub> receptors. Isolated from *Acorus gramineus*,  $\alpha$ -asarone has been successfully used in clinical practice to treat various epileptic seizures [78]. Detailed neurobiological analysis showed that  $\alpha$ -asarone combines potent antiseizure activity with neuroprotective, antipyretic, and analgesic effects, offering therapeutic potential against epilepsy, depression and anxiety [79]. Its antiepileptic effects are mediated via multitarget actions, including the enhancement of GABA<sub>A</sub> receptor activity, inhibition of glutamatergic drive mediated via NMDA receptors, and block of VGSC [80]. In PTZ and kainite mouse models,  $\alpha$ -asarone strongly inhibits the excitability of hippocampal neurons, prolonging the latency of clonic and tonic seizures and reducing animal mortality [78] (Table 1).

### Anticonvulsant herbals targeting neuroinflammation

Increasing preclinical and clinical data support the role of neuroinflammation in epileptogenesis [21, 81]. As part of the inflammatory response, glial cells release cytokines and

proinflammatory factors, exacerbating the disease process and promoting epileptic seizures [50, 82]. Lowering the level of cytokines and inflammatory factors is, therefore, expected to restore the physiological activity of neuronal networks and suppress epileptogenesis (Fig. 1).

Several medicinal plants rich in flavonoids were shown to counter inflammation by lowering proinflammatory factors and cytokines [51, 83] (Fig. 2, Table 1). Baicalein, a flavone from *Scutellaria baicalensis* and *Scutellaria lateriflora*, has ameliorative effects on seizure propensity and cognitive deficits in epilepsy models [52, 53]. In posttraumatic epilepsy, which is acquired epilepsy secondary to traumatic brain injury and has a significant inflammatory component, baicalein exerts neuroprotective and antiepileptic effects [53]. It also significantly reduced the number and scores of seizures and average seizure duration in an iron chloride (FeCl<sub>3</sub>)-induced posttraumatic epilepsy (PTE) mouse model. The neuroprotective effects of baicalein were also reported in ferric ammonium citrate (FAC)-induced HT22 hippocampal injury model, where baicalein reduced ferroptotic indices in vitro and suppressed the expression of 12/15-lipoxygenase (12/15-LOX) in an iron-induced HT22 cell damage model. These findings support baicalein's neuroprotective and anticonvulsive effects on PTE [53]. Likewise, in pilocarpine-induced epileptic rats, oral administration of baicalein dose-dependently decreased epilepsy symptoms, which was associated with inhibition of microglial proliferation, reduction in inflammatory markers, such as IL-1 $\beta$ , IL-6, and TNF- $\alpha$  in the brain [84].

Another flavonoid, quercetin, which is enriched in *Allium cepa* [85, 86] and several common fruits and vegetables, has shown potent anti-inflammatory action by inhibiting the TLR4/NF- $\kappa$ B signaling in animal models of epilepsy [85, 86]. In the same vein, in KA-injected rats, rutin from *Ruta graveolens*, attenuated the release of inflammatory molecules like IL-1 $\beta$ , IL-6, TNF- $\alpha$ , and high-mobility group box 1 (HMGB1) protein and suppressed the IL-1R1 and toll-like receptor 4 (TLR4) expression, preventing seizures [87]. A similar analysis of the effects of hesperidin, a flavonoid found in citrus fruits, showed longer latency with reduced intensity of PTZ-induced seizures in zebrafish larvae [88]. The anticonvulsant effects of HMGB1 were associated with changes in the expression of brain-derived neurotrophic factor (BDNF) and IL-10. Of note, hesperidin has a strong affinity for several receptors, including IL-10, as suggested by *in-silico* studies [88].

A recent analysis of the effects of methanolic extracts of *Otostegia limbata* on the expression of TNF- $\alpha$  and phosphorylated transcription factor nuclear factor kappa B (p-NF- $\kappa$ B) in the cortex and hippocampus of the PTZ-induced convulsion model showed significant anti-inflammatory effects associated with shorter duration and increased latency of seizures [56] (Fig. 2, Table 1). TNF- $\alpha$

and p-NF- $\kappa$ B expressions were strongly reduced compared to the vehicle-untreated control group. The authors suggested that the effects of *O. limbata* are attributed to high quantities of phenols and flavonoids, which demonstrated anticonvulsant effects in preclinical models of epilepsy.

### Anticonvulsant herbals targeting mTOR signaling

The mammalian target of the rapamycin (mTOR) signaling pathway, which controls cell growth, differentiation, proliferation, and metabolism [46, 54], is another putative target of antiepileptic herbal leads (Fig. 1). Pathological changes causing hyperactivity of mTOR signaling are known to lead to neuronal dysplasia with neurodevelopmental disorders, including autism and epilepsy [44, 89–91]. Other etiologies of epilepsy, such as brain injury and neonatal hypoxia–ischemia, have also been linked to hyperactivity of the mTOR pathway [92].

Several substances extracted from herbs have been reported to inhibit mTOR signaling [20, 93]. Morin, a phytochemical derived from a variety of plants, including *Maclura pomifera*, *M. tinctoria*, and *Psidium guajava*, has been shown to lower the vulnerability of preclinical models to epileptic seizures, an effect associated with reduction in expression of apoptotic molecules and inflammatory cytokines (Fig. 2, Table 1). In KA-induced seizure studies, morin caused significant inhibition of mTOR complex 1 (mTORC1) and decreased the propensity of mice to develop generalized seizures [94]. These effects were associated with reduced granule cell dispersion and suppression of mossy fiber sprouting in the hippocampus seven days after KA treatment [94]. Inhibition of mTOR signaling has also been implicated in the antiepileptic effects of SSAs in PTZ-induced rat models of epilepsy [95]. These oleanane-based glycosides, abundantly present in *Radix bupleuri* and several other medicinal plants, significantly reduce the severity and duration of seizures in PTZ models and increase the seizure latency [95]. The anticonvulsive effects of glycosides are associated with downregulation in p-mTOR, p-70S6K, L-1 $\beta$ , and TNF- $\alpha$  expression in hippocampal neurons of PTZ rat models [95]. Inhibition of mTOR activity and associated downregulation of the mitogen-activated kinase signaling also emerges to be involved in antiseizure effects of curcumin, an active phytochemical constituent of the medicinal herb *Curcuma longa* [46]. Quantitative real-time PCR analysis revealed that curcumin, but not rapamycin, reduced inflammatory markers IL-6 and COX-2 levels in cultured astrocytes challenged with IL-1 $\beta$ . In SH-SY5Y cells, curcumin reduced ROS levels, suggesting combined antioxidant and anti-inflammatory effects, with secondary antiepileptic outcomes. In the SE rat model, however, treatment with rapamycin or curcumin did not lower the inflammatory and oxidative stress markers up to one week after SE [46].

Resveratrol is another natural medicinal plant extract polyphenol with antiepileptic effects, curtailing the recurrence of seizures and SE in preclinical models [96, 97]. Its antiseizure effects seem to involve inhibiting NF- $\kappa$ B and suppressing proinflammatory cytokine synthesis, which can be influenced by upstream mTOR signaling [54, 98]. In pilocarpine-induced seizures in a rat model, resveratrol isolated from *Vitis vinifera* lowered COX-2, IL-1 $\beta$  and inducible NO synthase levels in the hippocampus 3 h after SE [93] (Fig. 2). Pretreatment of rats with resveratrol before pilocarpine-induced seizures also increased the expression of AMP-activated protein kinase, which was associated with a reduction in phosphor-mTOR and target phospho-S6 (pS6) protein, implying dual anti-inflammatory and antiepileptic effects, which involve AMPK/mTOR signaling pathway.

## Summary and conclusions

Nearly half of the pharmaceutical products used in medicinal practice today draw from traditional medicine and natural remedies, with many delivering life-saving solutions. The isolation of artemisinin from sweet wormwood by Tu and co-workers [99] to treat quinidine-resistant malaria presents an iconic example of a cure by natural remedies used in traditional medicine, saving millions of lives. The impact of this breakthrough has been recognized by the Nobel Prize for Physiology and Medicine of 2015 [100, 101], with the World Health Organization recommending artemisinin as the first- and second-line malaria treatment.

Unlike quinidine-resistant malaria treated nowadays by traditional medicine, many devastating conditions, which also include refractory epilepsy, are resistant to pharmacotherapies and have no natural solution. Research on herbal anticonvulsants offering relief from epileptic seizures, therefore, is viewed as one of the promising avenues towards enabling effective intervention. In this analysis, we considered the emerging data from in vitro and preclinical studies of herbal leads in animal models showing promise for managing epileptic seizures. In addition to engaging with ion channels and neurotransmitter receptors, the evolving data suggests herbal-induced improvements in mitochondrial functions with antioxidant effects, attenuation of neuroinflammatory response and immune processes, and alterations in protein synthesis and metabolism as mechanisms for beneficial outcomes (Fig. 1). Given the limited efficacy of frontline antiseizure pharmacotherapies acting upon neurotransmission and excitability of neurons in refractory epilepsy, engagement with additional targets is well warranted and likely to yield beneficial effects [4, 5, 61, 92, 102]. Discussed herein studies in neuronal cultures and preclinical models suggest that ameliorative outcomes of herbals may result from their use as monotherapy and in combination with conventional

antiepileptic pharmacotherapies. Both approaches necessitate rigorous dose–response, efficacy, and safety considerations, along with the review of the specifics related to individual variability and potential co-morbid conditions. In tandem with studies of clinical biomarkers and in silico analysis, these developments should facilitate understanding the neurobiological mechanisms of refractory epilepsy to guide the discovery of antiseizure leads with better efficacy, safety, and cost outcomes.

In conclusion, the future of herbal treatments of epilepsy might hold significant mechanistic and translational insights towards developing innovative primary therapies and adjuvants for integration with antiseizure medications. Guided by herbal leads, the drug discovery process will also likely facilitate the detection of synthetic analogues and modified leads to meet therapeutic needs through well-designed clinical trials and rigorous regulatory approval.

**Acknowledgements** S.V.O. acknowledges the critical comments of Prof. V.B. O’Leary on the final version of the manuscript.

**Author contribution** S.T., S.S., and S.V.O. conceived the study and designed the manuscript. S.T. and S.V.O. reviewed the literature and prepared the first draft of the study. S.T., S.S., and S.V.O. revised and commented on the manuscript and prepared for submission. All authors have reviewed and approved the final version of the manuscript for submission.

**Funding** SVO received funding from Innovation Fund Award 2023 from the University of Greenwich.

**Availability of data and material** This is not applicable. If accepted for publication, the material will be made freely available to the public.

## Declarations

**Ethics approval and consent to participate** Not applicable.

**Competing interests** The authors declare no competing interests.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## References

1. Collaborators GBDE (2019) Global, regional, and national burden of epilepsy, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet Neurol* 18:357–375



2. Fiest KM, Sauro KM, Wiebe S, Patten SB, Kwon CS, Dykeman J, Pringsheim T, Lorenzetti DL, Jette N (2017) Prevalence and incidence of epilepsy: a systematic review and meta-analysis of international studies. *Neurology* 88:296–303
3. Kwan P, Schachter SC, Brodie MJ (2011) Drug-resistant epilepsy. *N Engl J Med* 365:919–926
4. Zaitsev AV, Smolensky IV, Jorratt P, Ovsepian SV (2020) neurobiology, functions, and relevance of excitatory amino acid transporters (EAATs) to treatment of refractory epilepsy. *CNS Drugs* 34:1089–1103
5. Loscher W, Potschka H, Sisodiya SM, Vezzani A (2020) Drug resistance in epilepsy: clinical impact, potential mechanisms, and new innovative treatment options. *Pharmacol Rev* 72:606–638
6. Lawn ND, Bamlet WR, Radhakrishnan K, O'Brien PC, So EL (2004) Injuries due to seizures in persons with epilepsy: a population-based study. *Neurology* 63:1565–1570
7. Fazel S, Wolf A, Langstrom N, Newton CR, Lichtenstein P (2013) Premature mortality in epilepsy and the role of psychiatric comorbidity: a total population study. *Lancet* 382:1646–1654
8. Goldenberg MM (2010) Overview of drugs used for epilepsy and seizures: etiology, diagnosis, and treatment. *P T* 35:392–415
9. Fattorusso A, Matricardi S, Mencaroni E, Dell'Isola GB, Di Cara G, Striano P, Verrotti A (2021) The pharmacoresistant epilepsy: an overview on existant and new emerging therapies. *Front Neurol* 12:674483
10. Pack A (2005) Effects of treatment on endocrine function in patients with epilepsy. *Curr Treat Options Neurol* 7:273–280
11. Svalheim S, Sveberg L, Mochol M, Tauboll E (2015) Interactions between antiepileptic drugs and hormones. *Seizure* 28:12–17
12. Verrotti A, Laus M, Scardapane A, Franzoni E, Chiarelli F (2009) Thyroid hormones in children with epilepsy during long-term administration of carbamazepine and valproate. *Eur J Endocrinol* 160:81–86
13. Liu W, Ge T, Pan Z, Leng Y, Lv J, Li B (2017) The effects of herbal medicine on epilepsy. *Oncotarget* 8:48385–48397
14. Manchishi SM (2018) Recent advances in antiepileptic herbal medicine. *Curr Neuropharmacol* 16:79–83
15. Pearl PL, Drillings IM, Conry JA (2011) Herbs in epilepsy: evidence for efficacy, toxicity, and interactions. *Semin Pediatr Neurol* 18:203–208
16. Schachter SC (2009) Botanicals and herbs: a traditional approach to treating epilepsy. *Neurotherapeutics* 6:415–420
17. Kakooza-Mwesige A (2015) The importance of botanical treatments in traditional societies and challenges in developing countries. *Epilepsy Behav* 52:297–307
18. Mbuba CK, Ngugi AK, Newton CR, Carter JA (2008) The epilepsy treatment gap in developing countries: a systematic review of the magnitude, causes, and intervention strategies. *Epilepsia* 49:1491–1503
19. Fu Z, Zhao PY, Yang XP, Li H, Hu SD, Xu YX, Du XH (2023) Cannabidiol regulates apoptosis and autophagy in inflammation and cancer: a review. *Front Pharmacol* 14:1094020
20. He LY, Hu MB, Li RL, Zhao R, Fan LH, He L, Lu F, Ye X, Huang YL, Wu CJ (2021) Natural medicines for the treatment of epilepsy: bioactive components, pharmacology and mechanism. *Front Pharmacol* 12:604040
21. Mukhtar I (2020) Inflammatory and immune mechanisms underlying epileptogenesis and epilepsy: From pathogenesis to treatment target. *Seizure* 82:65–79
22. Challal S, Skiba A, Langlois M, Esguerra CV, Wolfender JL, Crawford AD, Skalicka-Wozniak K (2023) Natural product-derived therapies for treating drug-resistant epilepsies: From ethnopharmacology to evidence-based medicine. *J Ethnopharmacol* 317:116740
23. Abu-Sawwa R, Scutt B, Park Y (2020) Emerging Use of Epidiolex (Cannabidiol) in Epilepsy. *J Pediatr Pharmacol Ther* 25:485–499
24. Zhu Z, Dluzynski D, Hammad N, Pugalenti D, Walser SA, Mittal R, Samanta D, Brown ML, Asadi-Pooya AA, Kakooza-Mwesige A, Spalice A, Capponi M, Lekoubou A, Kumar A, Paudel S, Carney PR, Mainali G, Naik S (2023) Use of integrative, complementary, and alternative medicine in children with epilepsy: a global scoping review. *Children (Basel)* 10(4):713. <https://doi.org/10.3390/children10040713>. PMID: 37189961; PMCID: PMC10136598.
25. Lerche H, Shah M, Beck H, Noebels J, Johnston D, Vincent A (2013) Ion channels in genetic and acquired forms of epilepsy. *J Physiol* 591:753–764
26. Oyrer J, Maljevic S, Scheffer IE, Berkovic SF, Petrou S, Reid CA (2018) ion channels in genetic epilepsy: from genes and mechanisms to disease-targeted therapies. *Pharmacol Rev* 70:142–173
27. Ovsepian SV, LeBerre M, Steuber V, O'Leary VB, Leibold C, Oliver Dolly J (2016) Distinctive role of KV1.1 subunit in the biology and functions of low threshold K(+) channels with implications for neurological disease. *Pharmacol Ther* 159:93–101
28. Yogeewari P, Ragavendran JV, Thirumurugan R, Saxena A, Sriram D (2004) Ion channels as important targets for antiepileptic drug design. *Curr Drug Targets* 5:589–602
29. Errington AC, Stohr T, Lees G (2005) Voltage gated ion channels: targets for anticonvulsant drugs. *Curr Top Med Chem* 5:15–30
30. Radad K, Gille G, Liu L, Rausch WD (2006) Use of ginseng in medicine with emphasis on neurodegenerative disorders. *J Pharmacol Sci* 100:175–186
31. Kim S, Rhim H (2004) Ginsenosides inhibit NMDA receptor-mediated epileptic discharges in cultured hippocampal neurons. *Arch Pharm Res* 27:524–530
32. Wang G, Lemos JR (1995) Tetrandrine: a new ligand to block voltage-dependent Ca<sup>2+</sup> and Ca(+)-activated K<sup>+</sup> channels. *Life Sci* 56:295–306
33. Chen Y, Xiao X, Wang C, Jiang H, Hong Z, Xu G (2015) Beneficial effect of tetrandrine on refractory epilepsy via suppressing P-glycoprotein. *Int J Neurosci* 125:703–710
34. Hino H, Takahashi H, Suzuki Y, Tanaka J, Ishii E, Fukuda M (2012) Anticonvulsive effect of paeoniflorin on experimental febrile seizures in immature rats: possible application for febrile seizures in children. *PLoS ONE* 7:e42920
35. Bagal SK, Brown AD, Cox PJ, Omoto K, Owen RM, Pryde DC, Sidders B, Skerratt SE, Stevens EB, Storer RI, Swain NA (2013) Ion channels as therapeutic targets: a drug discovery perspective. *J Med Chem* 56:593–624
36. Pal R, Kumar B, Akhtar MJ, Chawla PA (2021) Voltage gated sodium channel inhibitors as anticonvulsant drugs: a systematic review on recent developments and structure activity relationship studies. *Bioorg Chem* 115:105230
37. Lin CH, Hsieh CL (2021) Chinese herbal medicine for treating epilepsy. *Front Neurosci* 15:682821
38. Shao H, Yang Y, Qi AP, Hong P, Zhu GX, Cao XY, Ji WG, Zhu ZR (2017) Gastrodin reduces the severity of status epilepticus in the rat pilocarpine model of temporal lobe epilepsy by inhibiting Nav1.6 sodium currents. *Neurochem Res* 42:360–374
39. Chen L, Liu X, Wang H, Qu M (2017) Gastrodin attenuates pentylentetrazole-induced seizures by modulating the mitogen-activated protein kinase-associated inflammatory responses in mice. *Neurosci Bull* 33:264–272
40. Shao H, Yang Y, Mi Z, Zhu GX, Qi AP, Ji WG, Zhu ZR (2016) Anticonvulsant effect of Rhynchophylline involved in the inhibition of persistent sodium current and NMDA receptor current in the pilocarpine rat model of temporal lobe epilepsy. *Neuroscience* 337:355–369

41. Lu CN, Yuan ZG, Zhang XL, Yan R, Zhao YQ, Liao M, Chen JX (2012) Saikosaponin a and its epimer saikosaponin d exhibit anti-inflammatory activity by suppressing activation of NF-kappaB signaling pathway. *Int Immunopharmacol* 14:121–126
42. Yu YH, Xie W, Bao Y, Li HM, Hu SJ, Xing JL (2012) Saikosaponin a mediates the anticonvulsant properties in the HNC models of AE and SE by inhibiting NMDA receptor current and persistent sodium current. *PLoS ONE* 7:e50694
43. Daneshkhah M, Setorki M (2019) Protective effects of *Artemisia persica* essential oil against pentylenetetrazol-induced seizure in male mice with emphasizing its mechanism of action. *Iran Red Crescent Med J* 21(2):1–8. <https://doi.org/10.5812/ircmj.85021>
44. Wong M (2013) Mammalian target of rapamycin (mTOR) activation in focal cortical dysplasia and related focal cortical malformations. *Exp Neurol* 244:22–26
45. Diniz TC, Silva JC, de Lima-Saraiva SR, Ribeiro FP, Pacheco AG, de Freitas RM, Quintans-Junior LJ, S. Quintans JDE, R.L. Mendes, J.R. Almeida, (2015) The role of flavonoids on oxidative stress in epilepsy. *Oxid Med Cell Longev* 2015:171756
46. Drion CM, van Scheppingen J, Arena A, Geijtenbeek KW, Kooijman L, van Vliet EA, Aronica E, Gorter JA (2018) Effects of rapamycin and curcumin on inflammation and oxidative stress in vitro and in vivo - in search of potential anti-epileptogenic strategies for temporal lobe epilepsy. *J Neuroinflammation* 15:212
47. Fritschy JM (2008) Epilepsy, E/I balance and GABA(A) receptor plasticity. *Front Mol Neurosci* 1:5
48. Chebib M, Johnston GA (1999) The 'ABC' of GABA receptors: a brief review. *Clin Exp Pharmacol Physiol* 26:937–940
49. Ovsepian SV, Vesselkin NP (2004) Dual effect of GABA on descending monosynaptic excitatory postsynaptic potential in frog lumbar motoneurons. *Neuroscience* 129:639–646
50. Kongsman JP (2022) Cytokines in the brain and neuroinflammation: we didn't starve the fire! *Pharmaceuticals (Basel)* 15(2):140. <https://doi.org/10.3390/ph15020140>. PMID: 35215252; PMCID: PMC8878213.
51. Al-Khayri JM, Sahana GR, Nagella P, Joseph BV, Alessa FM, Al-Mssallem MQ (2022) Flavonoids as potential anti-inflammatory molecules: a review. *Molecules* 27(9):2901. <https://doi.org/10.3390/molecules27092901>
52. Mao L, Wang K, Zhang Q, Wang J, Zhao Y, Peng W, Ding J (2022) Felt stigma and its underlying contributors in epilepsy patients. *Front Public Health* 10:879895
53. Li Q, Li QQ, Jia JN, Sun QY, Zhou HH, Jin WL, Mao XY (2019) Baicalein exerts neuroprotective effects in FeCl(3)-induced posttraumatic epileptic seizures via suppressing ferroptosis. *Front Pharmacol* 10:638
54. Lu X, Ma L, Ruan L, Kong Y, Mou H, Zhang Z, Wang Z, Wang JM, Le Y (2010) Resveratrol differentially modulates inflammatory responses of microglia and astrocytes. *J Neuroinflammation* 7:46
55. Beckhauser TF, Francis-Oliveira J, De Pasquale R (2016) Reactive oxygen species: physiological and pathophysiological effects on synaptic plasticity. *J Exp Neurosci* 10:23–48
56. Amin F, Tabassum S, Sarwar S, Qureshi R, Sohaib Khalid M, Riaz N, Al-Qahtani WH, Murtaza I (2022) Neuroprotective effect of *ostegia limbata* against PTZ-induced mice model of epilepsy by attenuated expression of p-NFkappaB and TNF-alpha. *Front Neurosci* 16:779681
57. Borowicz-Reutt KK, Czuczwar SJ (2020) Role of oxidative stress in epileptogenesis and potential implications for therapy. *Pharmacol Rep* 72:1218–1226
58. Parsons ALM, Bucknor EMV, Castroflorio E, Soares TR, Oliver PL, Rial D (2022) The interconnected mechanisms of oxidative stress and neuroinflammation in epilepsy. *Antioxidants (Basel)* 11(1):157. <https://doi.org/10.3390/antiox11010157>. PMID: 35052661; PMCID: PMC8772850.
59. Massaad CA, Klann E (2011) Reactive oxygen species in the regulation of synaptic plasticity and memory. *Antioxid Redox Signal* 14:2013–2054
60. Waldbaum S, Patel M (2010) Mitochondrial dysfunction and oxidative stress: a contributing link to acquired epilepsy? *J Bioenerg Biomembr* 42:449–455
61. Cardenas-Rodriguez N, Huerta-Gertrudis B, Rivera-Espinosa L, Montesinos-Correa H, Bandala C, Carmona-Aparicio L, Coballase-Urrutia E (2013) Role of oxidative stress in refractory epilepsy: evidence in patients and experimental models. *Int J Mol Sci* 14:1455–1476
62. Lee SG, Lee H, Nam TG, Eom SH, Heo HJ, Lee CY, Kim DO (2011) Neuroprotective effect of caffeoylquinic acids from *Artemisia princeps* Pampanini against oxidative stress-induced toxicity in PC-12 cells. *J Food Sci* 76:C250-256
63. Hosseinzadeh L, Malekshahi A, Ahmadi F, Emami SA, Hajialyani M, Mojarrab M (2018) The protective effect of different extracts of three artemisia species against H(2)O(2)-Induced oxidative stress and apoptosis in PC12 neuronal cells. *Pharmacognosy Res* 10:64–71
64. Waqar H, Khan HM, Anjum AA (2016) Antiepileptic potential of *silybum marianum* seeds in pentylenetetrazol-induced kindled mice, *Bangladesh Journal of Pharmacology* 11:603–609
65. Zhan HD, Zhou HY, Sui YP, Du XL, Wang WH, Dai L, Sui F, Huo HR, Jiang TL (2016) The rhizome of *Gastrodia elata* Blume - an ethnopharmacological review. *J Ethnopharmacol* 189:361–385
66. Liu Y, Gao J, Peng M, Meng H, Ma H, Cai P, Xu Y, Zhao Q, Si G (2018) A review on central nervous system effects of gastrodin. *Front Pharmacol* 9:24
67. Hsieh CL, Chiang SY, Cheng KS, Lin YH, Tang NY, Lee CJ, Pon CZ, Hsieh CT (2001) Anticonvulsive and free radical scavenging activities of *Gastrodia elata* Bl. in kainic acid-treated rats. *Am J Chin Med* 29:331–341
68. Bi D, Wen L, Wu Z, Shen Y (2020) GABAergic dysfunction in excitatory and inhibitory (E/I) imbalance drives the pathogenesis of Alzheimer's disease. *Alzheimers Dement* 16:1312–1329
69. Cherubini E, Di Cristo G, Avoli M (2021) Dysregulation of GABAergic signaling in neurodevelopmental disorders: targeting cation-chloride co-transporters to re-establish a proper E/I balance. *Front Cell Neurosci* 15:813441
70. Ovsepian SV, O'Leary VB (2016) Neuronal activity and amyloid plaque pathology: an update. *J Alzheimers Dis* 49:13–19
71. Ovsepian SV, O'Leary VB, Zaborszky L, Ntziachristos V, Dolly JO (2019) Amyloid plaques of Alzheimer's disease as hotspots of glutamatergic activity. *Neuroscientist* 25:288–297
72. Wong CG, Bottiglieri T, Snead OC 3rd (2003) GABA, gamma-hydroxybutyric acid, and neurological disease. *Ann Neurol* 54(Suppl 6):S3-12
73. Miles R, Blaesse P, Huberfeld G, Wittner L, Kaila K (2012) Chloride homeostasis and GABA signaling in temporal lobe epilepsy. In: Noebels JL, Avoli M, Rogawski MA, Olsen RW, Delgado-Escueta AV (eds) *Jasper's Basic Mechanisms of the Epilepsies*. Bethesda (MD)
74. Jager AK, Saaby L (2011) Flavonoids and the CNS. *Molecules* 16:1471–1485
75. Nilsson J, Sterner O (2011) Modulation of GABA(A) receptors by natural products and the development of novel synthetic ligands for the benzodiazepine binding site. *Curr Drug Targets* 12:1674–1688
76. Deng CX, Wu ZB, Chen Y, Yu ZM (2020) Pinellia total alkaloids modulate the GABAergic system in hippocampal formation on pilocarpine-induced epileptic rats. *Chin J Integr Med* 26:138–145

77. Khan I, Karim N, Ahmad W, Abdelhalim A, Chebib M (2016) GABA-A receptor modulation and anticonvulsant, anxiolytic, and antidepressant activities of constituents from *Artemisia indica* Linn. *Evid Based Complement Alternat Med* 2016:1215393
78. Huang C, Li WG, Zhang XB, Wang L, Xu TL, Wu D, Li Y (2013) alpha-asarone from *Acorus gramineus* alleviates epilepsy by modulating A-type GABA receptors. *Neuropharmacology* 65:1–11
79. Chellian R, Pandey V, Mohamed Z (2017) Pharmacology and toxicology of alpha- and beta-Asarone: A review of preclinical evidence. *Phytomedicine* 32:41–58
80. Wang ZJ, Levinson SR, Sun L, Heinbockel T (2014) Identification of both GABAA receptors and voltage-activated Na(+) channels as molecular targets of anticonvulsant  $\alpha$ -asarone. *Front Pharmacol* 5:40
81. Choi J, Koh S (2008) Role of brain inflammation in epileptogenesis. *Yonsei Med J* 49:1–18
82. Soltani Khaboushan A, Yazdanpanah N, Rezaei N (2022) Neuroinflammation and proinflammatory cytokines in epileptogenesis. *Mol Neurobiol* 59:1724–1743
83. Hamsalakshmi AM, Alex M, Arehally, Marappa S, Joghee S.B. Chidambaram (2022) Therapeutic benefits of flavonoids against neuroinflammation: a systematic review. *Inflammopharmacology* 30:111–136
84. Fu P, Yuan Q, Sun Y, Wu X, Du Z, Li Z, Yu J, Lv K, Hu J (2020) Baicalein ameliorates epilepsy symptoms in a pilocarpine-induced rat model by regulation of IGF1R. *Neurochem Res* 45:3021–3033
85. Ahmed T, Raza SH, Maryam A, Setzer WN, Braidy N, Nabavi SF, de Oliveira MR, Nabavi SM (2016) Ginsenoside Rb1 as a neuroprotective agent: a review. *Brain Res Bull* 125:30–43
86. Wu Y, Wei H, Li P, Zhao H, Li R, Yang F (2022) Quercetin administration following hypoxia-induced neonatal brain damage attenuates later-life seizure susceptibility and anxiety-related behavior: modulating inflammatory response. *Front Pediatr* 10:791815
87. Chang A, Chang Y, Wang SJ (2022) Rutin prevents seizures in kainic acid-treated rats: evidence of glutamate levels, inflammation and neuronal loss modulation. *Food Funct* 13:10401–10414
88. Sharma P, Kumari S, Sharma J, Purohit R, Singh D (2020) Hesperidin interacts with CREB-BDNF signaling pathway to suppress pentylenetetrazole-induced convulsions in zebrafish. *Front Pharmacol* 11:607797
89. Ryskalin L, Lazzeri G, Flaibani M, Biagioni F, Gambardella S, Frati A, Fornai F (2017) mTOR-dependent cell proliferation in the brain. *Biomed Res Int* 2017:7082696
90. Kutna V, O’Leary VB, Newman E, Hoschl C, Ovsepian SV (2021) Revisiting brain tuberous sclerosis complex in rat and human: shared molecular and cellular pathology leads to distinct neurophysiological and behavioral phenotypes. *Neurotherapeutics* 18:845–858
91. Granak S, Tuckova K, Kutna V, Vojtechova I, Bajkova L, Petrasek T, Ovsepian SV (2023) Developmental effects of constitutive mTORC1 hyperactivity and environmental enrichment on structural synaptic plasticity and behaviour in a rat model of autism spectrum disorder. *Eur J Neurosci* 57:17–31
92. Griffith JL, Wong M (2018) The mTOR pathway in treatment of epilepsy: a clinical update. *Future Neurol* 13:49–58
93. Wang SJ, Bo QY, Zhao XH, Yang X, Chi ZF, Liu XW (2013) Resveratrol pre-treatment reduces early inflammatory responses induced by status epilepticus via mTOR signaling. *Brain Res* 1492:122–129
94. Lee JM, Hong J, Moon GJ, Jung UJ, Won SY, Kim SR (2018) Morin prevents granule cell dispersion and neurotoxicity via suppression of mTORC1 in a kainic acid-induced seizure model. *Exp Neurobiol* 27:226–237
95. Ye M, Bi YF, Ding L, Zhu WW, Gao W (2016) Saikosaponin a functions as anti-epileptic effect in pentylenetetrazol induced rats through inhibiting mTOR signaling pathway. *Biomed Pharmacother* 81:281–287
96. Wu Z, Xu Q, Zhang L, Kong D, Ma R, Wang L (2009) Protective effect of resveratrol against kainate-induced temporal lobe epilepsy in rats. *Neurochem Res* 34:1393–1400
97. Shetty AK, Upadhy D (2016) GABA-ergic cell therapy for epilepsy: advances, limitations and challenges. *Neurosci Biobehav Rev* 62:35–47
98. Pallas M, Ortuno-Sahagun D, Andres-Benito P, Ponce-Regalado MD, Rojas-Mayorquin AE (2014) Resveratrol in epilepsy: preventive or treatment opportunities? *Front Biosci (Landmark Ed)* 19:1057–1064
99. Tu Y (2017, July 8) From *Artemisia annua* L. to Artemisinin. The discovery and development of artemisinins and anti-malarial agents, 1st edn. Academic Press. Hardback ISBN: 9780128116555, p 468.
100. Kong LY, Tan RX (2015) Artemisinin, a miracle of traditional Chinese medicine. *Nat Prod Rep* 32:1617–1621
101. Su XZ, Miller LH (2015) The discovery of artemisinin and the Nobel Prize in Physiology or Medicine. *Sci China Life Sci* 58:1175–1179
102. Terrone G, Balosso S, Pauletti A, Ravizza T, Vezzani A (2020) Inflammation and reactive oxygen species as disease modifiers in epilepsy. *Neuropharmacology* 167:107742

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.