



Circular RNAs in EMT-driven metastasis regulation: modulation of cancer cell plasticity, tumorigenesis and therapy resistance

Milad Ashrafizadeh^{1,2,3} · Jingyuan Dai⁴ · Pedram Torabian^{5,6} · Noushin Nabavi⁷ · Amir Reza Aref^{8,9} · Alaa A. A. Aljabali¹⁰ · Murtaza Tambuwala^{11,14}  · Minglin Zhu^{12,13}

Received: 5 December 2023 / Revised: 5 March 2024 / Accepted: 3 April 2024
© The Author(s) 2024

Abstract

The non-coding RNAs comprise a large part of human genome lack of capacity in encoding functional proteins. Among various members of non-coding RNAs, the circular RNAs (circRNAs) have been of importance in the pathogenesis of human diseases, especially cancer. The circRNAs have a unique closed loop structure and due to their stability, they are potential diagnostic and prognostic factors in cancer. The increasing evidences have highlighted the role of circRNAs in the modulation of proliferation and metastasis of cancer cells. On the other hand, metastasis has been responsible for up to 90% of cancer-related deaths in patients, requiring more investigation regarding the underlying mechanisms modulating this mechanism. EMT enhances metastasis and invasion of tumor cells, and can trigger resistance to therapy. The cells demonstrate dynamic changes during EMT including transformation from epithelial phenotype into mesenchymal phenotype and increase in N-cadherin and vimentin levels. The process of EMT is reversible and its reprogramming can disrupt the progression of tumor cells. The aim of current review is to understanding the interaction of circRNAs and EMT in human cancers and such interaction is beyond the regulation of cancer metastasis and can affect the response of tumor cells to chemotherapy and radiotherapy. The onco-suppressor circRNAs inhibit EMT, while the tumor-promoting circRNAs mediate EMT for acceleration of carcinogenesis. Moreover, the EMT-inducing transcription factors can be controlled by circRNAs in different human tumors.

Keywords RNA transcripts · circRNAs · cancer malignancy · EMT · cancer cell plasticity · Drug resistance

Introduction

Since cancer mortality is associated with metastasis, it is essential to emphasize the role of factors that regulate tumor invasion [1]. Therefore, improving treatment and the survival of patients rely on reducing invasion and the malignancy of tumors. Metastasis is one of the hallmarks of tumor and was coined in 1829 by Jean Claude [2]. The word “metastasis” has Greek roots and means “displacement”, with “meta” meaning next and “statis” meaning placement [3]. Metastasis refers to the process in which tumor cells migrate to another part of the body from their primary site to form new cancer cells, leading to death [4]. Cancer metastasis can be modulated via various mechanisms, such as EMT. Increasing evidence reveals that EMT mechanisms

have a potential role in tumor invasion and drug insensitivity (Fig. 1).

EMT causes a loss of cell-cell and cell-extracellular matrix adhesion, causing cells with a mesenchymal phenotype to detach from their primary site and separate from surrounding tissues. The process of EMT is not only vital for cancer metastasis but is essential for embryogenesis, tissue fibrosis, and wound healing [6]. Upon EMT induction, the motility of cancer cells increases, and epithelial cells lose their apical-basal polarity in order to transform into mesenchymal cells [7]. The generation of adherens junctions by E-cadherin is vital for cell adhesion and polarity in epithelia and leads to cell-cell attachment and recruitment of signaling complexes [8, 9]. The earliest step in the process of EMT is the loss of E-cadherin levels. EMT-TFs mediate epithelial reorganization via changing and decreasing

Extended author information available on the last page of the article

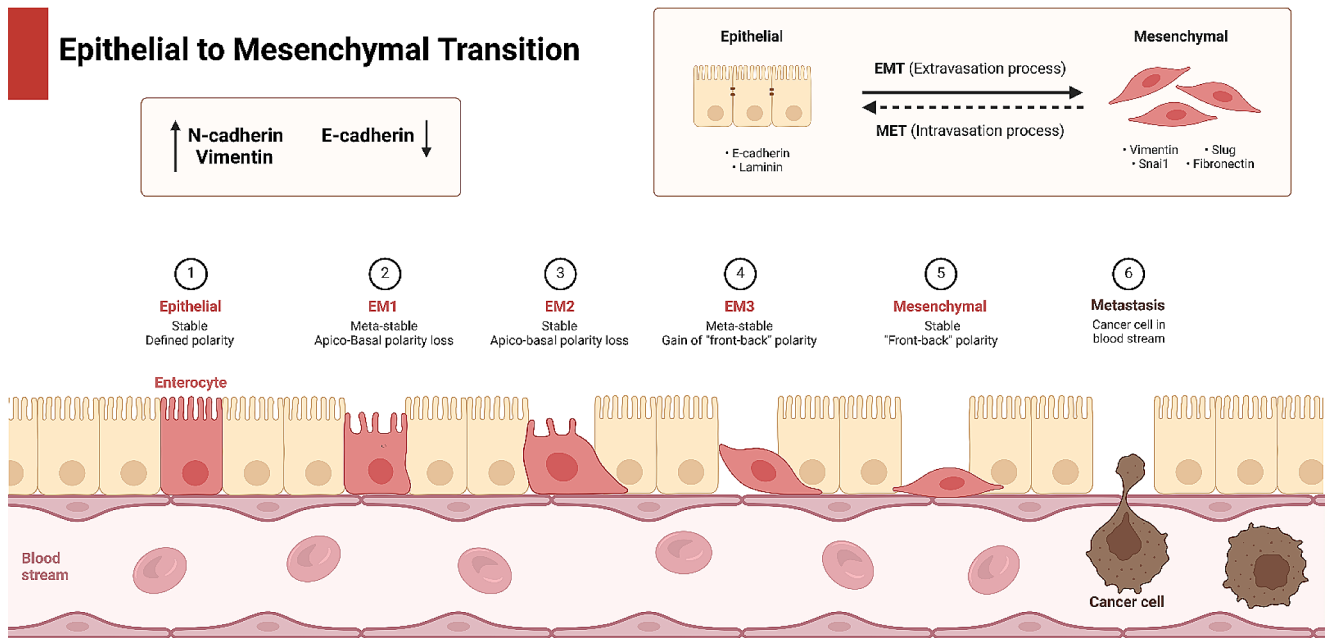


Fig. 1 A summary of EMT mechanism [5]. The EMT includes morphological and biochemical alterations that can occur simultaneously. The biochemical features include the downregulation of E-cadherin and upregulation of vimentin and N-cadherin that can participate in EMT induction. The epithelial cells are changed into mesenchymal

cells during EMT that demonstrate high levels of Slug, Snail1, vimentin and fibronectin. The occurrence of EMT can result in escape of a number of cells from other population of colony that diffuse into blood stream to reach to a new site for the establishment of a new colony, participating in the metastasis and progression of tumor

E-cadherin levels [10, 11]. EMT is associated with epithelial-mesenchymal transitions, changes in EMT-TF expression levels, including ZEB proteins, TGF- β , Twist and Snail, as well as Wnt/ β -catenin and Notch regulation. The induction of EMT and overexpression of PD-L1 by ERK signaling can cause drug resistance lung tumor. Suppressing PAR2 leads to inhibition of ERK-induced EMT to inhibit osimertinib resistance in lung cancer [12]. Also, Bakuchiol administration is critical for suppressing EMT and cancer invasion by downregulating TGF- β [13]. Low expression levels of ADRB2 by cardamonin are advantageous in inhibiting EMT and reducing the metastasis of colorectal tumor [14]. Besides, activation of Hedgehog signaling is vital for EMT induction in breast tumor [15]. Hence, the regulation of EMT in cancer occurs via various molecular pathways, and targeting inducers of EMT is therefore beneficial in reversing tumor invasion [16–18]. Interestingly, non-coding RNAs can control EMT mechanism [19], and the purpose of the present review is to shed some light on the function of circular RNAs (circRNAs) in regulating EMT and tumor invasion. The Figs. 2 and 3 demonstrate an overview of EMT-related pathways.

The function of EMT in tumorigenesis

Despite the valuable functions of EMT during the physiological and developmental stages, the function of EMT in cancer is oncogenic. Although EMT has been confirmed as a modulator of cancer migration, the increasing evidences have shown that EMT can also mediate the drug resistance [21, 22]. Therefore, the recent experiments have emphasized on highlighting the underlying mechanisms modulating EMT in human cancers. The androgen receptor has shown ability in increasing prostate cancer invasion. The androgen receptor stimulates eIF5A2 expression to enhance vimentin and N-cadherin levels for EMT induction in prostate tumor [23]. In pancreatic cancer, the induction of EMT can significantly increase cancer metastasis and mediate poor prognosis. The upregulation of MACC1 in nucleus and its interaction with SNAIL as EMT regulator can significantly enhance the metastasis of pancreatic tumor [24]. The lncRNA NRON has been shown as an inducer of metastasis and progression in bladder tumor. LncRNA NRON can upregulate EZH2 to stimulate EMT for bladder tumor invasion [25]. Noteworthy, the induction of EMT can also participate in the immunosuppression in human cancers. The ZEB1 is an inducer of EMT in which can stimulate exocytotic vesicular trafficking through release of Rab6A and Rab8A. Then, MMP14-mediated focal adhesion turnover occurs in lung cancer and induces CD8⁺T cell exhaustion

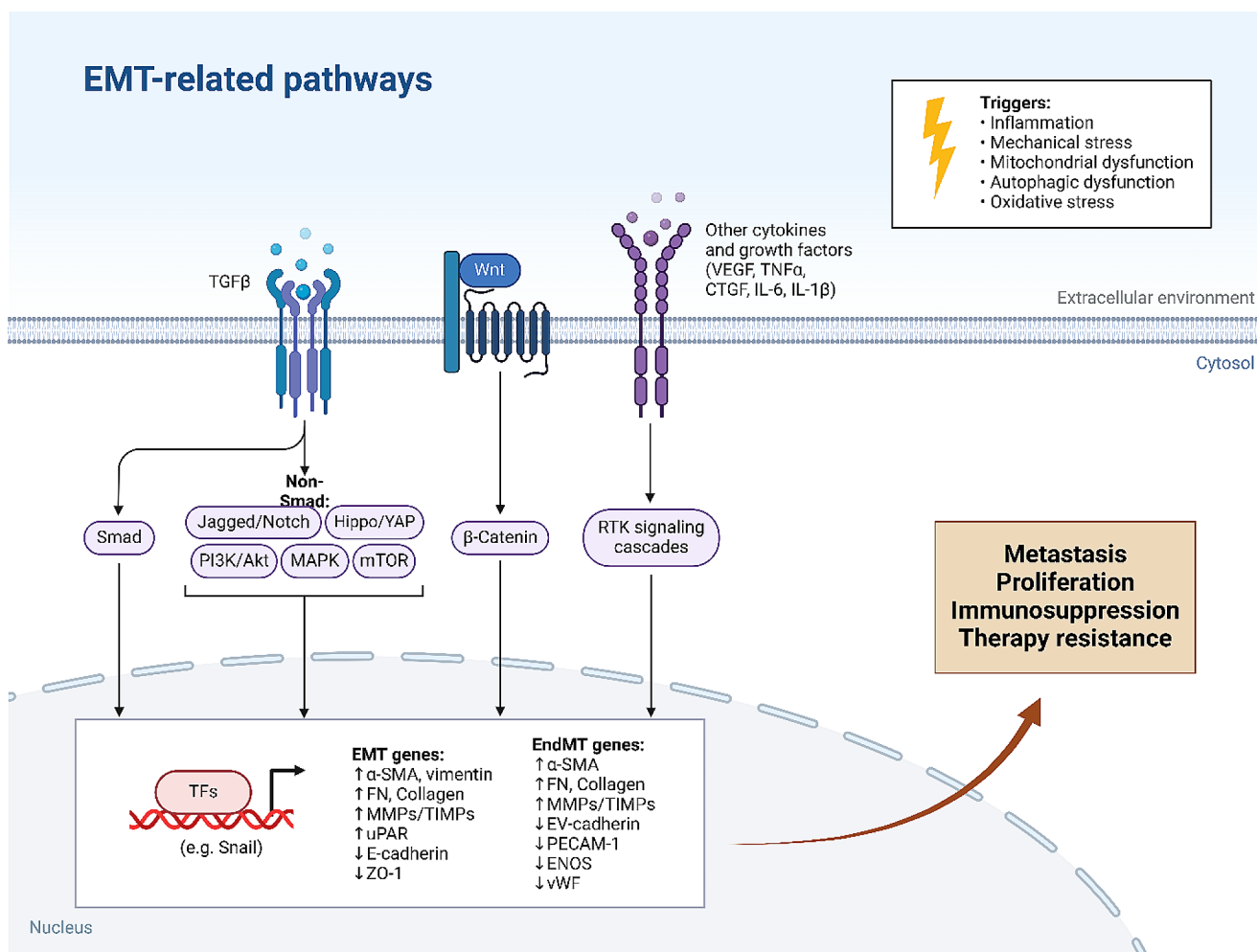


Fig. 2 An overview of a number of pathways regulating EMT. TGF- β , Wnt, PI3K/Akt, mTOR, YAP, MAPK and RTK are a number of molecular interactions contributing to the regulation of EMT. Such pathways are related to the proteins that finally translocate into the nucleus to

[26]. In cholangiocarcinoma, the upregulation of PLCB1 occurs to induce PI3K/Akt axis. Then, phosphorylation of GSK-3 β occurs to increase Snail levels for induction of EMT and increasing cancer progression. However, miR-26b-5p suppresses EMT through PLCB1 downregulation [27]. These experiments provide the insight that EMT has a versatile function in tumorigenesis and it is not limited to metastasis, while it can affect therapy response and impair the immune system.

CircRNAs: an overview in oncology

More than four decades ago, a new kind of RNA molecule without the capability of protein coding was recognized as circular RNA (circRNA). The first circRNA was identified by Sanger and colleagues with a covalently closed loop structure in plant viroids [28]. After that, circRNAs were

regulate expression levels of EMT-related genes. The process of EMT is vital for carcinogenesis, since EMT participates in metastasis, therapy resistance and immunosuppression in human cancers. (Created by Biorender.com)

recognized in eukaryotic cells and viruses [29, 30]. In 1991, the first mammalian circular transcript from the DCC gene was identified in cancerous and non-cancerous cells. Nevertheless, circRNAs were not easily accepted after their discovery and were initially dismissed. They were believed to be by-products of erroneous splicing and transcription errors, and their function was called into question [31]. Progress and efforts on the identification of these RNA molecules were stopped until recently with the discovery of high-throughput sequencing technology and bioinformatic tools. A high number of circRNAs, 32,000 human exonic circRNAs, have been discovered to date [32]. As stable non-coding RNA molecules, circRNAs are mainly found in the cytoplasm of eukaryotic cells, although a number of them, such as intronic circRNAs, can be present in the nucleus [33]. Linear RNA molecules, such as lncRNAs and miRNAs, have a 5' end cap and a 3' end poly(A) tail, but circRNAs have a unique structure that lacks both 5' and 3'

EMT-related pathways

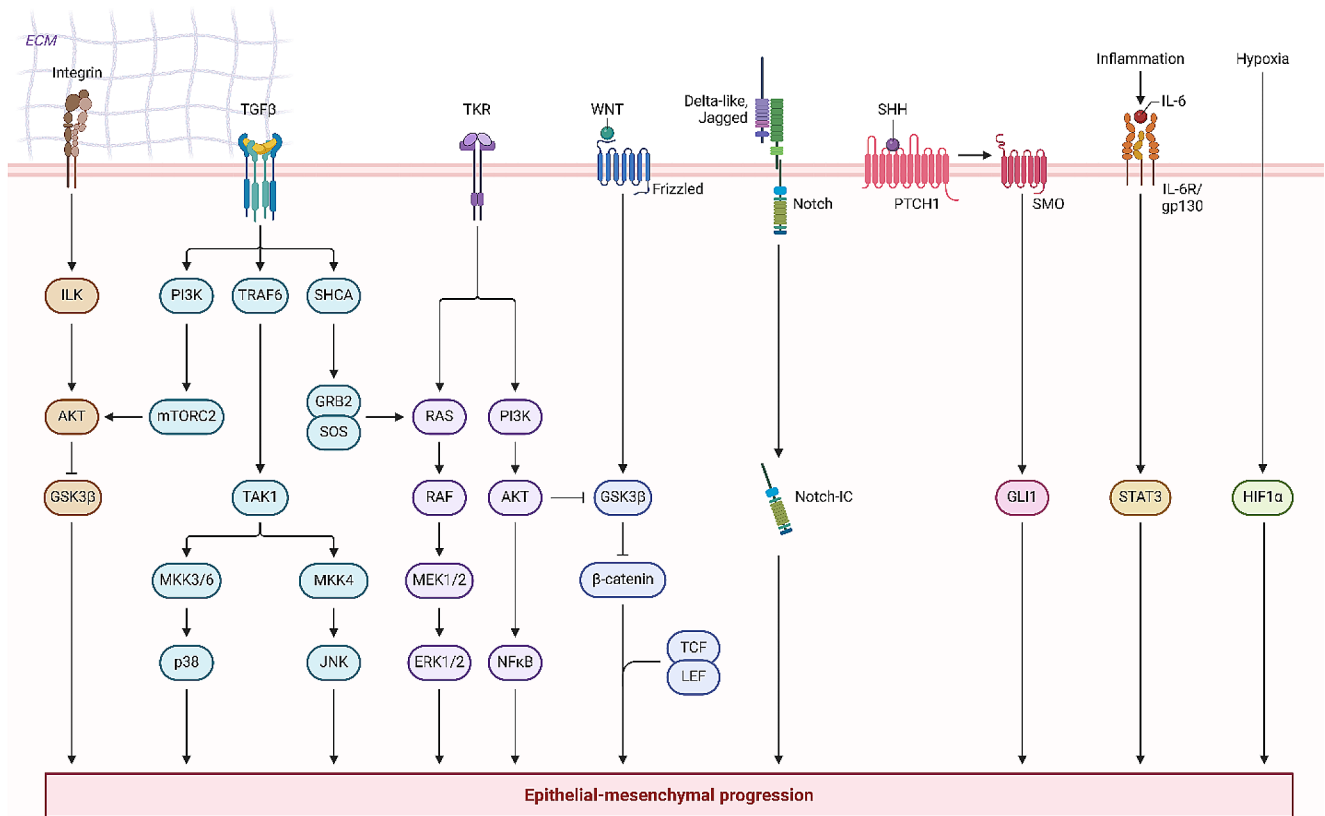


Fig. 3 The other mechanisms regulating EMT [20]. A number of these pathways demonstrate interaction in the regulation of EMT such as TGF-β that modulates mTORC2 and the integrin that regulates AKT.

Finally, mTORC2 can induce AKT. TKB, Wnt/β-catenin, Notch, GLI1, STAT3 and HIF-1α are among the other pathways regulating EMT. (Created by Biorender.com)

ends. Therefore, expression of circRNAs is stable, and their stability is high due to a lack of degradation by endonucleases [34]. CircRNAs are able to sponge miRNAs in the cytoplasm and accelerate the procedure of protein translation. Moreover, circRNAs regulate the transcription process by interacting with RBPs [35–37]. The action of circRNAs in affecting tumor progression is mainly performed via sponging miRNAs. For example, circCCDC85A sponges miR-550a-5p, increasing MOB1A levels and impairing breast tumor progression [38]. Tumor progression and proliferation are tightly regulated by the function of circRNAs [39]. Hsa-circ-0001013 upregulates TWSG1 via miR-136 inhibition, promoting gastric tumorigenesis [40]. The circMET contributes to tamoxifen resistance in breast tumor via increasing AHR level [41]. Furthermore, stabilization of ATP7A and upregulation of PHLPP2 by circPBX3 and circ-0001017, respectively, can lead to tumorigenesis [42, 43].

There are three subcategories of circRNAs based on the distribution and biogenesis [44]. The first subclass is exonic circRNAs (ecRNAs) that are generated through

back-splicing of the 5' splice site (splice donor site) to a 3' splice site (splice acceptor site) [45, 46]. The second subclass is intronic circRNAs (ciRNAs) that are originated from the intronic lariats precursors escaping from the debranching step of canonical linear splicing [47]. The third subclass is exon-intron circRNA (ElciRNAs) that increase the expression of parental genes in *cis* through circularizing with the retained intronic sequences among circularized exons [48]. Moreover, a new kind of circRNAs have been recognized known as mitochondria-encoded circRNAs (meccirNA) that their biogenesis occurs through a splicing-independent pathway and can function as molecular chaperons to accelerate the mitochondrial entry of nuclear-encoded proteins [49]. The biogenesis of circRNAs has been highlighted somehow and there are still many unknown aspects. The circRNAs have been shown to be derived from pre-mRNAs and they undergo canonical spliceosomal machinery [50]. The back-splicing reactions in circRNAs require the covalent linking of the 3' splice site with the 5' splice site, occurring during biogenesis of circRNAs and can be

accelerated through reverse complementary Alu repeats flanking the circularized exon [51].

The function of circRNAs occurs through four distinct mechanisms [52]. For influencing the expression of genes, the circRNAs affect the parental gene mRNA mediated through interaction with RNA binding proteins [53–55]. Notably, the function of circRNAs is also related to acting as ceRNAs to sponge miRNAs [56–58]. Moreover, the circRNAs have been shown to regulate the immune responses [59–61]. Although circRNAs are mainly believed as non-coding RNA factors, a variety of circRNAs have been shown to exert their function through encoding proteins [62–64]. The circRNAs are considered as potential factors in carcinogenesis. The circ-403,658 increases LDHA expression to induce glycolysis in bladder tumor [65]. Moreover, circ-103,809 facilitates the tumorigenesis of hepatocellular carcinoma through acting as tumor-promoting factor [66]. Table 1 summarizes the function of circRNAs in tumors. Table 2 summarizes the role of circRNAs in the controlling EMT.

Function of circRNAs as biomarkers

The distinct biochemical characteristics of circRNAs has led to the application of circRNAs as potential cancer biomarkers. The circRNAs possess high stability and they have cell-, tissue- and developmental-stage-specific paradigms of expression [90]. Moreover, the circRNAs are demonstrated to be highly conserved among the species and resistant to RNase R activity. The enrichment of circRNAs in exosomes can occur and they are found in body fluids including saliva, plasma and blood. The RNA sequencing has led to the recognition of high number of circRNAs in human peripheral whole blood cell [91]. In high number of cases, the level of circRNAs is higher compared to the corresponding linear RNA isoform [91]. According to this fact, a variety of experiments have emphasized on application of circRNAs as biomarkers in various human cancers [92]. The circ-002059 has a poor expression in gastric tumor compared to the healthy tissues and can influence the distal metastasis and TNM stage, highlighting its function as biomarker in gastric tumor [93]. The circRNA-PVT1 has upregulation in gastric tumor that can affect the overall survival and disease-free survival of patients and through sponging miR-125 family, it functions as tumor-promoting factor [94]. In addition to gastric tumor, the circRNAs have been considered as biomarkers in other kinds of tumors. The circ-0005075 has differential expression between hepatocellular carcinoma and health tissues and its expression can increase tumor size [95]. Moreover, the circ-0001649 has differential expression and shows low expression in hepatocellular carcinoma

Table 1 A summary of circRNAs function

CircRNA	Cancer type	Remark	Reference
Hsa_circ_0136666	Gastric cancer	Sponging miR-375-3p to increase PRKDC expression and enhance PD-L1 levels in immune evasion	[67]
Hsa_circ_0006401	Colorectal cancer	Downregulation of circRNA decreases growth and invasion of tumor cells	[68]
Circ_0004140	Lung adenocarcinoma	Circ_0004140 sponges miR-1184 to upregulate CCL22 in tumorigenesis	[69]
Hsa_circ_0000437	Gastric cancer	Increasing lymph node metastasis through HSPA2/ERK axis regulation	[70]
Hsa_circ_0003258	Prostate cancer	Sponging miR-653-5p and complexation with IGF2BP3 to enhance invasion	[71]
Circ_0000235	Bladder cancer	Downregulation of miR-330-5p to stimulate glycolysis	[72]
Circ_0042881	Breast cancer	EIF4A3 increases circ-0042881 levels to stimulate RAS pathway in tumorigenesis	[73]
Hsa_circ_0060467	Breast cancer	Hsa_circ_0060467 inhibits miR-1205 and mediates complexation with eIF4A3 to enhance liver invasion	[74]
Circ-hmRNPU	Gastric cancer	Suppression of NONO-mediated e-Myc transactivation and mRNA stabilization vital for glycolysation and tumorigenesis	[75]

[96]. Circ-100,876 has high expression in lung cancer and mediates lymph node metastasis and tumor staging, acting as prognostic factor [97].

CircRNA/EMT axis in cancer metastasis

In this section, we discuss the function of the circRNA/EMT in regulating the invasion of tumor. Based on the role of circRNAs, the interaction of circRNAs and EMT in cancers mediates metastasis. Oncogenic circRNAs induce EMT to increase tumorigenesis, while onco-suppressor circRNAs reduce cancer invasion via EMT inhibition. The field of biology and next-generation sequencing enables the identification of new circRNAs with important functional roles in cancer. The first complexity that arises from circRNAs is that their function in cancer varies and is context-dependent. For instance, circ-0008305 increases the progression of hepatocellular carcinoma via promoting TMED2 level and miR-186 inhibition [43]. Circ-0008305, on the other hand, acts as an oncosuppressor in lung cancer by regulating EMT. Circ-0008305, also known as circPTK2, interacts with miRNAs and EMT regulators such as TGF- β in lung tumor. TGF- β induced EMT in lung tumor promotes tumor cell progression and invasion. Notably, TIF1 γ deficiency is required for TGF- β -associated EMT induction in lung tumor [98]. According to these studies, as circRNA induces EMT, it may be capable of suppressing EMT in other cancers.

The expression of certain factors can significantly promote the metastasis, and IGF2BP3 is one of them. USP11 promotes the stability of IGF2BP3 to increase the colorectal cancer metastasis [99]. The upregulation of IGF2BP3 by linc 01305 can lead to overexpression of HTR3A at the mRNA level, which elevates the progression of esophageal tumor [100]. Hence, IGF2BP3 upregulation also increases invasion [101, 102]. IGF2BP3 is the target of circRNAs in regulating the EMT. CircIGHG elevates the malignancy of oral tumor via EMT induction. CircIGHG increases IGF2BP3 level via miR-142-5p inhibition to elevate the progression of oral cancer via an EMT mechanism [103]. Although IGF2BP3 is a target of circRNAs in increasing cancer metastasis, it is not the only mechanism. For instance, the function of circ-0003258 in prostate tumor has been shown to be oncogenic as it induces EMT to elevate cancer metastasis. Circ-0003258 affects two distinct molecular pathways to increase prostate tumor metastasis via EMT. Initially, circ-0003258 sponges miR-635-5p to upregulate ARHGAP5 expression, which then elevates cancer metastasis. Subsequently, circ-0003258 interacts with IGF2BP3 to upregulate HDAC4 and to induce ERK along with EMT [71].

Circ-0001666 impairs breast carcinogenesis via sponging miR-620 to upregulate WNK2 [104]. However,

circ-0001666 also enhances the progression of lung and thyroid cancers via regulating ETV4 and AGO1, among others [105, 106]. Circ-0001666 acts as an onco-suppressor to regulate the EMT mechanism in colorectal cancer.

“Once miR-576-5p is synthesized in the nucleus, it’s shuttled into the cytoplasm via Exportin5, where it acts to diminish PCDH10 expression, consequently promoting the progression of colorectal cancer. In contrast, the back-splicing event between exon 2 and exon 4 in FAM120B pre-mRNA leads to the creation of circ-0001666. This particular circRNA elevates PCDH10 levels by inhibiting miRNA-576-5p. The outcome is a decrease in the ‘stemness’ of colorectal cancer cells due to the suppression of EMT, which is facilitated by hindering Wnt/ β -catenin signaling pathways.” [79]. Circ-0001017 is another example of circRNAs with onco-suppressor function in gliomas, which reduce let-7 g-3p level to upregulate NDST3, suppressing EMT [107]. Drawing from these studies, it becomes clear that the circRNA/EMT axis serves as a key regulatory mechanism in cancer metastasis (Table 3; Fig. 4).

CircRNA/EMT axis in chemoresistance and radioresistance

Chemoresistance is an intriguing concept and can develop in two ways, including intrinsic and acquired chemoresistance. Although understanding drug resistance is a key focus in the past decade, the failure of therapy has not been appropriately solved. Non-coding RNAs are regulators of tumorigenesis, and interestingly, circRNAs can determine therapy response in cancer patients. Hsa-circ-0007883 interacts with FUS to increase the FOXR2 stability in developing paclitaxel resistance in ovarian cancer [114]. Exosomal circ-SFMBT2 upregulates TRIB1 via miR-136-5p sponges to induce docetaxel resistance in prostate cancer [115]. Therefore, the association of circRNAs with downstream targets can change the chemotherapy response in cancer [116].

Paclitaxel is an inhibitor of tumor progression and increases polymerization of microtubules to disturb their stability in inhibiting proliferation. COL5A1 upregulation causes paclitaxel resistance [117]. KHDRBS3 increases MIR17HG level to stimulate glycolysis and paclitaxel insensitivity in ovarian tumor [118]. Upregulation of gp96 and HIF-1 α can also result in paclitaxel resistance in cancer [119, 120]. Overexpression of circ-0007534 in endometrial tumor results in paclitaxel resistance. The circ-0007534 elevates ZEB2 level via miR-625 inhibition to accelerate EMT and accelerate paclitaxel insensitivity [121]. Docetaxel functions similarly to paclitaxel and prevents the depolymerization of microtubules to suppress cancer. Upregulation of circ-CRIM1 in nasopharyngeal cancer is

Table 2 The circRNAs-associated EMT control

CircRNA	Effect on EMT	Cancer type	Remarks	References
Hsa_circ_0094606	Induction	Prostate cancer	Hsa_circ_0094606 attaches to PRMT1 to mediate M2 polarization of macrophages EMT induction	[76]
Hsa_circ_0013561	Induction	Ovarian cancer	Hsa_circ_0013561 upregulates ANXA2 through miR-23b-3p suppression in EMT induction	[77]
Circ_0087429	Suppression	Cervical cancer	EIF4A3 inhibits circ-0087429 to mediate EMT	[78]
Hsa_circ_0001666	Suppression	Colorectal cancer	Hsa_circ_0001666 increases PCDH10 level through miR-576-5p disruption to inhibit EMT	[79]
Hsa_circ_0006732	Induction	Colorectal cancer	Hsa_circ_0006732 downregulates miR-127-3p to increase Rab3D levels in EMT induction	[80]
Circ_0000799	Induction	Colorectal cancer	Sponging miR-647	[81]
Hsa_circ_0000497 and hsa_circ_0000918	Induction	Ovarian cancer	Hsa_circ_0000497 and hsa_circ_0000918 regulate miRNAs to induce EMT in ascites	[82]
Circ_0001589	Induction	Cervical cancer	Circ_0001589 sponges miR-1248 to upregulate HMGB1 in EMT induction	[83]
Circ-OXCT1	Suppression	Gastric cancer	Circ-OXCT1 increases SMAD4 expression to promote N-cadherin and vimentin levels and reduce E-cadherin levels	[84]
Circ_0003221	Induction	Cervical cancer	Circ_0003221 increases S100A14 level by miR-139-3p inhibition	[85]
CircRNA circ_0001666	Induction	Pancreatic cancer	Downregulation of circRNA circ_0001666 increasing miR-1251 level, while it reduces SOX4 level to impair EMT	[86]
Circ_0058106	Induction	Hypopharyngeal squamous cell carcinoma	miR-185-3p downregulation to induce Wnt2b/ β -catenin/c-Myc axis	[87]
Circ-STK39	Induction	Pancreatic cancer	Circ-STK39 inhibits miR-140-3p to increase TRAM2 levels	[88]
Hsa_circ_0009092	Suppression	Colorectal cancer	Hsa_circ_0009092 sponges miR-665 to elevate NLK levels and impairing Wnt/ β -catenin axis	[89]

a positive indicator of docetaxel resistance in tumor cells. miR-422a decreases FOXQ1 level to inhibit the EMT and mediate docetaxel sensitivity. However, when expression of circ-CRIM1 increases, it sponges miR-422a to upregulate FOXQ1, induce EMT, and increase the docetaxel resistance [122]. The docetaxel potential in prostate tumor therapy increases upon upregulation of circ-Foxo3 and is related to Foxo3 and EMT suppression, thereby reducing the malignancy of cancer cells [123].

Circ-0003998 is another regulator of tumor progression and can enhance proliferation and invasion in lung tumor via miR-326 inhibition [124]. The function of circ-0003998 in triggering chemoresistance has been evaluated in hepatocellular carcinoma. Circ-0003998 stimulates the EMT to promote tumorigenesis. Additionally, Circ-0003998 reduces miR-218-5p level, which upregulates EIF5A2 [125]. Circ-0007022 is present on chromosome 19 and is formed as a result of back-splicing. Circ-0007022 suppresses miR-338-3p to cause radioresistance. Following the inhibition of miR-338-3p by circ-0007022, the levels of NRP1 increased. Two molecular pathways, including PI3K/Akt and EMT, are activated to increase tumor progression and mediate radioresistance [126]. However, only one experiment investigated the function of the circRNA/EMT axis in radioresistance, and more research is required to confirm and expand on the current findings (Fig. 5).

CircRNA/miRNA/EMT axis

Linear RNA molecules do not encode proteins with a length of less than 24 nucleotides, best known as microRNAs (miRNAs). MicroRNAs sequester mRNA to downregulate gene [127]. The expression level of miRNAs changes during cancer progression and can be used as biomarkers. Furthermore, miRNAs can be sponged by lncRNAs and circRNAs [128, 129]. miRNAs demonstrate dysregulation in tumorigenesis and are biomarkers. The malignancy of cervical tumor depends on the high level of circ-000322, a driver of tumor progression via EMT induction. Upregulation of S100A14 leads to EMT induction and cervical cancer progression. The role of circ-0003221 in enhancing cervical tumorigenesis is related to sponging miR-139-3p to stimulate EMT mechanisms via S100A14 upregulation. Knocking down the expression of circ-0003221 is beneficial for impairing cervical cancer progression via EMT suppression [85]. miR-370 is inhibited by SNHG15 in ovarian cancer to increase the progression of tumor cells [117]. miR-370 overexpression elevates the response of ovarian tumor to cisplatin chemotherapy [130].

Restoring expression of miR-185-5p is beneficial in suppressing osteosarcoma malignancy, as miR-185-5p reduces

CTSE expression [131]. PCAT6 downregulation of miR-185-5p in osteosarcoma can result in TGF- β signaling activation and tumor cell progression [132]. Circ-001569 is capable of sponging miR-185-5p in osteosarcoma, and this interaction results in upregulation of FLOT2, a factor involved in triggering EMT in cancer cells and thus enhancing tumor progression [133]. In esophageal cancer, the interaction of circRNA and miRNA is beneficial in determining the progression. Hsa-circ-0000277 undergoes overexpression in esophageal cancer, and by reducing miR-4766-5p expression, it paves the way for upregulation of LAMA1 to induce EMT [134]. However, not all circRNAs sponge miRNA to induce EMT and tumor metastasis. Sometimes, circRNAs sponge oncogenic miRNAs to suppress EMT mechanisms. Hsa-circ-001988 reduces miR-197-3p level to suppress EMT and the progression of gastric tumor [135]. Moreover, miR-503-5p overexpression can result in EMT in oral cancer as circ-0072387 inhibits miR-503-3p to decrease its expression, disrupt EMT, and minimize tumor progression [136]. More importantly, the interaction of circRNAs and miRNAs can be affected by upstream mediators in cancer. The biogenesis of circ-DOCK5 is inhibited in esophageal cancer by ZEB1. To prevent circ-DOCK5 biogenesis in esophageal cancer, ZEB1 downregulates EIF4E3 and DOCK5 mRNA levels to affect back-splicing between exon 49 and exon 50. When circ-DOCK5 is inhibited, miR-527-3p expression decreases, resulting in decreased TGF- β secretion. TGF- β overexpression then forms a positive feedback loop with ZEB1 to elevate esophageal cancer EMT induction and progression [137]. Table 4; Fig. 6 summarize the function of circRNA/miRNA interactions in the control of EMT mechanisms.

CircRNAs regulating EMT-TFs in cancers

ZEB proteins

The ZEB family of proteins, which includes ZEB1 and ZEB2, is one of the best-known modulators of EMT. Upregulation of ZEB1 in cancer can lead to tumor metastasis. YTHDF3 increases the expression of ZEB1 and promotes its stability at the mRNA level to enhance breast cancer invasion [146]. Induction of PI3K/Akt signaling by BAG4 leads to an increase in cancer metastasis via ZEB1 overexpression [147]. Furthermore, when the expression of ZEB1 increases, it represses E-cadherin to enhance the mobility of tumor cells [148]. Similarly, ZEB2 functions as an oncogenic factor. ZEB2 down-regulation by miR-518a-5p leads to inhibition of breast cancer progression [149]. Paeonol increases the expression level of miR-126-5p to downregulate ZEB2, thereby decreasing the invasion of lung

Table 3 CircRNA/EMT axis in cancers

CircRNA	Molecular pathway	Cancer type	Remark	Ref.
Circ-0082182	miR-411/miR-1205/Wnt	Colorectal cancer	Circ-0082182 stimulates Wnt/ β -catenin signaling via miR-411 and miR-1205 sponging in carcinogenesis	[108]
Circ-0001666	miR-1251/SOX4/EMT	Pancreatic cancer	Circ-0001666 upregulates SOX4 via miR-1251 inhibition in EMT acceleration	[86]
Circ-0087429	miR-5003-3p/OGN	Cervical cancer	Circ-0087429 overexpression occurs by EIF4A3 to increase OGN level by miR-5003-3p inhibition in EMT suppression	[78]
Circ-0004913	miR-184/HAMP	Hepatocellular carcinoma	Circ-0004913 increases HAMP level via miR-184 inhibition in suppressing tumor metastasis	[109]
Circ-0089153	miR-2467-3p/E2F6	Breast cancer	Circ-0089153 upregulates E2F6 via miR-2467-3p inhibition in accelerating tumorigenesis	[110]
Circ-GLIS2	GLIS2	Hepatocellular carcinoma	Circ-GLIS2 increases GLIS2 expression in reducing invasion and decreasing levels of EMT-related markers	[111]
Circ-0006732	miR-127-5p/RAB3D	Colorectal cancer	Circ-0006732 increases RAB3D via miR-127-5p inhibition in EMT induction	[112]
Circ-FOXM1	FOXM1/Wnt	Osteosarcoma	Circ-FOXM1 sponges miR-320a and -320b Upregulation of FOXM1 in Wnt signaling induction Inducing EMT	[113]

tumor cells [150]. Therefore, both ZEB1 and ZEB2 proteins are inducers of cancer metastasis, and notably, their expression level is controlled by circRNAs in cancer [151]. Circ-VANGL1 upregulation is in favor of enhancing the metastasis of thyroid cancer cells as it affects ZEB1 expression. Depleting circ-VANGL1 with siRNA is advantageous in reducing the metastasis of tumor cells and decreasing their malignancy. Circ-VANGL1 acts as a ceRNA, decreasing miR-194 expression while increasing ZEB1 expression. As EMT-TFs, ZEB1 stimulates EMT to promote the progression and metastasis of thyroid cancer cells [152]. The expression level of ZEB1 is tightly regulated by miRNAs in cancer. miR-429 is a negative regulator of ZEB1, leading to inhibition of hepatocellular carcinoma cells. In contrast, MAPKAP5-AS1 increases ZEB1 via adsorption of miR-429 to elevate tumor progression [153]. Moreover, miR-144-3p reduces ZEB1 expression to interfere with lung cancer invasion [154]. Therefore, miRNAs and ZEB1 closely interact to regulate cancer metastasis [155].

However, there are studies demonstrating that circRNAs can suppress ZEB1. Circ-ACAP2 is an inhibitor of EMT in head and neck cancer. Circ-ACAP2 sponges miR-21-5p to suppress STAT3 signaling in cancers, lowering ZEB1 expression and impairing EMT [156]. In addition, ZEB2 is regulated by circRNAs in cancer. Circ-0007534 stimulates EMT in endometrial cancer and, in this way, promotes the progression of tumor cells to mediate paclitaxel resistance. Circ-0007534 increases ZEB2 expression via miR-625 inhibition to trigger the EMT mechanism and paclitaxel resistance [121]. According to these studies, circRNAs regulate ZEB1 and ZEB2, and more research is needed to understand ZEB2 regulation by circRNAs in cancers.

Slug and Snail

Upregulation of Slug or Snail has been shown to be a driver for the progression and metastasis of cancer cells. Upon overexpression of Snail in cancer, EMT is induced to enhance metastasis. An experiment has shown that upregulation of circ-0023642 can result in a significant enhancement of metastasis in gastric cancer. Circ-0023642 promotes tumor invasion by increasing N-cadherin, vimentin, and Snail expression while decreasing E-cadherin expression [157]. As a result, circRNAs are regulators of snail and slug EMT mechanisms in cancer.

TGF- β

TGF- β is another important regulator of cancer progression, and abnormal expression can influence cancer metastasis and malignancy. Smad3 is one of the key players of TGF- β signaling. LHPP inhibits phosphorylation of Smad3 to impair

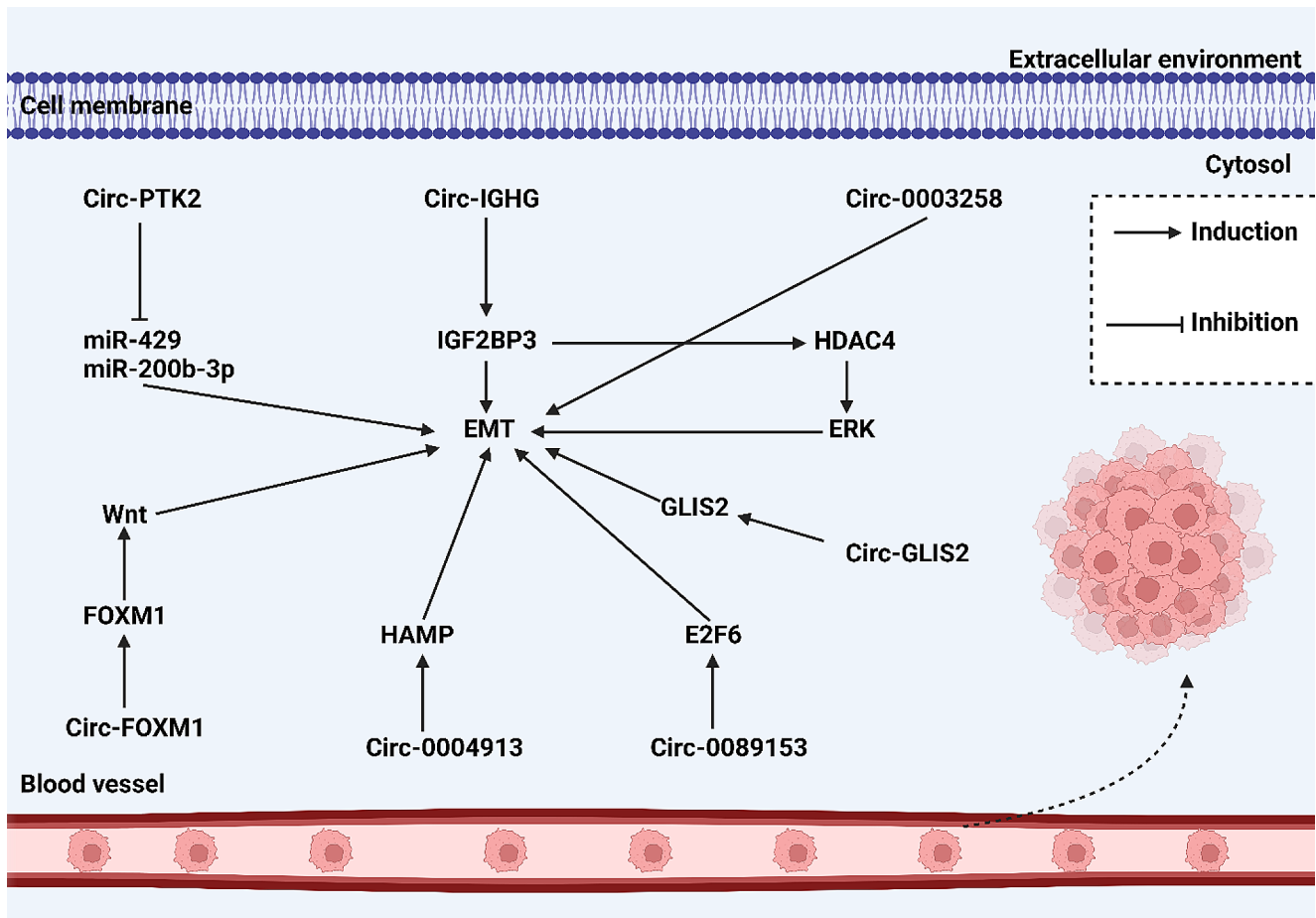


Fig. 4 CircRNA/EMT axis in modulating cancer invasion and metastasis. The circRNAs mainly induce EMT in increasing progression of tumor cells. In some cases, including circ-GLIS2, circ-0089153 and

circ-0004913, the circRNAs affect the proteins to regulate EMT. However, in other cases, such as circ-PTK2, the circRNAs sponge miRNAs to affect EMT.

the progression and invasion of colorectal cancer [158]. Furthermore, overexpression of TGF- β has been implicated in inhibiting apoptosis in tumor cells and increasing their survival rate [159]. TGF- β and c-Myc signaling pathways are both involved in gastric tumor cell progression and the induction of EMT mechanisms. The expression levels of TGF- β and c-Myc increase in gastric cancer. Notably, circ-CCDC66 stimulates EMT and accelerates the progression of gastric tumors by promoting TGF- β and c-Myc [160].

Circ-VANGL1 interacts with TGF- β to modulate tumorigenesis. miR-150-5p is an inhibitor of TGF- β signaling to prevent EMT mechanism in melanoma. Circ-VANGL1 sponges miR-150-5p to enhance TGF- β expression, resulting in EMT induction and enhancement of melanoma metastasis [161]. Furthermore, TGF- β can regulate expression of circRNAs to affect EMT mechanism. Activation of TGF- β signaling and phosphorylation of Smad2/3 can result in upregulation of circ-Akt1 which in turn increase Akt1 expression via miR-942-5p inhibition to then promote cancer metastasis and EMT induction [162]. Moreover,

TGF- β 2 promotes the expression level of circ-PRDM5 to enhance COL1A2 expression via miR-92b-3p inhibition, thereby triggering EMT mechanisms and elevating cancer invasion [163].

Twist

Another important regulator of the EMT mechanism is Twist. Upregulation of Twist can lead to the induction of cancer metastasis via EMT. Silencing EGFR leads to suppression of tamoxifen resistance in breast cancer via reducing Twist expression to impair the EMT mechanism [164]. Twist expression decreases when METTL14 is inhibited, resulting in a reduction in lung cancer progression and invasion [165]. Notably, non-coding RNA molecules can regulate Twist-mediated EMT in cancer [166]. CircRNAs are effective Twist regulators in cancer EMT mechanisms. CircRAB31P increases the migration and invasion of osteosarcoma cells. miR-580-3p is an inhibitor of Twist1, which reduces the malignancy of osteosarcoma cells. However,

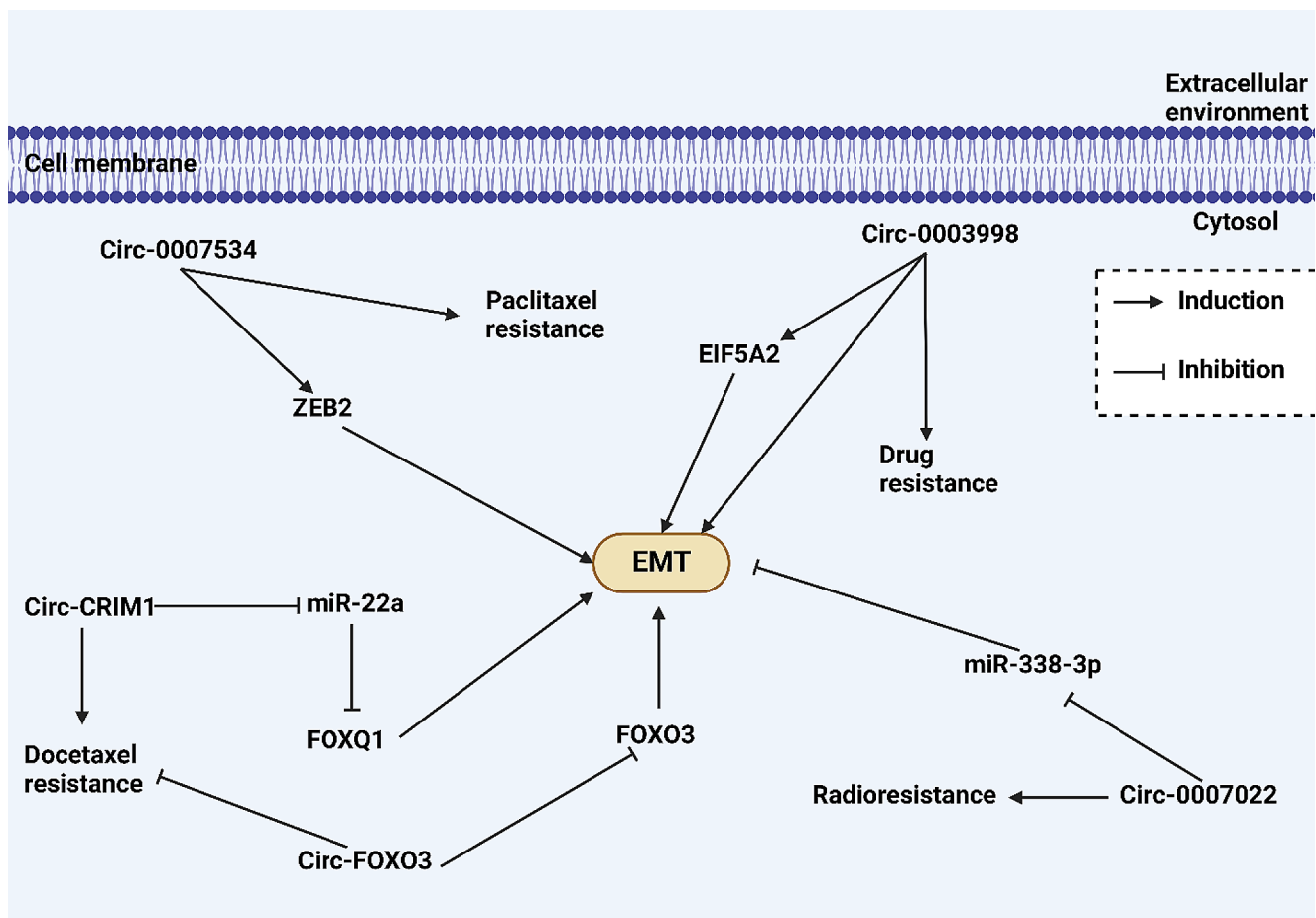


Fig. 5 CircRNA/EMT axis in drug resistance and radio-resistance. The upregulation of ZEB2 by circ-0007534 can induce EMT to mediate paclitaxel resistance. Moreover, circ-0003998 increases EIF5A2 levels

to mediate EMT-induced drug resistance. The circ-0007022 inhibits miR-338-3p to mediate EMT-induced radioresistance. The sponging of miR-22a by circ-CRIM1 can induce docetaxel resistance

circRAB3IP reduces miR-580-3p expression to upregulate Twist1 and promote osteosarcoma malignancy [167]. CircRNA PVT1 is upregulated in cancers and stimulates the Wnt4/ β -catenin axis to increase carcinogenesis [168]. Silencing PVT1 is advantageous for reducing the progression of lung cancer and enhancing sensitivity to cisplatin [169]. Reducing the expression level of oncogenic circRNAs is beneficial in impairing tumorigenesis. Upregulation of circ-0001681 results in Twist1 overexpression to enhance thyroid cancer metastasis. Silencing circ-0001681 impairs tumor invasion via EMT inhibition. Mechanistically, circ-0001681 increases Twist1 expression via miR-942-5p sponging to induce EMT and cancer invasion [170]. The interesting point is that the expression level of circRNAs can be increased by Twist during cancer. Circ-10,720 overexpression can lead to EMT induction in hepatocellular carcinoma. Twist1 increases the expression level of circ-10,720 to upregulate vimentin, trigger EMT, and enhance cancer invasion [171]. These studies highlight the fact that EMT-TFs are regulated by circRNAs in cancer, and

interconnected molecular pathways need to be highlighted in the near future.

CircRNA/EMT axis in different cancers

Brain cancers

One of the malignancies of the central nervous system is glioma, and due to its aggressive nature, the survival rate of patients is 12–14 months. Furthermore, other challenges, including resistance to therapy and recurrence, prevent effective strategies for treatment [172, 173]. Increasing evidence demonstrates the role of circRNAs in regulating the progression of gliomas. Circ-0000215 promotes glioma progression by activating the EMT mechanism, which mediates metastasis. Circ-0000215 reduces the expression level of miR-495-3p to upregulate CXCR2, induce EMT, and pave the way for glioma metastasis [174]. miR-599 is an inhibitor of EMT in esophageal cancer that reduces the progression of

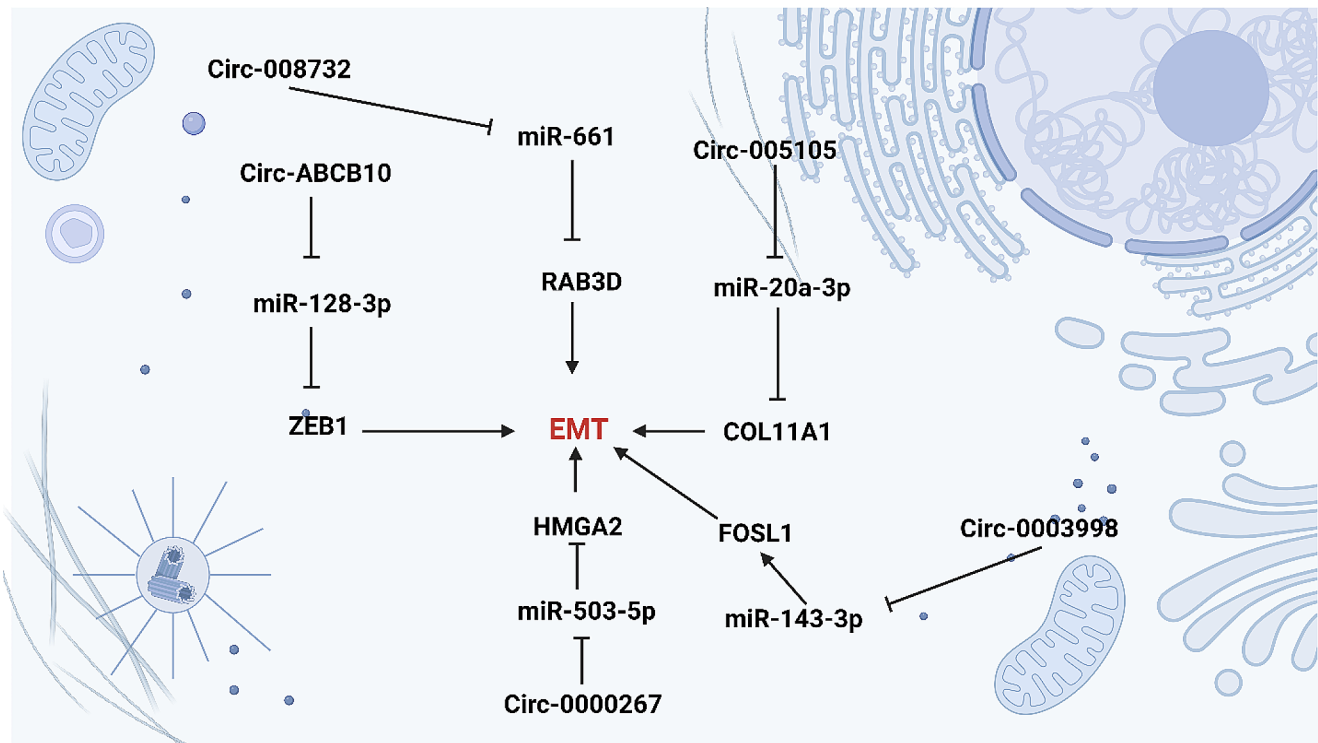


Fig. 6 CircRNA/miRNA/EMT axis in human cancers. The circRNAs mainly function as ceRNA to sponge miRNAs in the control of carcinogenesis. The circ-008732 sponges miR-661 to upregulate RAB3D in EMT induction. Moreover, circ-ABCB10 sponges miR-128-3p

to stimulate EMT via ZEB1 overexpression. The circ-0000267 suppresses miR-503-5p to increase HMG2 levels in EMT. Finally, circ-0003998 stimulates EMT through miR-143-3p inhibition

tumor cells. Circ-0030018 is an oncogenic factor in esophageal cancer and sponges miR-599 to increase the expression of ENAH. The positive association of circ-0030018 with ENAH expression is vital for inducing EMT and increasing tumor cell invasion and metastasis [175]. Like glioma, EMT induction poses a treatment challenge in glioblastoma. MICAL2 promotes glioblastoma progression by inducing EMT mechanisms via TGF- β upregulation [176]. GRP78 stabilization by UBE2T can lead to EMT induction, thereby increasing the invasion of glioblastoma cells [177]. Anti-cancer agents such as Eriodictyol have been used to reduce ZEB1 expression to suppress EMT in glioblastoma [178]. EMT is commonly induced in glioblastoma. Circ-0001801 stimulates EMT to increase glioblastoma progression. Circ-0001801 enhances the expression level of HMGB3 via miR-628-5p sponging to induce EMT in favor of glioblastoma malignancy [179]. Circ-0067934 also induces EMT in glioblastoma metastasis by inducing PI3K/Akt signaling to mediate EMT and enhance tumor invasion [180].

Gastrointestinal cancers

- Gastric cancer is a life-threatening disease and is the third leading cause of death [181]. Gastric tumor cells

show little response to chemotherapy and radiotherapy and manifest a metastatic nature. The malignancy of gastric cancer cells can be enhanced by circRNAs. An experiment has shown that upregulation of circ-101,882 is beneficial in promoting malignancy and the progression of gastric tumor cells. Circ-101,882 prevents apoptosis and enhances invasion of cancer cells. For this purpose, circ-101,882 promotes vimentin, N-cadherin, and Snail levels while decreasing E-cadherin expression to stimulate EMT [182]. One of the changes in the tumor microenvironment that can increase gastric cancer progression is hypoxia, which promotes CD36 expression to mediate peritoneal invasion [183]. The expression level of non-coding RNAs changes under hypoxia in gastric cancer [184, 185]. Hypoxia induces EMT in gastric cancer and promotes the progression of tumor cells. Circ-0081143 enhances the expression level of EGFR via miR-497-5p inhibition. Since miR-497-5p suppresses EMT, it reduces gastric cancer progression. Conversely, down-regulation of circ-0081143 can lead to EMT induction [186].

- The malignancy of the colon or rectum is known as colorectal cancer, which has high mortality rates [187–189]. The prognosis of colorectal cancer patients is poor despite advances in treatment [190, 191]. miR-338-5p

Table 4 Modulation of EMT mechanism by circRNA/miRNA axis in tumors

CircRNA	Molecular pathway	Cancer type	Remark	Ref
Circ-0088732	miR-661/RAB3D	Glioma	Circ-0088732 upregulates RAB3D via miR-661 sponging in EMT induction	[138]
Circ-ABCB10	miR-128-3p/ZEB1	Cervical cancer	Circ-ABCB10 upregulates ZEB1 by miR-128-3p inhibition in accelerating tumorigenesis and EMT	[139]
Circ-0000267	miR-503-5p/HMGGA2	Gastric cancer	Circ-0000267 upregulates HMGGA2 via miR-503-5p suppression in EMT stimulation	[140]
Circ-0000144	miR-217/EMT	Gastric cancer	Circ-0000144 sponges miR-217 in triggering EMT mechanism	[141]
Circ-CORO1C	miR-138-5p/KLFL12	Gastric cancer	Circ-CORO1C upregulates KLFL12 via miR-138-5p inhibition in EMT	[142]
Circ-03955	miR-3662/MTDH	Osteosarcoma	Circ-03955 promotes MTDH level via miR-3662 suppression in EMT induction	[143]
Circ-006100	miR-195/GPRC5A	Gastric cancer	Circ-006100 upregulates GPRC5A via miR-195 downregulation in EMT	[144]
Circ-0058106	miR-185-3p/Wnt2b/ β -catenin/c-Myc	Hypopharyngeal cancer	Circ-0058106 sponges miR-185-3p in inducing Wnt2b/ β -catenin/c-Myc in EMT induction	[145]

expression increases the progression of colorectal cancer and induces EMT. The expression level of circ-0137008 decreases in colorectal cancer samples, which leads to significant increases in the progression of tumor cells. Circ-0137008 suppresses EMT mechanisms in colorectal cancer via miR-338-5p down-regulation [192]. Silencing circ-0026416 can be considered an impediment to the progression of colorectal cancer. Circ-0026416 increases the progression of colorectal cancer via EMT induction. Circ-0026416 reduces miR-545-3p expression, which subsequently leads to an increased expression level of MYO6 to promote tumor metastasis [193].

Gynecological cancers

Cervical cancer is a gynecological malignant tumor with high incidence and death rates [194, 195]. The number of cervical cancer cases in developing countries is higher and comprises up to 85% of the global burden [196]. CircRNAs are considered key players in the progression of cervical cancer and the development of therapy resistance [197, 198]. Both proliferation and invasion of cervical tumor cells are enhanced by circ-MYBL2. However, the function of circ-MYBL2 in increasing cervical cancer metastasis is based on regulating EMT mechanisms. Circ-MYBL2 reduces miR-361-3p expression via sponging to enhance lymph node metastasis in cervical tumors [199]. On the other hand, miR-449a overexpression in cervical cancer can impair the progression of tumor cells. Notably, circ-CDK6 is an onco-suppressor factor, and by reducing miR-449a expression, it suppresses EMT in cervical tumors [200]. Due to the aggressive behavior of ovarian cancer cells and the unfavorable prognosis of patients, patients manifest higher death rates compared to cervical and endometrial cancers [201]. Similar to cervical cancer, the function of circRNAs in regulating the progression of ovarian cancer has been well-documented as they affect various molecular pathways [202–204]. A recent experiment has revealed the role of circ-0001756 in regulating ovarian cancer metastasis via affecting EMT mechanisms. Circ-0001756 promotes the expression level of RAB5A by inducing IGF2BP2 expression. Then, RAB5A stimulates the EGFR/MAPK axis to mediate EMT and increasing the progression and metastasis of ovarian tumor cells [205]. Similar to other tumor types, circRNAs regulate miRNA expression by affecting the progression of ovarian cancer. miR-361-5p is an inhibitor of ovarian cancer progression and reduces the expression levels of c-Met to inhibit the Akt/mTOR axis [206]. Circ-0061140 enhances the progression of ovarian cancer and induces EMT mechanisms. Circ-0061140 decreases miR-361-5p expression to promote RAB1A, which in turn

Table 5 The role of circRNA/EMT axis in regulating progression of cancers

CircRNA	Molecular pathway	Cancer type	Remark	Ref
Circ-0084904	miR-802/MAL2	Cervical cancer	Circ-0084904 promotes MAL2 expression by miR-802 inhibition in EMT induction	[229]
Circ-UBAP2	miR-361-3p/SOX4	Cervical cancer	Circ-UBAP2 sponges miR-361-3p to promote SOX4 expression in EMT stimulation	[230]
Circ-NFATC3	miR-9-5p/SDC2	Cervical cancer	Circ-NFATC3 promotes SDC2 expression by miR-9-5p inhibition in cancer progression development	[231]
Circ-OXCT1	miR-136/SMAD4	Gastric cancer	Circ-OXCT1 increases SMAD4 expression via miR-136 inhibition in EMT stimulation	[84]
Circ-0049447	miR-324-5p/EMT	Gastric cancer	Circ-0049447 inhibits EMT via miR-324-5p sponging in reducing tumor progression	[232]
Circ-0005230	miR-1299/RHOT1	Gastric cancer	Circ-0005230 increases RHOT1 expression by miR-1299 inhibition in triggering EMT mechanism	[233]
Circ-104,916	-	Gastric cancer	Low expression of circ-104,916 in gastric cancer	[234]
Circ-0029803	miR-216b-5p/SKIL	Colorectal cancer	Circ-104,916 inhibits EMT via E-cadherin upregulation and N-cadherin down-regulation	[235]
Circ-0053277	miR-2467-3p/MMP-14	Colorectal cancer	Circ-0029803 increases SKIL expression via miR-216b-5p sponging in EMT induction	[236]
Circ-0000395	miR-432-5p/MYH9	Colorectal cancer	Circ-0053277 promotes MMP-14 expression via miR-2467-3p inhibition in increasing cancer progression and EMT induction	[237]
Circ-0056618	miR-411-5p/PRRG4	Colorectal cancer	Circ-0000395 enhances MYH9 expression via miR-432-5p inhibition in EMT induction	[238]
Circ-0065378	miR-4701-5p/TUSC1	Colorectal cancer	EMT induction by Circ-0056618 via miR-411-5p sponging and upregulating PRRG4 expression	[239]
Circ-0001459	miR-6165/IGF1R	Hepatocellular carcinoma	Circ-0065378 reduces miR-4701-5p expression to upregulate TUSC1 expression in impairing EMT	[240]
Circ-0101145	miR-548c-3p/LAMC2	Hepatocellular carcinoma	Circ-0001459 increases IGF1R expression via miR-6165 inhibition in EMT induction	[241]
Circ-0003528	miR-421/MMP-3	Hepatocellular carcinoma	Circ-0101145 increases LAMC2 expression by miR-548c-3p inhibition in triggering EMT	[242]
Circ-PRKAR1B	miR-361-3p/FZD4	Osteosarcoma	Circ-0003528 increases MMP-3 expression via miR-421 inhibition in EMT induction	[243]
Circ-ITCH	miR-199a-5p/Klotho	Gastric cancer	EIF4A3 promotes Circ-PRKAR1B expression to sponge miR-361-3p in upregulating FZD4 expression and mediating EMT mechanism	[235]
Circ-HIPK3	miR-326	Breast cancer	Circ-ITCH promotes Klotho expression via miR-199a-5p inhibition in impairing EMT	[244]
			Circ-HIPK3 sponges miR-326 in increasing expression level of EMT markers for cancer progression	[244]

triggers EMT-mediated ovarian cancer invasion [207]. Based on these studies, circRNAs are important regulators of EMT in gynecological cancers.

Urological cancers

The lives of men are commonly threatened by a malignant disease known as prostate cancer, which causes 300,000 deaths annually [188]. Many deaths in prostate cancer patients result from metastasis [208]. Patients with metastasis have an overall survival of 42 months [209], whereas patients with progressive metastatic prostate cancer have a survival of 27 months [210]. New technologies have been developed for the treatment of prostate cancer patients, such as nanoplatforms, but prostate cancer is still an incurable disease. CircRNAs are potential regulators of prostate cancer progression, affecting various molecular pathways [71, 210, 211]. Furthermore, accumulating data emphasizes the role of EMT as an inducer of prostate cancer invasion [212]. The function of circ-0004296 significantly reduces the metastasis of prostate cancer cells. Since EMT is an inducer of prostate cancer invasion, it is regulated by circ-0004296. EIF4A3 interacts with circ-0004296 to suppress nuclear translocation of ETS1, leading to impairment of prostate cancer invasion via EMT suppression [213]. Overexpression of circ-0030586 in prostate cancer can lead to induction of EMT, which enhances the invasion of tumor cells [214]. On the other hand, induction of Akt signaling can result in an increase in the progression of prostate cancer [215, 216]. Circ-0030586 stimulates PI3K/Akt signaling to induce EMT and enhance the invasion and metastasis of prostate tumor cells. This is mediated by E-cadherin downregulation and Twist upregulation [214]. Therefore, the circRNA/EMT axis is a regulator of prostate cancer invasion. The delineation of molecular pathways that can be targeted for treatment of this malignant disease warrants further research [217, 218].

Another malignancy of the genitourinary tract is bladder cancer, with increasing morbidity and mortality around the world [201]. Despite using chemotherapy, radiotherapy, and surgery, the 5-year survival rate of bladder cancer patients is low and has a high economic burden [219, 220]. New therapeutic approaches such as nanotheranostics have been developed for bladder cancer. However, it is highly recommended to focus on the underlying molecular pathways involved in bladder cancer progression. The expression level of circRNAs changes during bladder cancer development and progression. Besides, the metastasis of bladder cancer cells is highly dependent on the activation of the EMT mechanism [221–223]. The interaction of circRNAs with EMT in bladder cancer is important. Circ-BIRC6 is involved in enhancing the progression and metastasis of bladder tumor

cells, and its depletion is key in reducing tumor malignancy. Circ-BIRC6 increases the expression level of XBP1 by inhibiting miR-495-3p to enhance the metastasis of bladder cancer cells via EMT induction. As a result, silencing circ-BIRC6 is advantageous in reversing EMT in bladder tumors [224]. In fact, the malignant behavior of bladder cancer cells is mediated via EMT induction. Circ-0006948 contributes to the progression of bladder tumor cells by reducing E-cadherin and increasing N-cadherin, vimentin, and β -catenin levels to stimulate EMT [224]. One of the key inducers of the EMT mechanism in cancer is HMGA2. HMGA2 stimulates Wnt/ β -catenin signaling is activated by HMGA2 to mediate EMT and promote the progression of gastric tumor cells [225]. In bladder cancer, upregulation of HMGA2 leads to EMT induction, which enhances the progression of tumor cells. Notably, circ-0000658 promotes the expression level of HMGA2 via miR-498 inhibition to facilitate tumorigenesis in bladder cancer [226]. Therefore, the circRNA/EMT axis is a promising target in bladder cancer therapy (Table 5) [227, 228].

Conclusion and remarks

Cancer metastasis involves the migration of tumors from their original location to different organs, where they establish new growths. This journey necessitates breaking away from adjacent tissue and entering the circulatory system. Elevated motility and aggressiveness in cancer cells correspond to heightened rates of metastasis. One of the problems of metastasis is related to the diffusion of cancer cells into distant parts of body, challenging eradication of cancer cells. EMT induction is one mechanism facilitating this spread. As detachment of a number of cancer cells from other parts of colony is required for metastasis, the EMT can increase motility of tumor cells to facilitate this process. This paper delves into the influence of EMT-associated pathways in cancer, particularly highlighting the role of epigenetic factors such as circRNAs in modulating EMT. The dysregulation of epigenetic factors in cancer is a common feature and the researchers have focused on understanding the association between epigenetic alterations and EMT in context of cancer progression and EMT. EMT significantly boosts the migratory capabilities of tumor cells, a fact supported by studies across various types of cancers such as those affecting the brain, gastrointestinal tract, reproductive system, urinary system, and blood. It should be noted that function of EMT in cancer is versatile and it is beyond regulation of metastasis and can affect cancer therapy resistance and mediate immunosuppression. Most crucially, circRNAs serve as powerful modulators of EMT pathways in these diverse cancer types. Targeting EMT directly or adjusting

the expression levels of circRNAs emerge as two effective approaches for cancer treatment, with the potential to reduce metastatic spread and improve both patient survival rates and prognoses. Since EMT is a reversible process, targeting related circRNAs can significantly change the progression of cancer cells. In exploring the relationship between circRNA and the EMT axis in cancer, two crucial dimensions emerge: metastasis and responsiveness to treatment. On one hand, circRNAs have the ability to either trigger or suppress EMT processes, thus influencing the invasive capabilities of cancer cells. On the other hand, the regulation of EMT by circRNAs plays a key role in determining how cancer cells react to therapeutic interventions. When circRNAs promote EMT, they essentially bolster the malignant properties of tumors, setting the stage for resistance to treatment. Conversely, suppressing EMT through circRNAs can increase a tumor sensitivity to both drug and radiation therapies. Yet, circRNAs are not confined to modulating EMT directly; they can also affect other signaling pathways. A primary function of circRNAs in this context is their ability to sponge miRNAs, thus affecting EMT processes in cancer. Consequently, the circRNA/EMT axis serves as a promising target for therapeutic interventions. EMT-related transcription factors such as ZEB proteins, TGF- β , Twist, and Slug can also be influenced by circRNAs, impacting both the metastasis and severity of cancer. Preliminary studies suggest that circRNAs are pivotal in controlling EMT in cancer, and future research may well center on their potential as biomarkers for treatment outcomes and long-term prognosis in cancer patients. Although significant efforts have been made in understanding the function of circRNAs in regulating EMT in cancers, there are still some limitations to be explored in the future studies. A number of anti-cancer compounds have shown potential in the inhibition of EMT. However, the studies should focus on the compounds regulating circRNA/EMT axis. Moreover, the genetic tools, specially CRISPR/Cas9 system should be introduced for circRNA/EMT axis regulation in human cancer therapy. One of the most promising limitations is lack of emphasizing on nanoparticles for targeting circRNA/EMT axis to impair metastasis and invasion of cancer cells.

The pre-clinical studies provide the valuable insights regarding the metastasis of tumor cells mediated by EMT and then, the function of circRNAs in the modulation of EMT-associated metastasis. Based on the estimates, up to 90% of cancer-related deaths in patients are due to the metastasis. In fact, the metastasis allows the tumor cells to diffuse into various parts of body and develop new colonies that further can proliferate and improve the survival of tumors. Therefore, the modulation of metastasis can be of high importance in clinical level. Since circRNAs can mainly affect metastasis through EMT regulation and their stability is high, they can

be utilized as diagnostic and prognostic tools in patients. The changes in the circRNAs in serum or exosomal serum can provide a non-invasive method for the metastasis prediction in patients. In addition to prognostic and diagnostic tool, the circRNAs regulating EMT mechanism can be considered as valuable targets in cancer therapy. However, the drugs have poor ability in the regulation of circRNA regulation and therefore, it is suggested to apply the genetic tools to regulate circRNA expression in cancer therapy.

Author contributions M.A and M.Z developed the idea, designed outline and collected papers. P.T, N.N, M.T, M.Z and M.A wrote the paper and prepared the first draft. A.R.A critically edited the paper. J.D revised the paper, drew the figures and improved English editing.

Declarations

Conflict of interest The authors declare no conflict of interest.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Babaei G, Aziz SG-G, Jaghi NZZ, EMT (2021) cancer stem cells and autophagy; the three main axes of metastasis. *Biomed Pharmacother* 133:110909. <https://doi.org/10.1016/j.biopha.2020.110909>
- Talmadge JE, Fidler IJ (2010) J.C.R. AACR centennial series: the biology of cancer metastasis: historical perspective. 70:5649–5669
- Baldawa P, Shirol P, Alur J, Kulkarni VV, JOMFP MP, Metastasis (2017) *fro* 21:463
- PSJNrc Steeg (2016) Targeting metastasis. 16:201–218
- Nieto MA, Huang RY-J, Jackson RA, Thiery JP (2016) EMT: 2016. *Cell* 166:21–45. <https://doi.org/10.1016/j.cell.2016.06.028>
- Lo U-G, Lee C-F, Lee M-S, Hsieh (2017) J.-T.J.I.j.o.m.s. The role and mechanism of epithelial-to-mesenchymal transition in prostate cancer progression. 18:2079
- Moreno-Bueno G, Portillo F, Cano A (2008) Transcriptional regulation of cell polarity in EMT and cancer. *Oncogene* 27:6958–6969. <https://doi.org/10.1038/onc.2008.346>
- Knust E, Bossinger OJS (2002) Composition and formation of intercellular junctions in epithelial cells. 298:1955–1959
- Perez-Moreno M, Jamora C, Fuchs EJC (2003) Sticky business: orchestrating cellular signals at adherens junctions. 112:535–548
- Peinado H, Portillo F, Cano (2004) A.J.I.J.o.D.B. Transcriptional regulation of cadherins during development and carcinogenesis. 48:365–375

11. Jeanes A, Gottardi C, Yap AJO (2008) Cadherins and cancer: how does cadherin dysfunction promote tumor progression? 27:6920–6929
12. Jiang Y, Zhuo X, Wu Y, Fu X, Mao C (2022) PAR2 blockade reverses osimertinib resistance in non-small-cell lung cancer cells via attenuating ERK-mediated EMT and PD-L1 expression. *Biochim et Biophys Acta Mol Cell Res* 1869:119144. <https://doi.org/10.1016/j.bbamcr.2021.119144>
13. Lee DE, Jang EH, Bang C, Kim GL, Yoon SY, Lee DH, Koo J, Na JH, Lee S, Kim JH (2021) Bakuchiol, main component of root bark of *Ulmus davidiana* var. *japonica*, inhibits TGF- β -induced in vitro EMT and in vivo metastasis. *Arch Biochem Biophys* 709:108969. <https://doi.org/10.1016/j.abb.2021.108969>
14. Lu T, Zheng C, Fan Z (2022) Cardamonin suppressed the migration, invasion, epithelial mesenchymal transition (EMT) and lung metastasis of colorectal cancer cells by down-regulating ADRB2 expression. *Pharm Biol* 60:1011–1021. <https://doi.org/10.1080/13880209.2022.2069823>
15. Bose C, Das U, Kuilya TK, Mondal J, Bhadra J, Banerjee P, Goswami RK, Sinha S (2022) Cananginone abrogates EMT in breast Cancer cells through hedgehog signaling. *Chem Biodivers* 19:e202100823. <https://doi.org/10.1002/cbdv.202100823>
16. Liu J, Chen X, Zhou X, Yi R, Yang Z, Zhao X (2021) *Lactobacillus fermentum* ZS09 mediates epithelial-mesenchymal transition (EMT) by regulating the transcriptional activity of the Wnt/ β -Catenin signalling pathway to inhibit Colon cancer activity. *J Inflamm Res* 14:7281–7293. <https://doi.org/10.2147/jir.S344564>
17. Block CJ, Mitchell AV, Wu L, Glassbrook J, Craig D, Chen W, Dyson G, DeGracia D, Polin L, Ratnam M et al (2021) RNA binding protein RBMS3 is a common EMT effector that modulates triple-negative breast cancer progression via stabilizing PRRX1 mRNA. *Oncogene* 40:6430–6442. <https://doi.org/10.1038/s41388-021-02030-x>
18. Kitz J, Lefebvre C, Carlos J, Lowes LE, Allan AL (2021) Reduced Zeb1 expression in prostate Cancer cells leads to an aggressive Partial-EMT phenotype Associated with altered global methylation patterns. *Int J Mol Sci* 22. <https://doi.org/10.3390/ijms222312840>
19. Lin XM, Wang ZJ, Lin YX, Chen H (2021) Decreased exosome-delivered mir-486-5p is responsible for the peritoneal metastasis of gastric cancer cells by promoting EMT progress. *World J Surg Oncol* 19:312. <https://doi.org/10.1186/s12957-021-02381-5>
20. Lamouille S, Xu J, Derynck R (2014) Molecular mechanisms of epithelial–mesenchymal transition. *Nat Rev Mol Cell Biol* 15:178–196. <https://doi.org/10.1038/nrm3758>
21. Xue W, Yang L, Chen C, Ashrafizadeh M, Tian Y, Sun R (2024) Wnt/ β -catenin-driven EMT regulation in human cancers. *Cell Mol Life Sci* 81:79. <https://doi.org/10.1007/s00018-023-05099-7>
22. Guo Z, Ashrafizadeh M, Zhang W, Zou R, Sethi G, Zhang X (2023) Molecular profile of metastasis, cell plasticity and EMT in pancreatic cancer: a pre-clinical connection to aggressiveness and drug resistance. *Cancer Metastasis Rev*. <https://doi.org/10.1007/s10555-023-10125-y>
23. Zheng Y, Li P, Huang H, Ye X, Chen W, Xu G, Zhang F (2021) Androgen receptor regulates eIF5A2 expression and promotes prostate cancer metastasis via EMT. *Cell Death Discovery* 7:373. <https://doi.org/10.1038/s41420-021-00764-x>
24. Zhang X, Luo Y, Cen Y, Qiu X, Li J, Jie M, Yang S, Qin S (2022) MACC1 promotes pancreatic cancer metastasis by interacting with the EMT regulator SNAI1. *Cell Death Dis* 13:923. <https://doi.org/10.1038/s41419-022-05285-8>
25. Xiong T, Huang C, Li J, Yu S, Chen F, Zhang Z, Zhuang C, Li Y, Zhuang C, Huang X et al (2020) LncRNA NRON promotes the proliferation, metastasis and EMT process in bladder cancer. *J Cancer* 11:1751–1760. <https://doi.org/10.7150/jca.37958>
26. Xiao GY, Tan X, Rodriguez BL, Gibbons DL, Wang S, Wu C, Liu X, Yu J, Vasquez ME, Tran HT et al (2023) EMT activates exocytotic rabs to coordinate invasion and immunosuppression in lung cancer. *Proc Natl Acad Sci USA* 120:e2220276120. <https://doi.org/10.1073/pnas.2220276120>
27. Liang S, Guo H, Ma K, Li X, Wu D, Wang Y, Wang W, Zhang S, Cui Y, Liu Y et al (2021) A PLCB1-PI3K-AKT Signaling Axis activates EMT to promote Cholangiocarcinoma Progression. *Cancer Res* 81:5889–5903. <https://doi.org/10.1158/0008-5472.can-21-1538>
28. Papatsirou M, Artemaki PI, Scorilas A, Kontos CK (2020) J.P.M. The role of circular RNAs in therapy resistance of patients with solid tumors. 17:469–490
29. Hsu M-T, Coca-Prados MJN (1979) Electron microscopic evidence for the circular form of RNA in the cytoplasm of eukaryotic cells. 280:339–340
30. Kos A, Dijkema R, Arnberg A, Van der Meide P, Schellekens (1986) H.J.N. The hepatitis delta (δ) virus possesses a circular RNA. 323:558–560
31. Nigro JM, Cho KR, Fearon ER, Kern SE, Ruppert JM, Oliner JD, Kinzler KW, Vogelstein (1991) B J C Scrambled Exons 64:607–613
32. Chen X, Han P, Zhou T, Guo X, Song X, Li YJ (2016) S.r. circRNADb: a comprehensive database for human circular RNAs with protein-coding annotations. 6:1–6
33. Yu J, Yang L, Lu H (2021) The emerging role of circular RNAs in common solid malignant tumors in children. *Cancer Cell Int* 21:309. <https://doi.org/10.1186/s12935-021-01998-6>
34. Jeck WR, Sharpless NE (2014) Detecting and characterizing circular RNAs. *Nat Biotechnol* 32:453–461. <https://doi.org/10.1038/nbt.2890>
35. Shen B, Sun K (2021) Exosomal circular RNAs: a new frontier in the metastasis of digestive system tumors. *Oncol Lett* 22. <https://doi.org/10.3892/ol.2021.13087>
36. Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, Damgaard CK, Kjems J (2013) Natural RNA circles function as efficient microRNA sponges. *Nature* 495:384–388. <https://doi.org/10.1038/nature11993>
37. Zheng Q, Bao C, Guo W, Li S, Chen J, Chen B, Luo Y, Lyu D, Li Y, Shi G et al (2016) Circular RNA profiling reveals an abundant circHIPK3 that regulates cell growth by sponging multiple miRNAs. *Nat Commun* 7. <https://doi.org/10.1038/ncomms11215>
38. Meng L, Chang S, Sang Y, Ding P, Wang L, Nan X, Xu R, Liu F, Gu L, Zheng Y et al (2022) Circular RNA circCCDC85A inhibits breast cancer progression via acting as a miR-550a-5p sponge to enhance MOB1A expression. *Breast Cancer Research: BCR* 24:1. <https://doi.org/10.1186/s13058-021-01497-6>
39. Wang R, Wang J, Chen Y, Chen Y, Xi Q, Sun L, Zhang X, Zhang G, Ding X, Shi T et al (2022) Circular RNA circDLDR facilitates cancer progression by altering the miR-30a-3p/SOAT1 axis in colorectal cancer. *Cell Death Discovery* 8. <https://doi.org/10.1038/s41420-022-01110-5>
40. Gao Z, Hu L, Chen F, He C, Hu B, Wang X (2022) Hsa_circular RNA_0001013 exerts oncogenic effects in gastric cancer through the microRNA-136-TWSG1 axis. *Am J Translational Res* 14:4948–4963
41. Liu J, Dai Z, Li M, Wang B, Zhang X, Li F, Zhang M, Zhang W (2022) Circular RNA circMET contributes to tamoxifen resistance of breast cancer cells by targeting miR-204/AHR signaling. *Biochem Biophys Res Commun* 627:200–206. <https://doi.org/10.1016/j.bbrc.2022.07.097>
42. Fu L, Zhang D, Yi N, Cao Y, Wei Y, Wang W, Li L, Circular (2022) RNA circPBX3 promotes cisplatin resistance of ovarian cancer cells via interacting with IGF2BP2 to stabilize ATP7A mRNA expression. *Hum Cell* 35:1560–1576. <https://doi.org/10.1007/s13577-022-00748-8>

43. Li H, Luo F, Jiang X, Zhang W, Xiang T, Pan Q, Cai L, Zhao J, Weng D, Li Y et al (2022) CircITGB6 promotes ovarian cancer cisplatin resistance by resetting tumor-associated macrophage polarization toward the M2 phenotype. *J Immunother Cancer* 10. <https://doi.org/10.1136/jitc-2021-004029>
44. Chen L, Shan G (2021) CircRNA in cancer: fundamental mechanism and clinical potential. *Cancer Lett* 505:49–57. <https://doi.org/10.1016/j.canlet.2021.02.004>
45. Chen L, Huang C, Wang X, Shan G (2015) Circular RNAs in eukaryotic cells. *Curr Genom* 16:312–318
46. Huang G, Li S, Yang N, Zou Y, Zheng D, Xiao T (2017) Recent progress in circular RNAs in human cancers. *Cancer Lett* 404:8–18
47. Zhang Y, Zhang X-O, Chen T, Xiang J-F, Yin Q-F, Xing Y-H, Zhu S, Yang L, Chen (2013) L.-L. circular intronic long noncoding RNAs. *Mol Cell* 51:792–806
48. Li Z, Huang C, Bao C, Chen L, Lin M, Wang X, Zhong G, Yu B, Hu W, Dai L (2015) Exon-intron circular RNAs regulate transcription in the nucleus. *Nat Struct Mol Biol* 22:256–264
49. Liu X, Wang X, Li J, Hu S, Deng Y, Yin H, Bao X, Zhang QC, Wang G, Wang B (2020) Identification of mecciRNAs and their roles in the mitochondrial entry of proteins. *Sci China Life Sci* 63:1429–1449
50. Ashwal-Fluss R, Meyer M, Pamudurti NR, Ivanov A, Bartok O, Hanan M, Evantal N, Memczak S, Rajewsky N, Kadener S (2014) circRNA biogenesis competes with pre-mRNA splicing. *Mol Cell* 56:55–66
51. Kristensen LS, Hansen TB, Venø MT, Kjems J (2018) Circular RNAs in cancer: opportunities and challenges in the field. *Oncogene* 37:555–565
52. Xue C, Li G, Zheng Q, Gu X, Bao Z, Lu J, Li L (2022) The functional roles of the circRNA/Wnt axis in cancer. *Mol Cancer* 21. <https://doi.org/10.1186/s12943-022-01582-0>
53. Du WW, Zhang C, Yang W, Yong T, Awan FM, Yang BB (2017) Identifying and characterizing circRNA-Protein Interaction. *Theranostics* 7:4183–4191. <https://doi.org/10.7150/thno.21299>
54. Zang J, Lu D, Xu A (2020) The interaction of circRNAs and RNA binding proteins: an important part of circRNA maintenance and function. *J Neurosci Res* 98:87–97. <https://doi.org/10.1002/jnr.24356>
55. Okholm TLH, Sathe S, Park SS, Kamstrup AB, Rasmussen AM, Shankar A, Chua ZM, Fristrup N, Nielsen MM, Vang S et al (2020) Transcriptome-wide profiles of circular RNA and RNA-binding protein interactions reveal effects on circular RNA biogenesis and cancer pathway expression. *Genome Med* 12. <https://doi.org/10.1186/s13073-020-00812-8>
56. Wang J, Zhao X, Wang Y, Ren F, Sun D, Yan Y, Kong X, Bu J, Liu M, Xu S (2020) circRNA-002178 act as a ceRNA to promote PDL1/PD1 expression in lung adenocarcinoma. *Cell Death Dis* 11. <https://doi.org/10.1038/s41419-020-2230-9>
57. Li H, Xu JD, Fang XH, Zhu JN, Yang J, Pan R, Yuan SJ, Zeng N, Yang ZZ, Yang H et al (2020) Circular RNA circRNA_000203 aggravates cardiac hypertrophy via suppressing miR-26b-5p and mir-140-3p binding to Gata4. *Cardiovascular Res* 116:1323–1334. <https://doi.org/10.1093/cvr/cvz215>
58. Hansen TB, Kjems J, Damgaard CK (2013) Circular RNA and miR-7 in cancer. *Cancer Res* 73:5609–5612. <https://doi.org/10.1158/0008-5472.can-13-1568>
59. Chen X, Yang T, Wang W, Xi W, Zhang T, Li Q, Yang A, Wang T (2019) Circular RNAs in immune responses and immune diseases. *Theranostics* 9:588–607. <https://doi.org/10.7150/thno.29678>
60. Yan L, Chen YG (2020) Circular RNAs in Immune Response and viral infection. *Trends Biochem Sci* 45:1022–1034. <https://doi.org/10.1016/j.tibs.2020.08.006>
61. Yang J, Cheng M, Gu B, Wang J, Yan S, Xu D (2020) CircRNA_09505 aggravates inflammation and joint damage in collagen-induced arthritis mice via miR-6089/AKT1/NF- κ B axis. *Cell Death Dis* 11:833. <https://doi.org/10.1038/s41419-020-03038-z>
62. Wu P, Mo Y, Peng M, Tang T, Zhong Y, Deng X, Xiong F, Guo C, Wu X, Li Y et al (2020) Emerging role of tumor-related functional peptides encoded by lncRNA and circRNA. *Mol Cancer* 19. <https://doi.org/10.1186/s12943-020-1147-3>
63. Wang Y, Liu B, Circular RNA (2020) Diseased Heart Cells 9. <https://doi.org/10.3390/cells9051240>
64. Wang J, Zhu S, Meng N, He Y, Lu R, Yan GR (2019) ncRNA-Encoded peptides or proteins and Cancer. *Mol Therapy: J Am Soc Gene Therapy* 27:1718–1725. <https://doi.org/10.1016/j.ymthe.2019.09.001>
65. Wei Y, Zhang Y, Meng Q, Cui L, Xu C (2019) Hypoxia-induced circular RNA has_circRNA_403658 promotes bladder cancer cell growth through activation of LDHA. *Am J Translational Res* 11:6838–6849
66. Zhan W, Liao X, Chen Z, Li L, Tian T, Yu L, Wang W, Hu Q (2020) Circular RNA hsa_circRNA_103809 promoted hepatocellular carcinoma development by regulating miR-377-3p/FGFR1/ERK axis. *J Cell Physiol* 235:1733–1745. <https://doi.org/10.1002/jcp.29092>
67. Miao Z, Li J, Wang Y, Shi M, Gu X, Zhang X, Wei F, Tang X, Zheng L, Xing Y (2023) Hsa_circ_0136666 stimulates gastric cancer progression and tumor immune escape by regulating the miR-375/PRKDC Axis and PD-L1 phosphorylation. *Mol Cancer* 22. <https://doi.org/10.1186/s12943-023-01883-y>
68. Zhang C, Zhou X, Geng X, Zhang Y, Wang J, Wang Y, Jing J, Zhou X, Pan W (2021) Circular RNA hsa_circ_0006401 promotes proliferation and metastasis in colorectal carcinoma. *Cell Death Dis* 12:443. <https://doi.org/10.1038/s41419-021-03714-8>
69. Liu Y, Zhang H, Zhang W, Xiang L, Yin Z, Xu H, Lu P, Ma Y, Xiong L, Zhang X et al (2022) circ_0004140 promotes LUAD tumor progression and immune resistance through circ_0004140/miR-1184/CCL22 axis. *Cell Death Discovery* 8:181. <https://doi.org/10.1038/s41420-022-00983-w>
70. Shen X, Kong S, Ma S, Shen L, Zheng M, Qin S, Qi J, Wang Q, Cui X, Ju S (2022) Hsa_circ_0000437 promotes pathogenesis of gastric cancer and lymph node metastasis. *Oncogene* 41:4724–4735. <https://doi.org/10.1038/s41388-022-02449-w>
71. Yu YZ, Lv DJ, Wang C, Song XL, Xie T, Wang T, Li ZM, Guo JD, Fu DJ, Li KJ et al (2022) Hsa_circ_0003258 promotes prostate cancer metastasis by complexing with IGF2BP3 and sponging miR-653-5p. *Mol Cancer* 21. <https://doi.org/10.1186/s12943-021-01480-x>
72. Zhong J, Xu A, Xu P, Su M, Wang P, Liu Z, Li B, Liu C, Jiang N (2023) Circ_0000235 targets MCT4 to promote glycolysis and progression of bladder cancer by sponging miR-330-5p. *Cell Death Discovery* 9:283. <https://doi.org/10.1038/s41420-023-01582-z>
73. Ju C, Zhou M, Du D, Wang C, Yao J, Li H, Luo Y, He F, He J (2023) EIF4A3-mediated circ_0042881 activates the RAS pathway via miR-217/SOS1 axis to facilitate breast cancer progression. *Cell Death Dis* 14. <https://doi.org/10.1038/s41419-023-06085-4>
74. Zeng Y, Du W, Huang Z, Wu S, Ou X, Zhang J, Peng C, Sun X, Tang H (2023) Hsa_circ_0060467 promotes breast cancer liver metastasis by complexing with eIF4A3 and sponging miR-1205. *Cell Death Discovery* 9. <https://doi.org/10.1038/s41420-023-01448-4>
75. Li H, Jiao W, Song J, Wang J, Chen G, Li D, Wang X, Bao B, Du X, Cheng Y et al (2023) circ-hnRNPU inhibits NONO-mediated c-Myc transactivation and mRNA stabilization essential for glycosylation and cancer progression. *J Experimental Clin Cancer Research: CR* 42. <https://doi.org/10.1186/s13046-023-02898-5>
76. Zhang Y, Wang K, Yang D, Liu F, Xu X, Feng Y, Wang Y, Zhu S, Gu C, Sheng J et al (2023) Hsa_circ_0094606 promotes malignant

- progression of prostate cancer by inducing M2 polarization of macrophages through PRMT1-mediated arginine methylation of ILF3. *Carcinogenesis* 44:15–28. <https://doi.org/10.1093/carcin/bgac091>
77. Lv J, Zhang Y, Yang M, Qiao L, Wang H, Jiang H, Fu M, Qin J, Xu S (2024) Hsa_circ_0013561 promotes epithelial-mesenchymal transition and tumor progression by regulating ANXA2 via miR-23b-3p in ovarian cancer. *Cancer Gene Ther* 31:108–118. <https://doi.org/10.1038/s41417-023-00686-z>
 78. Yang M, Hu H, Wu S, Ding J, Yin B, Huang B, Li F, Guo X, Han L (2022) EIF4A3-regulated circ_0087429 can reverse EMT and inhibit the progression of cervical cancer via miR-5003-3p-dependent upregulation of OGN expression. *J Experimental Clin cancer Research: CR* 41. <https://doi.org/10.1186/s13046-022-02368-4>
 79. Zhou J, Wang L, Sun Q, Chen R, Zhang C, Yang P, Tan Y, Peng C, Wang T, Jin C et al (2021) Hsa_circ_0001666 suppresses the progression of colorectal cancer through the miR-576-5p/PCDH10 axis. *Clin Translational Med* 11:e565. <https://doi.org/10.1002/ctm2.565>
 80. Yang T, Sun J, Wang W, Li D, Yang X, Jia A, Ma Y, Fan Z (2022) Hsa_circ_0006732 regulates colorectal cancer cell proliferation, invasion and EMT by miR-127-5p/RAB3D axis. *Mol Cell Biochem* 477:2751–2760. <https://doi.org/10.1007/s11010-022-04458-5>
 81. Geng CH, Zhang XS, He M, Gao P, Zhao HW (2022) Circ_0000799 promotes proliferation and invasion in colorectal cancer and epithelial-mesenchymal transition. *J Gastrointest Oncol* 13:3090–3099. <https://doi.org/10.21037/jgo-22-1176>
 82. Luo N, Sulaiman Z, Wang C, Ding J, Chen Y, Liu B, Cheng Z, Liu S (2022) Hsa_circ_0000497 and hsa_circ_0000918 contributed to peritoneal metastasis of ovarian cancer via ascites. *J Translational Med* 20:201. <https://doi.org/10.1186/s12967-022-03404-9>
 83. Ma T, Guo J, Han J, Li L, Ren Y, Huang J, Diao G, Zheng X, Zheng Y (2023) Circ_0001589/miR-1248/HMGB1 axis enhances EMT-mediated metastasis and cisplatin resistance in cervical cancer. *Mol Carcinog* 62:1645–1658. <https://doi.org/10.1002/mc.23605>
 84. Liu J, Dai X, Guo X, Cheng A, Mac SM, Wang Z (2020) Circ-OXCT1 suppresses gastric Cancer EMT and metastasis by attenuating TGF- β pathway through the Circ-OXCT1/miR-136/SMAD4 Axis. *OncoTargets Therapy* 13:3987–3998. <https://doi.org/10.2147/ott.s239789>
 85. Chi X, Gu X, Chen S, Shen X (2022) Circ_0003221 downregulation restrains cervical Cancer Cell Growth, Metastasis and Angiogenesis by governing the miR-139-3p/S100A14 pathway. *Reproductive Sci (Thousand Oaks Calif)* 29:1822–1835. <https://doi.org/10.1007/s43032-021-00815-9>
 86. Zhang R, Zhu W, Ma C, Ai K (2021) Silencing of circRNA circ_0001666 represses EMT in pancreatic Cancer through upregulating miR-1251 and downregulating SOX4. *Front Mol Biosci* 8:684866. <https://doi.org/10.3389/fmolb.2021.684866>
 87. Li C, Li W, Cao S, Xu J, Qian Y, Pan X, Lei D, Wei D (2021) Circ_0058106 promotes proliferation, metastasis and EMT process by regulating Wnt2b/ β -catenin/c-Myc pathway through miR-185-3p in hypopharyngeal squamous cell carcinoma. *Cell Death Dis* 12. <https://doi.org/10.1038/s41419-021-04346-8>
 88. Li C, Cai J, Liu W, Gao Z, Li G (2023) Downregulation of circ-STK39 suppresses pancreatic cancer progression by sponging miR-140-3p and regulating TRAM2-mediated epithelial-mesenchymal transition. *Apoptosis: Int J Program cell Death* 28:1024–1034. <https://doi.org/10.1007/s10495-023-01813-9>
 89. Song J, Liu Q, Han L, Song T, Huang S, Zhang X, He Q, Liang C, Zhu S, Xiong B (2023) Hsa_circ_0009092/miR-665/NLK signaling axis suppresses colorectal cancer progression via recruiting TAMs in the tumor microenvironment. *J Experimental Clin cancer Research: CR* 42. <https://doi.org/10.1186/s13046-023-02887-8>
 90. Verduci L, Strano S, Yarden Y, Blandino G (2019) The circ RNA–micro RNA code: emerging implications for cancer diagnosis and treatment. *Mol Oncol* 13:669–680
 91. Memczak S, Papavasileiou P, Peters O, Rajewsky N (2015) Identification and characterization of circular RNAs as a New Class of putative biomarkers in human blood. *PLoS ONE* 10:e0141214. <https://doi.org/10.1371/journal.pone.0141214>
 92. Meng S, Zhou H, Feng Z, Xu Z, Tang Y, Li P, Wu M (2017) CircRNA: functions and properties of a novel potential biomarker for cancer. *Mol Cancer* 16:94. <https://doi.org/10.1186/s12943-017-0663-2>
 93. Li P, Chen S, Chen H, Mo X, Li T, Shao Y, Xiao B, Guo J (2015) Using circular RNA as a novel type of biomarker in the screening of gastric cancer. *Clin Chim Acta* 444:132–136. <https://doi.org/10.1016/j.cca.2015.02.018>
 94. Chen J, Li Y, Zheng Q, Bao C, He J, Chen B, Lyu D, Zheng B, Xu Y, Long Z et al (2017) Circular RNA profile identifies circPVT1 as a proliferative factor and prognostic marker in gastric cancer. *Cancer Lett* 388:208–219. <https://doi.org/10.1016/j.canlet.2016.12.006>
 95. Shang X, Li G, Liu H, Li T, Liu J, Zhao Q, Wang C (2016) Comprehensive circular RNA profiling reveals that hsa_circ_0005075, a New Circular RNA biomarker, is involved in Hepatocellular Crcinoma Development. *Medicine* 95:e3811. <https://doi.org/10.1097/md.0000000000003811>
 96. Qin M, Liu G, Huo X, Tao X, Sun X, Ge Z, Yang J, Fan J, Liu L, Qin W (2016) Hsa_circ_0001649: a circular RNA and potential novel biomarker for hepatocellular carcinoma. *Cancer Biomark A* 16:161–169. <https://doi.org/10.3233/cbm-150552>
 97. Yao JT, Zhao SH, Liu QP, Lv MQ, Zhou DX, Liao ZJ, Nan KJ (2017) Over-expression of CircRNA_100876 in non-small cell lung cancer and its prognostic value. *Pathol Res Pract* 213:453–456. <https://doi.org/10.1016/j.prp.2017.02.011>
 98. Wang L, Yang H, Lei Z, Zhao J, Chen Y, Chen P, Li C, Zeng Y, Liu Z, Liu X et al (2016) Repression of TIF1 γ by SOX2 promotes TGF- β -induced epithelial–mesenchymal transition in non-small-cell lung cancer. *Oncogene* 35:867–877. <https://doi.org/10.1038/onc.2015.141>
 99. Huang YY, Zhang CM, Dai YB, Lin JG, Lin N, Huang ZX, Xu TW (2021) USP11 facilitates colorectal cancer proliferation and metastasis by regulating IGF2BP3 stability. *Am J Translational Res* 13:480–496
 100. Huang GW, Chen QQ, Ma CC, Xie LH, Gu J (2021) linc01305 promotes metastasis and proliferation of esophageal squamous cell carcinoma through interacting with IGF2BP2 and IGF2BP3 to stabilize HTR3A mRNA. *Int J Biochem Cell Biol* 136. <https://doi.org/10.1016/j.biocel.2021.106015>
 101. Xu Y, Guo Z, Peng H, Guo L, Wang P (2022) IGF2BP3 promotes cell metastasis and is associated with poor patient survival in nasopharyngeal carcinoma. *J Cell Mol Med* 26:410–421. <https://doi.org/10.1111/jcmm.17093>
 102. Jiang W, Cheng X, Wang T, Song X, Zheng Y, Wang L (2020) LINC00467 promotes cell proliferation and metastasis by binding with IGF2BP3 to enhance the mRNA stability of TRAF5 in hepatocellular carcinoma. *J Gene Med* 22:e3134. <https://doi.org/10.1002/jgm.3134>
 103. Liu J, Jiang X, Zou A, Mai Z, Huang Z, Sun L, Zhao J (2021) circIGHG-Induced epithelial-to-mesenchymal transition promotes oral squamous cell carcinoma progression via miR-142-5p/IGF2BP3 signaling. *Cancer Res* 81:344–355. <https://doi.org/10.1158/0008-5472.Can-20-0554>
 104. Su N, Liu L, He S, Zeng L (2021) Circ_0001666 affects miR-620/WNK2 axis to inhibit breast cancer progression. *Genes Genomics* 43:947–959. <https://doi.org/10.1007/s13258-021-01114-y>
 105. Wang X, Li R, Feng L, Wang J, Qi Q, Wei W, Yu Z (2022) Hsa_circ_0001666 promotes non-small cell lung cancer migration

- and invasion through miR-1184/miR-548I/AGO1 axis. *Mol Therapy Oncolytics* 24:597–611. <https://doi.org/10.1016/j.omto.2022.02.011>
106. Qi Y, He J, Zhang Y, Wang L, Yu Y, Yao B, Tian Z (2021) Circular RNA hsa_circ_0001666 sponges miR-330-5p, miR-193a-5p and miR-326, and promotes papillary thyroid carcinoma progression via upregulation of ETV4. *Oncol Rep* 45. <https://doi.org/10.3892/or.2021.8001>
 107. Li T, Li Y, Zhang W, Zhang J (2021) Hsa_circ_0001017 inhibits proliferation and metastasis via regulating the let-7 g-3p/NDST3 axis in glioma. *Folia Neuropathol* 59:174–188. <https://doi.org/10.5114/fn.2021.107115>
 108. Liu R, Deng P, Zhang Y, Wang Y, Peng C (2021) Circ_0082182 promotes oncogenesis and metastasis of colorectal cancer in vitro and in vivo by sponging miR-411 and miR-1205 to activate the Wnt/ β -catenin pathway. *World J Surg Oncol* 19. <https://doi.org/10.1186/s12957-021-02164-y>
 109. Wu M, Sun T, Xing L (2020) Circ_0004913 inhibits cell growth, metastasis, and glycolysis by absorbing miR-184 to regulate HAMP in Hepatocellular Carcinoma. *Cancer Biother Radiopharm*. <https://doi.org/10.1089/cbr.2020.3779>
 110. Gao SL, Fan Y, Liu XD, Liu W, Zhao M (2022) circ_0089153 exacerbates breast cancer cells proliferation and metastasis via sponging miR-2467-3p/E2F6. *Environ Toxicol* 37:1458–1471. <https://doi.org/10.1002/tox.23498>
 111. Xing W, Zhou PC, Zhang HY, Chen LM, Zhou YM, Cui XF, Liu ZG (2022) Circular RNA circ_GLIS2 suppresses hepatocellular carcinoma growth and metastasis. *Liver International: Official J Int Association Study Liver* 42:682–695. <https://doi.org/10.1111/liv.15097>
 112. Yang T, Sun J, Wang W, Li D, Yang X, Jia A, Ma Y, Fan Z (2022) Hsa_circ_0006732 regulates colorectal cancer cell proliferation, invasion and EMT by miR-127-5p/RAB3D axis. *Mol Cell Biochem*. <https://doi.org/10.1007/s11010-022-04458-5>
 113. Zhang H, Zhou Q, Shen W (2022) Circ-FOXO1 promotes the proliferation, migration and EMT process of osteosarcoma cells through FOXO1-mediated wnt pathway activation. *J Orthop Surg Res* 17:344. <https://doi.org/10.1186/s13018-022-03207-0>
 114. Liang YX, Zhang LL, Yang L (2022) circANKRD17(hsa_circ_0007883) confers paclitaxel resistance of ovarian cancer via interacting with FUS to stabilize FOXR2. *Mol Cell Biochem*. <https://doi.org/10.1007/s11010-022-04548-4>
 115. Tan X, Song X, Fan B, Li M, Zhang A, Pei L (2022) Exosomal circRNA scm-like with four malignant brain tumor domains 2 (circ-SFMBT2) enhances the docetaxel resistance of prostate cancer via the microRNA-136-5p/tribbles homolog 1 pathway. *Anticancer Drugs* 33:871–882. <https://doi.org/10.1097/cad.0000000000001365>
 116. Hashemi M, Ghadyani F, Hasani S, Olyaei Y, Raei B, Khodadadi M, Ziyarani MF, Basti FA, Tavakolpournegari A, Matinmehdi AJJoDDS; et al. (2022) Nanoliposomes for doxorubicin delivery: reversing drug resistance, stimuli-responsive carriers and clinical translation. 104112
 117. Zhang J, Zhang J, Wang F, Xu X, Li X, Guan W, Men T, Xu G (2021) Overexpressed COL5A1 is correlated with tumor progression, paclitaxel resistance, and tumor-infiltrating immune cells in ovarian cancer. *J Cell Physiol* 236:6907–6919. <https://doi.org/10.1002/jcp.30350>
 118. Wu X, Qiu L, Feng H, Zhang H, Yu H, Du Y, Wu H, Zhu S, Ruan Y, Jiang H (2022) KHDRBS3 promotes paclitaxel resistance and induces glycolysis through modulated MIR17HG/CLDN6 signaling in epithelial ovarian cancer. *Life Sci* 293. <https://doi.org/10.1016/j.lfs.2022.120328>
 119. Mirzaei S, Zarrabi A, Asnaf SE, Hashemi F, Zabolian A, Hushmandi K, Raei M, Goharizi MASB, Makvandi P, Samarghandian SJ (2021) L.s. The role of microRNA-338-3p in cancer: growth, invasion, chemoresistance, and mediators. 268:119005
 120. Tian T, Han J, Huang J, Li S, Pang H (2021) Hypoxia-Induced Intracellular and Extracellular Heat shock protein gp96 increases Paclitaxel-Resistance and facilitates Immune Evasion in breast Cancer. *Front Oncol* 11:784777. <https://doi.org/10.3389/fonc.2021.784777>
 121. Yi H, Han Y, Li S (2022) Oncogenic circular RNA circ_0007534 contributes to paclitaxel resistance in endometrial cancer by sponging miR-625 and promoting ZEB2 expression. *Front Oncol* 12:985470. <https://doi.org/10.3389/fonc.2022.985470>
 122. Hong X, Liu N, Liang Y, He Q, Yang X, Lei Y, Zhang P, Zhao Y, He S, Wang Y et al (2020) Circular RNA CRIM1 functions as a ceRNA to promote nasopharyngeal carcinoma metastasis and docetaxel chemoresistance through upregulating FOXQ1. *Mol Cancer* 19. <https://doi.org/10.1186/s12943-020-01149-x>
 123. Shen Z, Zhou L, Zhang C, Xu J (2020) Reduction of circular RNA Foxo3 promotes prostate cancer progression and chemoresistance to docetaxel. *Cancer Lett* 468:88–101. <https://doi.org/10.1016/j.canlet.2019.10.006>
 124. Yu W, Jiang H, Zhang H, Li J (2018) Hsa_circ_0003998 promotes cell proliferation and invasion by targeting miR-326 in non-small cell lung cancer. *OncoTargets Therapy* 11:5569–5577. <https://doi.org/10.2147/ott.S174750>
 125. Li X, He J, Ren X, Zhao H, Zhao H (2020) Circ_0003998 enhances doxorubicin resistance in hepatocellular carcinoma by regulating miR-218-5p/EIF5A2 pathway. *Diagn Pathol* 15:141. <https://doi.org/10.1186/s13000-020-01056-1>
 126. Zhang J, Yu Y, Yin X, Feng L, Li Z, Liu X, Yu X, Li BA (2022) Circ-0007022/miR-338-3p/Neuropilin-1 Axis reduces the radiosensitivity of esophageal squamous cell carcinoma by activating Epithelial-To-Mesenchymal transition and PI3K/AKT pathway. *Front Genet* 13:854097. <https://doi.org/10.3389/fgene.2022.854097>
 127. Chipman LB, Pasquinelli A (2019) E.J.T.i.g. miRNA targeting: growing beyond the seed. 35, 215–222
 128. Liu Y, Chen T, Zheng G (2021) Exosome-transmitted circ-CARD6 facilitates posterior capsule opacification development by miR-31/FGF7 axis. *Exp Eye Res* 207:108572. <https://doi.org/10.1016/j.exer.2021.108572>
 129. Noman MZ, Parpal S, Van Moer K, Xiao M, Yu Y, Arakelian T, Viklund J, De Milito A, Hasmim M, Andersson M (2020) Inhibition of Vps34 reprograms cold into hot inflamed tumors and improves anti-PD-1/PD-L1 immunotherapy. *Sci Adv* 6:eaax7881
 130. Chen XP, Chen YG, Lan JY, Shen ZJ (2014) MicroRNA-370 suppresses proliferation and promotes endometrioid ovarian cancer chemosensitivity to cDDP by negatively regulating ENG. *Cancer Lett* 353:201–210. <https://doi.org/10.1016/j.canlet.2014.07.026>
 131. Wu Y, Zhou W, Yang Z, Li J, Jin Y (2022) Mir-185-5p represses cells growth and metastasis of Osteosarcoma via Targeting cathepsin E. *Int J Toxicol* 41:115–125. <https://doi.org/10.1177/10915818211069270>
 132. Zhu C, Huang L, Xu F, Li P, Li P, Hu F (2020) LncRNA PCAT6 promotes tumor progression in osteosarcoma via activation of TGF- β pathway by sponging miR-185-5p. *Biochem Biophys Res Commun* 521:463–470. <https://doi.org/10.1016/j.bbrc.2019.10.136>
 133. Xiao B, Zhang X, Li X, Zhao Z (2020) Circ_001569 regulates FLOT2 expression to promote the proliferation, migration, invasion and EMT of osteosarcoma cells through sponging miR-185-5p. *Open life Sci* 15:476–487. <https://doi.org/10.1515/biol-2020-0050>
 134. Zhou PL, Wu Z, Zhang W, Xu M, Ren J, Zhang Q, Sun Z, Han X (2021) Circular RNA hsa_circ_0000277 sequesters mir-4766-5p to upregulate LAMA1 and promote esophageal

- carcinoma progression. *Cell Death Dis* 12. <https://doi.org/10.1038/s41419-021-03911-5>
135. Sun D, Wang G, Xiao C, Xin Y (2021) Hsa_circ_001988 attenuates GC progression in vitro and in vivo via sponging miR-197-3p. *J Cell Physiol* 236:612–624. <https://doi.org/10.1002/jcp.29888>
 136. Han L, Cheng J, Li A, hsa_circ_0072387 (2021) Suppresses Proliferation, Metastasis, and glycolysis of oral squamous cell carcinoma cells by downregulating miR-503-5p. *Cancer Biother Radiopharm* 36:84–94. <https://doi.org/10.1089/cbr.2019.3371>
 137. Meng L, Zheng Y, Liu S, Ju Y, Ren S, Sang Y, Zhu Y, Gu L, Liu F, Zhao Y et al (2021) ZEB1 represses biogenesis of circ-DOCK5 to facilitate metastasis in esophageal squamous cell carcinoma via a positive feedback loop with TGF- β . *Cancer Lett* 519:117–129. <https://doi.org/10.1016/j.canlet.2021.06.026>
 138. Jin T, Liu M, Liu Y, Li Y, Xu Z, He H, Liu J, Zhang Y, Ke Y (2020) Lcn2-derived circular RNA (hsa_circ_0088732) inhibits cell apoptosis and promotes EMT in Glioma via the miR-661/RAB3D Axis. *Front Oncol* 10. <https://doi.org/10.3389/fonc.2020.00170>
 139. Feng W, Guo R, Zhang D, Zhang R (2021) Circ-ABC10 knock-down inhibits the malignant progression of cervical cancer through microRNA-128-3p/ZEB1 axis. *Biol Procedures Online* 23. <https://doi.org/10.1186/s12575-021-00154-8>
 140. Cai X, Nie J, Chen L, Yu F (2020) Circ_0000267 promotes gastric cancer progression via sponging MiR-503-5p and regulating HMGA2 expression. *Mol Genet Genom Med* 8:e1093. <https://doi.org/10.1002/mgg3.1093>
 141. Ji F, Lang C, Gao P, Sun H (2021) Knockdown of Circ_0000144 suppresses cell proliferation, Migration and Invasion in Gastric Cancer Via sponging MiR-217. *J Microbiol Biotechnol* 31:784–793. <https://doi.org/10.4014/jmb.2102.02005>
 142. Ma G, Li G, Fan W, Xu Y, Song S, Guo K, Liu Z (2021) Circ-0005105 activates COL11A1 by targeting miR-20a-3p to promote pancreatic ductal adenocarcinoma progression. *Cell Death Dis* 12:656. <https://doi.org/10.1038/s41419-021-03938-8>
 143. Wang Z, Deng M, Chen L, Wang W, Liu G, Liu D, Han Z, Zhou Y, Circular RNA (2020) Circ-03955 promotes epithelial-mesenchymal transition in Osteosarcoma by regulating miR-3662/Metadherin Pathway. *Front Oncol* 10. <https://doi.org/10.3389/fonc.2020.545460>
 144. Liang M, Huang G, Liu Z, Wang Q, Yu Z, Liu Z, Lin H, Li M, Zhou X, Zheng Y (2019) Elevated levels of hsa_circ_006100 in gastric cancer promote cell growth and metastasis via miR-195/GPRC5A signalling. *Cell Prolif* 52:e12661. <https://doi.org/10.1111/cpr.12661>
 145. Fan Y, Liu M, Liu A, Cui N, Chen Z, Yang Q, Su A (2021) Depletion of circular RNA circ_CORO1C suppresses gastric Cancer Development by modulating miR-138-5p/KLF12 Axis. *Cancer Manage Res* 13:3789–3801. <https://doi.org/10.2147/cmar.S290629>
 146. Lin Y, Jin X, Nie Q, Chen M, Guo W, Chen L, Li Y, Chen X, Zhang W, Chen H et al (2022) YTHDF3 facilitates triple-negative breast cancer progression and metastasis by stabilizing ZEB1 mRNA in an m(6)A-dependent manner. *Annals Translational Med* 10:83. <https://doi.org/10.21037/atm-21-6857>
 147. Jiang L, Chen Y, Min G, Wang J, Chen W, Wang H, Wang X, Yao N (2021) Bcl2-associated athanogene 4 promotes the invasion and metastasis of gastric cancer cells by activating the PI3K/AKT/NF- κ B/ZEB1 axis. *Cancer Lett* 520:409–421. <https://doi.org/10.1016/j.canlet.2021.08.020>
 148. López-Moncada F, Torres MJ, Lavanderos B, Cerda O, Castellón EA, Contreras HR (2022) SPARC induces E-Cadherin repression and enhances Cell Migration through Integrin α β 3 and the transcription factor ZEB1 in prostate Cancer cells. *Int J Mol Sci* 23. <https://doi.org/10.3390/ijms23115874>
 149. Wang L, Guo Z, Zhang S, Zhang X, Sang M, Liu Y, Shan B MIR-518a-5p Targets ZEB2 to Suppress the Migration and Invasion of Breast-cancer Cells. *Alternative therapies in health and medicine* 2022
 150. Lv J, Zhu S, Chen H, Xu Y, Su Q, Yu G, Ma W (2022) Paeonol inhibits human lung cancer cell viability and metastasis in vitro via miR-126-5p/ZEB2 axis. *Drug Dev Res* 83:432–446. <https://doi.org/10.1002/ddr.21873>
 151. Du Y, Liu X, Zhang S, Chen S, Guan X, Li Q, Chen X, Zhao Y (2021) CircCRIM1 promotes ovarian cancer progression by working as ceRNAs of CRIM1 and targeting miR-383-5p/ZEB2 axis. *Reproductive Biology Endocrinology: RB&E* 19:176. <https://doi.org/10.1186/s12958-021-00857-3>
 152. Xiang Y, Wang W, Gu J, Shang J, Circular (2022) RNA VANGL1 Facilitates Migration and Invasion of Papillary Thyroid Cancer by Modulating the miR-194/ZEB1/EMT Axis. *Journal of oncology* 2022, 4818651, <https://doi.org/10.1155/2022/4818651>
 153. Peng Z, Ouyang X, Wang Y, Fan Q (2022) MAPKAPK5-AS1 drives the progression of hepatocellular carcinoma via regulating miR-429/ZEB1 axis. *BMC Mol cell Biology* 23. <https://doi.org/10.1186/s12860-022-00420-x>
 154. Liu SW, Yang P, Li FN, Dou RG, Liu JX, Liu GJ (2022) LncRNA B4GALT1-AS1 promotes non-small cell lung cancer cell growth via increasing ZEB1 level by sponging miR-144-3p. *Translational cancer Res* 11:538–547. <https://doi.org/10.21037/tcr-22-296>
 155. Wu H, Dong D, Wang J, Yin S, Gong Y, Yang C, Bai Y, Wang J, Du Y (2022) LncRNA NEAT1 promotes the malignant progression of Colorectal Cancer by Targeting ZEB1 via miR-448. *Technol Cancer Res Treat* 21:15330338221085348. <https://doi.org/10.1177/15330338221085348>
 156. Ma C, Shi T, Qu Z, Zhang A, Wu Z, Zhao H, Zhao H, Chen H (2020) CircRNA_ACAP2 suppresses EMT in Head and Neck squamous cell carcinoma by targeting the miR-21-5p/STAT3 Signaling Axis. *Front Oncol* 10. <https://doi.org/10.3389/fonc.2020.583682>
 157. Zhou LH, Yang YC, Zhang RY, Wang P, Pang MH, Liang LQ (2018) CircRNA_0023642 promotes migration and invasion of gastric cancer cells by regulating EMT. *Eur Rev Med Pharmacol Sci* 22:2297–2303. https://doi.org/10.26355/eurev_201804_14818
 158. Hou B, Li W, Xia P, Zhao F, Liu Z, Zeng Q, Wang S, Chang D (2021) LHPP suppresses colorectal cancer cell migration and invasion in vitro and in vivo by inhibiting Smad3 phosphorylation in the TGF- β pathway. *Cell Death Discovery* 7:273. <https://doi.org/10.1038/s41420-021-00657-z>
 159. Liu F, Shi Z, Bao W, Zheng J, Chen K, Lin Z, Song HN, Luo X, Dong Q, Jiang L et al (2022) ZIC2 promotes colorectal cancer growth and metastasis through the TGF- β signaling pathway. *Exp Cell Res* 415:113118. <https://doi.org/10.1016/j.yexcr.2022.113118>
 160. Xu G, Chen Y, Fu M, Zang X, Cang M, Niu Y, Zhang W, Zhang Y, Mao Z, Shao M et al (2020) Circular RNA CCDC66 promotes gastric cancer progression by regulating c-Myc and TGF- β signaling pathways. *J Cancer* 11:2759–2768. <https://doi.org/10.7150/jca.37718>
 161. Zhou H, Wu J, Leng S, Hou C, Mo L, Xie X, Wang L, Xu Y (2021) Knockdown of circular RNA VANGL1 inhibits TGF- β -induced epithelial-mesenchymal transition in melanoma cells by sponging miR-150-5p. *J Cell Mol Med* 25:10837–10845. <https://doi.org/10.1111/jcmm.16887>
 162. Ou R, Mo L, Tang H, Leng S, Zhu H, Zhao L, Ren Y, Xu Y (2020) circRNA-AKT1 sequesters mir-942-5p to Upregulate AKT1 and promote cervical Cancer progression. *Mol Therapy Nucleic Acids* 20:308–322. <https://doi.org/10.1016/j.omtn.2020.01.003>
 163. Huang P, Hu Y, Duan Y (2022) TGF- β 2-induced circ-PRDM5 regulates migration, invasion, and EMT through the miR-92b-3p/

- COL1A2 pathway in human lens epithelial cells. *J Mol Histol* 53:309–320. <https://doi.org/10.1007/s10735-021-10053-7>
164. Takeda T, Tsubaki M, Matsuda T, Kimura A, Jinushi M, Obana T, Takegami M, Nishida S (2022) EGFR inhibition reverses epithelial–mesenchymal transition, and decreases tamoxifen resistance via snail and twist downregulation in breast cancer cells. *Oncol Rep* 47. <https://doi.org/10.3892/or.2022.8320>
 165. Yang F, Yuan WQ, Li J, Luo YQ (2021) Knockdown of METTL14 suppresses the malignant progression of non-small cell lung cancer by reducing twist expression. *Oncol Lett* 22:847. <https://doi.org/10.3892/ol.2021.13108>
 166. Tzeng HE, Tang CH, Tsai CH, Chiu CH, Wu MH, Yen Y (2021) ET-1 promotes epithelial-mesenchymal transition in oral squamous cell carcinoma cells via the microRNA-489-3p / TWIST Axis. *Oncotargets Therapy* 14:5005–5018. <https://doi.org/10.2147/ott.S294312>
 167. Tang G, Liu L, Xiao Z, Wen S, Chen L, Yang P (2021) CircRAB3IP upregulates twist family BHLH transcription factor (TWIST1) to promote osteosarcoma progression by sponging miR-580-3p. *Bioengineered* 12:3385–3397. <https://doi.org/10.1080/21655979.2021.1948487>
 168. Yu F, Lin Y, Ai MM, Tan GJ, Huang JL, Zou ZR (2021) Knockdown of circular RNA hsa_circ_PVT1 inhibited Laryngeal Cancer Progression via preventing wnt4/ β -Catenin signaling pathway activation. *Front cell Dev Biology* 9. <https://doi.org/10.3389/fcell.2021.658115>
 169. Cao L, Zhou X, Ding X, Gao D (2021) Knockdown of circ-PVT1 inhibits the progression of lung adenocarcinoma and enhances the sensitivity to cisplatin via the miR-429/FOXK1 signaling axis. *Mol Med Rep* 24. <https://doi.org/10.3892/mmr.2021.12323>
 170. Wang T, Huang Y (2021) Downregulation of hsa_circ_0001681 suppresses epithelial-mesenchymal transition in thyroid carcinoma via targeting to miR-942-5p/TWIST1 signaling pathway. *J Bioenerg Biomembr* 53:609–620. <https://doi.org/10.1007/s10863-021-09907-2>
 171. Meng J, Chen S, Han JX, Qian B, Wang XR, Zhong WL, Qin Y, Zhang H, Gao WF, Lei YY et al (2018) Twist1 regulates Vimentin through Cul2 circular RNA to promote EMT in Hepatocellular Carcinoma. *Cancer Res* 78:4150–4162. <https://doi.org/10.1158/0008-5472.Can-17-3009>
 172. Frosina G (2013) Development of therapeutics for high grade gliomas using orthotopic rodent models. *Curr Med Chem* 20:3272–3299. <https://doi.org/10.2174/0929867311320260011>
 173. Hansen TB, Wiklund ED, Bramsen JB, Villadsen SB, Statham AL, Clark SJ, Kjems J (2011) miRNA-dependent gene silencing involving Ago2-mediated cleavage of a circular antisense RNA. *EMBO J* 30:4414–4422. <https://doi.org/10.1038/emboj.2011.359>
 174. Mutalifu N, Du P, Zhang J, Akbar H, Yan B, Alimu S, Tong L, Luan X (2020) Circ_0000215 increases the expression of CXCR2 and promoted the Progression of Glioma Cells by sponging miR-495-3p. *Technol Cancer Res Treat* 19:1533033820957026. <https://doi.org/10.1177/1533033820957026>
 175. Wang C, Tang D, Wang H, Hu G, Hu S, Li L, Min B, Wang Y (2019) Circular RNA hsa_circ_0030018 acts as a sponge of miR-599 to aggravate esophageal carcinoma progression by regulating ENAH expression. *J Cell Biochem*. <https://doi.org/10.1002/jcb.29507>
 176. Pu B, Zhang X, Yan T, Li Y, Liu B, Jian Z, Mahgoub OK, Gu L, Xiong X, Zou N (2021) MICAL2 promotes Proliferation and Migration of Glioblastoma cells through TGF- β /p-Smad2/EMT-Like Signaling Pathway. *Front Oncol* 11. <https://doi.org/10.3389/fonc.2021.735180>
 177. Huang P, Guo Y, Zhao Z, Ning W, Wang H, Gu C, Zhang M, Qu Y, Zhang H, Song Y (2020) UBE2T promotes glioblastoma invasion and migration via stabilizing GRP78 and regulating EMT. *Aging* 12:10275–10289. <https://doi.org/10.18632/aging.103239>
 178. Lv F, Du Q, Li L, Xi X, Liu Q, Li W, Liu S (2021) Eriodictyol inhibits glioblastoma migration and invasion by reversing EMT via downregulation of the P38 MAPK/GSK-3 β /ZEB1 pathway. *Eur J Pharmacol* 900:174069. <https://doi.org/10.1016/j.ejphar.2021.174069>
 179. Chen WL, Jiang L, Wang JS, Liao CX (2019) Circ-0001801 contributes to cell proliferation, migration, invasion and epithelial to mesenchymal transition (EMT) in glioblastoma by regulating miR-628-5p/HMGB3 axis. *Eur Rev Med Pharmacol Sci* 23:10874–10885. https://doi.org/10.26355/eurrev_201912_19791
 180. Xin J, Zhang XY, Sun DK, Tian LQ, Xu P (2019) Up-regulated circular RNA hsa_circ_0067934 contributes to glioblastoma progression through activating PI3K-AKT pathway. *Eur Rev Med Pharmacol Sci* 23:3447–3454. https://doi.org/10.26355/eurrev_201904_17709
 181. Siegel RL, Miller KD, Jemal AJ (2019) *C.a.c.j.f.c.* Cancer statistics, 2019, 69, 7–34
 182. Yin GH, Gao FC, Tian J, Zhang WB (2019) Hsa_circ_101882 promotes migration and invasion of gastric cancer cells by regulating EMT. *J Clin Lab Anal* 33:e23002. <https://doi.org/10.1002/jcla.23002>
 183. Aoki T, Kinoshita J, Munesue S, Hamabe-Horiike T, Yamaguchi T, Nakamura Y, Okamoto K, Moriyama H, Nakamura K, Harada S et al (2022) Hypoxia-Induced CD36 expression in gastric Cancer cells promotes peritoneal metastasis via fatty acid uptake. *Ann Surg Oncol*. <https://doi.org/10.1245/s10434-022-12465-5>
 184. Yang H, Hu Y, Weng M, Liu X, Wan P, Hu Y, Ma M, Zhang Y, Xia H, Lv K (2022) Hypoxia inducible lncRNA-CBSLR modulates ferroptosis through m6A-YTHDF2-dependent modulation of CBS in gastric cancer. *J Adv Res* 37:91–106. <https://doi.org/10.1016/j.jare.2021.10.001>
 185. Li H, Cao B, Zhao R, Li T, Xu X, Cui H, Deng H, Gao J, Wei B (2022) circDNMT1 promotes malignant progression of gastric Cancer through Targeting miR-576-3p/Hypoxia Inducible Factor-1 alpha Axis. *Front Oncol* 12. <https://doi.org/10.3389/fonc.2022.817192>
 186. Tang J, Zhu H, Lin J, Wang H (2021) Knockdown of Circ_0081143 mitigates Hypoxia-Induced Migration, Invasion, and EMT in gastric Cancer cells through the miR-497-5p/EGFR Axis. *Cancer Biother Radiopharm* 36:333–346. <https://doi.org/10.1089/cbr.2019.3512>
 187. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D (2011) Global cancer statistics. *Cancer J Clin* 61:69–90. <https://doi.org/10.3322/caac.20107>
 188. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F (2015) Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer* 136:E359–386. <https://doi.org/10.1002/ijc.29210>
 189. Fitzmaurice C, Allen C, Barber RM, Barregard L, Bhutta ZA, Brenner H, Dicker DJ, Chimed-Orchir O, Dandona R, Dandona L et al (2017) Global, Regional, and National Cancer incidence, mortality, years of Life Lost, Years lived with disability, and disability-adjusted life-years for 32 Cancer groups, 1990 to 2015: a systematic analysis for the global burden of Disease Study. *JAMA Oncol* 3:524–548. <https://doi.org/10.1001/jamaoncol.2016.5688>
 190. Peng HX, Yang L, He BS, Pan YQ, Ying HQ, Sun HL, Lin K, Hu XX, Xu T, Wang S (2017) Combination of preoperative NLR, PLR and CEA could increase the diagnostic efficacy for I-III stage CRC. *31:e22075K.J.J.o.c.l.a*
 191. Brody HJS (2015) <https://doi.org/10.521S1a>. Colorectal cancer. *Nature* 521
 192. Yang Z, Zhang J, Lu D, Sun Y, Zhao X, Wang X, Zhou W, He Q, Jiang Z (2020) Hsa_circ_0137008 suppresses the malignant phenotype in colorectal cancer by acting as a microRNA-338-5p

- sponge. *Cancer Cell Int* 20. <https://doi.org/10.1186/s12935-020-1150-1>
193. Zhang B, Li Y, Wu Q, Xie L, Barwick B, Fu C, Li X, Wu D, Xia S, Chen J et al (2021) Acetylation of KLF5 maintains EMT and tumorigenicity to cause chemoresistant bone metastasis in prostate cancer. *Nat Commun* 12. <https://doi.org/10.1038/s41467-021-21976-w>
 194. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A (2015) Global cancer statistics, 2012. *Cancer J Clin* 65:87–108. <https://doi.org/10.3322/caac.21262>
 195. Franco EL, Schlecht NF, Saslow D (2003) The epidemiology of cervical cancer. *Cancer J (Sudbury Mass)* 9:348–359. <https://doi.org/10.1097/00130404-200309000-00004>
 196. Muñoz N (2000) Human papillomavirus and cancer: the epidemiological evidence. *J Clin Virology: Official Publication Pan Am Soc Clin Virol* 19:1–5. [https://doi.org/10.1016/s1386-6532\(00\)00125-6](https://doi.org/10.1016/s1386-6532(00)00125-6)
 197. Jiao J, Zhang T, Jiao X, Huang T, Zhao L, Ma D, Cui B (2020) hsa_circ_0000745 promotes cervical cancer by increasing cell proliferation, migration, and invasion. *J Cell Physiol* 235:1287–1295. <https://doi.org/10.1002/jcp.29045>
 198. Cen Y, Zhu T, Zhang Y, Zhao L, Zhu J, Wang L, Xu J, Ding T, Xie X, Wang X et al (2022) hsa_circ_0005358 suppresses cervical cancer metastasis by interacting with PTBP1 protein to destabilize CDCP1 mRNA. *Mol Therapy Nucleic Acids* 27:227–240. <https://doi.org/10.1016/j.omtn.2021.11.020>
 199. Wang J, Li H, Liang Z (2019) circ-MYBL2 serves as a sponge for mir-361-3p promoting Cervical Cancer cells Proliferation and Invasion. *OncoTargets Therapy* 12:9957–9964. <https://doi.org/10.2147/ott.S218976>
 200. Zhong P, Guo A, Wang L, Lin X, Feng M, Circular RNA CDK6 suppresses cervical cancer proliferation and metastasis by sponging miR-449a. *Bioengineered* 13:4885–4897. <https://doi.org/10.1080/21655979.2022.2036898>
 201. Siegel RL, Miller KD, Jemal A, *Cancer Statistics (2017) CA: a cancer journal for clinicians* 2017, 67, 7–30. <https://doi.org/10.3322/caac.21387>
 202. Hou W, Zhang Y (2021) Circ_0025033 promotes the progression of ovarian cancer by activating the expression of LSM4 via targeting miR-184. *Pathol Res Pract* 217:153275. <https://doi.org/10.1016/j.prp.2020.153275>
 203. Chen S, Wu W, Li QH, Xie BM, Shen F, Du YP, Zong ZH, Wang LL, Wei XQ, Zhao Y (2021) Circ-NOLC1 promotes epithelial ovarian cancer tumorigenesis and progression by binding ESRP1 and modulating CDK1 and RhoA expression. *Cell Death Discovery* 7. <https://doi.org/10.1038/s41420-020-00381-0>
 204. Wang X, Yao Y, Jin M (2020) Circ-0001068 is a novel biomarker for ovarian cancer and inducer of PD1 expression in T cells. *Aging* 12:19095–19106. <https://doi.org/10.18632/aging.103706>
 205. Ji J, Li C, Wang J, Wang L, Huang H, Li Y, Fang J (2022) Hsa_circ_0001756 promotes ovarian cancer progression through regulating IGF2BP2-mediated RAB5A expression and the EGFR/MAPK signaling pathway. *Cell Cycle (Georgetown Tex)* 21:685–696. <https://doi.org/10.1080/15384101.2021.2010166>
 206. Ma J, Jing X, Chen Z, Duan Z, Zhang Y (2018) MiR-361-5p decreases the tumorigenicity of epithelial ovarian cancer cells by targeting at RPL22L1 and c-Met signaling. *Int J Clin Exp Pathol* 11:2588–2596
 207. Zhang Y, Di Q, Chen J, Chang M, Ma Y, Yu J (2022) Circ_0061140 contributes to the malignant progression in Ovarian Cancer cells by mediating the RAB1A level through sponging miR-361-5p. *Biochem Genet*. <https://doi.org/10.1007/s10528-022-10200-z>
 208. Center MM, Jemal A, Lortet-Tieulent J, Ward E, Ferlay J, Brayley O, Bray F (2012) International variation in prostate cancer incidence and mortality rates. *Eur Urol* 61:1079–1092. <https://doi.org/10.1016/j.eururo.2012.02.054>
 209. James ND, Spears MR, Clarke NW, Dearnaley DP, De Bono JS, Gale J, Hetherington J, Hoskin PJ, Jones RJ, Laing R et al (2015) Survival with newly diagnosed metastatic prostate Cancer in the Docetaxel era: data from 917 patients in the control arm of the STAMPEDE trial (MRC PR08, CRUK/06/019). *Eur Urol* 67:1028–1038. <https://doi.org/10.1016/j.eururo.2014.09.032>
 210. Chowdhury S, Bjartell A, Lumen N, Maroto P, Paiss T, Gomez-Veiga F, Birtle A, Kramer G, Kalinka E, Spaeth D et al (2020) Real-world outcomes in First-Line treatment of metastatic castration-resistant prostate Cancer: the prostate Cancer Registry. *Target Oncol* 15:301–315. <https://doi.org/10.1007/s11523-020-00720-2>
 211. Kong M, Li H, Yuan W, Mao L, Chen J (2021) The role of Circ_PRKCI/miR-24-3p in the metastasis of prostate cancer. *J B U : Official J Balkan Union Oncol* 26:949–955
 212. Ren D, Yang Q, Dai Y, Guo W, Du H, Song L, Peng X (2017) Oncogenic mir-210-3p promotes prostate cancer cell EMT and bone metastasis via NF-κB signaling pathway. *Mol Cancer* 16:117. <https://doi.org/10.1186/s12943-017-0688-6>
 213. Mao S, Zhang W, Yang F, Guo Y, Wang H, Wu Y, Wang R, Maskey N, Zheng Z, Li C et al (2021) Hsa_circ_0004296 inhibits metastasis of prostate cancer by interacting with EIF4A3 to prevent nuclear export of ETS1 mRNA. *J Experimental Clin cancer Research: CR* 40. <https://doi.org/10.1186/s13046-021-02138-8>
 214. Luo GC, Chen L, Fang J, Yan ZJ (2021) Hsa_circ_0030586 promotes epithelial-mesenchymal transition in prostate cancer via PI3K-AKT signaling. *Bioengineered* 12:11089–11107. <https://doi.org/10.1080/21655979.2021.2008217>
 215. Chen H, Zhou L, Wu X, Li R, Wen J, Sha J, Wen X (2016) The PI3K/AKT pathway in the pathogenesis of prostate cancer. *Front Bioscience (Landmark Edition)* 21:1084–1091. <https://doi.org/10.2741/4443>
 216. Yi J, Zhu J, Wu J, Thompson CB, Jiang X (2020) Oncogenic activation of PI3K-AKT-mTOR signaling suppresses ferroptosis via SREBP-mediated lipogenesis. *Proc Natl Acad Sci USA* 117:31189–31197. <https://doi.org/10.1073/pnas.2017152117>
 217. Chen SS, Zhang YW, Wang YL, Gao Y, Wang C, Hu Q, Wu TG, Tao SC, Zhao JX, Chen M (2021) [Hsa_circ_0005221 promotes prostate cancer progression through the miR-339-5p/STAT5a pathway]. *Zhonghua Nan Ke Xue = Natl J Androl* 27:301–308
 218. Ren X, Cheng J, Zhu M, Chen X, Jiang M, Hu X, Lu Y (2022) Circular RNA circ_0062019 exerts oncogenic properties in prostate cancer via mediating miR-1253/NRBP1 axis. *Andrologia* 54:e14343. <https://doi.org/10.1111/and.14343>
 219. Antoni S, Ferlay J, Soerjomataram I, Znaor A, Jemal A, Bray F (2017) Bladder Cancer incidence and mortality: A global overview and recent trends. *Eur Urol* 71:96–108. <https://doi.org/10.1016/j.eururo.2016.06.010>
 220. Ploeg M, Aben KK, Kiemeny LA (2009) The present and future burden of urinary bladder cancer in the world. *World J Urol* 27:289–293. <https://doi.org/10.1007/s00345-009-0383-3>
 221. Zhang M, Du H, Wang L, Yue Y, Zhang P, Huang Z, Lv W, Ma J, Shao Q, Ma M et al (2020) Thymoquinone suppresses invasion and metastasis in bladder cancer cells by reversing EMT through the Wnt/β-catenin signaling pathway. *Chemico-Biol Interact* 320. <https://doi.org/10.1016/j.cbi.2020.109022>
 222. Zhang N, Hua X, Tu H, Li J, Zhang Z, Max C (2021) Isorhapontigenin (ISO) inhibits EMT through FOXO3A/METTL14/VIMENTIN pathway in bladder cancer cells. *Cancer Lett* 520:400–408. <https://doi.org/10.1016/j.canlet.2021.07.041>
 223. Tan S, Kang Y, Li H, He HQ, Zheng L, Wu SQ, Ai K, Zhang L, Xu R, Zhang XZ et al (2021) circST6GALNAC6 suppresses bladder cancer metastasis by sponging miR-200a-3p to modulate the STMN1/EMT axis. *Cell Death Dis* 12. <https://doi.org/10.1038/s41419-021-03459-4>
 224. Zhou L, Wang B, Zhang Y, Yao K, Liu B (2021) Silencing circ-BIRC6 inhibits the proliferation, invasion, migration and

- epithelial–mesenchymal transition of bladder cancer cells by targeting the miR-495-3p/XBP1 signaling axis. *Mol Med Rep* 24. <https://doi.org/10.3892/mmr.2021.12451>
225. Zha L, Zhang J, Tang W, Zhang N, He M, Guo Y, Wang Z (2013) HMGA2 elicits EMT by activating the Wnt/ β -catenin pathway in gastric cancer. *Dig Dis Sci* 58:724–733. <https://doi.org/10.1007/s10620-012-2399-6>
226. Qiu F, Liu Q, Xia Y, Jin H, Lin Y, Zhao X (2022) Circ_0000658 knockdown inhibits epithelial-mesenchymal transition in bladder cancer via miR-498-induced HMGA2 downregulation. *J Experimental Clin cancer Research: CR* 41:22. <https://doi.org/10.1186/s13046-021-02175-3>
227. Tong L, Yang H, Xiong W, Tang G, Zu X, Qi L (2021) circ_100984-miR-432-3p axis regulated c-Jun/YBX-1/ β -catenin feedback loop promotes bladder cancer progression. *Cancer Sci* 112:1429–1442. <https://doi.org/10.1111/cas.14774>
228. Li G, Guo BY, Wang HD, Lin GT, Lan TJ, Ying H, Xu J (2021) CircRNA hsa_circ_0014130 function as a miR-132-3p sponge for playing oncogenic roles in bladder cancer via upregulating KCNJ12 expression. *Cell Biol Toxicol*. <https://doi.org/10.1007/s10565-021-09668-z>
229. Chen L, Li H, Yao D, Zou Q, Yu W, Zhou L (2022) The novel circ_0084904/miR-802/MAL2 axis promotes the development of cervical cancer. *Reprod Biol* 22:100600. <https://doi.org/10.1016/j.repbio.2021.100600>
230. Meng L, Jia X, Yu W, Wang C, Chen J, Liu F, Circular (2020) RNA UBAP2 contributes to tumor growth and metastasis of cervical cancer via modulating miR-361-3p/SOX4 axis. *Cancer Cell Int* 20:357. <https://doi.org/10.1186/s12935-020-01436-z>
231. Ma N, Li X, Wei H, Zhang H, Zhang S, Circular RNA (2021) circNFATC3 acts as a miR-9-5p sponge to promote cervical cancer development by upregulating SDC2. *Cell Oncol (Dordrecht)* 44:93–107. <https://doi.org/10.1007/s13402-020-00555-z>
232. Tang KW, Guo ZX, Wu ZH, Zhou C, Sun J, Wang X, Song YX, Wang ZN (2021) Circ_0049447 acts as a tumor suppressor in gastric cancer through reducing proliferation, migration, invasion, and epithelial-mesenchymal transition. *Chin Med J* 134:1345–1355. <https://doi.org/10.1097/cm9.0000000000001494>
233. Peng YY, Sun D, Xin Y (2022) Hsa_circ_0005230 is up-regulated and promotes gastric cancer cell invasion and migration via regulating the miR-1299/RHOT1 axis. *Bioengineered* 13:5046–5063. <https://doi.org/10.1080/21655979.2022.2036514>
234. Li J, Zhen L, Zhang Y, Zhao L, Liu H, Cai D, Chen H, Yu J, Qi X, Li G (2017) Circ-104916 is downregulated in gastric cancer and suppresses migration and invasion of gastric cancer cells. *OncoTargets Therapy* 10:3521–3529. <https://doi.org/10.2147/ott.S136347>
235. Huang L, Zhu L, Pan S, Xu J, Xie M, Wang W, Xia G (2021) Circ_0029803 serves as the sponge of miR-216b-5p to promote the progression of colorectal cancer by regulating SKIL expression. *World J Surg Oncol* 19:268. <https://doi.org/10.1186/s12957-021-02368-2>
236. Xiao H, Liu M (2020) Circular RNA hsa_circ_0053277 promotes the development of colorectal cancer by upregulating matrix metalloproteinase 14 via miR-2467-3p sequestration. *J Cell Physiol* 235:2881–2890. <https://doi.org/10.1002/jcp.29193>
237. Fan L, Li W, Jiang H (2022) Circ_0000395 promoted CRC Progression via elevating MYH9 expression by sequestering miR-432-5p. *Biochem Genet*. <https://doi.org/10.1007/s10528-022-10245-0>
238. Zhang B, Cao W, Liu Y, Zhao Y, Liu C, Sun B (2022) Circ_0056618 enhances PRRG4 expression by competitively binding to miR-411-5p to promote the malignant progression of colorectal cancer. *Mol Cell Biochem*. <https://doi.org/10.1007/s11010-022-04525-x>
239. Yan D, Liu W, Liu Y, Zhu X (2022) Circular RNA circ_0065378 upregulates tumor suppressor candidate 1 by competitively binding with miR-4701-5p to alleviate colorectal cancer progression. *J Gastroenterol Hepatol* 37:1107–1118. <https://doi.org/10.1111/jgh.15862>
240. Shen D, Zhao H, Zeng P, Ge M, Shrestha S, Zhao W (2022) Circular RNA circ_0001459 accelerates hepatocellular carcinoma progression via the miR-6165/IGF1R axis. *Ann N Y Acad Sci* 1512:46–60. <https://doi.org/10.1111/nyas.14753>
241. Jin J, Liu H, Jin M, Li W, Xu H, Wei F (2020) Silencing of hsa_circ_0101145 reverses the epithelial-mesenchymal transition in hepatocellular carcinoma via regulation of the miR-548c-3p/LAMC2 axis. *Aging* 12:11623–11635. <https://doi.org/10.18632/aging.103324>
242. Zhang B, Zhou J (2022) CircSEC24A (hsa_circ_0003528) interference suppresses epithelial-mesenchymal transition of hepatocellular carcinoma cells via miR-421/MMP3 axis. *Bioengineered* 13:9049–9062. <https://doi.org/10.1080/21655979.2022.2057761>
243. Feng ZH, Zheng L, Yao T, Tao SY, Wei XA, Zheng ZY, Zheng BJ, Zhang XY, Huang B, Liu JH et al (2021) EIF4A3-induced circular RNA PRKAR1B promotes osteosarcoma progression by miR-361-3p-mediated induction of FZD4 expression. *Cell Death Dis* 12:1025. <https://doi.org/10.1038/s41419-021-04339-7>
244. Luo N, Liu S, Li X, Hu Y, Zhang K, Circular RNA (2021) circHIPK3 promotes breast cancer progression via sponging MiR-326. *Cell Cycle (Georgetown Tex)* 20:1320–1333. <https://doi.org/10.1080/15384101.2021.1939476>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Authors and Affiliations

Milad Ashrafizadeh^{1,2,3} · Jingyuan Dai⁴ · Pedram Torabian^{5,6} · Noushin Nabavi⁷ · Amir Reza Aref^{8,9} · Alaa A. A. Aljabali¹⁰ · Murtaza Tambuwala^{11,14}  · Minglin Zhu^{12,13}

✉ Jingyuan Dai
S571358@nwmissouri.edu

✉ Murtaza Tambuwala
mtambuwala@lincoln.ac.uk

✉ Minglin Zhu
zhuminglin@whu.edu.cn

¹ Department of Radiation Oncology, Shandong Provincial Key Laboratory of Radiation Oncology, Shandong Cancer Hospital and Institute, Shandong First Medical University, Shandong Academy of Medical Sciences, Jinan, Shandong 250000, China

² Department of General Surgery and Integrated Chinese and Western Medicine, Institute of Precision Diagnosis and Treatment of Gastrointestinal Tumors, Carson International Cancer Center, Shenzhen University General Hospital, Shenzhen University, Shenzhen, Guangdong 518060, China

³ Shanghai Institute of Cardiovascular Diseases, Zhongshan Hospital, Fudan University, Shanghai 200032, China

⁴ School of computer science and information systems, Northwest Missouri State University, Maryville, MO 64468, USA

⁵ Cumming School of Medicine, Arnie Charbonneau Cancer Research Institute, University of Calgary, Calgary, AB T2N 4Z6, Canada

⁶ Department of Medical Sciences, University of Calgary, Calgary, AB T2N 4Z6, Canada

⁷ Department of Urologic Sciences and Vancouver Prostate Centre, University of British Columbia, Vancouver, BC V6H3Z6, Canada

⁸ Belfer Center for Applied Cancer Science, Dana-Farber Cancer Institute, Harvard Medical School, Boston, MA, USA

⁹ Department of Translational Sciences, Xspera Biosciences Inc. Boston, Boston, MA, USA

¹⁰ Faculty of Pharmacy, Department of Pharmaceutics and Pharmaceutical Technology, Yarmouk University, Irbid, Jordan

¹¹ Lincoln Medical School, University of Lincoln, Brayford Pool Campus, Lincoln LN6 7TS, UK

¹² Department of Cardiovascular Surgery, Zhongnan Hospital of Wuhan University, Wuhan, Hubei, China

¹³ Hubei Provincial Engineering Research Center of Minimally Invasive Cardiovascular Surgery, Wuhan, Hubei 430071, China

¹⁴ College of Pharmacy, Ras Al Khaimah Medical and Health Sciences University, Ras Al Khaimah, United Arab Emirates