**Conclusion:** L'anesthésie au propofol est associée à une baisse de Vi alors que sous anesthésie à l'halothane Vi ne change pas. Les paramètres de l'effort et du synchronisme ventilatoires du groupe P différaient de ceux du groupe H.

Reports of apnea during propofol anaesthesia suggest that it may modify ventilatory control mechanisms.<sup>1,2</sup> In this study, an analysis was made of the inspiratory flow waveform during spontaneous ventilation and the pressure waveform during an occluded inspiration, to characterize the effect of propofol on ventilatory control in terms of (1) the Timing of the respiratory cycle, (2) the Amplitude of the respiratory neural drive and (3) the Shape of the intra-breath flow profiles in order to further characterize this apparent effect on Ventilatory Control.<sup>3,4</sup> Similar measurements were made in a second group of subjects during halothane anaesthesia.

## Methods

### Patient preparation

With Institutional ethics committee approval, informed parental consent was obtained for 20 children presenting for dental restoration under general anaesthesia. All patients were fasting, ASA grade I and were unpremedicated. Patients were randomised to receive either propofol (Group P) or halothane (Group H), for induction and maintenance of anaesthesia.

Preliminary trials showed that propofol alone did not provide the level of anaesthesia required for the dental procedure and the anaesthetic technique required a background of nitrous oxide and ketorolac (900  $\mu g \cdot k g^{-1}$ ). Anaesthesia was induced in Group P with propofol 2.5 mg · kg<sup>-1</sup>. Anaesthesia was induced in Group H patients with an inspired halothane concentration (FiH) of 2%. All patients received atropine (20  $\mu g \cdot k g^{-1}$ ) and vecuronium (100  $\mu g \cdot k g^{-1}$ ) iv. Naso-tracheal intubation was performed with an uncuffed Sheridan endotracheal tube (ETT) with a distal sideport for CO<sub>2</sub> sampling. A throat pack was inserted in all children. The leak around the ETT was detected at 15 cm H<sub>2</sub>O. The fresh gas flow of 70% N<sub>2</sub>O:O<sub>2</sub> was delivered through a Mapleson D breathing system with a fresh gas flow of 12 L · min<sup>-1</sup>. Muscle paralysis was reversed after 30 min with a neostigmine/atropine mixture (50  $\mu$ g  $\cdot$  kg<sup>-1</sup> and 20  $\mu$ g · kg<sup>-1</sup> respectively).

Anaesthesia was maintained with propofol or halothane using a washout of anaesthetic agent. Flow ( $\dot{V}$ ), airway pressure (Pao) and end tidal carbon dioxide concentration (PETCO<sub>2</sub>) were recorded at three concentrations of halothane (FiH): 2, 1 and 0%; and five propofol infusion rates (RivP): 18, 15, 12, 9 and 0

 $mg \cdot kg^{-1} \cdot hr^{-1}$ . Data were collected from spontaneously breathing patients after 30 min at each set FIH or RivP. Although 30 min may not represent a true pharmacological steady state, we considered it a reasonable compromise between the time constraints imposed by pharmacological and ethical considerations. (Ventilation was assisted between periods of data collection.)

At the end of the procedure, the anaesthetic agents were discontinued and data were recorded continuously until arousal (spontaneous movement) at which point the behavioral response necessitated tracheal extubation. The FETH corresponding to FiH 0%, 1%, and 2% were  $0.191 \pm 0.01\%$ ,  $0.857 \pm 0.03\%$  and  $1.38 \pm 0.06\%$ . Preemergence occlusion data in Group H were collected at FETH 0.3%. The FiH 0% and RivP 0 mg  $\cdot$  kg<sup>-1</sup>  $\cdot$  hr<sup>-1</sup> values were collected in the last 30 sec as the preemergence values in unstimulated patients. Preemergence occlusion data in Group P was collected 10 min after stopping the RivP. Since the children were unresponsive at the time of airway occlusion, a residual effect of propofol was probably present.

In each patient, body temperature (rectal) was maintained within  $0.5^{\circ}$ C of the post induction value.

#### Signal measurement

As previously reported<sup>4,5</sup> airway pressure (Pao) was measured through a sideport in the ETT connector using a piezoresistive differential pressure transducer (Microswitch #162PC01D, Honeywell, Scarborough, Ontario) referenced to the atmosphere. Flow (V) was measured by a heated Fleisch pneumotachograph (#0). The pressure drop across the pneumotachograph was measured with another peizoresistive differential pressure transducer (Microswitch #163PCO1036). The dead space of this pneumotachograph with connector was 15 ml. Calibration of this device was performed using the flow rotameters. Volume calibration was performed with a precision 100 ml syringe and the gas mixture used for the study. Pressure was calibrated with a pressure manometer. The stability of the calibration with time was verified for each patient.

Halothane and PETCO<sub>2</sub> were measured with a Nellcor N-2500 multi-function pulse oximeter with agent analyzer (Nellcor Inc., Hayward, CA). The accuracy of the  $CO_2$  analyzer was verified with a known concentration of  $CO_2$ . The accuracy of the halothane analyzer was checked periodically with a known concentration of halothane.

A two-way valve with an inflatable balloon in the inspiratory limb (Hans Rudolph #2384B) was positioned between the pneumotachograph and the fresh gas flow inlet to allow interruption of the circuit during expiration, so that the next inspiration could be occluded. Even with rapid respiratory rates, the airway could be reliably occluded at end-expiration.

## Data analysis

A detailed description of the data analysis is provided in previous publications.<sup>4,5</sup> The beginning and end of inspiration and expiration were identified from the zero crossing points of the signal. Volume ( $\dot{V}$ ) was obtained by numerical integration of  $\dot{V}$ .

From the V signal we determined tidal volume (VT), minute ventilation ( $\dot{V}i = 60 \cdot VT \cdot Ttot^{-1}$ ) and the parameters of breath Amplitude: mean inspiratory flow (VT/Ti); breath Timing: total cycle time (Ttot) and inspiratory time (Ti); and breath Shape: the inspiratory flow centroid (Ci/Ti) and the inspiratory duty cycle (Ti/Ttot). The mathematical equation used to calculate Ci/Ti is given in reference 4 and reflects the skewness of the flow waveform. Finally, end tidal CO<sub>2</sub> (PETCO<sub>2</sub>) as obtained as the highest value of the smoothed expiratory PCO<sub>2</sub> signal from each breath.

During expiration the airway was occluded and the occluded pressure waveform of the occluded inspiration was recorded. The breath Amplitude parameter  $dP/dt_{0.1}$ , a correlate of  $P_{0.1}$ , was obtained from the occluded Pao waveforms as the slope of the line fit (by linear regression) to the first 100 msec of the occluded inspiration.<sup>5</sup> The coefficient of determination for the regressions was >0.97. Values for dP/dt<sub>0.1</sub> at all FtH and RivP were obtained from at least three occluded pressure waveforms.

Finally, the ratio between the occluded and unoccluded inspiratory times ( $Ti^{occ}/Ti$ ) was determined, where  $Ti^{occ}$  is the occluded inspiratory time defined as the time taken for Pao to decrease to its minimum value from baseline.

# Statistical analysis

Group data are presented as box plots. Intra and intergroup sample means were assessed with ANOVA for repeated measures.<sup>6</sup> Mean parameter values obtained at FiH 1% and 2% were compared with the pre-emergence (FiH 0%) values by Dunnett t test for repeated measures.<sup>7</sup> Mean parameter values obtained at RivP 18, 15, 12 and 9 mg·kg<sup>-1</sup>·min<sup>-1</sup> were compared with the preemergence (RivP 0 mg·kg<sup>-1</sup>·min<sup>-1</sup>) values by Dunnett t test for repeated measures.<sup>7</sup> Intergroup differences for the preemergence values were assessed with an unpaired t test. A *P* value of 0.05 was considered statistically significant.

### Results

We studied 10 patients in Group P (aged  $4.5 \pm 0.4$  yr, weighing  $17.5 \pm 1.2$  kg and whose body surface area



FIGURE 1 Effect of washout of halothane on the ventilatory parameters Vi, PETCO<sub>2</sub>, VT and Ttot. Data are presented as open box plots (showing 10th and 90th percentiles, central mean and the 5th and 95th error bars and outliers.) Vi was unchanged in Group H. PETCO<sub>2</sub> increased during halothane anaesthesia whereas VT and Ttot decreased. Intragroup differences from FiH 0% are shown (\*P < 0.05).

(BSA) was  $0.707 \pm 0.03$  m<sup>2</sup>) and 10 patients in Group H (aged 4.0 ± 0.3 yr, weighing 17.1 ± 1.4 kg, whose BSA was  $0.686 \pm 0.04$  m<sup>2</sup>). Body temperature in Group P was  $35.8 \pm 0.2^{\circ}$ C and in Group H it was  $35.6 \pm 0.2^{\circ}$ C. Three patients in Group P had nonpurposeful movement to surgical stimulus at RivP 9 mg·kg<sup>-1</sup>·r<sup>-1</sup>. In one, this necessitated an increase of RivP to 12 mg·kg<sup>-1</sup>·hr<sup>-1</sup> and representative data at RivP 9 mg·kg<sup>-1</sup>·hr<sup>-1</sup> could not be obtained. Satisfactory occluded inspiratory pressure waveforms were obtained for ten patients in Group P and nine patients in Group H.

There were no differences between groups in age, weight, BSA or body temperature. There were no differences between groups in the pre-emergence values of Vi, VT, VT/Ti or PETCO<sub>2</sub> (Figures 1, 2 and Table I) nor were there differences in the Shape parameters Ti/Ttot or Ci/Ti (Table II). There were no differences in the pre-emergence values for Respiratory Mechanics [PMAX/VT and (VT/Ti)/dP/dt<sub>0.1</sub>].

Figures 1 and 2 summarize the pattern of breathing during a washout of anaesthetic in Group P and Group H. The Vi, VT, Ttot increased during a washout of propofol and PETCO<sub>2</sub> decreased. In contrast the Vi did not change in Group H during a washout of halothane. The PETCO<sub>2</sub> however did increase during halothane anaesthesia, in fact since the mean Te in Group H was less (0.71 sec) than in Group P (1.12 sec), the expiratory



FIGURE 2 Effect of washout of propofol (hatched box plots) on the ventilatory parameters  $\dot{V}_1$ , PETCO<sub>2</sub>, VT and Ttot. The scales of the Y-axis are identical for ease of comparison with Figure 1. The  $\dot{V}_1$ VT and Ttot increased in a dose response fashion during the washout of propofol, from a RivP of 18 mg  $\cdot$  kg<sup>-1</sup> · hr<sup>-1</sup> to 0 mg  $\cdot$  kg<sup>-1</sup> · hr<sup>-1</sup>, whereas PETCO<sub>2</sub> decreased. Intragroup differences from RivP 0 mg  $\cdot$  kg<sup>-1</sup> · hr<sup>-1</sup> are shown (\**P* < 0.05).

plateau on the  $CO_2$  waveform may have underestimated the arterial  $CO_2$  in Group H. The pattern of breathing in Group P was rapid and shallow as both VT and Ttot decreased in both groups.

At high values of F1H and RivP the Drive parameter VT/Ti decreased (Table II). However the Drive parameter dP/dt<sub>0.1</sub> did not change in either group. The preemergence dP/dt<sub>0.1</sub> was lower in Group P, indeed dP/dt<sub>0.1</sub> in Group H was roughly twice that of Group P at all RivP (Figure 3) (P < 0.05). The Shape parameters Ti/Ttot and Ci/Ti showed inconsistent changes during the washout of anaesthetic agents (Table II). There were group differences in the pre-emergence parameters of Timing (Ttot, Ti and Ti<sup>occ</sup>/Ti) with Group H having lower values. Whereas Ti<sup>occ</sup>/Ti decreased during halothane anaesthesia, it did not change during propofol anaesthesia (Figure 4). In Group P PMAX/VT and (VT/Ti)/dP/dt<sub>0.1</sub> did not change during propofol administration, whereas in Group H (VT/Ti)/dP/dt<sub>0.1</sub> decreased.

### Discussion

Although the procedure required the addition of  $N_2O$ and ketorolac to the anaesthetic, we made the assumption that the major effect on ventilation during the period of study was due to the anaesthetic agent – propofol or halothane. Minute ventilation did not change during halothane anaesthesia and this is in agreement with previous work<sup>5</sup> which showed that  $\dot{V}i$  was maintained at

TABLE I Mean values (X ± SEM) for the parameters presented in Figures 1 and 2. Intragroup differences from FtH 0% or RivP 0 mg·kg<sup>-1</sup>·hr<sup>-1</sup> are shown \*(P < 0.05). Intergroup differences between FtH 0% and RivP 0 mg·kg<sup>-1</sup>·hr<sup>-1</sup> are shown †(P < 0.05).

$RivP$ $(mg \cdot kg^{-l} \cdot hr^{-l})$ $(n = 10)$	Vi (L · min <sup>-1</sup> · m <sup>-2</sup> )	VT (ml·kg⁻¹)	Ttot (s)	Ti (s)	PETCO2 (mmHg)
18	2.4*	3.6*	2.18*	0.97*	54.9*
	(0.2)	(0.3)	(0.12)	(0.03)	(2.7)
15	2.5*	4.2*	2.39*	1.09	52.6
	(0.2)	(0.4)	(0.13)	(0.06)	(2.8)
12	2.8*	4.7*	2.45*	1.09	49.6
	(0.2)	(0.4)	(0.12)	(0.05)	(2.4)
9	2.8*	4.8*	2.5*	1.13	49.7
	(0.2)	(0.4)	(0.08)	(0.05)	(1.6)
0	3.2	6.2	2.82	1.16	44.8
	(0.2)	(0.4)	(0.09)	(0.03)	(0.7)
$F_{iH}(\%)(n = 1)$	0)				
2	2.8	2.5*	1.28*	0.57*	58.0*
	(0.4)	(0.4)	(0.05)	(0.02)	(2.1)
1	3.7	4.0*	1.63*	0.70*	46.7*
	(0.3)	(0.3)	(0.08)	(0.03)	(1.9)
0	3.6	5.5	2.24†	0.94†	42.7
	(0.2)	(0.3)	(0.08)	(0.04)	(0.9)

TABLE II Mean values (X  $\pm$  SEM) for VT/Ti, Ti/Ttot, Ci/Ti, PMAX/VT and (VT/Ti)/dP/dt<sub>0.1</sub>. Intragroup differences from FiH 0% or RivP 0 mg  $\cdot$  kg<sup>-1</sup> · hr<sup>-1</sup> are shown (\**P* < 0.05).

RivP (mg·kg <sup>→</sup> ·hr <sup>→</sup> )	VT/Ti ml·m <sup>−2</sup> ·sec <sup>−1</sup>	Ti/Ttot	Ci/Ti	Рмах/Vт cm · ml <sup>⊣</sup>	(VT/Ti)/ dP/dt <sub>0.1</sub> ml · cm <sup>-1</sup>
18	88.2*	0.463*	0.485	0.174	3.75
	(8.3)	(0.01)	(0.01)	(0.2)	(0.36)
15	92.8	0.456	0.503	0.183	4.83
	(8.7)	(0.01)	(0.01)	(0.02)	(1.02)
12	107	0.445	0.497	0.164	4.52
	(8.9)	(0.01)	(0.02)	(0.02)	(0.68)
9	104	0.451	0.507	0.164	4.64
	(7.4)	(0.01)	(0.02)	(0.02)	(0.51)
0	132	0.413	0.518	0.175	4.73
	(8.7)	(0.01)	(0.01)	(0.02)	(1.01)
F1H (%)					
2	106*	0.448*	0.468*	0.215	1.85*
	(14.8)	(0.01)	(0.01)	(0.03)	(0.25)
1	144	0.432	0.47*	0.189	2.05*
	(13.6)	(0.01)	(0.01)	(0.03)	(0.26)
0	152	0.418	0.519	0.19	2.61
	(11)	(0.02)	(0.02)	(0.03)	(0.36)

FiH 2% in oxygen in a study of similarly aged children. In contrast, the decrease in  $\dot{V}i$  with propofol anaesthesia occurred at an low RivP 9 mg·kg<sup>-1</sup>·hr<sup>-1</sup> even though this was below anaesthetic levels in two patients. Therefore at 2 MAC halothane  $\dot{V}i$  was maintained,





FIGURE 3 Values of  $dP/dt_{0.1}$  for Group H (Open box plots) and Group P (Hatched Box plots). The value for  $dP/dt_{0.1}$  did not change with either anaesthetic. In Group H the value was higher than in Group P (P < 0.05).



FIGURE 4 Ratios Ti<sup>occ</sup>/Ti for Group H (Open box plots) and Group P (Hatched box plots). The pre-emergence value for Group H was less than Group P. During halothane anaesthesia it decreased whereas in Group P it did not change (P < 0.05). See text for discussion.

whereas even at light levels of propofol anaesthesia depression in Vi occurred.

The PETCO<sub>2</sub> increased in both groups supporting the notion of respiratory depression during both halothane and propofol anaesthesia. Both agents have been shown to depress the ventilatory response to carbon dioxide.<sup>8,9</sup> In Group H the rapid and shallow pattern of breathing probably contributed to an increase in dead space ventilation predisposing to the increase in PETCO<sub>2</sub> even though Vi did not change. The pattern of ventilation in Group P seemed less rapid and shallow although comparison at equivalent anaesthetic depth was problematic. Unlike halothane anaesthesia, propofol was associated with a dose response decrease in Vi which may also have contributed to the dose response increase in PETCO<sub>2</sub>. As a parameter of breath Drive,  $dP/dt_{0,1}$  has an advantage over VT/Ti because measured during airway occlusion, it is minimally affected by the mechanics of the respiratory system.<sup>10</sup> Although the VT/Ti decreased during anaesthesia in both groups, the dP/dt<sub>01</sub> did not change. For Group H this finding agrees with a previous investigation in children of a similar age.<sup>5</sup> A striking finding was the intergroup difference in the value of  $dP/dt_{0.1}$ , with values in Group H being roughly twice that of Group P for all RivP and FiH. Does this represent a difference in the effect of halothane and propofol on ventilatory drive?

An inherent flaw in the study design which makes interpretation of these results difficult is that control values for dP/dt<sub>0.1</sub> were not available. In Group H, preemergence values were recorded at FETH 0.3% and in Group P there was an unknown concentration of propofol. Therefore preemergence values for dP/dt<sub>0.1</sub> in both groups did not represent unanaesthetized values. The expected value for dP/dt<sub>0.1</sub> may well be intermediate between the preemergence value of Group H, with halothane tending to increase dP/dt<sub>0.1</sub>, and propofol tending to depress it. Certainly Canet suggested that halothane increased P<sub>0.1</sub>, a dP/dt<sub>0.1</sub> correlate, from awake values.<sup>11</sup>

Interpretation of  $dP/dt_{0.1}$  as a drive parameter is complicated by the potential effect of lung volume on the shape of the diaphragm and its force-length characteristics.<sup>10</sup> A decrease in functional residual capacity (FRC), as occurs during anaesthesia,<sup>12</sup> might enhance the force generating characteristics of the diaphragm. If FRC decreased to a greater extent during halothane anaesthesia then this might account for the difference in  $dP/dt_{0.1}$ . Although tracheal intubation was facilitated with nondepolarizing muscle relaxants which were subsequently antagonized, no FRC restoring measures were taken. Therefore, the decrease in FRC should have been similar in both groups, indeed inductive plethysmography has been used in adults to demonstrate a fall in FRC with induction of propofol anaesthesia.<sup>2</sup> Therefore, *a priori*, a difference in lung volume should not account for the difference in  $dP/dt_{0,1}$ .

The ratio (VT/Ti)/dP/dt has been used as an index of the efficiency of the respiratory pump in generating flow. It has been shown to decrease during methoxyflurane<sup>13</sup> and halothane anaesthesia<sup>6</sup> and may indicate a mechanical impairment of the respiratory pump. The fact that it did not change in Group P suggests that respiratory efficiency may be maintained during propofol anaesthesia.

The ratio Ti<sup>occ</sup>/Ti is an index of the Hering Breuer reflex (HBR) assessing the influence of lung inflation on inspiratory duration.<sup>14,15</sup> The ratio Ti<sup>occ</sup>/Ti decreased during halothane anaesthesia in Group H which is consistent with studies in halothane anaesthetized adults.<sup>12,16,17</sup> The decrease in Ti<sup>occ</sup>/Ti suggests a central mechanism for the tachypnea associated with halothane anaesthesia, a notion which is consistent with experimental data.<sup>18,19</sup> In Group P the fact that the ratio Ti<sup>occ</sup>/Ti did not change suggests that during propofol anaesthesia pulmonary input remained an important determinant of the duration of inspiration.

The Shape parameters in Group H changed in that Ti/Ttot decreased during a washout of halothane and Ci/Ti increased. In a previous report of similarly aged children anaesthetized with halothane in air:oxygen we showed no change in the Shape parameters Ci/Ti and Ti/Ttot.<sup>5</sup> Similarly Jonson<sup>20</sup> showed that the Shape of the flow waveform was not changed with halothane administration. It may be that N<sub>2</sub>O can produce an effect on the Shape parameters since the Ti/Ttot also decreased during the washout of propofol and Ci/Ti tended to increase although this trend did not reach statistical significance.

In summary, PETCO<sub>2</sub> increased during anaesthesia in both groups. We speculate that the increase in Group H was due to the more rapid shallow pattern of breathing, whereas in Group P it was associated with the decrease in Vi. There was a twofold difference in the breath drive parameter dP/dt<sub>0.1</sub> between Group H and Group P which suggests that ventilatory drive was lower in the children anaesthetized with propofol compared with those anaesthetized with halothane, although the potential effect of lung volume on dP/dt<sub>0.1</sub> cannot be excluded. The decrease in the Timing parameter Ti<sup>occ</sup>/Ti during halothane anaesthesia and the lack of change in Ti<sup>occ</sup>/Ti suggests that the duration of inspiration during halothane anaesthesia was primarily determined by central events whereas during propofol anaesthesia pulmonary input remained an important determinant of inspiratory duration.

# Acknowledgement

We thank Ms. Roula Cacolyris for help with the preparation of the manuscript. Gratitude is also expressed to the Montreal Children's Hospital departments of Dentistry and Nursing for cooperating with the study.

#### References

- 1 Webster AC, Reid WD, Siebert LF, Taylor MD. Laryngeal mask airway for anaesthesia for cryopexy in low birth weight infants. Can J Anaesth 1995; 41: 361–2.
- 2 Grounds RM, Maxwell DL, Taylor MB, Aber V, Royston D. Acute ventilatory changes during i.v. induction of anaesthesia with thiopentone or propofol in man. Br J Anaesth 1987; 59: 1098–102.
- 3 Feldman JL, Smith JC, Ellenberger HH, et al. Neurogenesis of respiratory rhythm and pattern: emerging concepts. Am J Physiol 1990; 259: R879–86.
- 4 Reich O, Brown K, Bates JH. Breathing patterns in infants and children under halothane anesthesia: effect of dose and CO<sub>2</sub>. J Appl Physiol 1994; 76: 79-85.
- 5 Brown KA, Reich O, Bates JH. Ventilatory depression by halothane in infants and children. Can J Anaesth 1995; 42: 588–96.
- 6 Bourke GF, Daly LE, McGilvray J. Interpretation and Uses of Medical Statistics, 3rd ed. Oxford: Blackwell Scientific Publications, 1985.
- 7 Dunnett CW. A multiple comparison procedure for comparing several treatments with a control. Journal of American Statistical Association 1955; 50: 1096–121.
- 8 Knill RL, Gelb AW. Ventilatory responses to hypoxia and hypercapnia during halothane sedation and anesthesia in man. Anesthesiology 1978; 49: 244–51.
- 9 Blouin RT, Seifert HA, Babenco HD, Conard PF, Gross JB. Propofol depresses the hypoxic ventilatory response during conscious sedation and isohypercapnia. Anesthesiology 1993; 79: 1177-82.
- Cherniack NS, Lederer DH, Altose MD, Kelsen SG.
   Occlusion pressure as a technique in evaluating respiratory control. Chest 1976; 70: 137–41.
- 11 Canet J, Sanchis J, Zegrí A, Liorente C, Navajas D, Casan P. Effects of halothane and isoflurane on ventilation and occlusion pressure. Anesthesiology 1994; 81: 563-71.
- 12 Dobbinson TL, Nisbet HIA, Pelton DA, Levison H. Functional residual capacity (FRC) and compliance in anaesthetized paralysed children. Part II. Clinical results. Can Anaesth Soc J 1973; 20: 322–4.
- 13 Derenne JP, Couture J, Iscoe S, Whitelaw A, Milic-Emili J. Occlusion pressures in men rebreathing CO<sub>2</sub> under methoxyflurane anesthesia. J Appl Physiol 1976; 40: 805–14.
- 14 Von Euler C. On the central pattern generator for the basic breathing rhythmicity. J Appl Physiol 1983; 55: 1647–59.
- 15 Rabbette PS, Fletcher ME, Dezateux CA, Soriano-Brucher

Kulkarni and Brown: PROPOFOL AND VENTILATION

*H*, *Stocks J*. Hering-Breuer reflex and respiratory system compliance in the first year of life: a longitudinal study. J Appl Physiol 1994; 76: 650–6.

- 16 Kochi T, Izumi Y, Isono S, Ide T, Mizuguchi T. Breathing pattern and occlusion pressure waveform in humans anesthetized with halothane or sevoflurane. Anesth Analg 1991; 73: 327–32.
- 17 Drummond GB. Effect of airway occlusion on respiratory timing during anaesthesia with enflurane or halothane. Br J Anaesth 1984; 56: 215-21.
- 18 Gautier H, Bonora M, Zaoui D. Influence of halothane on control of breathing in intact and decerebrated cats. J A ppl Physiol 1987; 63: 546–53.
- 19 Berkenbosch A, deGoede J, Olievier CN, Quanjer H. Sites of action of halothane on respiratory pattern and ventilatory response to CO<sub>2</sub> in Cats. Anesthesiology 1982; 57: 389–98.
- 20 Jonsson LO, Zetterstrom H. Flow pattern and respiratory characteristics during halothane anaesthesia. Acta Anaesthesiol Scand 1985; 29: 309–14.