

Interaction between succinylcholine and cimetidine in rats

Yogendra Mishra M Pharm, Iqbal Ramzan PhD

The hypothesis that histamine H₂ receptor blockade adversely affects neuromuscular function was tested, in vivo, in rats anaesthetised with urethane during mechanical pulmonary ventilation. Succinylcholine was administered as a bolus and constant-rate infusion to maintain 49.2% (± 1.5 SEM) twitch suppression in 19 rats. Cimetidine iv, 3.2, 7.5, 10, 17.8, 23.7, 31.6, or 56.2 mg · kg⁻¹ was then administered in groups of two to three rats. Cimetidine produced an immediate potentiation of twitch suppression followed by a transient reversal and then a continued potentiation. Peak potentiation occurred within 19.0 (± 2.7) sec and was maintained in 11 rats at steady-state. Reversal was evident 4.1 (± 0.4) min after cimetidine administration. There was a good relationship between peak potentiation and serum cimetidine concentration with 50% potentiation occurring at 46.5 (± 4.6) $\mu\text{g} \cdot \text{ml}^{-1}$. Potentiation at steady-state was not correlated to serum cimetidine concentration but there was a weak relationship between reversal and serum cimetidine concentration. These results support reports from patients of an interaction between cimetidine and succinylcholine.

L'hypothèse voulant que le blocage de récepteurs histaminiques de type H₂ affecte la fonction neuromusculaire a été vérifiée in vivo sur des rats anesthésiés avec de l'uréthane et ventilés artificiellement. Chez 19 rats, un bolus de succinylcholine suivi d'une perfusion constante ont été administrés de façon à maintenir une dépression du « twitch » de 49,2% ($\pm 1,5$ SEM). Chez des groupes deux ou trois rats, de la cimétidine a alors été administrée par voie intraveineuse à raison de 3,2, 7,5, 10, 17,8, 23,7, 31,6, ou 56,2 mg · kg⁻¹. La cimétidine a produit une

potentialisation immédiate de la dépression du « twitch » suivie d'une récupération transitoire puis d'une potentialisation soutenue. La potentialisation maximale est survenue en 19,0 ($\pm 2,7$) sec et s'est maintenue à un niveau stable chez 11 rats. La récupération était évidente 4,1 ($\pm 0,4$) min après l'administration de cimétidine. Il y avait une bonne relation entre la potentialisation maximale et la concentration sérique de cimétidine avec une potentialisation de 50% survenant à 46,5 ($\pm 4,6$) $\mu\text{g} \cdot \text{ml}^{-1}$. Le degré de potentialisation soutenue à un niveau stable n'était pas corrélatif à la concentration sérique de cimétidine mais il y avait une faible relation entre la récupération et la concentration sérique de cimétidine. Ces résultats supportent les rapports d'une interaction entre la cimétidine et la succinylcholine chez les humains.

There are widespread indications for the use of H₂ receptor antagonists in anaesthetic practice. These agents are used routinely to increase pH and decrease the volume of gastric contents before induction of anaesthesia.¹ Also, it has been suggested that H₂ (and H₁) antagonists be used to prevent cardiovascular changes occurring during the administration of drugs which release histamine, e.g., certain neuromuscular blockers.²

Cimetidine, the prototypic H₂ antagonist, has been the most commonly used agent as premedication to reduce aspiration pneumonitis.³ For this purpose, cimetidine, 150 to 300 mg is administered the night before surgery and one to two hours before induction of anaesthesia. Since cimetidine exhibits diverse cholinergic effects including an anti-cholinesterase and a neuromuscular blocking effect,⁴ there is the potential for interaction between cimetidine and the neuromuscular blocking drugs administered to facilitate intubation and to maintain muscle relaxation during surgery. The effect of cimetidine on the action of the depolarizing (non-competitive) neuromuscular blocker, succinylcholine, has not attracted detailed systematic attention but one study has shown that cimetidine prolongs the duration of action of this neuromuscular blocker.⁵ However, two other studies, also in patients, did not reveal any changes in the duration of action of succinylcholine in the presence of cimetidine.^{6,7} Therefore, the present study was designed to examine the potential interaction between

Key words

NEUROMUSCULAR RELAXANTS: succinylcholine;

HISTAMINE: cimetidine;

PHARMACOLOGY: interaction.

From the Pharmacy Department, The University of Sydney, New South Wales, Australia.

Address correspondence to: Dr. I. Ramzan, Pharmacy Department, The University of Sydney, New South Wales 2006, Australia.

This project was supported in part by financial assistance from the Pharmacy Research Trust of New South Wales.

Accepted for publication 6th December, 1991.

succinylcholine and cimetidine and to test the hypothesis that H₂ receptor blockade with cimetidine adversely affects neuromuscular function during anaesthesia.

Methods

Anaesthesia and surgical preparation

The protocol was reviewed and approved by the Animal Care and Use Committee of The University of Sydney. Twenty-three male Sprague-Dawley rats (University of Sydney Animal Services) weighing 304–556 g were used. Anaesthesia was induced with diethyl ether and maintained throughout the experiment with *iv* urethane, 1.25 g · kg⁻¹ in total, given as six divided doses during the surgical preparation of the animal. Rectal temperature was servo-controlled to about 38° C with a heated surgical table and a heat lamp.

The surgical preparation consisted of cannulation of both jugular veins and a carotid artery. The right jugular vein was for administration of urethane and cimetidine and the left for the administration of the neuromuscular blocker. Blood samples for drug assay were obtained from the carotid artery cannula. Following the vascular cannulations, a tracheostomy was performed and the lungs were ventilated mechanically with room air. A sciatic nerve in the thigh was then isolated and stimulated electrically using supramaximal stimuli of 2 volts with a duration of 0.2 msec at a frequency of 0.1 Hz. The mechanical twitch response of the isolated tibialis anterior muscle was then measured using a force-displacement transducer and a polygraph recorder.

Experimental protocol

The preparation was allowed to stabilize for 15 min during which baseline twitch measurements were performed. Succinylcholine was then administered as an *iv* bolus of 1 mg · kg⁻¹, followed by an infusion at approximately 50 µg · kg⁻¹ · min⁻¹ and titrated to maintain approximately 50% suppression of the baseline twitch response. The concentration of the drug in the infusion solution was such as to allow infusion of about 1.5 ml of infusate every hour. After a further 20 min of stabilization, cimetidine *iv* was administered into groups of two to three rats in a random order in doses of 3.2, 7.5, 10, 17.8, 23.7, 31.6 or 56.2 mg · kg⁻¹. The selected doses were considered to be sufficient to show either zero or maximum effect and were chosen based on the concept of equally spaced log doses.⁸ Two blood samples, 0.35 ml, were withdrawn, one as close as possible to the time of the peak potentiation by cimetidine and the other when the potentiation by cimetidine had stabilized. After the observation of the entire cholinergic effect of cimetidine, approximately 20–25 min after cimetidine dosing, the infusion of suc-

cynylcholine was discontinued to record the spontaneous recovery of the muscle twitch response. In another four rats, either 10 or 100 mg · kg⁻¹ cimetidine was given *iv* but without any previous succinylcholine to determine whether cimetidine alone altered neuromuscular transmission.

Measurements and calculations

Cimetidine serum concentrations were determined using a modification (protein precipitation rather than liquid extraction) of a published reversed phase liquid chromatographic procedure.⁹ The coefficient of variation of the assay was 3.3 and 0.7% at 5 and 150 µg · ml⁻¹ respectively.

Cimetidine potentiation and reversal of succinylcholine-induced suppression of the twitch response was calculated as:

$$\text{Potentiation of twitch suppression (\%E)} = \frac{\text{Pre-cimetidine twitch height} - \text{Post-cimetidine twitch height (at peak or plateau potentiation)}}{\text{Pre-cimetidine twitch height}} \times 100$$

and

$$\text{Reversal of twitch suppression (\%E)} = \frac{\text{Pre-cimetidine paralysis} - \text{Post-cimetidine paralysis (at reversal)}}{\text{Pre-cimetidine paralysis}} \times 100$$

These effects (either peak or plateau potentiation or reversal) were related to either the cimetidine dose administered or the cimetidine serum concentration measured using either a linear, log-linear, an E_{max} or a sigmoid E_{max} (Hill) equation¹⁰ using the personal computer based non-linear regression program PCNONLIN (Version 3.0, SCI Software, USA). For the reversal effect-serum concentration relationship this could only be done by use of the cimetidine concentration taken at peak potentiation since no concentrations were available at peak reversal. F-ratio tests were used to examine if the relationship between cimetidine's effect(s) and its serum concentration was statistically significant, realizing that this is not an entirely unbiased procedure when used with nonlinear equations.¹¹ The appropriate equation that best described the effect of cimetidine was chosen by use of the F-ratio test, the Akaike Information Criteria, the Schwarz test and the Ip parameter.¹² *P* < 0.05 was considered statistically significant for the linear equations but an exact significance value was not computed for the nonlinear equations.¹¹ All data are presented as mean ± SEM.

Results

Succinylcholine mean (±SEM) infusion rate of 37.5 ± 2.3

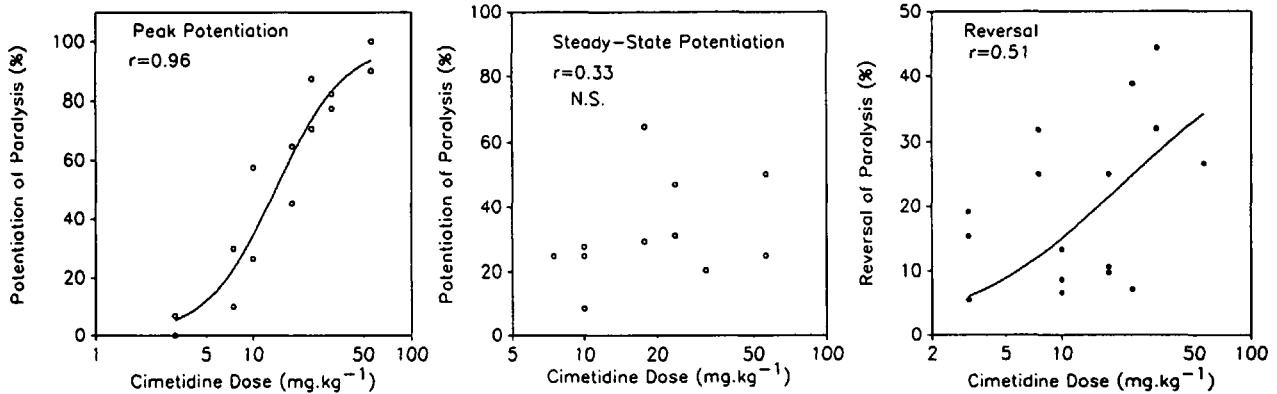


FIGURE 1 Muscle twitch recording in three rats illustrating the varied effects of *iv* cimetidine. 1 = baseline steady-state twitch suppression, 2 = peak potentiation, 3 = transient reversal, 4 = steady-state potentiation. At the first arrow cimetidine was administered and the succinylcholine infusion was discontinued at the second arrow.

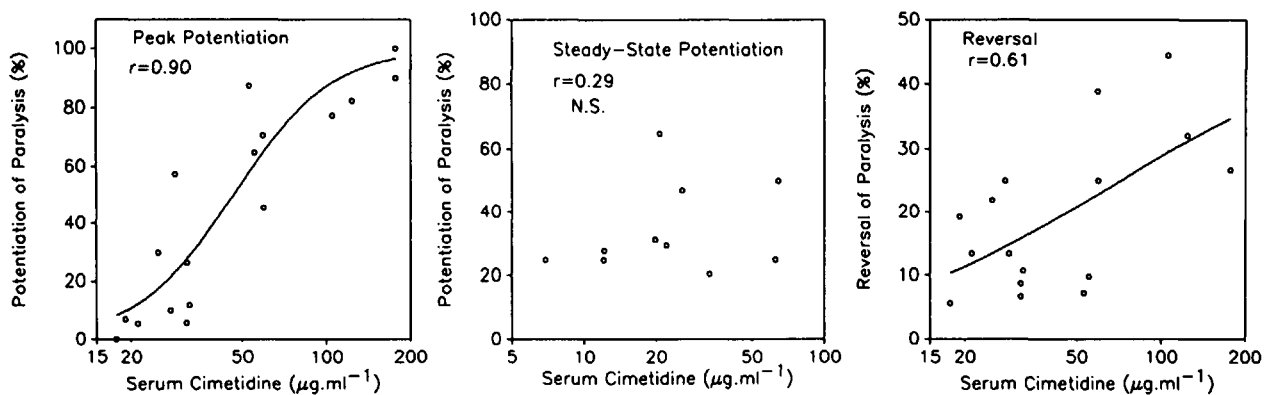


FIGURE 2 Relationship between cimetidine *dose* and its effect on succinylcholine-induced twitch suppression in rats. The relationship between peak potentiation and dose was most appropriately characterized by use of a sigmoidal E_{max} (Hill) equation while that between reversal and dose was described best by an E_{max} model. There was no significant relationship between steady-state potentiation and dose.

$\mu\text{g} \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ produced $49.2 \pm 1.5\%$ twitch suppression in 19 rats. When *iv* cimetidine was administered it produced an immediate potentiation of twitch suppression in all but two of the three rats receiving the lowest ($3.2 \text{ mg} \cdot \text{kg}^{-1}$) dose of cimetidine. This was followed by a transient reversal of paralysis in all but two rats who received either a dose of 56.2 or $23.7 \text{ mg} \cdot \text{kg}^{-1}$. A second phase of potentiation was then noted in eleven rats which persisted at steady-state or plateaued. This steady-state potentiation ranged from 9 to 65% and occurred in rats who received cimetidine doses ranging from 7.5 to $56.2 \text{ mg} \cdot \text{kg}^{-1}$. The onset of peak potentiation occurred in $19.0 \pm 2.7 \text{ sec}$ after cimetidine dosing and the peak reversal was observed $4.1 \pm 0.4 \text{ min}$ after the doses of cimetidine. In the 11 rats where the potentiation persisted, it took $23.4 \pm 1.5 \text{ min}$ to stabilize. The first blood sample for cimetidine assay was taken $75 \pm 6 \text{ sec}$ after the cimetidine dose. Due to technical constraints, this sampling time was

delayed by approximately one minute from the time of peak potentiation. Unfortunately, no cimetidine concentrations were available at peak reversal. The second cimetidine concentration was measured when the potentiation by cimetidine had stabilized (Figure 1).

There was a good relationship between the degree of peak potentiation of twitch suppression by cimetidine and its dose and serum concentration respectively and the sigmoid E_{max} (or Hill) equation was the most appropriate for characterizing these relationships. Potentiation at steady-state was not related to either the cimetidine dose or its serum concentration. However, there was an apparent relationship between the reversal of paralysis and the dose and serum cimetidine concentration respectively, although these relationships were not strong (Figures 2 and 3).

The cimetidine dose-producing 50% potentiation during its peak effect was $13.9 \pm 1.2 \text{ mg} \cdot \text{kg}^{-1}$ and this

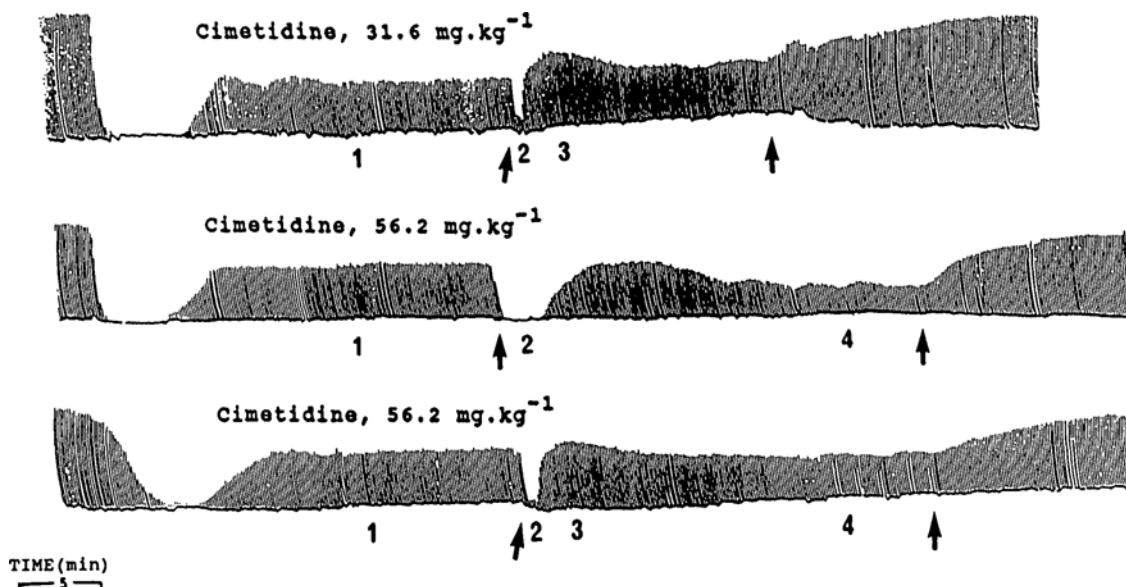


FIGURE 3 Relationship between serum cimetidine concentration and its effect on succinylcholine-induced twitch suppression in rats. The relationship between peak potentiation and concentration was most appropriately characterized by use of a sigmoidal E_{max} (Hill) equation while that between reversal and concentration was described best by an E_{max} model. There was no significant relationship between steady-state potentiation and concentration.

effect was associated with a cimetidine serum concentration of $46.5 \pm 4.6 \mu\text{g} \cdot \text{ml}^{-1}$ (or $184 \pm 18.2 \mu\text{M}$). Maximal reversal, of pre-cimetidine paralysis, was $46.8 \pm 17.5\%$ and the concentration producing half maximal reversal was $62.5 \pm 46.6 \mu\text{g} \cdot \text{ml}^{-1}$ ($248 \pm 185 \mu\text{M}$). When cimetidine 10 or 100 $\text{mg} \cdot \text{kg}^{-1}$ was administered alone to rats who had not received any succinylcholine, there was no effect on the normal (baseline) twitch response.

Discussion

Cimetidine produces three distinct cholinergic effects against the neuromuscular paralysis induced with the depolarizing agent succinylcholine. Peak potentiation of paralysis is followed by a rapid reversal which precedes plateau or steady-state potentiation. The predominant effect of cimetidine, however, is to potentiate the neuromuscular paralysis and the reversal is transient. This is supported by the observation that the relationship between peak potentiation and cimetidine serum concentration was easily discernable even though these concentrations were measured approximately one minute after peak potentiation. In contrast, reversal of paralysis was only weakly correlated to serum concentrations, although these concentrations were not determined exactly at the time of peak reversal. The results of this study support the hypothesis that *iv* cimetidine adversely affects neuromuscular function during anaesthesia, at least in rats.

The mechanism(s) responsible for the observed potentiation and transient reversal of neuromuscular paralysis by

cimetidine is not clear from this study but may be a result of either a neuromuscular blocking effect of cimetidine itself or an anti-cholinesterase effect of the drug. Cimetidine has been shown previously *in vitro* to produce neuromuscular blockade but at very high ($500\text{--}2000 \mu\text{g} \cdot \text{ml}^{-1}$) bath concentrations.¹³ However, in the present study, cimetidine by itself at $100 \text{mg} \cdot \text{kg}^{-1}$ failed to affect neuromuscular transmission. This is not surprising since serum concentrations of cimetidine generated at this dose were only about $200 \mu\text{g} \cdot \text{ml}^{-1}$. Cimetidine also exhibits cholinesterase inhibition *in vitro* but again at high concentrations between 25 and $2500 \mu\text{g} \cdot \text{ml}^{-1}$ (100 to $10000 \mu\text{M}$).¹⁴ Since succinylcholine depends on hydrolysis by cholinesterases for termination of its action,¹⁵ inhibition of this family of enzymes would either potentiate or reverse the action of succinylcholine. Potentiation by cimetidine would only be observed of course if cimetidine was able to inhibit the cholinesterase that breaks down succinylcholine more than the cholinesterase that metabolizes the endogenous transmitter acetylcholine itself. In this respect, it is important to note that cimetidine is a more potent inhibitor of acetylcholinesterase (AChE) than pseudo-cholinesterase (PChE) or butyrylcholinesterase (BuChE),¹⁴ the former being more specific for acetylcholine breakdown while the latter hydrolyzes succinylcholine.¹⁶ The potentiation of succinylcholine's effect by cimetidine may therefore result from an intrinsic neuromuscular blocking effect of cimetidine since inhibition of PChE or BuChE is weak. The transient reversal of paralysis by cimetidine may be

mediated via AChE inhibition provided there is a mixed or phase II block present since succinylcholine is a competitive agent. Phase II (non-depolarizing or competitive) block would be reversed by increased acetylcholine concentrations resulting from AChE inhibition by cimetidine. The presence of a phase II block by succinylcholine could not be confirmed since the single-twitch response used here is unable to distinguish between a phase I and a phase II neuromuscular blockade.

Potential of succinylcholine by cimetidine may have clinical implications but in quantitative terms this interaction is not likely to be pronounced and probably will be unnoticed in anaesthesia practice since cimetidine doses used in patients are substantially lower than in this study. The serum concentrations (18 to 177 $\mu\text{g} \cdot \text{ml}^{-1}$) that produce potentiation in rats are about 5 to 24 times higher than that noted (3.5 to 7.43 $\mu\text{g} \cdot \text{ml}^{-1}$) in patients.¹⁷ In addition, with clinical premedication doses, cimetidine does not affect PChE or BuChE enzyme activity *in vivo*¹⁸ although higher than normal clinical concentrations (150–1500 $\mu\text{g} \cdot \text{ml}^{-1}$) decrease the PChE activity and succinylcholine hydrolysis rate respectively *in vitro*.^{18,19} Thus, caution will need to be exercised in patients who are otherwise at risk of displaying prolonged paralysis, for example, those with abnormal or low plasma cholinesterase enzyme activity or those receiving drugs that depress neuromuscular transmission. Patients with atypical cholinesterases may also be more susceptible to phase II block²⁰ which may add a further complication.

In conclusion, this study demonstrates that *iv* cimetidine potentiates the neuromuscular blocking action of succinylcholine *in vivo* in rats. These results support reports of prolongation of succinylcholine blockade with cimetidine in patients.

References

- 1 *Stoelting RK*. Gastric fluid pH in patients receiving cimetidine. *Anesth Analg* 1978; 57: 675–7.
- 2 *Scott RPF, Savarese JJ, Basta SJ, et al*. Atracurium: clinical strategies for preventing histamine release and attenuating the haemodynamic response. *Br J Anaesth* 1985; 57: 550–3.
- 3 *Coombs DW, Hooper D, Colton T*. Pre-anesthetic cimetidine alteration of gastric fluid volume and pH. *Anesth Analg* 1979; 58: 183–8.
- 4 *Gwee MCE, Cheah LS*. Actions of cimetidine and ranitidine at some cholinergic sites: implications in toxicology and anaesthesia. *Life Sci* 1986; 39: 383–8.
- 5 *Kambam JR, Dymond R, Krestow M*. Effect of cimetidine on duration of action of succinylcholine. *Anesth Analg* 1987; 66: 191–2.
- 6 *Stirt JA, Sperry RJ, DiFazio CA*. Cimetidine and succinylcholine: potential interaction and effect on neuromuscular blockade in man. *Anesthesiology* 1988; 69: 607–8.
- 7 *Woodworth GE, Sears DH, Grove TM, Ruff RH, Kosek PS, Katz RL*. The effect of cimetidine and ranitidine on the duration of action of succinylcholine. *Anesth Analg* 1989; 68: 295–7.
- 8 *Van Rossum JM*. Cumulative dose-response curves II. Technique for the making of dose-response curves in isolated organs and the evaluation of drug parameters. *Arch Int Pharmacodyn Ther* 1963; 143: 299–330.
- 9 *Kaka JS*. Rapid method for cimetidine and ranitidine determination in human and rat plasma by HPLC. *J Liquid Chromatogr* 1988; 11: 3447–56.
- 10 *Holford NHG, Sheiner LB*. Understanding the dose-effect relationship: clinical application of pharmacokinetic-pharmacodynamic models. *Clin Pharmacokinet* 1981; 6: 429–53.
- 11 *Draper NR, Smith H*. *Applied Regression Analysis*. 2nd ed., New York: John Wiley & Sons Inc., 1981; 484.
- 12 *Imbimbo BP, Martinelli P, Rocchetti M, Ferrari G, Bassotti G, Imbimbo E*. Efficiency of different criteria for selecting pharmacokinetic multiexponential equations. *Biopharm Drug Dispos* 1991; 12: 139–47.
- 13 *Galatulas I, Bossa R, Benvenuti C*. Cimetidine induced neuromuscular blockade 'in vitro': antagonism by calcium. *IRCS J Med Sci* 1980; 8: 874.
- 14 *Hansen WE, Bertl S*. The inhibition of acetylcholinesterase and pseudocholinesterase by cimetidine. *Arzneimittel-Forschung (Drug Research)* 1983; 33: 161–3.
- 15 *Chagas C, Sollero L, Suarez-Kurtz G*. Synthetic neuromuscular blocking agents: absorption – distribution – metabolism – excretion. In: *Cheymol J (Ed)*. *Neuromuscular Blocking and Stimulating Agents*, Volume 1, 1st ed., Oxford: Pergamon Press Ltd., 1972; 409–23.
- 16 *Foldes FF*. Enzymes of acetylcholine metabolism. In: *Foldes FF (Ed)*. *Enzymes in Anesthesiology*, New York: Springer-Verlag Inc., 1978; 110.
- 17 *Walkenstein SS, Dubb JW, Randolph WC, Westlake WJ, Stote RM, Intoccia AP*. Bioavailability of cimetidine in man. *Gastroenterology* 1978; 74: 360–5.
- 18 *Kambam JR, Franks JJ*. Cimetidine does not affect plasma cholinesterase activity. *Anesth Analg* 1988; 67: 69–70.
- 19 *Cook DR, Stiller RL, Chakravorti S, Mannenhira T*. Cimetidine does not inhibit plasma cholinesterase activity. *Anesth Analg* 1988; 67: 375–6.
- 20 *Savarese JJ, Ali HH, Murphy JD, Padget C, Lee C, Ponitz J*. Train of four stimulation in the management of prolonged neuromuscular blockade following succinylcholine. *Anesthesiology* 1975; 42: 106–11.