nuclei to the culture, which may obscure metaphase spreads and possibly may interfere with the mitotic stimulation. In addition this separates leukocytes from any lipid component in the blood that may be significant, particularly in situations where control over feeding may not be possible (e.g., wild raptor species). The amount of blood required is similar to that used by Zartman (4), being much less than earlier methods (2,3). Finally, the culture conditions more closely approximate the average physiological state of birds than any previously reported method. In our limited experience of material from birds of 10 different orders, the use of phytohemagglutinin is far superior to other mitotic stimulants. Pokeweed mitogen, as suggested by Zartman (4), seems to be less suitable for most avian species, with the possible exception of the Falconiformes (11).

Inasmuch as leukocyte cultures are perhaps the easiest and most convenient method of obtaining chromosome preparations, the method reported here should facilitate cytogenetic investigations of avian species, particularly at the population level. In addition, it may find application in the genetic sexing of monotypic birds for captive propagation programs.

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## ERRATUM

Modified Nutrient Medium MCDB 151, Defined Growth Factors, Cholera Toxin, Pituitary Factors, and Horse Serum Support Epithelial Cell and Suppress Fibroblast Proliferation in Primary Cultures of Rat Ventral Prostate Cells. Wallace L. McKeehan, Pamela S. Adams, and Mary P. Rosser

Vol. 18, No. 2, page 88, line 27. "All culture media contained 100  $\mu$ g/ml of kanamycin sulfate" rather than "100 mg/ml".