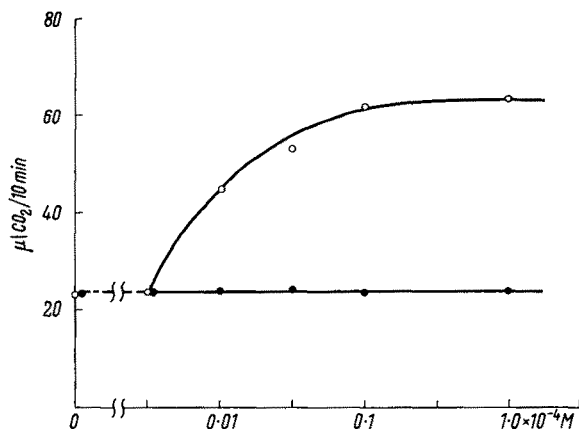


The Interaction Between Pyridoxal-5-Phosphate and Arginine Apodecarboxylase

In previous papers¹, the interaction between pyridoxal-5-phosphate (Py-5-P) and potassium cyanide was described. The resulting addition product, namely a cyanohydrine, which has been characterized by us both spectrophotometrically and ionophoretically, appeared unable to activate the glutamic-oxalacetic apotransaminase prepared and almost completely resolved according to the procedure of O'KANE and GUNSALUS². On the other hand, no competitive action with Py-5-P was observed.

These results suggested that the 4-aldehyde group of Py-5-P is more likely involved in the interaction between Py-5-P and the specific protein portion (apotransaminase) rather than in the formation of a Schiff base between Py-5-P and the aminoacid substrate, as assumed by SCHLENK and FISHER³ in their well known scheme.



Activity curves of apodecarboxylase preparation after addition of Py-5-P (○—○) and of its cyanohydrine (●—●). The first curve is not influenced by previous or simultaneous addition of the cyanohydrine.

In order to verify whether such a rôle of the 4-aldehyde group of Py-5-P is essential in the activation process of apotransaminase only or of other apoenzymes (whose coenzyme is Py-5-P) as well, we have investigated the interaction between Py-5-P and arginine apodecarboxylase.

The enzyme preparation used in our experiments has been obtained from *Esch. coli*, Mac Leod ATCC 10536, according to TAYLOR and GALE⁴. The resolution obtained was about 65%. The decarboxylation reaction was followed in Warburg manometers containing 0.5 ml enzyme preparation, 1.5 ml M/5 acetate buffer pH 5.25 and 0.5 ml M/15 L(+)arginine. Experiments were carried out at 30°C and the evolution of CO₂ was studied for 10 min after addition of substrate (from the side bulb) following equilibration. The specific activity of the arginine apodecarboxylase preparation (in presence of 0.5 × 10⁻⁴ M Py-5-P) was 102, in terms of µl CO₂ liberated in 10 min/mg protein.

While KCN inhibits arginine apodecarboxylase activity as reported by TAYLOR and GALE⁴, the cyanohydrine of Py-5-P does not modify the residual activity of the partially resolved enzyme. Thus, the enzyme preparation previously incubated for 30 min with Py-5-P cyanohydrine, even at a concentration 10⁻⁴ M, shows the same activity as the enzyme preparation not preincubated.

The activation curve of the arginine apodecarboxylase by Py-5-P is not influenced by the previous incubation of the enzyme with Py-5-P cyanohydrine, independently of the concentration of this latter. Further, the Py-5-P cyanohydrine does not interfere with Py-5-P in the activation process even when added to the apoenzyme simultaneously with Py-5-P (Figure).

As it appears from our results, the cyanohydrine of Py-5-P neither activates apodecarboxylase nor exerts a competitive action in regards to Py-5-P itself, so that the 4-aldehyde group of Py-5-P seems to be involved in the linkage to the apoenzyme.

It can be assumed that such a rôle is probably played by the 4-aldehyde-group in the case of any apoenzyme, whose coenzyme is Py-5-P.

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Riassunto

Vengono descritti i risultati di indagini sull'interazione tra piridossal-5-fosfato ed apodecarbossilasi arginina.

La cianidrina del piridossal-5-fosfato non attiva l'apoenzima né riduce l'effetto attivante del coenzima naturale. Si presume che il gruppo aldeidico del piridossal-5-fosfato sia interessato nell'attacco all'apoenzima.

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Breakdown of Adenine Nucleotides in Beef Muscle *post mortem* and its Relation to the Content of Ammonia

During the breakdown of adenosine nucleotides in muscle tissue *post mortem*, ammonia and inosinic acid (IMP) is released from adenylic acid (AMP). During autolysis of beef muscle at various temperatures, a different production of ammonia was found¹. It was less during autolysis at + 37°C than at + 5°C, taking into consideration the increased temperature. To what degree the content of ammonia in muscle *post mortem* is released from them, is studied in this article.

Musculus psoas maior from 4 well nourished young bulls was used. One part of each was hung in a thermostat at + 37°C (100% relative humidity) and irradiated with ultraviolet rays. The greater part of each muscle was placed in a box at + 5°C (70% relative humidity)². The first sample from the muscles was processed 3 h after slaughtering for the chromatography of free nucleotides. The other specimens were taken at different intervals of time. For chromatography, the tissue was extracted three times with 10% trichloroacetic acid, which was then ex-

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