from the 3 strains were observed. However, females of all the 3 strains had consistently higher protein contents.

Differences in the protein contents of diapausing and nondiapausing larvae have been reported within the same strain of pink bollworm^{6,11}. Lower protein contents in diapausing larvae was explained by its probable convertion to fat¹¹. Since the pink bollworm larvae do not show a

- uniform diapause intensity^{7,8}, the present comparison of a diapause and a non-diapause strain yields more meaningful results than diapause and non-diapause larvae from the same strain. The comparison of the IS and NAS strains shows that reduction of the protein concentration is not necessary for the induction of diapause.
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Evidence that the prostaglandin-like substances from Propionibacterium acnes are not identical with PGE,

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Summary. The prostaglandin-like substances (PLS) isolated from P. acnes were investigated by reversed phase chromatography and gas chromatography-mass spectrometry. These analyses demonstrated that PLS were not identical with PGE2, which supports a concept of PLS as a potential mediator of the inflammatory process in acne vulgaris.

The involvement of Propionibacterium acnes metabolites as mediators of inflammation has received increased attention during recent years. Our studies on P. acnes lipids, were to characterize the properties of the prostaglandin-like substances (PLS) found, as potent biologically active compound(s), in the lipid fraction of these bacteria¹. Bioassays on gerbil colon² and isolated human vessels³ revealed that PLS mimic the effects of prostaglandins of the E-type. Similarly, in vivo (hamster cheek pouch), these compounds induced a PGE-like microvascular response⁴. In addition, PLS raised cyclic AMP levels (rat ovary) approximately 2-fold⁵. On silicic acid chromatography PLS is eluted with the same solvent mixture as E-prostaglandins (i.e. with ethyl acetate: toluene 6:4, v/v). Therefore, we decided to investigate whether PGE₂ could be a constituent of PLS.

P. acnes were cultured anaerobically or aerobically for 1 week on a solid, artificial substrate. The recovery of the bacterial mass as well as the extraction procedure was reported earlier¹. About 1 µg of $(3,3,4,4-2\hat{H}_4)-(17,18^3H_2)$ labelled PGE₂ was added to the resulting lipid extract. (These carrier and tracer molecules will co-chromatograph with a possible PGE₂ generated by the bacteria.) The mixture was further purified and analyzed by gas chromatography-mass spectrometry using the accelerating voltage alternator technique^{6,7}. This analysis demonstrated that less than 1 ng of PGE₂ (=lower limit of safe detection) could be present in the bacterial extract. However, the biological activity of this sample corresponded to about 250 ng of PGE₂ equivalent in a gerbil colon bioassay.

In another experiment a sample (biological activity approximately 640 ng PGE₂ equivalent, gerbil colon bioassay) was subjected to reversed phase partition chromatography together with 50,000 dpm of 17,18-3H₂-PGE₂ (50 mCi/µmole) as described earlier⁸. No biological activity could be detected (gerbil colon bioassay) in those fractions where tritium-labelled PGE₂ appeared.

Thus, the present work demonstrates that PLS from P. acnes, despite their biological resemblance to E-prostaglandins, do not contain PGE2. This corresponds with a recent bioassay experiment on the human utero-tubal junction, where the effect of PLS was similar, although not identical with that of PGE₂⁹. Moreover, PLS possess potent chemotactic properties, which was not the case for PGE₂¹⁰. According to a recent suggestion¹¹, mediator functions may be associated with other prostaglandin-type compounds (e.g. endoperoxides), while modulator functions are associated with the end-products, i.e. prostaglandins themselves. This idea supports the concept of PLS in P. acnes as a potential inflammatory mediator, involved in the pathophysiology of acne vulgaris.

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