the present studies the contents of all cystic stomach implants were at pH 7-8. If acid were being produced by the implant, then buffering by proteins in the cyst contents or leakage of hydrogen ions across a faulty gastric mucosal barrier as postulated by Davenport[2] might be responsible for the neutral pH. It would be worthwhile to study acid-secretion in stomach implants using more sophisticated techniques and the use of gastrointestinal hormones and pharmacologically active substances.

In some implantation experiments we used very young fetal stomachs of age 13-14 days, when no canalicular parietal cell antigen can be demonstrated. The appearance of this antigen after 5-10 days implantation confirms the finding of other authors[12,14] that parietal cells can develop out of immature undifferentiated mucosal cells.

The decrease in expression of the canalicular parietal antigen in allogeneic implantations and the observed inflammatory and atrophic changes in the mucosa could be compared with the loss of parietal cells and humoral and cellular immune response to parietal cell antigen in chronic atrophic gastritis in humans.

1 Ceredig, R., Henderson, D.C., and Nairn, R.C., Experimental model of ulcerative colitis. Nature *266* (1977) 74-75.
2 Davenport, H.W., Physiological parameter of the gastric mucosal barrier. Am. J. dig. Dis. *21* (1976) 141-143.
3 Ferguson, A., and Parrott, D.M.V., Growth and development of 'antigen-free' grafts of foetal mouse intestine. J. Path. *106* (1972) 95-101.
4 Ferguson, A., and Parrott, D.M.V., Histopathology and time course fo rejection of allografts of mouse small intestine. Transplantation *15* (1973) 546-554.
5 Ferguson, A., Implantation of tissue under the kidney capsule, in: Handbook of Experimental Immunology, vol.3, 2nd edn, pp.A3, 12-13. Ed. D.M. Weir. Blackwell Scientific Publications, Oxford 1974.
6 Fisher, J.M., and Taylor, K.B., The intracellular localization of Castle's intrinsic factor by an immunofluorescent technique using antibodies. Immunology *16* (1969) 779-784.
7 Harrington, J.T., and Stastny, P., Macrophage migration from an agarose droplet: development of a micro-method for assay of delayed hypersensitivity. J. Immun. *110* (1973) 752-759.
8 Holden, R.J., and Ferguson, A., Histopathology of cell mediated immune reaction in mouse colon - allograft rejection. Gut *17* (1976) 661-670.
9 Lowry, O.H., Rosebrough, N.J., Farr, A.L., and Randall, R.J., Protein measurement with the folin reagent. J. biol. Chem. *193* (1951) 265-275.
10 MacDonald, T.T., and Ferguson, A., Hypersensitivity reactions in the small intestine. 2. Effects of allograft rejection on mucosal architecture and lymphoid cell infiltrate. Gut *17* (1976) 81-91.
11 McCoy, J.L., Dean, J.H., and Herberman, R.B., Direct and indirect agarose microdroplet migration inhibition assays for detection of cell-mediated immunity to human tumor-associated antigens, in: In vitro methods in cell-mediated and tumor immunity, pp.621-628. Eds B.R. Bloom, and J.R. David. Academic Press, New York 1976.
12 Matsuyama, M., and Suzuki, H., Differentiation of immature mucous cells into parietal, argyrophil, and chief cells in stomach grafts. Science *169* (1970) 385-386.
13 Pipan, N., Die Feinstruktur von Belegzellen der Maus während der Differenzierung. Cytobiologie *2* (1970) 33-46.
14 Ragins, H., Wincze, F., Liu, S.M., and Dittbrenner, M., The origin and survival of gastric parietal cells in the mouse. Anat. Rec. *162* (1968) 99-110.
15 Rugh, R., The Mouse (its Reproduction and Development). Burgess Publishing Company, Minneapolis 1968.

0014-4754/83/060558-09$1.50+0.20/0

Correction

Structure of three isomeric host-specific toxins from *Helminthosporium sacchari,* Experientia *39*/4 (1983) 343-347.

V. Macko, W. Acklin, C. Hildenbrand, F. Weibel and D. Arigoni

Boyce Thompson Institute for Plant Research at Cornell University, Ithaca (N. Y. 14853, USA), and Laboratorium für Organische Chemie, Eidgenössische Technische Hochschule Zürich, CH-8092 Zürich (Switzerland)

We regret a misprinting in table 3 of this article. The table should have read as follows:

Table 3. ^{1}H-NMR data[a] for aglycone moieties

Carbon No.	Isomer A		Isomer B		Isomer C	
	H_α	H_β	H_α	H_β	H_α	H_β
C-1	1.40	1.80	1.28	1.8	1.34	1.88
C-2	-	?	-	3.88	-	4.20
C-3	2.04	2.40	1.97	2.77	5.50	
C-5	-	-	1.92	-	2.10	-
C-6	2.73	2.24	1.8	1.63	1.99	1.52
C-7	-	2.61	-	2.62	-	2.64
C-8	1.64	2.00	1.8	1.8	1.83	1.93
C-9	1.37	1.37	1.43	1.32	1.45	1.29
C-12	5.11; 5.20		5.19; 5.30		5.24; 5.33	
C-13	4.03; 4.23		~4.07; 4.22		4.05; 4.23	
C-14	1.70		4.64; 4.95		1.72	
C-15	1.15		0.78		0.92	

[a] δ-Values in ppm relative to internal DSS = 0.