

### Interference of Sodium Ethylenediaminetetraacetate in the Determination of Proteins and its Elimination

In a course of investigations concerning purification of viruses, it became necessary to determine the concentration of proteins in samples containing sodium ethylenediaminetetraacetate ( $\text{Na}_2\text{EDTA}$ ). Since it has been found in preliminary experiments that  $\text{Na}_2\text{EDTA}$  reduces the Folin-Ciocalteu phenol reagent (Figure 1) in the absence of proteins, experiments have been undertaken to eliminate this reaction by binding EDTA with appropriate metallic ions. The presence of  $\text{Ca}^{++}$  ions has been found to decrease appreciably the development of blue color due to the reduction of phosphomolybdate (Figure 2). Results of further experiments show that it is possible to deter-

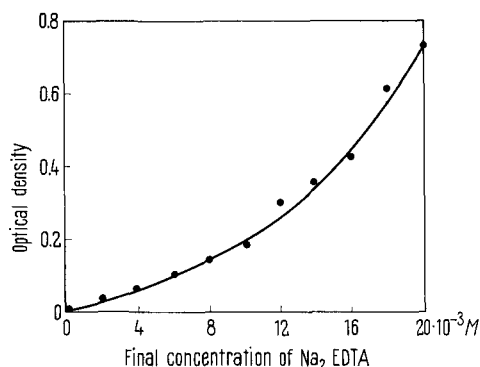


Fig. 1. Reduction of the Folin-Ciocalteu phenol reagent by EDTA. 0.5 ml of solutions containing various amounts of  $\text{Na}_2\text{EDTA}$  were mixed with 5 ml of reagent C<sup>1</sup>, and 10 min later 0.25 ml of the phenol reagent (Fisher Scientific Co., Philadelphia, Pa.) was added. After 30 min the optical density of the samples was read at 750 nm on the Beckman DU spectrophotometer.

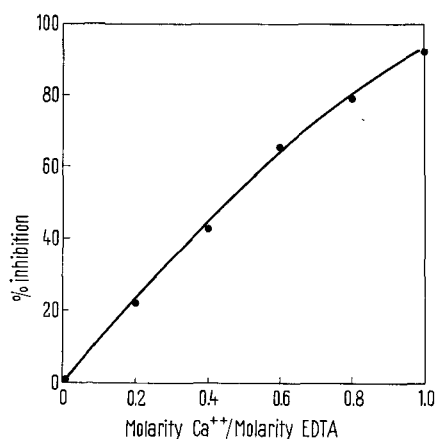


Fig. 2. Inhibition by  $\text{Ca}^{++}$  ions of the reduction of the Folin reagent by EDTA. 0.5 ml volumes of 0.1M  $\text{Na}_2\text{EDTA}$  were mixed with 0–0.05 ml of 1M  $\text{CaCl}_2$  and then the procedures given under Figure 1 were used.

**Zusammenfassung.** Es wird die Interferenz von Natrium Ethylendiaminetetraacetat bei der Eiweissbestimmung nach der Methode von LOWRY et al.<sup>1</sup> und deren Verhinderung gezeigt. Die Anwesenheit von Natrium Ethylendiaminetetraacetat interferiert bei der Eiweissbestimmung nach der Methode von LOWRY, da sie selbst zur

mine proteins in the presence of  $\text{Na}_2\text{EDTA}$  after eliminating its effect by addition of an excess of  $\text{CaCl}_2$  to the sample to be analyzed (Figure 3). The color development in the presence of  $\text{CaEDTA}$  is delayed (Figure 4), but higher final optical density (O.D.) readings are obtained in comparison with control samples without  $\text{CaEDTA}$  and the same protein content. Evidently the presence of  $\text{CaEDTA}$  does not impair the enhancing effect of  $\text{Cu}^{++}$  ions on the color development<sup>1</sup>, though the formation constant of  $\text{CuEDTA}$  is nearly  $10^5$  times higher than that of  $\text{CaEDTA}$ <sup>2</sup>.

<sup>1</sup> O. H. LOWRY, N. J. ROSEBROUGH, A. L. FARR, and R. J. RANDALL, *J. biol. Chem.* 193, 265 (1951).

<sup>2</sup> F. J. WELCHER, *The Analytical Uses of Ethylenediaminetetraacetic Acid* (D. Van Nostrand Co., Inc., Princeton, N.J. 1957).

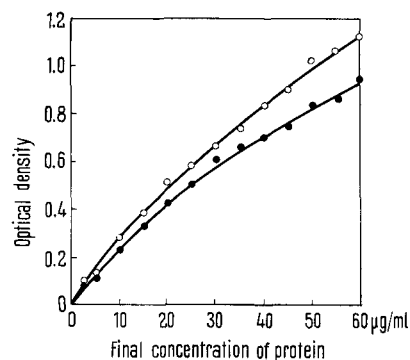


Fig. 3. Calibration curves for bovine serum albumin in the presence (—○—) and absence (—●—) of EDTA. 0.5 ml samples containing different amounts of protein were mixed with 0.5 ml of 0.1M  $\text{Na}_2\text{EDTA}$  + 0.06 ml of 1M  $\text{CaCl}_2$  (—○—) (= 20% excess) or 0.56 ml of distilled water (—●—) and then with the appropriate reagents. The samples containing  $\text{Ca}^{++}$  were centrifuged before measuring the optical density to remove the precipitate formed after addition of reagent C<sup>1</sup> (=  $\text{CaCO}_3$ ).

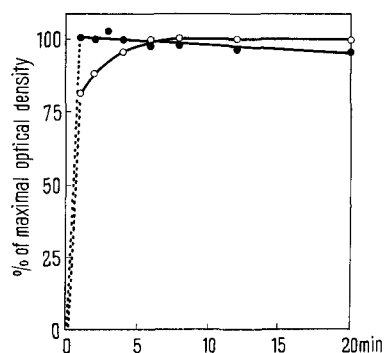


Fig. 4. Color development in the presence (—○—) and absence (—●—) of EDTA. The conditions were identical with those given under Figure 3. 0.5 ml samples of a 0.12% solution of bovine serum albumin were used.

Reduktion des Folin-Reagens führt. Es gelingt, diese Interferenz durch Zusatz von  $\text{CaCl}_2$  zu verhindern.

A. R. NEURATH

Wyeth Laboratories Inc., Radnor (Pa., USA),  
November 25, 1965.