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Erratum

Rapid HLA-DPB typing using enzymatically amplified DNA and nonradioactive sequence-specific oligonucleotide probes

Teodorica L. Bugawan, Ann B. Begovich, and Henry A. Erlich

We wish to clarify some of the issues related to cloning polymerase chain raction (PCR) products amplified with the HLA-DPB1 primers UG19 and UG21 described in Bugawan and co-workers (1990). These primers were designed with restriction sites added to their 5′ ends in order to facilitate cloning. UG19 (the left primer) has one strand of the *Pst* I site and UG21 (the right primer) has one strand of the *Bam* HI site. However, there is an internal *Pst* I site (codons 89 and 90) within the DPB1 second exon amplified by these primers so that complete *Bam* HI and *Pst* I digestion creates a 261-bp *Pst* I fragment. Consequently, cloning a *Bam* HI-*Pst* I digested PCR product into a *Bam* HI-*Pst* I cut vector results in very low cloning efficiency. One solution to this problem is to clone *Pst* I digested PCR products amplified with UG19 and UG21 into a *Pst* I digested vector that has been treated with bacterial alkaline phosphatase to minimize self-ligation of the vector. Alternatively, the PCR products can be blunt-end cloned or alternative DPB1 primers (e. g., DB01, CAGGGATCCGCAGAGAATTAC, described in Bugawan and co-workers (1988); a "left" primer with a *Bam* HI site) can be used. We regret any inconvenience experienced in cloning DPB1 sequences amplified with these primers and hope this note clarifies the situation.

Reference

Bugawan, T. L., Horn, G. T., Long, C. M., Mickelson, E., Hansen, J. A., Ferrara, G. B., Angelini, G., and Erlich, H. A.: Analysis of HLA-DP allelic sequence polymorphism using the in vitro enzymatic DNA amplification of DPα and DPβ loci. *J Immunol 141:* 4024–4030, 1988