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Contact Infection of Mink with Influenza A Viruses of Avian and Mammalian Origin

Brief Report

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Summary

Avian influenza A virus Hav 7 N 2 was transmitted to mink by contact. Other avian influenza A viruses, Hav 4 Nav 1 and Hav 6 Nav 5, were not transmitted, and human, swine and equine influenza A viruses were transmitted to mink by a similar contact.

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Recently, attention has been focused on avian influenza viruses which have been considered to be the origin of a 'New' pandemic influenza virus (8, 9). Mammalian infection due to avian influenza viruses by intranasal inoculation has been reported in mink (*Mustela vison*) (5); therefore, it is possible that mink and related species of mammals play a role in the epidemiology of human influenza. However, contact infection with influenza viruses was experimentally evidenced in mink only with H3N2 (5). In the present study, avian as well as mammalian influenza A viruses were examined whether they could be transmitted to mink by contact.

The viruses used were A/duck/Hokkaido/5/77 (Hav 7 N 2) (2), A/budgerigar/Hokkaido/1/77 (Hav 4 Nav 1) (4), A/dunling/Hokkaido/1015/79 (Hav 6 Nav 5) (Submitted for publication), A/Hokkaido/3/78 (H 1 N 1) (Unpublished data), A/New Jersey/8/76 (Hsw 1 N 1) and A/equine/Miami/63 (Heq 2 Neq 2). These viruses were propagated in the allantoic cavity of 11-day-old embryonated chicken eggs for 3 days at 35° C. Allantoic fluids which were harvested and stored at —80° C were used as inocula. The inocula had 108 · 0 EID₅₀ per 0.5 ml.

Female Sapphire mink, 4- to 9-months-old and weighing 600 to 900 g, were used. Before inoculation, the sera were tested for antibodies to the 6 influenza viruses used; no antibodies were detected. Skin temperature was taken twice daily

by electrothermometer. The mink were housed individually in separate cages before contact. Experimental contact infections were carried out as described previously (5); the mink were anaesthetized with ether and received 0.25 ml of the inoculum in each nostril by means of a metallic catheter inserted about 1.0 cm into the nostrils. Each uninfected mink was paired in the same cage with a mink which had been inoculated intranasally with each virus 24 hours before contact.

Nasal swabs were taken daily until 14 days after contact and dropped into 1 ml broth, pH 7.4, containing antibiotics. After mixing, 0.2 ml of the broth was inoculated into the allantoic cavity of 2 11-day-old embryonated eggs. The recovered viruses were identified by the hemagglutination inhibition (HI) test, which was done by micromethods (7).

Avian influenza A virus Hav 7N2 was recovered from 3 (Nos. 14, 16 and 18) of the 4 contact mink (Table 1). The virus recovery from the nasal swabs of the contact mink was positive 2—9 days after contact. Antibody response was positive in these mink at the second week after contact; the maximum HI titers until the third week after contact was 1:64. The remaining mink (No. 20) was not infected. Other avian influenza A viruses, Hav 4 Nav 1 and Hav 6 Nav 5, were not transmitted to mink by contact.

Mammalian influenza A viruses were recovered from contact mink without exception. The virus recovery from the nasal swabs of the contact mink was positive 1—9 days (H1N1), 2—10 days (Hsw1N1) and 2—11 days (Heq2Neq2) after contact. Antibody response against these viruses was positive at the second week; the maximum HI titers until the third week were 1:256 (H1N1 and Hsw1N1) and 1:64 (Heq2Neq2). Clinical signs such as sneezing and nasal discharge were seen only in the mink infected by contact with H1N1 and Hsw1N1 from the second to third days after contact.

Virus	Mink No.	Virus recovery (Days after contact with each donor)	Antibody response
Hav 7 N 2	14 16 18 20	5—9 5—7 2, 4—6, 8 No virus recovered	$egin{array}{lll} + & (64)^a \ + & (64) \ + & (64) \ - & \end{array}$
Hav 4 Nav 1	$egin{array}{c} 22 \\ 24 \\ 26 \\ \end{array}$	No virus recovered No virus recovered No virus recovered	
Hav 6 Nav 5	$\frac{28}{30}$	No virus recovered No virus recovered	
H1N1	$\frac{2}{4}$	3, 5—9 1—9	+ (128) + (256)
Hsw 1 N 1	6 8	2, 3, 6—8 3—10	$^{+}$ (256) $^{+}$ (256)
${ m Heq}2{ m Neq}2$	10 12	2—4, 4—6 3, 5—11	$^{+}$ (64) $^{+}$ (64)

Table 1. Virus recovery and antibody response in contact mink

^a Maximum HI titer until the third week after contact

In all the mink which were intranasally inoculated with the avian and mammalian influenza A viruses, virus recovery was positive from nasal swabs of the inoculated mink 3—11 days after inoculation.

We considered that contact infection resembles the natural transmission of influenza viruses more than any other infection route. This study showed that avian influenza virus Hav7N2 was transmitted to mink by contact; this result may be the first evidence of contact infection of an avian influenza virus in mammals.

Of the three avian origin viruses, Hav7N2, Hav4Nav1, and Hav6Nav5, only Hav7N2 was transmitted to the mink by contact. Mammalian influenza viruses H3N2 (5), Heq2Neq2, H1N1, and Hsw1N1 were transmitted to mink by contact. The subtype of the hemagglutinin Hav7 was antigenically similar to H3 of the Hong Kong virus (3). Recently, Hav7 was grouped into a new H3 group together with H3 and Heq2 (1, 6). It is interesting that only the avian virus possessing Hav7 could infect the mink by contact.

Although natural infection of mammals by avian influenza viruses has not been proved by virus isolation, the present experiment may suggest that Hav7 (or H3) may be circulated easily between mammals and birds.

References

- 1. Hinshaw, V. S., Webster, R. G., Rodriguez, R. J.: Influenza A viruses: Combinations of hemagglutinin and neuraminidase subtypes isolated from animals and other sources. Arch. Virol. **62**, 281—290 (1979).
- Kida, H., Yanagawa, R.: Isolation and characterization of influenza A viruses from wild free-flying ducks in Hokkaido, Japan. Zbl. Bakt. Hyg., I. Abt. Orig. A244, 135—143 (1979).
- 3. LAVER, W. G., WEBSTER, R. G.: Studies on the origin of pandemic influenza. III. Evidence implicating duck and equine influenza viruses as possible progenitors of the Hong Kong strain of human influenza. Virology 51, 383—391 (1973).
- 4. Matsuoka, Y., Kida, H., Yanagawa, R.: Isolation of an influenza virus subtype Hav4Nav1 from a budgerigar. Microbiol. Immunol. 23, 35—38 (1979).
- 5. MATSUURA, Y., YANAGAWA, R., NODA, H.: Experimental infection of mink with influenza A viruses. Arch. Virol. 62, 71—76 (1979).
- SCHILD, G. C., NEWMAN, R. W., WEBSTER, R. G., MAJOR, D., HINSHAW, V. S.: Antigenic analysis of influenza A virus surface antigens: Considerations for the nomenclature of influenza virus. Arch. Virol. 63, 171—184 (1980).
- Sever, J. L.: Application of a microtechnique to viral serological investigations. J. Immunol. 88, 320—329 (1962).
- 8. WEBSTER, R. G.: On the origin of pandemic influenza viruses. Curr. Top. Microbiol. Immunol. 59, 75—105 (1972).
- WEBSTER, R. G., LAVER, W. G.: Studies on the origin of pandemic influenza.
 I. Antigenic analysis of A₂ influenza viruses isolated before and after the appearance of Hong Kong influenza using antisera to the isolated hemagglutinin subunits. Virology 48, 433—444 (1972).

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