Reproducibility of glycaemic thresholds for activation of counterregulatory hormones and hypoglycaemic symptoms in healthy subjects

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Summary. Nine healthy subjects were studied on two separate occasions, at least two weeks apart, using the glucose clamp technique to produce a gradual hypoglycaemia. Glucose thresholds for neuroendocrine and symptom responses varied up to 1.5 mmol/l between subjects. There was a significant correlation between individual glucose thresholds on day 1 and 2 for adrenaline (p = 0.0008), growth hormone (p = 0.007) and pancreatic polypeptide (p = 0.02), and for autonomic (p = 0.018) and neuroglycopoenic (p = 0.023) symptoms, whereas no significant correlations were found

Regulation of glycaemic thresholds for activation of counterregulatory hormones and symptoms is not well understood, but it appears that daily glucose levels are of importance. Accordingly, Amiel et al. [1] reported that a period of intensive insulin therapy significantly increased the glycaemic thresholds (i.e. lower blood glucose was needed to elicit the hypoglycaemic responses) in a group of young Type 1 (insulin-dependent) diabetic patients, and Mokan et al. [2] found the thresholds to be at a plasma glucose level approximately 1 mmol/l higher in diabetic patients with poor compared to those with good glycaemic control. Similarly, in a case report on a patient with an insulinoma [3], the glucose threshold for adrenaline changed from 2.6 to 3.7 mmol/l when the blood glucose level was normalized following removal of the tumour.

However, average daily blood glucose levels cannot be the only factor regulating the threshold for hypoglycaemic responses. In a group of patients with functional hypoglycaemia, normal HbA_{1c} and normal blood glucose levels on self-testing, the hypoglycaemic threshold was found to be lower than in control subjects in a study by Snorgaard et al. [4].

When testing whether the rate of glucose fall affected counterregulation, Amiel et al. [5] found that healthy subjects exhibited a wide range in glucose thresholds for adrenaline release. Whether this apparent variation in hypoglycaemic thresholds between healthy subjects truly for glucagon and cortisol. The mean intra-individual differences in glucose thresholds between day 1 and 2 were 0.22 mmol/l for the hormones and 0.25 mmol/l for the symptoms. We conclude that healthy subjects differ in hypoglycaemic thresholds, and that the difference reflects individual variation.

Key words: Autonomic symptoms, counterregulatory hormones, glucose thresholds, hypoglycaemia, neuroglycopoenia.

reflects individual differences, or is simply the result of inaccurate methods, is not known.

The purpose of this study was, therefore, to look at the reproducibility of hypoglycaemic thresholds for activation of neuroendocrine responses and symptoms in healthy subjects by comparing two consecutive studies using the glucose clamp technique to produce a gradual decline in blood glucose levels.

Subjects and methods

Nine healthy subjects were studied on two occasions. Both studies were performed identically, at least 2 weeks apart. Data on age, sex, BMI, smoking, and fasting blood glucose levels are given in Table 1. None of the subjects were taking any medication. The subjects were admitted after an overnight fast to the metabolic research laboratory. A cannula for constant infusion of rapidacting insulin $(1.5 \text{ mU} \cdot \text{kg}^{-1} \cdot \text{min}^{-1})$ and 24 % glucose was placed in the antecubital vein of the left arm. A second cannula for blood sampling was inserted into a dorsal hand vein on the right arm. This arm was placed in a heated box (63 °C) to arterialize the venous blood and the intravenous line was kept patent with 0.9 % NaCl (0.5 ml/min).

After a resting and stabilizing period of approximately 30 min, the tests were started between 09.00 and 10.30 hours. Initially the plasma glucose level was stabilized between 4.5 and 5.5 mmol/l for 30 min and subsequently allowed to fall gradually during 240 min to 2.0 mmol/l (giving a rate of decline of 0.5 mmol/l per 40 min). All subjects were informed that the blood glucose level would be lowered, without knowing the level at any stage. If serious clinical symptoms of hypoglycaemia (falling asleep, diplopia, slurred and inH. Vea et al.: Reproducibility of hypoglycaemic thresholds

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Subject	Age (years)	Sex	BMI (kg/m²)	Smoking	Fasting blood glucose Study day 1/day 2 (mmol/l)
1	26	М	20.5	no	5.4/4.9
2	20	М	20.7	no	4.6/5.4
3	23	F	23.4	no	5.2/5.5
4	23	Μ	19.4	no	5.3/5.2
5	27	Μ	21.2	no	5.5/5.5
6	29	Μ	26.0	no	5.5/5.1
7	25	F	21.5	no	5.5/4.9
8	27	F	19.1	yes	4.7/4.7
9	27	F	20.3	no	4.5/4.8

Table 1. Clinical characteristics of the participants

coordinate speech etc.) occurred before the plasma glucose had fallen to 2.0 mmol/l, the insulin infusion was stopped and the blood glucose normalized by glucose infusion. Plasma glucose was measured repeatedly, and the desired glucose level achieved by adjusting the glucose infusion rate accordingly.

Blood samples were drawn and a symptom questionnaire completed at 5-min intervals during the first 30 min and thereafter every 10 min.

Written informed consent was obtained from each subject before participation. The protocol was approved by the regional ethical committee.

Clinical measurements

Plasma glucose was measured at the bedside by the glucose oxidase method on a Beckman glucose analyser (Fullerton, Calif., USA). Blood for plasma adrenaline was collected in tubes containing glutathione, centrifuged at 4 °C, frozen at -70 °C, and later analysed by HPLC [6], with a detection limit of 5.5 pmol/l. Serum growth hormone was analysed by immuno-radiometric assay (IRMA) developed by the Hormone Laboratory, Aker University Hospital, Oslo, Norway. This assay has a coefficient of variation (CV) of 10.0% [7]. Determination of serum cortisol was based on enhanced luminescence (Amerlite Diagnostics Ltd, Amersham, Buckinghamshire, UK), with a CV of 4.3%. Blood for determination of plasma glucagon was collected in ice-chilled tubes containing 10% trasylol/EDTA, centrifuged at 4 °C, frozen, and later analysed by RIA (CV, 12.0%) [8]. Serum pancreatic polypeptide (PP) was frozen and later measured by RIA (CV, 12.0%) [9].

Symptoms

Prior to the study the subjects were given instructions for completing the visual scale analogue symptom questionnaire. Eight symptoms (dizziness, weakness, tiredness, difficulty concentrating, trembling, sweating, pounding heart, anxiety) were scored on a line from 0 (no symptoms) to a maximum of 15 cm. Consistent with the categorization recommended by Hepburn et al. [10] the first four symptoms were considered neuroglycopoenic and the other four were considered autonomic. The order of the symptoms was systematically changed on each questionnaire. The symptoms in each of the two groups were added together to give an autonomic and a neuroglycopoenic score.

Statistical analysis

Glucose thresholds for each hormone or symptom are given as the mean between the plasma glucose level where responses were more than 2 SD above baseline, followed by an unequivocal, sustained increase, and the preceding plasma glucose. In a few cases the preceding plasma glucose level was lower than the mean between the two, and in these cases regarded as the threshold value. Some patients had an initial decrease or spike in the first hormone samples in which case the values which followed were chosen as baseline. Correlations between the glucose thresholds day 1 and day 2 were performed by ordinary least squares regression. Thresholds are given as mean \pm SEM.

Differences in glucose thresholds for hormones and symptoms were evaluated with two-tailed Wilcoxon's rank sum tests and p < 0.05 was considered statistically significant.

Results

A comparable and gradual decrease in plasma glucose was obtained on both study days in all nine subjects (Fig. 1).

Except for growth hormone in three subjects and PP and cortisol in a fourth subject, glucose thresholds for all hormones could be assigned on both days. Similarly, glucose thresholds for autonomic symptoms were found in eight, and thresholds for neuroglycopoenic symptoms in seven of the nine subjects (Fig. 2).

There was a significant correlation between glucose thresholds day 1 and 2 for adrenaline (r = 0.91, p = 0.0008), growth hormone (r = 0.93, p = 0.007) and PP (r = 0.79, p = 0.02), and for autonomic (r = 0.80, p = 0.018) and neuroglycopoenic symptoms (r = 0.82, p = 0.023). There was a non-significant correlation for glucagon (r = 0.62, p = 0.074) and cortisol (r = 0.40, p = 0.33). The mean difference between the glycaemic thresholds day 1 and 2 was 0.22 ± 0.03 mmol/l for the hormones and 0.25 ± 0.05 mmol/l for the symptoms.

Dividing the group by sex still demonstrated a positive correlation between thresholds day 1 and 2 for all hormones and symptoms.

No correlations were seen between initial plasma glucose levels and glucose thresholds, or between body mass index and glucose thresholds.

The thresholds for the hormones differed, being $3.8 \pm 0.11 \text{ mmol/l}$ for adrenaline, $3.6 \pm 0.08 \text{ mmol/l}$ for cortisol, $3.7 \pm 0.12 \text{ mmol/l}$ for growth hormone, $3.7 \pm 0.10 \text{ mmol/l}$ for glucagon and $3.5 \pm 0.10 \text{ mmol/l}$ for PP (the two days combined). The mean cortisol and PP thresholds on both days were at a significantly (p < 0.02) lower plasma glucose level than the adrenaline threshold.

The mean threshold for neuroglycopoenic symptoms $(3.4 \pm 0.14 \text{ mmol/l})$ was not at a significantly lower plasma glucose level than that for the autonomic symptoms $(3.6 \pm 0.11 \text{ mmol/l})$.

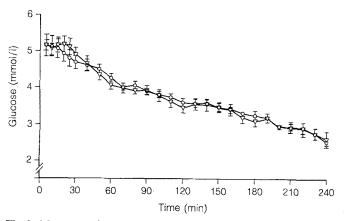


Fig.1. Mean arterialized venous plasma glucose concentrations $(\pm SEM)$ during the hyperinsulinaemic clamp in nine diabetic patients on day $1(\bigcirc -\bigcirc)$ and day $2(\bigtriangledown -\neg \bigtriangledown)$

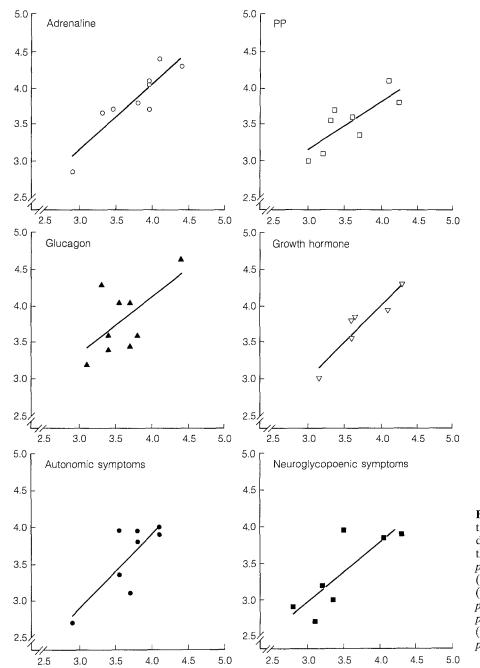


Fig. 2. Correlation between hypoglycaemic thresholds (plasma glucose level in mmol/l) day 1 (along the abscissa) and day 2 (along the coordinate) for adrenaline (r = 0.91, p = 0.0008), pancreatic polypeptide (PP) (r = 0.79, p = 0.02), growth hormone (r = 0.93, p = 0.007), glucagon (r = 0.62, p = 0.074), autonomic symptoms (r = 0.80, p = 0.018) and neuroglycopoenic symptoms (r = 0.82, p = 0.023). Cortisol (r = 0.40, p = 0.33) is not illustrated

Discussion

To our knowledge this is the first study examining the reproducibility of the glucose clamp technique to determine glucose thresholds for hypoglycaemic responses.

The method used, a constant insulin infusion and adjusted glucose infusion after blood glucose measurements every 5 to 10 min, should ideally allow plasma glucose to fall gradually, as does a straight line. Of course, this is not always the case, and is highly dependent on the expertise of the investigator monitoring the clamp. Usually a decline in plasma glucose of approximately 0.1 mmol/l per 10 min is obtained, but occasional levelling off or sharp drops do occur. In our hands the method gave reproducible results with a mean difference in glycaemic thresholds between the two separate tests of 0.2 mmol/l. The problems associated with trying to achieve an ideal blood glucose fall is the most likely cause for this intra-individual difference.

The possible influence of hormone assay variation appears to be minor, since at the threshold point the increase in hormone levels usually far exceeded the baseline plus 2 SD. Thus, the modified glucose clamp technique is a suitable tool for further studies on factors affecting the hypoglycaemic thresholds.

Furthermore, healthy individuals do differ in the glycaemic thresholds both for counterregulatory hormone release and recognition of hypoglycaemic symptoms. These differences should not be ascribed to method inaccuracy.

The reason for this individual variation is difficult to explain. Firstly, individual differences as the result of study group heterogenicity should be ruled out. After our study was completed, two abstracts have been published showing that males have hypoglycaemic thresholds at

H. Vea et al .: Reproducibility of hypoglycaemic thresholds

higher glucose levels than females (regardless of menstrual phase) [11, 12]. The same tendency towards thresholds at lower glucose levels in females was also seen in our study, and thus may have influenced the level of correlation between the two days. However, a possible sex difference does not alter our conclusion, since a positive correlation between thresholds on day 1 and 2 was found for all the hormones and symptoms even after dividing the group into males and females.

Secondly, differences in daily blood glucose levels could, as in diabetic patients [1,2], possibly result in different glycaemic thresholds. Although fasting blood glucose levels showed no correlation with the glycaemic thresholds, multiple blood glucose determinations for several days would be needed to rule out this possibility.

Our slow fall clamp technique results in glucose thresholds where the effect of time to elicit a response has not been taken into account. Thus, if the blood glucose had been clamped for some time at a level above the "threshold", a significant response might have occurred. Furthermore, the slow glucose fall may possibly cause cerebral adaptation to the hypoglycaemia giving higher thresholds that what might have been found with a more rapid blood glucose fall. However, the thresholds found in our study are similar, or even occur at somewhat higher plasma glucose levels than those reported by others [13, 14], and the above-mentioned factors are probably not of major importance when determining the glucose thresholds.

The insulin concentration may affect the symptoms and hormonal responses to hypoglycaemia as shown by Kerr et al. [15]. In their study the two insulin doses used to show this effect differed three-fold and did not affect the adrenaline response. Unfortunately, insulin levels were not measured in our study. The insulin dose was given according to weight and therefore should result in fairly similar circulating levels. Although we cannot rule out an effect of different insulin levels on the inter-individual threshold differences, we believe this effect to have been minor, and for adrenaline non-existant.

The clamp technique used in the present study is not primarily designed to evaluate the magnitude of the neuroendocrine responses. Usually this is tested by injection of a standard dose of insulin [16, 17], or by clamping the blood glucose at a fixed, low level [18]. Since the insulin sensitivity and the hypoglycaemic threshold differs between individuals, these methods will give a hypoglycaemic stimulus that is very differently experienced. Accordingly, when comparing groups for neuroendocrine responses to hypoglycaemia with these methods, the hypoglycaemic thresholds should be similar if the comparison is to be valid.

In conclusion, we have found the present application of the glucose clamp technique to give reproducible results, revealing individual differences in glycaemic thresholds of neuroendocrine responses and symptoms to hypoglycaemia.

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