

Review

Acute and chronic effects of hyperglycaemia on glucose metabolism

H. Yki-Järvinen

Second Department of Medicine, Helsinki University, Helsinki, Finland

Summary. In normal man, several hormonal and metabolic adjustments allow the maintenance of the blood glucose concentration within narrow limits. Hyperglycaemia participates in this regulation via stimulation of glucose disposal and inhibition of glucose production. The effects are mediated, in addition to changes in insulin and glucagon secretion, by the mass-action effect of glucose. In both Type 1 (insulin-dependent) and Type 2 (non-insulin-dependent) diabetic patients, hyperglycaemia, by mass-action abnormally elevates the basal glucose utilization rate but compensates for reduced postprandial insulin-stimulated glucose disposal. When exposed to chronic hyperglycaemia, the body tissues seem to protect themselves, at least partly, against excessive glucose utilization. These protective mechanisms include both a reduction in insulin stimulated glucose disposal and insulin secretion. Chronic hyperglycaemia may also reduce non-in-

sulin-dependent glucose utilization, at least in rats. In Type 1 diabetic patients with normal peripheral insulin concentrations, chronic hyperglycaemia per se could be a major cause of insulin resistance. In Type 2 diabetic patients, insulin resistance is often already present before the development of overt fasting hyperglycaemia. At the diabetic stage, hyperglycaemia could, however, maintain a self-perpetuating cycle, where the deleterious effects of high glucose concentrations on insulin action and secretion cause further deterioration of glycaemic control. The biochemical basis for hyperglycaemia-induced insulin resistance is still far from clear, but could involve changes in the glucose transporter number and gene expression.

Key words: Glucose, insulin, diabetes, glucose transport, insulin resistance.

In 1985, in this journal, Unger and Grundy proposed that chronic hyperglycaemia could be an inducer as well as a consequence of both impaired Beta cell function and insulin resistance in diabetic patients [1]. In recent years, this hypothesis has been experimentally tested in studies carried out both in vivo and in vitro. In contrast to the inhibitory effects of chronic hyperglycaemia on both glucose utilization and insulin secretion, acute hyperglycaemia stimulates, by the mass-action effect of glucose, both of these processes. Stimulation of glucose utilization by hyperglycaemia has important implications with respect to the understanding of mechanisms maintaining hyperglycaemia in diabetic patients. The ensuing discussion is focussed on reviewing our present knowledge of the effects of acute and chronic hyperglycaemia on glucose metabolism. The clinical significance of these data for the treatment of Type 1 (insulin-dependent) and Type 2 (non-insulin-dependent) diabetic patients will also be addressed.

Acute effects of hyperglycaemia on glucose metabolism

Glucose utilization

In normal subjects, the effect of hyperglycaemia per se on glucose utilization can be studied, in the virtual absence of insulin and at different insulin concentrations, by em-

ploying simultaneous infusions of glucose, insulin, and somatostatin. Somatostatin blocks endogenous insulin release as well as growth hormone and glucagon secretion [2]. When measured during inhibition of endogenous insulin release by somatostatin, whole body glucose utilization is stimulated in a glucose-concentration dependent manner [3, 4] both in the absence and presence of insulin (Fig. 1). Within the physiological range of insulin concentrations, doubling the plasma glucose concentration will approximately double glucose utilization (Fig. 1). The ability of glucose to promote glucose utilization depends on the insulin concentration; an increase in the plasma glucose concentration from 5 to 9 mmol/l at insulin concentrations of 9 (fasting), 50 (postprandial), 160 and 1800 mU/l will increase glucose utilization by ~1.8, ~2.7, ~6.6 and ~11.5 mg/kg fat free mass · min in normal subjects [3]. The glucose-induced increases in glucose utilization are quantitatively significant compared to those induced by insulin alone under normoglycaemic conditions (Fig. 1). The increments in glucose utilization in response to acute increments in plasma glucose appear normal in Type 1 diabetic patients, in whom the insulin-induced increase in glucose utilization is reduced [5]. The normal glucose-mediated glucose uptake in these patients has been interpreted to suggest normal function of glucose transporters with the decrease in insulin-mediated glucose uptake being due to a reduced total number of glu-

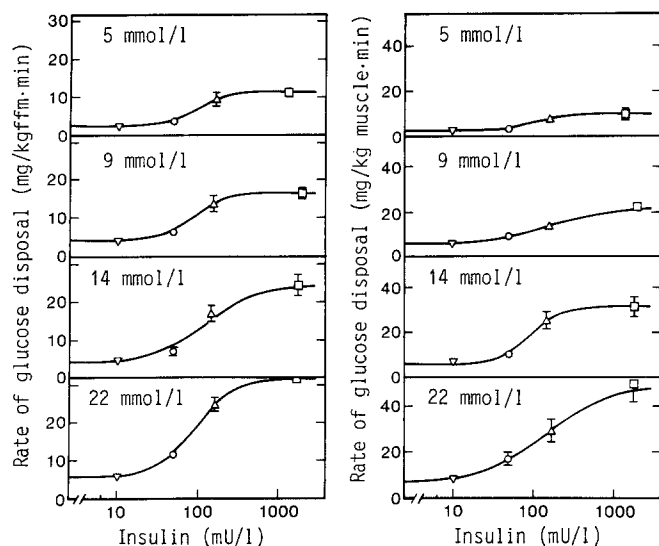


Fig. 1. Effect of glucose and insulin on whole body (left) and forearm glucose uptake (right) in normal subjects [3]

glucose transporters [5], impaired mobilization of transporters from an intracellular pool to the cell surface membrane [6], or perhaps to impaired transporter activation in the plasma membrane of the cell [7].

Under intravenously maintained normoglycaemic hyperinsulinaemic conditions, muscle tissue is responsible for most (60–80%) of the glucose utilization [3, 8]. In response to hyperglycaemia, glucose uptake in muscle increases in a similar dose-dependent fashion as at the whole body level (Fig. 1). Hyperglycaemia induced by intravenous glucose is also capable of enhancing splanchnic glucose uptake in man [9, 10]. This increase is explained by increased delivery of glucose to the splanchnic area rather than an increase in the intrinsic ability of the splanchnic bed to retain glucose [10]. It has been reported that insulin is required for stimulation of splanchnic glucose uptake by hyperglycaemia and that net splanchnic glucose uptake during intravenously induced hyperglycaemia accounts for only ~5% of total glucose uptake by all tissues in the body [9, 10].

After hyperglycaemia induced by oral glucose, net splanchnic glucose uptake is ~6-fold higher than after intravenous glucose, when peripheral glucose concentrations are maintained identical [10]. This difference is not explained by differences in the portal or peripheral insulin concentrations, but may be related to activation of the autonomic nervous system, and/or to a greater portal-arterial glucose concentration difference during oral than intravenous glucose [11].

Hyperglycaemia stimulates both oxidative and non-oxidative glucose disposal in normal subjects [12]. When compared at matched rates of glucose utilization, induced either by hyperinsulinaemia or hyperglycaemia, glucose is less potent than insulin in stimulating glucose oxidation under physiological conditions [12]. The superiority of insulin in promoting glucose oxidation seems to be related to its antilipolytic effect [12]. Even a small increment in serum insulin promptly suppresses plasma non-esterified fatty acids and lipid oxidation and this can increase glu-

ucose oxidation [13]. On the other hand, hyperglycaemia probably has no antilipolytic effect *in vivo* [12]. At higher matched rates of glucose disposal (e.g. insulin 50 mU/l and glucose 24 mmol/l vs insulin 1700 mU/l and glucose 5 mmol/l), glucose oxidation becomes entirely dependent on intracellular glucose availability and then hyperglycaemia and hyperinsulinaemia become equipotent stimulators of glucose oxidation [12].

Intracellularly, the acute stimulatory effect of hyperglycaemia on muscle glucose utilization through oxidative and non-oxidative pathways could be explained by simple mass-action. In contrast to insulin, which stimulates muscle glucose oxidation and glycogen synthesis via changes in the phosphorylation state of key regulatory enzymes (e.g. dephosphorylation of glycogen synthase and pyruvate dehydrogenase, [14, 15]), hyperglycaemia increases muscle glycogen content without dephosphorylation of glycogen synthase in humans [14, 16]. In the perfused rat liver, however, hyperglycaemia does stimulate glycogen synthase activity and inhibit glycogen phosphorylase activity, independent of, and even in the absence of insulin [17].

Non-oxidative glycolysis, as determined from net muscle lactate release, is not stimulated by hyperglycaemia [18–20]. However, the plasma lactate concentration is highly correlated with the rate of whole body glucose disposal under conditions where hepatic glucose production is suppressed, and can be increased by hyperglycaemia, independent of insulin [18]. The most plausible explanation for these data is that hyperglycaemia stimulates non-oxidative glycolysis in tissues other than muscle [18].

Glucose production

Hyperglycaemia, independent of insulin and glucagon, inhibits hepatic glucose production [3, 9]. Half-maximal inhibition of hepatic glucose production is achieved at a glucose concentration of 9 mmol/l in normal subjects [3]. This glucose concentration could be an underestimation due to problems inherent to isotopic measurement of glucose production [21, 22]. Whether the sensitivity of glycogenolysis and gluconeogenesis to inhibition by hyperglycaemia are similar is presently unclear. In the study of Bell et al. [23], the rate of glucose recycling (a qualitative index of gluconeogenesis), remained unchanged in response to an increase in plasma glucose from 5.3 to 9.7 mmol/l [23]. The rate of glucose recycling at the 9.7 mmol/l glucose concentration was similar to the total rate of hepatic glucose production suggesting that hyperglycaemia was more effective in suppressing glycogenolysis than gluconeogenesis [23]. The rate of the futile cycle at the level of glucose/glucose-6-phosphate is not altered by hyperglycaemia [23, 24].

Consequences of the acute stimulatory effect of hyperglycaemia on glucose disposal in Type 1 and Type 2 diabetic patients

Increased glucose utilization after an overnight fast

The fasting plasma glucose concentration is directly proportional to the rate of endogenous glucose production in both Type 1 and Type 2 diabetic patients [25, 26]. As a

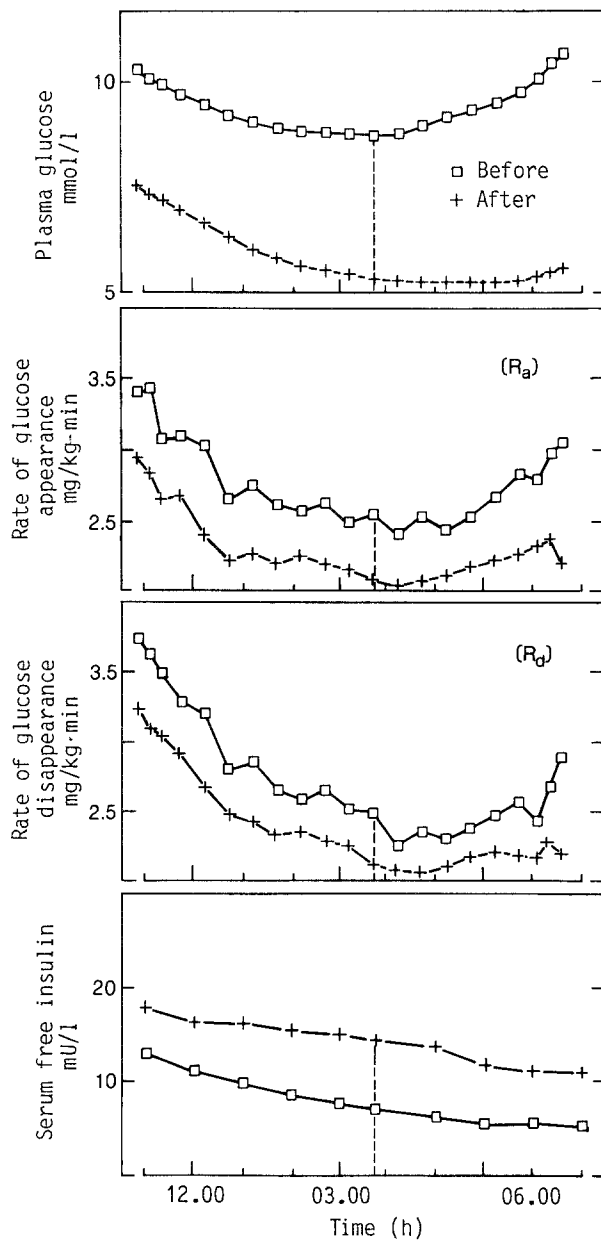


Fig. 2. Example of the mass-action effect of glucose during treatment of patients with Type 2 (non-insulin-dependent) diabetes with insulin. Plasma glucose and serum free insulin concentrations, and rates of glucose appearance and disappearance were measured before (\square) and after ($+$) treatment of Type 2 diabetic patients with evening insulin therapy [28]. Despite a significant increase in the serum free insulin concentration (bottom panel), the rate of glucose disappearance decreased significantly as a consequence of the decrease in plasma glucose caused by the decrease in glucose production

consequence of the mass-action effect of glucose, fasting glycaemia also is positively correlated with the rate of glucose utilization [25, 26]. The rate of basal glucose utilization is increased in both types of diabetes [25, 27]. The effect of evening insulin therapy on glucose production and utilization in Type 2 diabetic patients demonstrates the practical consequence of the glucose mass-action effect. Although insulin stimulates glucose utilization, the

rate of overnight glucose utilization is decreased rather than increased by insulin therapy because inhibition of glucose production decreases the plasma glucose concentration, and the rate of glucose utilization (Fig. 2, [28]). Thus, the inhibitory effect of reduced glucose mass-action on glucose utilization overrides the stimulatory effect of insulin on glucose utilization. These data also imply that fasting hyperglycaemia is the consequence of increased hepatic glucose production rather than impaired glucose utilization. Therefore, therapeutic measures aimed at reducing fasting hyperglycaemia should inhibit hepatic glucose production rather than further increase glucose utilization.

Although in the basal state, the absolute rate of glucose utilization is increased, an impairment in the mass-action effect of glucose could contribute to maintenance of hyperglycaemia in diabetic patients [29, 30]. However, in this context it is important to remember that at low insulin concentrations, 50–60% of glucose is utilized at a fixed rate by the brain [31]. Since the brain does not increase its glucose uptake in response to hyperglycaemia [32], whole body glucose clearance i. e. the rate of glucose utilization divided by the plasma glucose concentration will decrease as the plasma glucose concentration increases [3, 4, 27].

Hyperglycaemia compensates for insulin resistance under postprandial conditions

When measured under normoglycaemic conditions using the insulin clamp technique [33], the ability of insulin to increase total, oxidative and non-oxidative glucose disposal is reduced in both Type 1 [25] and Type 2 [26] diabetic patients. However, if the rate of glucose disposal is determined in Type 1 [13] or Type 2 [34] diabetic patients at glucose concentrations characterizing these patients during their everyday life, glucose utilization is normal. Hyperglycaemia also normalizes the flux through oxidative and non-oxidative pathways in both Type 1 [16] and Type 2 [34] diabetic patients. In Type 1 diabetic patients, normalization of muscle glucose uptake by hyperglycaemia corrects the defect in muscle glycogen synthesis found in these patients under normoglycaemic conditions [16]. This occurs without a change in muscle free glucose or glucose-6-phosphate concentrations indicating that hyperglycaemia overcomes by mass-action a rate-limiting defect at the level of glucose transport [16].

Total body and muscle glucose disposal rates also are normal in Type 2 diabetic patients during an oral glucose tolerance test [33], and in Type 1 diabetic patients after ingestion of a mixed meal [34] and oral glucose [35–37]. In the latter studies, postprandial hyperglycaemia resulted from impaired suppression of hepatic glucose production [35–37]. These studies indicate that hyperglycaemia and the enhanced glucose utilization by glucose mass-action compensate for the reduction in the efficiency of glucose disposal due to insulin resistance in hyperglycaemic Type 1 and Type 2 diabetic patients.

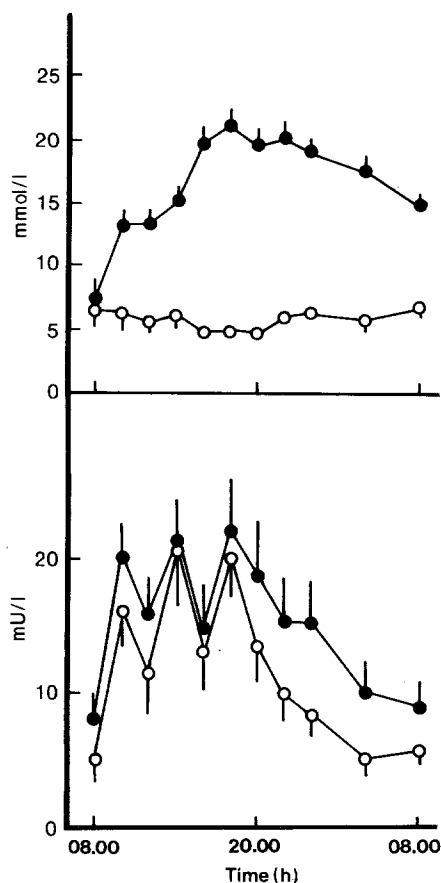


Fig. 3. Plasma glucose (top panel) and free insulin concentrations (bottom panel) during induction of hyperglycaemia by an intravenous glucose infusion in 10 Type 1 diabetic patients treated with continuous subcutaneous insulin infusion therapy. During the control day, the patients received identical diets and insulin doses as during the hyperglycaemic day, with being infused instead of glucose [3]

Chronic effects of hyperglycaemia on glucose metabolism

In 1987, evidence was first presented for the ability of hyperglycaemia to induce insulin resistance in Type 1 diabetic patients [38]. Similar conclusions were reached from studies in partially pancreatectomized rats [39].

Type 1 diabetic patients

Insulin-stimulated glucose uptake was measured in 10 Type 1 diabetic patients treated with continuous subcutaneous insulin infusion on two occasions, after 24 h of hyperglycaemia induced by an intravenous glucose infusion (mean diurnal glucose 16 mmol/l) and after 24 h of normoglycaemia (6 mmol/l, [38], Fig. 3). After 24 h of hyperglycaemia, the rate of insulin-stimulated glucose uptake was consistently lower than after the normoglycaemic period demonstrating that short-term hyperglycaemia can induce insulin resistance in humans ([38], Fig. 4).

Diabetic rats

In the study by Rossetti et al. [39] diabetes was induced in rats by removing 90% of the pancreas. The diabetic state was associated with a 30% reduction in insulin-mediated glucose uptake. Hyperglycaemia was treated (without altering the circulating insulin concentrations) by phlorizin, which inhibits glucose reabsorption by the proximal tubuli. Treatment with phlorizin normalized insulin sensitivity in the diabetic rats. The hyperglycaemia induced insulin resistance seemed restricted to impaired stimulation of glucose uptake and did not include suppression of glucose production by insulin [39]. On the other hand, hepatic insulin resistance cannot be conclusively excluded, as insulin sensitivity was measured using an insulin dose which caused near-maximal suppression of glucose production.

Mechanisms of hyperglycaemia-induced insulin resistance

The exact biochemical mechanism by which chronic hyperglycaemia reduces glucose uptake is unclear. In vitro, pre-exposure of cultured rat skeletal muscle myocytes and myotubes [40, 41] and 3T3-L1 adipocytes [42] to high glucose concentrations, reduces 2-deoxyglucose uptake. The decrease in glucose uptake is due to a reduction in the maximal velocity rather than the affinity of the glucose transport system [40, 42]. Downregulation of the transport system in cultured 3T3-L1 fat cells can be induced, in addition to glucose, by methylglucose and mannose, which are taken up in competition with 2-deoxyglucose but not with the non-competing hexoses galactose and fructose [42]. These data suggest that transport but not further metabolism is required for downregulation of the glucose transport system by hexoses in cultured fat cells [42]. High glucose concentrations do not change muscle ATP and free intracellular glucose concentrations [40, 42]. The glucose-induced downregulation appears to be restricted to the glucose transport system, at least α -aminobutyric acid uptake is not altered by glucose [40].

Over the past 5 years, major advances have been made in understanding the molecular basis for cellular glucose transport [43]. At least four glucose transporter-like proteins have been identified by cDNA cloning techniques. The effect of hyperglycaemia on the glucose transporter gene expression has been studied using cDNA probes recognizing the mRNA of the adipocyte/muscle and the HepG2/brain type glucose transporter. The adipocyte/muscle glucose transporter mRNA is expressed exclusively in insulin-sensitive tissues (muscle, adipose tissue), while the HepG2/brain type glucose transporter is found both in insulin-dependent and insulin-independent tissues [43].

In cultures of the rat brain glial cells, glucose starvation induces increases in glucose transport, glucose transporter protein and the mRNA for the rat brain glucose transporter [44]. Opposite changes are observed when glucose starved cells are exposed to glucose [44]. Chronic hyperglycaemia has also been shown to decrease the glucose transport capacity of the rat blood-brain barrier [45]. In cultured rat skeletal muscle cells, exposure to high glu-

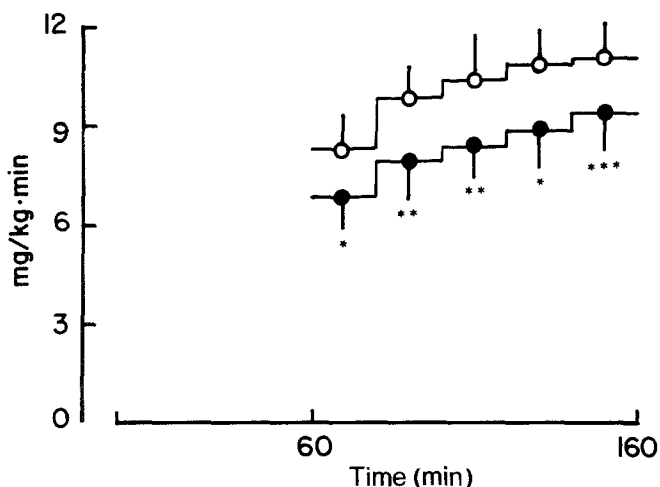


Fig. 4. Rates of glucose metabolism, measured during maintenance of similar glucose and insulin concentrations, after 24 h of hyperglycaemia (●) and after 24 h of normoglycaemia (○, Fig. 3) in 10 Type 1 diabetic patients. The rate of glucose metabolism was significantly lower after hyper- than normoglycaemia, demonstrating induction of insulin resistance by a 24 h exposure to hyperglycaemia

glucose concentrations decreases both glucose uptake, the transporter protein and the mRNA for the rat brain glucose transporter [41]. The increase in glucose transport by insulin is also lower in cells cultured in the presence rather than in the absence of glucose [41]. Preliminary data would suggest that hyperglycaemia does not change the mRNA concentration for the adipose/muscle glucose transporter in cultured rat skeletal muscle cells or 3T3-L1 adipocytes [41].

Contribution of hyperglycaemia-induced insulin resistance to insulin resistance in Type 1 and Type 2 diabetes

Type 1 diabetic patients

Before directly proving that hyperglycaemia can induce insulin resistance in Type 1 diabetic patients, several lines of indirect evidence had suggested this could be the case. First, in both a small ($n = 18$, [46]), and a larger ($n = 53$, [25]) group of insulin-treated C-peptide negative Type 1 diabetic patients, an inverse relationship between long-term glycaemic control as measured by glycosylated haemoglobin (HbA_{1c}) and insulin sensitivity was observed. Since mean peripheral insulin concentrations in insulin-treated Type 1 diabetic patients are usually not lower than those in non-diabetic subjects [47], peripheral hypoinsulinaemia cannot explain reduced insulin-stimulated glucose uptake by peripheral tissues. In support of this, the mean insulin dose/body weight, insulin antibodies, or residual endogenous insulin secretion did not correlate with insulin action in insulin-treated Type 1 diabetic patients [25]. In addition, previous studies in Type 1 diabetic patients demonstrated that glycaemic control and insulin sensitivity could be improved by continuous subcutaneous insulin infusion therapy without having to increase

the total insulin dose [48, 49]. Combining these indirect data and the direct demonstration of the ability of hyperglycaemia to impair insulin action, it can be postulated that hyperglycaemia per se is a major cause of insulin resistance in Type 1 diabetic patients.

As no portal/peripheral insulin gradient exists in Type 1 diabetic patients, the liver is underinsulinized compared to normal subjects, and therefore overproduces glucose [49]. From these data a pathogenic scheme for insulin resistance in these patients can be postulated: fasting and postprandial portal hypoinsulinization leads to overproduction of glucose by the liver, chronic hyperglycaemia and insulin resistance.

Type 2 diabetic patients

Recent prospective [50–53] and cross-sectional [54, 55] studies have provided evidence that hyperinsulinaemia and insulin resistance precede and predict the subsequent development of Type 2 diabetes. In these studies the defect in insulin action appears to be detectable already in subjects with normal glucose tolerance [50–55]. Therefore, chronic hyperglycaemia is unlikely to be the cause of the initial insulin resistance. Once diabetes has developed, the progressive increase in fasting plasma glucose concentrations seems to be better explained by deterioration of insulin secretion than by worsening of insulin resistance [50]. The progressive deterioration of insulin secretion could be both a cause and a consequence of chronic hyperglycaemia [50, 56]. It has also been proposed that the progression of glucose tolerance from impaired to diabetic could be due to effects of mild chronic hyperglycaemia on beta-cell function [50, 56]. This proposal is, however, presently far from proven and awaits localization of the primary cause of Type 2 diabetes.

Regarding the significance of chronic hyperglycaemia on glucose metabolism in established Type 2 diabetes, one could intuitively assume that it impairs glycaemic control by worsening the pre-existing (inherited?, [57]) insulin resistance. At present, only indirect evidence exists to support this proposal. Several modes of diabetic therapy, including insulin, oral agents, weight loss, and exercise improve both glycaemic control, insulin secretion and insulin sensitivity. However, as all of these treatments may improve insulin sensitivity also by mechanisms not mediated via changes in the blood glucose concentration [26], it is unclear to what extent insulin sensitivity is improved by reduction of chronic hyperglycaemia per se.

References

1. Unger RH, Grundy S (1985) Hyperglycaemia as an inducer as well as a consequence of impaired islet cell function and insulin resistance: implications for the management of diabetes. *Diabetologia* 28: 119–121
2. Gerich JE (1981) Somatostatin. In: Brownlee M, (ed) *Handbook of diabetes mellitus*, vol 1. New York: Garland STRM Press, pp 297–354

3. Yki-Järvinen H, Young AA, Lamkin C, Foley JE (1987) Kinetics of glucose disposal in whole body and across the forearm in man. *J Clin Invest* 79: 1713–1719
4. Gottesman I, Mandarino L, Verdonk C, Rizza R, Gerich J (1982) Insulin increases the maximum velocity for glucose uptake without altering the Michaelis constant in man. *J Clin Invest* 70: 1310–1314
5. Hansen IL, Cryer PE, Rizza A (1985) Comparison of insulin-mediated and glucose disposal in patients with insulin-dependent diabetes mellitus and in nondiabetic subjects. *Diabetes* 34: 751–766
6. Garvey WT, Huecksteadt TP, Matthaël S, Olefsky JM (1988) Role of glucose transporters in the cellular insulin resistance of type II non-insulin-dependent diabetes mellitus. *J Clin Invest* 81: 1528–1536
7. Baly DL, Horuk R (1987) Dissociation of insulin-stimulated glucose transport from the translocation of glucose carriers in rat adipose cells. *J Biol Chem* 262: 21–24
8. DeFronzo RA, Jacot E, Jequier E, Maeder E, Wahren J, Felber JP (1981) The effect of insulin on the disposal of intravenous glucose. Results from indirect calorimetry and hepatic and femoral venous catheterization. *Diabetes* 30: 1000–1007
9. DeFronzo RA, Ferrannini E, Hendler R, Felig R, Wahren J (1983) Regulation of splanchnic and peripheral glucose uptake by insulin and hyperglycemia in man. *Diabetes* 32: 35–45
10. DeFronzo RA, Ferrannini E (1987) Regulation of hepatic glucose production in humans. *Diabetes/Metabolism Reviews* 3: 415–459
11. Cherrington AD, Stewenson RW, Steiner KE, Davis MA, Myers SR, Adkins BA, Abumrad NN, Williams PE (1987) Insulin, glucagon, and glucose as regulators of hepatic glucose uptake and production in vivo. *Diabetes/Metabolism Reviews* 3: 307–332
12. Yki-Järvinen H, Bogardus C, Howard BV (1987) Hyperglycemia stimulates carbohydrate oxidation in humans. *Am J Physiol* 253: E376–E382
13. Groop LC, Bonadonna RC, Del Prato S, Ratheiser K, Zyck K, Ferrannini E, DeFronzo RA (1989) Glucose and free fatty acid metabolism in noninsulin dependent diabetes mellitus: evidence for multiple sites of insulin resistance. *J Clin Invest* 84: 205–213
14. Yki-Järvinen H, Mott D, Young AA, Stone K, Bogardus C (1987) Regulation of glycogen synthase and phosphorylase activities by glucose, insulin and basal enzyme activity in human skeletal muscle. *J Clin Invest* 80: 95–100
15. Mandarino LJ, Wright KS, Verity LS, Nichols J, Bell JM, Kolterman OG, Beck-Nielsen H (1987) Effects of insulin infusion on human skeletal muscle pyruvate dehydrogenase, phosphofructokinase, and glycogen synthase. Evidence for their role in oxidative and nonoxidative glucose metabolism. *J Clin Invest* 80: 655–663
16. Yki-Järvinen H, Sahlin K, Ren JM, Koivisto VA (1990) Localization of rate-limiting defect for glucose disposal in skeletal muscle of insulin-resistant type 1 diabetic patients. *Diabetes* 39: 157–167
17. Buschiazzo H, Exton JH, Park CR (1970) Effects of glucose on glycogen synthetase, phosphorylase, and glycogen deposition in the perfused rat liver. *Proc Natl Acad Sci* 65: 383–387
18. Yki-Järvinen H, Bogardus C, Foley JE (1990) The rate of glucose disposal in extramuscular tissues: The key regulator of plasma lactate concentration in resting human subjects. *Metabolism* 39: (in press)
19. Kelley D, Mitrakou A, Marsh H, Schwenk F, Benn J, Sonnenberg G, Arcangeli M, Aoki T, Sorensen J, Berger M, Sonksen P, Gerich J (1988) Skeletal muscle glycolysis, oxidation, and storage of an oral glucose load. *J Clin Invest* 81: 1563–1571
20. Jackson R, Roshnia R, Hawa M, Sim B, DiSilvio L (1986) Impact of glucose ingestion on hepatic and peripheral glucose metabolism in man: an analysis based on simultaneous use of the forearm and double isotope techniques. *J Clin Endocrinol Metab* 63: 541–549
21. Bergman R, Finegood DT, Ader M (1985) Assessment of insulin-sensitivity in vivo. *Endocrine Rev* 6: 45–86
22. Yki-Järvinen H, Consoli A, Nurjahan N, Young AA, Gerich JE (1989) Mechanism for underestimation of isotopically determined glucose disposal. *Diabetes* 38: 744–751
23. Bell POM, Firth RG, Rizza RA (1986) Effects of hyperglycemia on glucose production and utilization in humans. Measurement with (^2H)-, (^3H)-, and (^6C)glucose. *Diabetes* 35: 642–648
24. Efendic S, Wajngot A, Vranic M (1985) Increased activity of the glucose cycle in liver: early characteristic of type II diabetes. *Proc Natl Acad Sci USA* 82: 2965–2969
25. Yki-Järvinen H, Koivisto VA (1986) Natural course of insulin resistance in type 1 diabetes. *N Engl J Med* 315: 224–230
26. DeFronzo RA, Ferrannini E, Koivisto VA (1983) New concepts in the pathogenesis and treatment of noninsulin-dependent diabetes mellitus. *Am J Med* 74: 52–81
27. Gerich JE, Mitrakou A, Kelley D, Mandarino L, Nurjhan N, Reilly J, Jenssen T, Veneman T, Consoli A (1990) Contribution of impaired muscle glucose clearance to reduced postabsorptive systemic glucose clearance in NIDDM. *Diabetes* 39: 211–216
28. Yki-Järvinen H, Helve E, Sane T, Taskinen M-R (1989) Insulin inhibition of overnight glucose production and gluconeogenesis from lactate in NIDDM. *Am J Physiol* 256: E732–E739
29. DeFronzo RA, Ferrannini E, Simonson DC (1989) Fasting hyperglycemia in non-insulin-dependent diabetes mellitus: contributions of excessive hepatic glucose production and impaired glucose uptake. *Metabolism* 38: 387–395
30. Chen Y-D, Jeng CY, Hollenbeck C, Wu NS, Reaven G (1988) Relationship between plasma glucose and insulin concentration, glucose production, and glucose disposal in normal subjects and patients with noninsulin-dependent diabetes. *J Clin Invest* 82: 21–25
31. Scheinberg P, Stead EA Jr (1949) Cerebral blood flow in male subjects as measured by the nitrous oxide technique: normal values for blood flow, glucose utilization, and peripheral resistance, with observations of the effects of tilting and anxiety. *J Clin Invest* 28: 1168–1171
32. Brooks D, Gibbs J, Sharp P, Herold S, Turton D, Luthra S, Kohner E, Bloom S, Jones T (1986) Regional cerebral glucose transport in insulin-dependent diabetic patients studied using (^{11}C)-3-O-methyl-D-glucose and position emission tomography. *J Cereb Blood Flow Metab* 6: 240–244
33. DeFronzo RA, Tobin JD, Andres R (1979) Glucose clamp technique: a method for quantifying insulin secretion and resistance. *Am J Physiol* 237: E214–E224
34. Henry RR, Gumbiner B, Flynn T, Thorburn AW (1990) Metabolic effects of hyperglycemia and hyperinsulinemia on fate of intracellular glucose in NIDDM. *Diabetes* 39: 149–156
35. Firth R, Bell P, Marsh H, Hanse I, Rizza R (1986) Postprandial hyperglycemia in patients with noninsulin-dependent diabetes mellitus: roles of hepatic and extrahepatic tissues. *J Clin Invest* 77: 1525–1532
36. Pehling G, Tessari P, Gerich JE, Haymond MW, Service FJ, Rizza RA (1984) Abnormal meal carbohydrate disposition in insulin-dependent diabetes. Relative contributions of endogenous glucose production and initial splanchnic uptake and effect of intensive insulin therapy. *J Clin Invest* 74: 985–991
37. Benn JJ, Bozzard SJ, Kelley D, Mitrakou A, Aoki T, Sorensen J, Gerich J, Sonksen PH (1989) Persistent abnormalities of the metabolism of an oral glucose load in insulin-treated type I diabetics. *Metabolism* 38: 1047–1055
38. Yki-Järvinen H, Helve E, Koivisto VA (1987) Hyperglycemia decreases glucose uptake in type I diabetes. *Diabetes* 36: 892–896
39. Rossetti L, Smith D, Shulman GI, Papachristou D, DeFronzo RA (1987) Correction of hyperglycemia with phlorizin normalized tissue sensitivity to insulin in diabetic rats. *J Clin Invest* 79: 1510–1515
40. Sasson S, Cerasi E (1986) Substrate regulation of the glucose transport system in rat skeletal muscle. Characterization and kinetic analysis in isolated soleus muscle and skeletal muscle cells in culture. *J Biol Chem* 261: 16827–16833

41. Walker PS, Ramlal T, Donovan JA, Doering TP, Sandra A, Klip A, Pessin JE (1989) Insulin and glucose-dependent regulation of the glucose transport system in the rat L6 skeletal muscle cell line. *J Biol Chem* 264: 6587–6595
42. van Putten JPM, Krans HMJ (1985) Glucose as a regulator of insulin-sensitive hexose uptake in 3T3 adipocytes. *J Biol Chem* 260: 7996–8001
43. Mueckler M (1990) Family of glucose-transporter genes. Implications for glucose homeostasis and diabetes. *Diabetes* 39: 6–11
44. Walker PS, Donovan JA, van Ness BG, Fellows RE, Pessin JE (1988) Glucose-dependent regulation of glucose transport activity, protein, and mRNA in primary cultures of rat brain glial cells. *J Biol Chem* 263: 15 594–15 601
45. Gjedde A, Crone C (1981) Blood-brain glucose transfer: repression in chronic hyperglycemia. *Science* 214: 456–457
46. Yki-Järvinen H, Taskinen M-R, Kiviluoto T, Hilden H, Helve E, Koivisto VA, Nikkilä EA (1984) Site of insulin resistance in type 1 diabetes: insulin-mediated glucose disposal in vivo in relation to insulin binding and action in adipocytes in vitro. *J Clin Endocrinol Metab* 59: 1183–1192
47. Rizza RA, Gerich JE, Haymond MW, Westland RE, Hall LD, Clemens AH, Service FJ (1980) Control of blood sugar in insulin-dependent diabetes: Comparison of an artificial pancreas, continuous subcutaneous insulin infusion, and intensified conventional insulin therapy. *N Engl J Med* 303: 1313–1318
48. Lager I, Lonnroth P, von Schenk H, Smith U (1983) Reversal of insulin resistance in type I diabetes with continuous subcutaneous insulin infusion. *Br Med J* 287: 1661–1664
49. Yki-Järvinen H, Koivisto VA (1984) Continuous subcutaneous insulin infusion therapy decreases insulin resistance in type I diabetes. *J Clin Endocrinol Metab* 58: 659–666
50. Lillioja S, Mott DM, Howard BV, Bennett PH, Yki-Järvinen H, Freymond D, Nyomba BL, Zurlo Z, Swinburn B, Bogardus C (1988) Impaired glucose tolerance as a disorder of insulin action. Longitudinal and cross-sectional studies in Pima Indians. *N Engl J Med* 318: 1217–1225
51. Saad MF, Knowler EC, Pettitt DJ, Nelson RG, Mott DM, Bennett PH (1988) The natural history of impaired glucose tolerance in the Pima Indians. *N Engl J Med* 319: 1500–1506
52. Sicree RA, Zimmet PZ, King HOM, Coventry JS (1987) Plasma insulin response among Nauruans. Prediction of deterioration in glucose tolerance over 6 yr. *Diabetes* 36: 179–186
53. Warram J, Martin B, Soeldner JS (1988) Diminished insulin sensitivity in normoglycemic offspring of two non-insulin-dependent diabetic parents. *Diabetes* 37: 9A (Abstract)
54. Haffner SM, Stern MP, Hazuda HP, Mitchell BD, Patterson JK (1988) Increased insulin concentrations in nondiabetic offspring of diabetic parents. *N Engl J Med* 319: 1297–1301
55. Eriksson J, Franssila-Kallunki A, Ekstrand A, Saloranta C, Widen E, Schalin C, Groop L (1989) Early metabolic defects in persons at increased risk for non-insulin-dependent diabetes mellitus. *N Engl J Med* 321: 337–343
56. Leahy JL, Cooper HE, Deal DA, Weir GC (1986) Chronic hyperglycemia is associated with impaired glucose influence on insulin secretion. A study in normal rats using chronic in vivo glucose infusions. *J Clin Invest* 77: 908–915
57. Bogardus C, Lillioja S, Nyomba BL, Zurlo F, Swinburn B, Espósito-Del Puente A, Knowler WC, Ravussin E, Mott DM, Bennett PH (1989) Distribution of in vivo insulin action in Pima Indians as mixture of three normal distributions. *Diabetes* 38: 1423–1432

H. Yki-Järvinen, M. D.
Second Department of Medicine
Helsinki University
Haartmaninkatu 4
SF-00290 Helsinki
Finland