Transcription factors HSF and NF-κB as targets for cytoprotective eicosanoids: a new strategy for therapeutic intervention

M. G. SANTORO

The observation that an increase in temperature of a few degrees above the physiological level induces the synthesis of a small number of proteins in *Drosophila* salivary glands led to the discovery of a universal protective mechanism which prokaryotic and eukaryotic cells utilize to preserve cellular function and homeostasis¹. This complex physiological defence mechanism, known as the heat shock response, involves the rapid induction of a specific set of genes encoding cytoprotective proteins (heat shock protein, HSP). In mammalian cells HSP synthesis is induced in a wide variety of toxic conditions, including extreme temperatures, oxidative stress, exposure to heavy metals or cytotoxic drugs, glucose deprivation and virus infection¹. Whereas HSP induction was at first interpreted as a signal for detection of physiological stress, it is now well-documented that HSPs are utilized by the cells in the repair process following different types of injury, to prevent damage resulting from the accumulation and aggregation of non-native proteins².

Induction requires the activation and translocation to the nucleus of a transregulatory protein, the heat shock transcription factor (HSF). In mammalian cells, HSF exists as an inactive non-DNA binding form, which is rapidly converted to the DNA-binding form upon exposure to heat shock or other stimuli³. HSF activation is a multistep process and requires oligomerization, acquisition of DNA-binding activity, localization to the nucleus and phosphorylation by an as yet unknown kinase³. In the nucleus HSF binds to specific promoter elements (HSE) located upstream of heat shock (hs) genes, activating transcription. Several HSFs (HSF1–HSF4) have been identified in vertebrate cells ³; however, the molecular mechanisms responsible for signal transduction which leads to HSF activation and phosphorylation have not been completely elucidated.

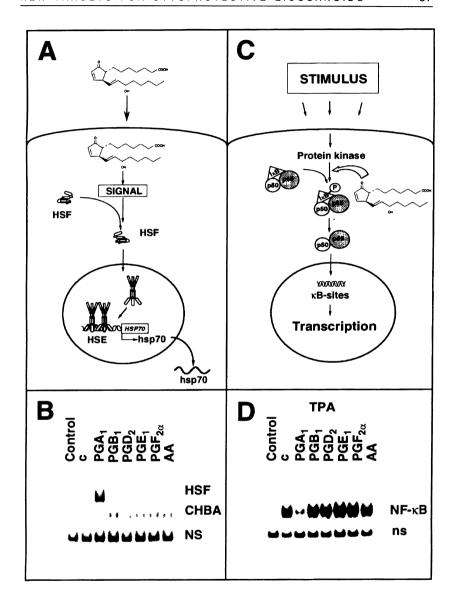
HSP can be divided into five families, depending on their molecular weight. In eukaryotic cells 70 kDa heat shock proteins (hsp 70), which are encoded by a large multigene family, include the constitutively expressed hsc70, the major inducible hsp70, the inducible hsp72, the glucose regulated grp78/BiP and the mitochondrial P75³. In mammalian cells, several HSP that function as molecular chaperones and are essential for a correct folding, assembly and intracellular translocation of proteins, are expressed

during normal growth conditions and can be induced by biologically active molecules such as haemin⁴ and prostaglandins⁵.

CYTOPROTECTIVE ROLE OF HEAT SHOCK PROTEINS IN HUMAN DISEASES

In the last few years, the cytoprotective role of HSP has been described in several human diseases, including metabolic disorders⁶, inflammation⁷, infection⁸ and ischaemia⁹. In cardiac tissues a wide variety of insults, including myocardial ischaemia, trauma and hyperthermia, results in the synthesis of HSP, which have been shown to play a pivotal role in restoring normal cardiac function after injury, possibly by removal of denatured cardiac proteins and re-establishment of normal cardiac protein synthesis. A correlation has been shown between the amount of hsp70 and the degree of myocardial protection in vitro as well as in animal models^{9,10}. Moreover, transfected heart-derived cells overexpressing hsp70 and hsp70-transgenic mice show enhanced resistance to ischaemic stress, providing evidence for a direct role of this protein in cytoprotection after this type of injury^{9,11}. A pharmacological approach to hsp70 induction and cardiac protection is suggested by the recently described hsp70-induced protection by simulated ischaemia in rat neonatal cardiomyocytes treated with the hsp70 inducer herbimycin-A¹². In the case of inflammation, HSP have been shown to protect mammalian cells from tumour necrosis factors α- and β-mediated cytotoxicity¹³, as well as to suppress astroglial-inducible nitric oxide synthase expression¹⁴. Moreover, in a rodent model for adult respiratory distress syndrome (ARDS), pre-exposure of animals to heat shock, leading to massive induction of hsp70 within

Figure 1 Activation of the heat-shock response and inhibition of NF-xB by cyclopentenone prostaglandins. (A) Synthesis of cytoprotective heat-shock proteins is induced by PG via the activation of heat-shock transcription factor HSF. HSF converts from a monomeric non-DNA-binding form to an oligomeric form that translocates to the nucleus and binds to specific promoter elements (HSE) located upstream of heat-shock genes (e.g. HSP70). (B) Effect of arachidonic acid metabolites on HSF activation. Wholecell extracts of Jurkat cells treated with 24 μ M arachidonic acid (AA), PGA, PGB, PGD, PGE, PGF2 α or diluent control (c) for 6 h were incubated with a ³²P-labelled HSE probe, followed by analysis of DNA-binding activity by EMSA (21). Position of HSF-DNA binding complex (HSF), constitutive HSEbinding activity (CHBA) and non-specific protein -DNA interactions (NS) are shown. (C) Inhibition of NF-kB activation. NF-kB normally exists in an inactive cytoplasmic complex, whose predominant form is a heterodimer composed of p50 and p65 subunits, bound to the inhibitory protein kBa. Cyclopentenone prostaglandins act by inhibiting $k B \alpha$ phosphorylation and degradation. (D) Whole-cell extracts from Jurkat cells treated with AA, PGA1, PGB₁, PGD₂, PGE₁, PGF₂a or diluent control (c) for 2 h and then stimulated with 12-O-tetradecanoylphorbol 13-acetate (TPA) for 6 h were incubated with a 32P-labelled kB DNA probe, followed by EMSA. Position of NF-kB-DNA complex (NF-kB) and non-specific binding (ns) are indicated. Control cells are neither stimulated with TPA nor treated with prostaglandins



the lung, is associated with decreased pulmonary inflammation and prevention of death¹⁵. Increased expression of hsp70 has also been associated with inhibition of virus replication during acute infection^{8,16}. These observations suggest new therapeutic strategies relying upon the development of drugs which selectively turn on heat shock genes.

EICOSANOIDS AND THE HEAT SHOCK RESPONSE

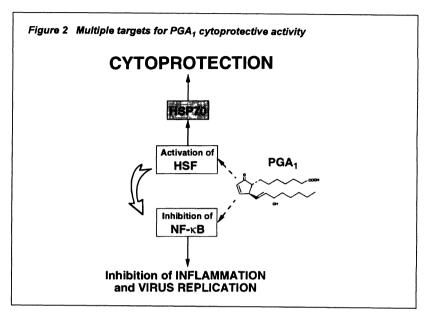
We have shown that cyclooxygenase cyclopentenone metabolites such as prostaglandins of the A and J type (PGAs and PGJs) induce the synthesis of hsp70 in a non-stressful situation in a wide variety of human and mammalian cells (reviewed in 17,18), whereas leukotrienes, and in particular LTB₄, do not activate HSF nor stimulate HSP expression in human and murine cells (Elia G, Santoro MG, unpublished results). The mechanism of hsp70 induction by cyclopentenone prostaglandins via cycloheximide-sensitive activation of HSF1 (Figure 1) and the kinetics of hsp70 gene transcription and hsp70 protein synthesis have been well characterized in human cells¹⁹. The PGJ metabolite 15-deoxy-Δ^{12,14}PGJ₂, which was shown to be the natural ligand for the adipocyte determination factor PPARy inducing adipocyte differentiation²⁰, is also a potent inducer of HSF activation²¹. We have also shown that PGA₁ and PGJ₂ induce the synthesis of the oxidative stress protein haeme oxygenase in murine myoblasts²² and in human monocytes from healthy donors (Elia et al., in preparation). Structure-activity relationship studies have recently shown that HSP induction requires the presence of a reactive α,β -unsaturated carbonyl group in the cyclopentane ring (cyclopentenone), which renders this portion of the molecule able to form Michael's adducts with cellular nucleophilics, and to covalently bind to cysteine residues of proteins²³.

Induction of hsp70 by cyclopentenone prostaglandins is associated with a cytoprotective effect during hyperthermia and virus infection: cyclopentenone prostaglandins induce a thermotolerant state in human cells and protect cells from subsequent lethal injury²⁴. Cyclopentenone prostaglandins are also characterized by a potent antiviral activity against a wide variety of DNA and RNA viruses, including the human immunodeficiency virus (HIV-1), in several experimental models in vitro and in vivo (reviewed in 25). A long-acting synthetic analogue of PGA₂ (di-M-PGA₃) has antiviral activity in a mouse model infected with influenza A virus²⁶. In negative strand RNA virus models prostaglandins of the A and J type provoke a selective and dramatic block of viral protein synthesis^{8,27}. This block is exerted at the translational level and is dependent on hsp70 expression in infected cells. A possible model has been hypothesized in which HSP and virus messages, both of which can be translated in conditions where cellular protein synthesis is impaired, could possess similar mechanisms for preferential translation, and could then compete with each other²⁵. In the case of HIV-1 infection, a single treatment with PGA₁ or PGJ₂ is cytoprotective in lymphoblastoid CEM-SS cells, and causes a more than 1000-fold reduction in infectious virus yield, due to a selective block of HIV-1 mRNA transcription²⁸. Inhibition of HIV-1 RNA transcription appears to be mediated by the recently discovered ability of cyclopentenone PGs to inhibit the activation of nuclear factor- $\kappa B (NF-\kappa B)^{21}$.

NUCLEAR FACTOR K-B

NF-κB is an inducible eukaryotic transcription factor of the rel family, which normally exists in an inactive cytoplasmic complex, whose predominant form is a heterodimer composed of p50 and p65 (Rel A) subunits, bound to inhibitory proteins of the IkB family, usually IκBα, and is activated in response to primary (viruses, bacteria, UV) or secondary (inflammatory cytokines) pathogenic stimuli^{29,30}. Stimulation triggers rapid phosphorylation and degradation of IκBα, resulting in NF-κB translocation to the nucleus, where it binds to DNA at specific kB-sites, rapidly inducing a variety of genes encoding signalling proteins. Target genes include cyclooxygenase-2, nitricoxide synthase (iNOS), several inflammatory and chemotactic cytokines, cytokine receptors, cell adhesion molecules as well as viral genes²⁹. NF-κB is a critical regulator of the immediate early pathogen response and activation of the immune system, and is involved in many pathological events, including progression of AIDS by enhancing HIV-1 transcription³¹. A possible role for NF-κB in atherosclerosis and reoxygenation damage has also been suggested^{32,33}. Consequently NF-kB is an attractive therapeutic target for novel anti-inflammatory, antiviral and cytoprotective drugs. and the need for the development of effective NF-kB inhibitors with therapeutic efficacy is widely recognized.

Several arachidonic acid metabolites appear to interfere with NF-κB activation. 12(R)-hydroxyeicosatrienoic acid [12(R)-HETrE], but not the enantiomer 12(S)-HETrE, was found to markedly activate NF-κB DNA-binding activity in coronary microvessel endothelial cells³⁴. 5-Lipoxygenase inhibitors (nordihydroguaiaretic acid and AA861), but not cyclooxygenase inhibitors, were reported to block IL-1βinduced expression of cell surface adhesion molecules (VCAM-1), via inhibition of NF-κB in human cells³⁵. Leukotriene B₄ was also reported to activate NF-κB and NF-κB-controlled interleukin-6 expression in human blood monocytes³⁶. However, we were unable to detect any significant increase in the constitutive level of DNA-binding activity of NF-kB in human peripheral blood monocytes in the presence of concentrations of LTB₄ ranging from 10⁻¹¹ to 10⁻⁶ M (Elia G, Santoro MG, unpublished data). On the other hand, we have recently reported that cyclopentenone prostaglandins are potent inhibitors of NF-kB activation in human cells, and of NF-kB-dependent HIV-1 transcription in long terminal repeat-chloramphenicol acetyl transferase transient transfection experiments²¹. These eicosanoids act by inhibiting phosphorylation and preventing degradation of the NF- κ B inhibitor $I\kappa B\alpha^{21}$. Inhibition does not require protein synthesis and is dependent on the presence of a reactive cyclopentenonic moiety. Interestingly, the NF-kB inhibitory effect is tightly associated with HSF activation in human cells21. In fact activation of HSF1 by different cyclopentenone prostaglandins, by other chemical inducers, such as sodium arsenite, or by hyperthermia itself was found to prevent or suppress NF-kB activation, indicating a link between the regulatory pathways of these factors and suggesting the possibility that triggering of HSF1 could render cells unresponsive to stimulation of NF-kB. This hypothesis is very attractive considering the opposite roles of HSF and NF-κB in cytoprotection and cell injury respectively, as in the case of inflammation¹⁴ and viral infection^{8, 25, 31} (Figure 2).



PERSPECTIVES AND CONCLUSIONS

These observations encourage the search for novel prostanoids with the ability to induce HSP and inhibit NF- κ B simultaneously, which could be utilized as cytoprotective and antiviral drugs. Prostaglandins are used clinically in the treatment of several diseases, including gastric ulcers and congenital heart disease, and to facilitate labour, and are generally effective and well-tolerated ³⁷. Administration of PGE₁ has also been shown to be beneficial in patients with fulminant viral hepatitis ³⁸. In studies on volunteers with hypertension, infusion with PGA₁ had beneficial effects on blood pressure, while no deleterious effects on kidney function or other significant side-effects were found ³⁹. On the other hand, the fact that different types of PGs are synthesized in different tissues throughout the body and have multiple effects on blood pressure, inflammatory and immune responses, poses several questions on the possible use of natural prostaglandins as cytoprotective compounds.

We have recently reported that one component of the PGA molecule, 2-cyclopenten-1-one, is able to activate HSF and to trigger rapid and selective transcription and translation of the hsp70 gene, leading to the accumulation of high levels of hsp70 protein in human cells²³. As previously shown for other HSP inducers, including antiviral prostaglandins, sodium arsenite, cadmium and hyperthermia itself, 2-cyclopenten-1-one-induced hsp70 synthesis is associated with a selective inhibition of virus protein synthesis, and viral replication²³. The α,β -unsaturated carbonyl group present in 2-cyclopenten-1-one is the molecular structure responsible for HSF activation, since even high concentrations of cyclopentanone (a similar molecule with a saturated carbonyl), or cyclopentene, which contains a double bond, but not a carbonyl

group were unable to induce HSF DNA-binding activity. These results indicate that a class of novel molecules, characterized by the ability to activate HSF while inhibiting NF- κ B, and devoid of the pleiotropic effects of natural PGs could be designed, opening new perspectives for therapeutic intervention in inflammatory and infectious diseases.

Acknowledgements

This work was supported by a grant from the Italian Ministry of Public Health, 1997 AIDS Research project and Multiple Sclerosis project, and CNR P.S. 'Stress Proteins'.

References

- 1. Linquist S, Craig EA. The heat shock proteins. Annu Rev Genet. 1988; 22: 631-77.
- Feige U, Morimoto RI, Yahara I, Polla BS. Stress-Inducible Cellular Responses. Basel-Boston-Berlin: Birkhäuser-Verlag, 1996.
- Morimoto RI, Sarge KD, Abravaya K. Transcriptional regulation of heat shock genes. J Biol Chem. 1992; 267: 21987–90.
- Sistonen L, Sarge KD, Phillips B, Abravaya K, Morimoto RI. Activation of heat shock factor 2 during hemin-induced differentiation of human erythroleukemia cells. Mol Cell Biol. 1992; 12: 4104–11.
- Santoro MG, Garaci E, Amici C. Prostaglandins with antiproliferative activity induce the synthesis of a heat shock protein in human cells. Proc Natl Acad Sci USA. 1989; 86: 8407–11.
- Williams RS, Thomas JA, Fina M, German Z, Benjamin IJ. Human heat shock protein 70 (hsp70) protects murine cells from injury during metabolic stress. J Clin Invest. 1993; 92: 503-8.
- Polla BS, Cossarizza A. Stress proteins in inflammation. In: Feige U, Morimoto RI, Yahara I, Polla BS, editors. Stress-Inducible Cellular Responses. Basel–Boston–Berlin: Birkhäuser-Verlag, 1996: 375–91.
- 8. Amici C, Giorgi C, Rossi A, Santoro MG. Selective inhibition of virus protein synthesis by prostaglandin A: A translational block associated with HSP70 synthesis. J Virol. 1994; 68: 6890-9.
- 9. Mestril R, Chi SH, Sayen MR, O'Reilly K, Dillmann WH. Expression of inducible stress protein 70 in rat heart myogenic cells confers protection against stimulated ischemia-induced injury. J Clin Invest. 1994; 93: 759-67.
- Marber MS, Walker JM, Latchman DS, Yellon DM. Myocardial protection after whole body heat stress in the rabbit is dependent on metabolic substrate and is related to the amount of the inducible 70-KD heat stress protein. J Clin Invest. 1994; 93: 1087-94.
- Plumier J, Ross BM, Currie RW, et al. Transgenic mice expressing the human heat shock protein 70 have improved post-ischemic myocardial recovery. J Clin Invest. 1995; 95: 1854-60
- 12. Morris SD, Cumming DV, Latchman DS, Yellon DM. Specific induction of the 70-kD heat stress protein by the tyrosine kinase inhibitor herbimycin-A protects rat neonatal cardiomyocytes. J Clin Invest. 1996; 97: 706–12.
- 13. Iaattela M, Wissing D, Bauer PA, Li GC. Major heat shock protein hsp70 protects tumor cells from tumor necrosis factor cytotoxicity. EMBO J. 1992; 11: 3507–12.
- Feinstein DL, Galea E, Aquino DA, Li GC, Xu H, Reis DJ. Heat shock protein 70 suppresses astroglial-inducible nitric-oxide synthase expression by decreasing NF-κB activation. J Biol Chem. 1996; 271: 17724–32.
- 15. Villar J, Edelson JD, Post M, Brendan J, Mullen M, Slutsky AS. Induction of heat stress proteins is associated with decreased mortality in an animal model of acute lung injury. Am Rev Resp Dis. 1993; 147 177-81
- Santoro MG. Viral infection. In: Feige U, Morimoto RI, Yahara I, Polla BS, editors. Stress-Inducible Cellular Responses. Basel–Boston–Berlin: Birkhäuser-Verlag, 1996 337–57.

 Santoro MG, Garaci E, Amici C. Induction of HSP70 by prostaglandins. In: Schlesinger MJ, Santoro MG, Garaci E, editors. Stress Proteins: Induction and Function. Berlin: Springer-Verlag, 1990: 27–44.

- 18. Santoro MG. Heat shock proteins and virus replication: Hsp70s as mediators of the antiviral effects of prostaglandins. Experientia. 1994; 50: 1039–47.
- Amici C, Sistonen L, Santoro MG, Morimoto RI. Antiproliferative prostaglandins activate heat shock transcription factor. Proc Natl Acad Sci USA. 1992; 89: 6227–31.
- Kliewer SA, Lenhard JM, Wilson TM, Patel I, Morris DC, Lehmann JM. A prostaglandin J₂ metabolite binds peroxisome proliferator-activated receptor γ and promotes adipocyte differentiation. Cell 1995; 83: 813–19.
- Rossi A, Elia G, Santoro MG. Inhibition of nuclear factor κB by prostaglandin A₁: An
 effect associated with heat shock transcription factor activation. Proc Natl Acad Sci USA.
 1997; 94: 746–50.
- Rossi A, Santoro MG. Induction by prostaglandin A₁ of haem oxygenase in myoblastic cells: An effect independent of expression of the 70 kDa heat shock protein. Biochem J. 1995; 308: 455-63.
- 23. Rossi A, Elia G, Santoro MG. 2-Cyclopenten-1-one, a new inducer of heat shock protein 70 with antiviral activity. J Biol Chem. 1996; 271: 32192-6.
- 24. Amici C, Palamara AT, Santoro MG. Induction of thermotolerance by prostaglandin A in human cells. Exp Cell Res. 1993; 207: 230–4.
- Santoro MG. Antiviral activity of cyclopentenone prostanoids. Trends Microbiol. 1997; 5: 276–81.
- Santoro MG, Favalli C, Mastino A, Jaffe BM, Esteban M, Garaci E. Antiviral activity of a synthetic analog of prostaglandin A in mice infected with influenza A virus Arch Virol. 1988; 99: 89–100.
- 27. Pica F, De Marco A, De Cesare F, Santoro MG. Inhibition of vesicular stomatitis virus replication by Δ^{12} -prostaglandin J_2 is regulated at two separate levels and is associated with induction of stress protein synthesis Antiviral Res. 1993; 20: 193–208.
- Rozera C, Carattoli A, De Marco A, Amici C, Giorgi C, Santoro MG. Inhibition of HIV-1 replication by cyclopentenone prostaglandins in acutely infected human cells. J Clin Invest. 1996; 97: 1795–803.
- Baldwin AS. The NF-κB and IκB proteins: New discoveries and insights. Annu Rev Immunol. 1996; 14: 649-81.
- 30. Bauerle PA, Baltimore D. NF-κB: Ten years after. Cell. 1996; 87: 13-20.
- 31. Nabel G, Baltimore D. An inducible transcription factor activates expression of human immunodeficiency virus in T cells Nature. 1987; 326: 711-13.
- 32. Brand K, Page S, Rogler G, et al. Activated transcription factor nuclear factor-kappa B is present in the atherosclerotic lesion. J Clin Invest. 1996; 97: 1715–22.
- Rupec RA, Baeuerle PA. The genomic response of tumor cells to hypoxia and reoxygenation. Differential activation of transcription factors AP-1 and NF-κB. Eur J Biochem. 1995; 234: 632-40.
- Stoltz RA, Abraham NG, Laniado-Schwartzman M. The role of NF-kappa B in the angiogenic response of coronary microvessel endothelial cells. Proc Natl Acad Sci USA. 1996; 93: 2832–7.
- 35 Lee S, Felts KA, Parry GC, Armacost LM, Cobb RR. Inhibition of 5-lipoxygenase blocks IL-1 beta-induced vascular adhesion molecule-1 gene expression in human endothelial cells. J Immunol. 1997; 158: 3401–7.
- Brach MA, de Vos S, Arnold C, Gruß HJ, Mertelsmann R, Herrmann F. Leukotriene B₄ transcriptionally activates interleukin-6 expression involving NF-κB and NF-IL6. Eur J Immunol. 1992; 22: 2705–11.
- 37. Shield MJ. Novel applications of misoprostol. Pharmacol Ther. 1995; 65: 125-47.
- 38. Sinclair SB, Greig PD, Blendis LM, et al. Biochemical and clinical response of fulminant viral hepatitis to administration of prostaglandin E. J Clin Invest 1989; 84: 1063–9.
- Lee J, Kannegiesser H, O'Toole J, Westura E. Hypertension and the renomedullary prostaglandins: A human study of the antihypertensive effects of PGA₁. Ann NY Acad Sci. 1971; 180: 218–40.