REVIEW



Current Prophylactic Medications for Migraine and Their Potential Mechanisms of Action

Till Sprenger 1 . M. Viana 2 · C. Tassorelli 2,3

Published online: 18 April 2018

© The American Society for Experimental NeuroTherapeutics, Inc. 2018

Abstract

A relatively high number of different medications is currently used for migraine prevention in clinical practice. Although these compounds were initially developed for other indications and differ in their mechanisms of action, some general themes can be identified from the mechanisms at play. Efficacious preventive drugs seem to either suppress excitatory nervous signaling via sodium and/or calcium receptors, facilitate GABAergic inhibition, reduce neuronal sensitization, block cortical spreading depression and/or reduce circulating levels of CGRP. We here review such mechanisms for the different compounds.

Keywords Migraine prevention · Mechanism of actions · Cortical spreading depression · β blockers · Antiepileptic drugs · Antidepressants · Botulinum toxin

Introduction

Migraine is a disorder characterized by recurrent attacks of headache with moderate to severe intensity, often accompanied by photophobia, phonophobia, and nausea. The headache can aggravate with physical activity, is often unilateral, and may have a throbbing character. The underlying pathophysiology of migraine is only partly understood to date. Malfunctioning of brain areas and channels, which modulate the excitability of nociceptive brain circuits as well as central sensitization are thought to play a role [1], the latter especially in the transformation from episodic to chronic migraine [2]. Migraine aura is thought to relate to cortical spreading depression (CSD), a slow wave of depolarization propagating across the cortex, and some authors have suggested that CSD is also implicated in the pathogenesis of migraine headache [3], although this remains controversial [4]. Calcitonin gene-related peptide (CGRP) is a neuropeptide,

☐ Till Sprenger till.sprenger@helios-gesundheit.de

- Department of Neurology, DKD Helios Klinik Wiesbaden, Aukammallee 33, 65191 Wiesbaden, Germany
- Headache Science Centre, IRCCS Mondino Foundation, 27100 Pavia, Italy
- Department of Brain and Behavioral Sciences, University of Pavia, 27100 Pavia, Italy

which appears firmly connected to migraine pathogenesis. Systemic levels of this strong vasodilator rise during migraine attacks and while they fall thereafter, they appear to remain elevated even interictally as compared to healthy subjects [5]. Interestingly, endogenous CGRP release has been shown also during experimentally induced CSD in cortical rat slices. Intravenous injection of CGRP can trigger migraine attacks.

Patients affected by frequent attacks may need preventive treatment in order to reduce the frequency and severity of attacks. A number of mostly oral pharmacological treatments have been evaluated and are recommended for migraine prevention [6, 7]. Thereof, propranolol, metoprolol, flunarizine, topiramate, and valproate were considered first line treatments in the EFNS guideline for the prevention of migraine [7].

One can generally expect a 50% reduction in attack frequency in every second patient with these compounds, if they are taken regularly. Just as our understanding of migraine pathophysiology is limited in general, the mechanisms of action of such preventive compounds are also understood only in part and for the currently licensed compounds, the clinical development did typically not go from bench to bedside, but rather the opposite route. The majority of available treatments were repurposed for migraine prevention after initial development for other indications such as arterial hypertension, epilepsy, and depression.

In this review, we will discuss potential mechanisms of action of currently available treatment options for migraine prevention and cover agents for both episodic as well as



chronic migraine. Because of space limitations, the focus will be on agents that are either licensed for migraine specifically or recommended in international treatment guidelines [7]. Novel compounds specifically targeting the CGRP pathway will be discussed in another dedicated article in this special issue (Reuter et al. Neurotherapeutics). Nonpharmacological migraine prevention strategies are very important in clinical practice [8, 9] but not covered in this article. We refer the interested reader to the article by Puledda and Shields in this issue (Puledda and Shields. Non-pharmacological approaches for migraine. Neurotherapeutics).

General Preventive Treatment Principles and Mechanisms of Action

The recommendations for when to use pharmacological migraine prevention vary greatly between published treatment guidelines. As a general principle, migraine prevention should be considered when attacks affect quality of life and is indicated in roughly one third of migraine patients [10]. Comorbidities should be taken into account when considering and selecting a migraine preventive.

Although the different compounds used for migraine prevention clearly act at different receptor sites or channels, it has been suggested that they may modulate common pathways or mechanisms implicated in migraine pathogenesis. For example, several migraine preventives such as propranolol, topiramate, valproate, amitriptyline [11], and flunarizine [12], but also CGRP antagonists [13] have been shown to suppress CSD. Other compounds have been suggested to reduce neuronal sensitization [14, 15]. Moreover, some of the available migraine preventives may reduce levels of circulating neuropeptides such as CGRP [16] and of "hyperalgesic" cytokines such as tumor necrosis factor α , interleukin-1 β , and interleukin 6 [17], although it remains unclear whether such effects directly relate to the medication or indirectly relate to the reduction of attacks.

In the following, we will summarize the current understanding of mechanisms of action of specific agents or groups of agents (Table 1).

β-Blockers

Although all beta-adrenergic blockers (BABs) are competitive inhibitors of β -receptors, they may differ with regard to receptor binding (i.e., β 1-selectivity and partial agonistic activity) and pharmacokinetic properties [18]. BABs with intrinsic sympathomimetic activity (e.g., acebutolol, alprenolol, oxprenolol, and pindolol) are not effective for migraine prevention [19]. Clinical studies support the efficacy of propranolol (80–240 mg/day), timolol (20–30 mg/day), bisoprolol

(5 mg/day), and metoprolol (200 mg/day) in migraine preventive treatment [20–22]. Atenolol and nadolol also have a moderate effect in reducing migraine attack frequency [19, 23].

The mechanisms of action of BABs in migraine prevention are not completely understood. Inhibition of β1-mediated effects could be considered the main mechanism of action. Indeed, blockade of \(\beta 1 \) receptors could inhibit noradrenaline (NA) release and tyrosine hydroxylase activity, the rate-limiting step in NA synthesis [24]. Moreover, propranolol reduces the neuronal firing rate of noradrenergic neurons of the locus coeruleus [25]. Interestingly, BABs also regulate the firing rate of periaqueductal gray matter (PAG) neurons via a GABA-mediated action [26]. Both these effects may contribute to the antimigraine action of BABs. Recent findings in an animal model of trigeminovascular activation showed that propranolol exerts its prophylactic action, at least in part, by interfering with the chronic sensitization processes in the rostral ventromedial medulla and locus coeruleus, and by counteracting the facilitation of trigeminovascular transmission within the trigeminocervical complex [15].

Some BABs may also interact with the serotonergic system by blocking 5-HT_{2C} and 5-HT_{2B} receptors [27–29]. Since 5-HT plays a pivotal role in the pathophysiology of migraine, the BABs-induced effects on this neurotransmitter could account for their prophylactic action on migraine. Moreover, PET studies have supported the possibility that BABs also affect 5-HT synthesis, which is altered in migraine patients [30].

Propranolol inhibits nitric oxide production by blocking inducible nitric oxide synthase (NOS). Propranolol also inhibits kainite-induced currents and is synergistic with N-methyl-D-aspartate blockers, which reduce neuronal activity and have membrane-stabilizing properties [31].

In a double-blind randomized study, it was shown that propranolol and metoprolol decreased VEP amplitude in migraine patients. This action was associated with a better clinical response to the prophylactic treatment [32]. Therefore, it could be hypothesized that BABs have a significant effect on the excitability of the visual system (and probably more generally the cortex) in migraine patients. In migraine patients, the effect of BABs on auditory evoked cortical potentials was correlated with a reduction of migraine attack frequency [33]. This effect of BABs on auditory evoked cortical potential could be due to a modulatory action on serotonergic transmission [34].

Other neurophysiological studies analyzed the contingent negative variation, a slow event-related potential measuring cortical information processing. This potential is increased in amplitude, lacks habituation and is normalized by BABs in association with a clinical improvement in migraine [35]. All these clinical neurophysiological studies support the hypothesis that BABs exert their prophylactic action in migraine by influencing cortical information processing and cortical excitability possibly via NA and 5-HT systems [34, 36, 37].



 Table 1
 Preventive treatments used in migraine and their potential mechanisms of action

Drug	Dose	Potential mechanisms of action
β-Blocker		
Propranolol	40 mg to 120 mg b.d.	Inhibition of nitric oxide synthase Interaction with the serotonergic system
Metoprolol	25-100 mg twice daily	Inhibition of thalamic relay neurons Block of central sensitization
Anticonvulsants		
Valproate	400-600 mg twice daily	Block of voltage-dependent sodium channels Block of low-threshold T-type calcium ion channels Suppression of proteinkinase C (glumatatergic neurotransmission) Inhibition of the NF-kB pathway (in an animal Downregulation of CGRP expression
Topiramate Calcium channel blocker	50-200 mg/day	Block of voltage-dependent sodium channels and high-voltage-activated L-type calcium channels Inhibition of glutamate-mediated excitatory neurotransmission Facilitation of GABA-A-mediated inhibition. Inhibition of carbonic anhydrase activity Reduction of CGRP secretion from trigeminal neurons
Flunarizine	5 15 mg daily	Plack of voltage geted codium shannels
Fiunalizine	5-15 mg daily	Block of voltage-gated sodium channels Reduction of neuronal excitability and normalization cortical hyperexcitability D2 antagonism
Antidepressants		
Amitriptyline	25 mg to 75 mg nocte (typically start with 10 mg)	Inhibition of serotonin/noradrenaline reuptake pump Facilitation of endogenous pain control mechanisms
Fluoxetine	20 mg daily	Block of serotonin reuptake reversible 5-HT2C receptor blocker
Onabotulinumtoxin A	155 units at fixed sites (+ additional units "follow the pain")	Block of peripheral and central sensitization Relaxation of head and neck muscles
Renin angiotensin system (RAS) r	nodulating compounds	
Lisinopril	20 mg daily	Modulation of vasoreactivity, alteration of sympathetic tone, and promotion of degradation of proinflammatory factors Modulation of endogenous opioid system Reduction of angiotensin-mediated
Candasartan	16 mg daily	Vasoconstriction Increase of sympathetic discharge Release of adrenal catecholamines Modulation of cerebral RAS influencing neuronal, astrocytic, and endothelial cell activity
Nutraceuticals		
Riboflavin	400 mg daily	Effect on mitochondria?
Coenzyme Q10	100 mg three times daily	Effect on mitochondria?
Butterbur	or 75 mg twice daily 50 to 75 mg twice daily	Anti-inflammatory action through inhibition of cyclooxygenase-2 and
Feverfew	6.25 mg three times daily	Vasodilatory effects through inhibition of L-type voltage-gated calcium channels Inhibition of Fos-induced activation of trigeminal nucleus caudalis Partial agonist activity at TRPA1 channels
		Antinociceptive and anti-inflammatory effects Effect on enzymatic function/mitochondria?



Cortical spreading depression (CSD) could also represent a target for BABs in migraine. In an experimental model, it was observed that treatment with propranolol suppressed retinal spreading depression [38]. Moreover, treatment with propranolol blocked CSD in rats, without altering regional cerebral blood flow and systemic arterial blood pressure [39].

Finally, it has also been hypothesized that BABs exert some of their therapeutic effects in migraine through an action at the ventroposteromedial thalamic nucleus, which represents a relay of trigeminal sensory input to the primary somatosensory cortex. Considering the complex and widespread nature of the sensory disturbance in migraine, and neurophysiological findings, a possible thalamic involvement in the mechanisms of action of BABs represents a fascinating hypothesis [40, 41].

Antiepileptics

Several antiepileptic drugs (AEDs) acting at different pre- and post-synaptic sites have been studied and proven effective for migraine prevention with the best clinical trial evidence in terms of migraine preventive action being available for topiramate and valproate [7]. However, clearly not all antiepileptics are similarly efficacious in migraine as in epilepsy. Those compounds that possess a rather specific/narrow mechanism of action typically seem to be less efficacious in migraine. As an example, oxcarbazepine was not superior to placebo in reducing the number of migraine attacks in a double-blind trial [42]. Interestingly, oxcarbazepine did also not reduce the frequency of CSD in animal experiments [43], while topiramate and valproate (and also other effective migraine preventives) did [11], suggesting this might be useful as a predictive model for evaluating novel migraine treatments.

Topiramate has been proven efficacious both in episodic as well as chronic migraine and is licensed for migraine prophylaxis in many countries. Topiramate is a "dirty drug" and blocks multiple channels such as voltage-dependent sodium channels and high-voltage-activated L-type calcium channels [44]. It has also been shown to inhibit glutamate-mediated excitatory neurotransmission and facilitate GABA-A-mediated inhibition. Topiramate also inhibits carbonic anhydrase activity [45]. Moreover, it has been shown that topiramate can reduce CGRP secretion from trigeminal neurons in response to depolarizing stimuli [46]. It is currently unclear which of the noted mechanisms is key for topiramate's effect in migraine prevention. Interestingly, CGRP plasma levels were shown to be unaltered by low-dose (50 mg per day) topiramate treatment in a small migraine trial, providing some preliminary evidence that topiramate effect may be independent from the CGRP pathway in humans [47].

Just like topiramate, there are multiple mechanisms at play by which valproate may improve migraine frequency. Similar to topiramate, valproate also enhances GABAergic inhibition and blocks excitatory ion channels [44]. The exact mechanisms by which this is achieved differ between the two drugs, though. Valproate blocks the degradation of GABA, hence increasing GABA levels in axons and glia cells. It blocks voltage-dependent sodium and low-threshold T-type calcium ion channels [44]. Furthermore, valproate can suppress proteinkinase C, a regulator of the glutamatergic neurotransmission [48]. A more recent study identified the inhibition of the NF-kB pathway (in an animal model of migraine using nitroglycerin triggering) as an additional potential mechanism of action in migraine [49]. In the same preclinical study, valproate also downregulated the expression of CGRP in brain tissue.

Calcium Antagonists

Several calcium antagonists have been used for migraine prevention since the 1980s [50]. In this regard, flunarizine (usually used at daily doses of 5–10 mg [15]) is the best studied compound and licensed for this indication in many countries (although not available in the USA). Verapamil and the antihistaminergic drug cinnarizine are alternatives (e.g., in refractory migraine cases or when flunarizine is not available) that also act at calcium channels. However, both latter drugs are off-label for migraine treatment and the evidence in terms of efficacy is much more scarce than for flunarizine.

Flunarizine is a nonselective calcium antagonist, which has also been shown to block voltage-gated sodium channels [51, 52]. Action at these two sites may reduce neuronal excitability and normalize cortical hyperexcitability in migraine. Furthermore, flunarizine acts as a D2 dopamine antagonist and has in this regard been shown to possess (at high doses) comparable antipsychotic efficacy as haloperidol [53]. It has also been shown that flunarizine can reduce the number and duration of CSD waves [12]. Furthermore, it has been suggested that flunarizine can alleviate CSD-induced mitochondrial injury [12]. Finally, flunarizine has also been shown to increase leptin levels [17], which have been suggested to reflect leptin resistance, which may potentially explain the treatment-related weight gain [54].

Antidepressants

There are more than three placebo-controlled trials each supporting the superiority of the tricyclic antidepressant amitriptyline and of the selective serotonin reuptake inhibitor (SSRI) fluoxetine over placebo for migraine prevention [55]. This effect occurs in the absence of depression and antimigraine effects are typically observed at doses lower than in depression. Less evidence is available for other antidepressants, although at least other tricyclics and the selective



serotonin-norepinephrine reuptake inhibitor venlafaxine [56] are probably also effective for migraine prophylaxis.

Amitriptyline is a mixed serotonin-norepinephrine reuptake pump inhibitor and thereby thought to facilitate descending noxious inhibition, i.e., endogenous pain control mechanisms descending from the brainstem to the trigeminal nucleus caudalis and the spinal cord. Alpha2-adrenoceptor blockade has been shown to block the antinociceptive effect of amitriptyline [57] and hence at least part of amitriptyline's efficacy is thought to be mediated by α 2-agonism, but multiple other channel and receptor effects of amitriptyline are known. As such, amitriptyline is thought to act as a sodium channel blocker and also has antimuscarinic and antihistaminic effects. There is also an interaction with the endogenous adenosine system and it has also been shown to suppress cortical spreading depression [11]. As with other preventive migraine medications, it remains unclear which mechanism is key and probably the multiplicity of synergistic effects in multiple pathways explains the clinical efficacy (as well as the broad side-effect profile).

Fluoxetine is a more targeted substance that (relatively) selectively blocks serotonin reuptake from the synaptic cleft leading to increased serotonin levels. Noradrenaline reuptake is inhibited only at higher doses. Furthermore, fluoxetine is a competitive, reversible 5-HT2C receptor blocker [58]. The 5-HT2C mechanism is shared with the old antiheadache drug methysergide and a receptor gene polymorphisms of 5-HT2C has been shown to modulate migraine susceptibility (in a Turkish population) [59]. Hence this receptor blockade may explain (part) of the efficacy of fluoxetine.

Onabotulinumtoxin A

Trials investigating the efficacy of onabotulinumtoxin A in episodic migraine and chronic daily headache have yielded inconclusive results, partly because of methodological limitations regarding the injection paradigm and the dose [60–64]. The efficacy of onabotulinumtoxin A has been demonstrated for chronic migraine in the clinical programme called REsearch Evaluating Migraine Prophylaxis Therapy (PREEMPT) [65–67]. In the two phase III trials of the PREEMPT program (PREEMPT1 and 2), a minimum dose of 155 U (up to a maximum allowed dose of 195 U) of onabotulinumtoxin A was injected into head and neck muscles of each patient. Nowadays, onabotulinumtoxin A is marketed in several countries with an indication for chronic migraine.

Onabotulinum toxin A (BoNT-A) is a protein complex produced by the gram-positive, anaerobic bacterium *Clostridium botulinum* [68]. There are seven serotypes of botulinum toxin [68], the serotype A has been used in human therapy since 1980 when Scott proposed BoNT-A injection into extraocular muscles as an alternative to strabismus surgery [69]. The first evidence for an effect of BoNT on migraine was the

serendipitous observation of an improvement of migraine in patients treated with BoNT for hyperfunctional lines of the face made in the late 1990s by a plastic surgeon [70]. In the subsequent years, several exploratory studies with BoNT-A using heterogeneous doses and injection paradigms were carried out in migraine and tension-type headache and chronic daily headache with inconclusive and mostly negative results [62, 63, 71–74]. A subgroup analysis of 228 CDH patients enrolled in the randomized placebo-controlled study by Mathew et al. showed that the patients without prophylactic medication at the date of study enrolment experienced a significant reduction in the number of headaches in a 30-day period. So the authors concluded that onabotulinumtoxin A was effective in the treatment of patients with CDH, who do not receive other prophylactic medication [75].

Based on these preliminary studies, a series of studies called the Placebo-controlled phase III Research Evaluating Migraine Prophylaxis Therapy (PREEMPT), treated patients with chronic migraine who would receive BoNT-A at fixed sites (procerus, corrugator, frontalis, temporalis, occipitalis, cervical paraspinal muscles and trapezius) in fixed doses (5 units per site for a total of 155 units), including those with or without medication overuse. The studies revealed a significant reduction in headache and migraine days, moderate/severe headache days and cumulative headache hours on headache days as well as in patients' functioning, vitality, psychological distress, and overall health-related quality of life [65–67].

The exact mechanism by which BoNT-A reduces migraine days in chronic migraineurs is not yet fully understood, although an increasing body of evidence suggests the dominant hypothesis that the toxin exerts its antinociceptive action via peripheral mechanisms [76]. While it was initially thought that BoNT-A may exert positive effects in primary headache disorders simply by the relaxation of muscles in the head and neck region, it turned out that BoNT-A is rather ineffective in disorders, where muscle tension may play a key role such as tension-type headache. It is today believed that BoNT-A acts in migraine by directly inhibiting peripheral sensitization by attenuating neuropeptide and neurotransmitter exocytosis from peripheral sensory neurons, thereby indirectly reducing central sensitization, the hallmark of chronic migraine [76]. The effectiveness of BoNT-A in chronic migraine, as opposed to negative findings in episodic migraine, may relate to the fact that sensitization phenomena play a more important role in chronic versus episodic migraine.

Neuronal stimulation initiates a series of intracellular events that leads to the fusion of a neuropeptide and/or neurotransmitter containing vesicle with the nerve cell membrane. This process is facilitated by interaction between proteins on the vesicle, the so-called vesicle-associated membrane protein (VAMP/synaptobrevin) and on the internal membrane surface, the synaptosomal associated protein (SNAP-25), which together form the soluble N-ethylmaleimide–sensitive factor



attachment protein receptor (SNARE) complex [77–79]. The SNARE complex is instrumental to vesicular trafficking and to vesicle fusion with the membrane. BoNT-A adheres to the nerve cells and enters inside through the endocytosis mechanism and inhibits fusion of intracellular vesicles with the nerve membrane [80] by cleaving SNAP-25 [81]. Once intraneuronal vesicular fusion is impaired because of BONT-A, neuropeptide release is inhibited and receptors are downregulated [77, 79]. A large number of experiments in vitro and in vivo showed that [82] botulinum toxin could also inhibit the release of a variety of neurotransmitters, including glutamate, GABA, aspartate, catecholamines, dopamine, and monoamine neurotransmitters [83-85]. All these neurotransmitters and neuropeptides are key signaling molecules in chronic migraine and it is therefore likely that, altogether, these mechanisms are important in the current notion of migraine prevention because they are capable of disrupting the cascade of events that leads to peripheral and central sensitization, mainly via the block of release of inflammatory neuropeptides from stimulated trigeminal sensory neurons [76–78, 86]. This mechanism of action is supported by clinical studies demonstrating suppression of cutaneous allodynia, an indicator of central sensitization, after onabotulinumtoxin A injection in the periorbital skin [87, 88].

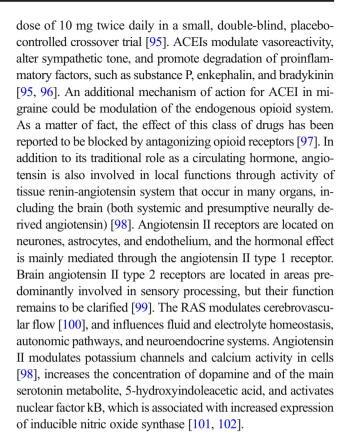
In this framework, it is relevant to note that, in an animal model, administration of BoNT-A in close proximity of sensory fibers that bifurcate from parent axons of intracranial meningeal nociceptors and reach extracranial tissues by crossing the calvarial bones through the sutures [89–91] proved more effective in inhibiting the responses of nociceptors to topical capsaicin than when it was injected into muscles and sutures [92].

Other Compounds

ACE and ARB Agents

There are two randomized controlled studies suggesting an effect of candesartan at a dose of 16 mg per day in migraine prevention with comparable efficacy to propranolol. Candesartan is an angiotensin II receptor, type 1 blocking (ARB) agent and thus it reduces the effect of angiotensin, which has several effects that may be relevant to migraine, such as increased sympathetic discharge, and adrenal medullary catecholamine release. Molecular biology confirmed the components of the cerebral renin–angiotensin system (RAS) and their location in the brain [93]. RAS may play a role also in migraine pathogenesis. Brain RAS could act independently of the peripheral RAS, influencing neuronal, astrocytic, and endothelial cell activity [94].

Lisinopril, an angiotensin-converting enzyme inhibitor (ACEI), has been shown to be prophylactically effective at a



Feverfew (*Tanacetum parthenium L.*)

Double-blind placebo-controlled studies suggested efficacy of feverfew in migraine prophylaxis, although it is usually not considered first choice as antimigraine preventive drug for tolerability (and safety) reasons [103–105]. The most frequent adverse events reported were sore mouth and tongue for the presence of ulcers, swollen lips, loss of taste, abdominal pain, and gastrointestinal disturbances [105]. Moreover, the safety of long-term use of feverfew needs to be established [106].

Feverfew is a member of the Asteracee family, empirically used as herbal remedy for migraine. Parthenolide, a sesquiterpene lactone, appears to be feverfew's active ingredient and has multiple actions in the central nervous system. Several of its properties suggest a potential mechanism of action in migraine prevention, including evidence to suggest that parthenolide inhibits Fos-induced activation of the nucleus trigeminalis caudalis [107], a nucleus central to migraine pathogenesis, and evidence for partial agonist activity of parthenolide at TRPA1 channels [108], which have been implicated in migraine pathogenesis [109]. It has also been demonstrated that oral administration of feverfew extract leads to significant antinociceptive and anti-inflammatory effects in experimental models [110]. This action could be explained by the regulation of degradation of the inhibitory protein of NF-kB and of expression of inducible NOS by parthenolide, the active constituent of feverfew [101].



Moreover, there is evidence that feverfew inhibits human blood platelet aggregation *in vitro*, probably through the protein kinase C pathway [111]. Feverfew could also exert modulation of 5-HT neurotransmission, antagonizing the effects mediated by 5-HT2B and 5-HT2A receptors implicated in the plasticity changes underlying chronic headache [112–114].

Butterbur (Petasites hybridus)

Some promising results support the efficacy of this extract in reducing migraine attacks [115–117]. Nevertheless, a recent systematic review shows that there is only moderate evidence for its effectiveness as an antimigraine drug [118].

Butterbur is a shrub with a long history of use for medicinal purposes. It demonstrates a variety of properties that render it a candidate for migraine prophylaxis, including anti-inflammatory action through inhibition of cyclooxygenase-2 [119]. Moreover, the co-active ingredients *petasins* could exert a block of Ca2+ channels [120–123].

Riboflavin (Vitamin B2)

A placebo-controlled, double-blind trial showed that a high dose of riboflavin can be effective in migraine prophylaxis [124]. This positive result was confirmed by an open-label study [125]. Riboflavin was comparable to propranolol after 3 and 6 months of administration in an RCT assessing its efficacy for adult migraine prophylaxis [126].

Riboflavin is a vitamin that plays an important role in cellular energy production through its two active coenzyme forms that are involved in oxidation-reduction reactions during a variety of cellular processes [127]. Brain energy metabolism has been found to be abnormal in migraine headache [128]. Riboflavin catalyzes the activity of flavoenzymes in the mitochondrial respiratory chain and improves the clinical and biochemical abnormalities in patients with inborn errors of mitochondrial metabolism [129, 130]. As mitochondrial deficits are suspected in some patients with migraine headache, this could reflect a mechanism of action in migraine prevention.

Magnesium

While a role for magnesium-related mechanisms can be hypothesized in migraine pathogenesis, limited evidence has indicated efficacy in migraine prophylaxis. In fact, several studies suggested a deficiency of Mg2+ in the central nervous system of migraineurs [131–133]. A Mg2+ deficit in migraine was also described at a peripheral level, in serum and blood cells [134, 135].

Clinical trials evaluated the potential usefulness of oral Mg2+ in migraine prevention, showing conflicting results [136, 137]. Three separate RCTs with varying methodologies,

magnesium dosages, and formulations have found oral magnesium to be effective for migraine prophylaxis in adults [138–140]. Another RCT contradicted these findings and found oral magnesium to be no different than placebo on interim analysis in a sample of refractory migraine patients and therefore halted recruitment prior to achieving the planned sample size [141]. A possible effect of Mg2+ supplementation was demonstrated for short-term prophylaxis of menstrual migraine [142].

Magnesium is found ubiquitous in the human body and is an essential cofactor for more than 350 enzymes. It plays an important role in a multitude of biological processes, some of which might be linked to migraine pathogenesis [143]. It could modulate mitochondrial oxidative phosphorylation, 5-HT neurotransmission, and the NO system [144]. Moreover, it could regulate the uptake of glutamate into astrocytes and interfere with NMDA receptor function [144] and thereby improve migraine.

Conclusions

Several compounds with heterogenous mechanisms of action have been studied for the prevention of migraine during the past decades with some of them being licensed for this indication. Potential mechanisms of action include a reduction of excitatory glutamatergic neurotransmission by inhibition of sodium and calcium channels, facilitation of inhibitory GABAergic neurotransmission, reduction of central sensitization, and suppression of cortical spreading depression, but a number of other mechanism may be at play. While the migraine preventive properties of the available compounds were mostly discovered post hoc by circumstantial evidence, the advent of CGRP antibodies will for the first time allow a treatment tailored to the disease after systematic drug development from bench to bedside (Reuter et al. Neurotherapeutics, in this issue).

Required Author Forms Disclosure forms provided by the authors are available with the online version of this article.

References

- Sprenger T, Goadsby PJ. Migraine pathogenesis and state of pharmacological treatment options. BMC Med 2009;7:71.
- Aurora SK, Brin MF. Chronic migraine: an update on physiology, imaging, and the mechanism of action of two available pharmacologic therapies. Headache 2017;57(1):109–25.
- Bolay H, Reuter U, Dunn AK, Huang Z, Boas DA, Moskowitz MA. Intrinsic brain activity triggers trigeminal meningeal afferents in a migraine model. Nat Med 2002;8(2):136–42.
- Wolthausen J, Sternberg S, Gerloff C, May A. Are cortical spreading depression and headache in migraine causally linked? Cephalalgia 2009;29(2):244–9.



- Niazi AK, Andelova M, Sprenger T. Is the migrainous brain normal outside of acute attacks? Lessons learned from psychophysical, neurochemical and functional neuroimaging studies. Expert Rev Neurother 2013;13(9):1061–7.
- Goadsby PJ, Sprenger T. Current practice and future directions in the prevention and acute management of migraine. Lancet Neurol 2010;9(3):285–98.
- Evers S, Afra J, Frese A, Goadsby PJ, Linde M, May A, et al. EFNS guideline on the drug treatment of migraine–revised report of an EFNS task force. Eur J Neurol 2009;16(9):968–81.
- 8. Kropp P, Meyer B, Dresler T, Fritsche G, Gaul C, Niederberger U, et al. [Relaxation techniques and behavioural therapy for the treatment of migraine: guidelines from the German Migraine and Headache Society]. Schmerz 2017.
- Puledda F, Goadsby PJ. An update on non-pharmacological neuromodulation for the acute and preventive treatment of migraine. Headache 2017;57(4):685–91.
- Lipton RB, Bigal ME, Diamond M, Freitag F, Reed ML, Stewart WF, et al. Migraine prevalence, disease burden, and the need for preventive therapy. Neurology 2007;68(5):343–9.
- Ayata C, Jin H, Kudo C, Dalkara T, Moskowitz MA. Suppression of cortical spreading depression in migraine prophylaxis. Ann Neurol 2006;59(4):652–61.
- Li F, Qiu E, Dong Z, Liu R, Wu S, Yu S. Protection of flunarizine on cerebral mitochondria injury induced by cortical spreading depression under hypoxic conditions. J Headache Pain 2011;12(1): 47–53.
- Tozzi A, de Iure A, Di Filippo M, Costa C, Caproni S, Pisani A, et al. Critical role of calcitonin gene-related peptide receptors in cortical spreading depression. Proc Natl Acad Sci U S A 2012;109(46):18985–90.
- Dolly JO, Aoki KR. The structure and mode of action of different botulinum toxins. Eur J Neurol 2006;13 Suppl 4:1–9.
- Boyer N, Signoret-Genest J, Artola A, Dallel R, Monconduit L. Propranolol treatment prevents chronic central sensitization induced by repeated dural stimulation. Pain 2017;158(10):2025-34.
- Cernuda-Morollon E, Ramon C, Martinez-Camblor P, Serrano-Pertierra E, Larrosa D, Pascual J. OnabotulinumtoxinA decreases interictal CGRP plasma levels in patients with chronic migraine. Pain 2015;156(5):820–4.
- Hirfanoglu T, Serdaroglu A, Gulbahar O, Cansu A. Prophylactic drugs and cytokine and leptin levels in children with migraine. Pediatr Neurol 2009;41(4):281–7.
- Bangalore S, Messerli FH, Kostis JB, Pepine CJ. Cardiovascular protection using beta-blockers: a critical review of the evidence. J Am Coll Cardiol 2007;50(7):563–72.
- Limmroth V, Michel MC. The prevention of migraine: a critical review with special emphasis on beta-adrenoceptor blockers. Br J Clin Pharmacol 2001;52(3):237–43.
- Tfelt-Hansen P. Efficacy of beta-blockers in migraine. A critical review. Cephalalgia 1986;6 Suppl 5:15–24.
- Schellenberg R, Lichtenthal A, Wohling H, Graf C, Brixius K. Nebivolol and metoprolol for treating migraine: an advance on beta-blocker treatment? Headache 2008;48(1):118–25.
- van de Ven LL, Franke CL, Koehler PJ. Prophylactic treatment of migraine with bisoprolol: a placebo-controlled study. Cephalalgia 1997;17(5):596–9.
- Buchanan TM, Ramadan NM. Prophylactic pharmacotherapy for migraine headaches. Semin Neurol 2006;26(2):188–98.
- Hanbauer I, Kopin IJ, Guidotti A, Costa E. Induction of tyrosine hydroxylase elicited by beta adrenergic receptor agonists in normal and decentralized sympathetic ganglia: role of cyclic 3',5' adenosine monophosphate. J Pharmacol Exp Ther 1975;193(1): 95–104.

- Hieble JP. Adrenoceptor subclassification: an approach to improved cardiovascular therapeutics. Pharm Acta Helv 2000;74(2-3):163-71.
- Xiao C, Zhou C, Atlas G, Delphin E, Ye JH. Labetalol facilitates GABAergic transmission to rat periaqueductal gray neurons via antagonizing beta1-adrenergic receptors—a possible mechanism underlying labetalol-induced analgesia. Brain Res 2008;1198: 34–43.
- Koella WP. CNS-related (side-)effects of beta-blockers with special reference to mechanisms of action. Eur J Clin Pharmacol 1985:28 Suppl:55–63.
- Ablad B, Dahlof C. Migraine and beta-blockade: modulation of sympathetic neurotransmission. Cephalalgia 1986;6 Suppl 5:7–13.
- Kalkman HO. Is migraine prophylactic activity caused by 5-HT2B or 5-HT2C receptor blockade? Life Sci 1994;54(10):641–4.
- Chugani DC, Niimura K, Chaturvedi S, Muzik O, Fakhouri M, Lee ML, et al. Increased brain serotonin synthesis in migraine. Neurology 1999;53(7):1473–9.
- 31. Ramadan NM. Prophylactic migraine therapy: mechanisms and evidence. Curr Pain Headache Rep 2004;8(2):91–5.
- Diener HC, Scholz E, Dichgans J, Gerber WD, Jack A, Bille A, et al. Central effects of drugs used in migraine prophylaxis evaluated by visual evoked potentials. Ann Neurol 1989;25(2):125–30.
- Sandor PS, Afra J, Ambrosini A, Schoenen J. Prophylactic treatment of migraine with beta-blockers and riboflavin: differential effects on the intensity dependence of auditory evoked cortical potentials. Headache 2000;40(1):30–5.
- Hegerl U, Juckel G. Intensity dependence of auditory evoked potentials as an indicator of central serotonergic neurotransmission: a new hypothesis. Biol Psychiatry 1993;33(3):173–87.
- Siniatchkin M, Andrasik F, Kropp P, Niederberger U, Strenge H, Averkina N, et al. Central mechanisms of controlled-release metoprolol in migraine: a double-blind, placebo-controlled study. Cephalalgia 2007;27(9):1024

 –32.
- Aston-Jones G, Chen S, Zhu Y, Oshinsky ML. A neural circuit for circadian regulation of arousal. Nat Neurosci 2001;4(7):732–8.
- Berridge CW, Waterhouse BD. The locus coeruleus-noradrenergic system: modulation of behavioral state and state-dependent cognitive processes. Brain research Brain Res Rev 2003;42(1):33–84.
- Wiedemann M, de Lima VM, Hanke W. Effects of antimigraine drugs on retinal spreading depression. Naunyn Schmiedeberg's Arch Pharmacol 1996;353(5):552–6.
- Richter F, Mikulik O, Ebersberger A, Schaible HG. Noradrenergic agonists and antagonists influence migration of cortical spreading depression in rat-a possible mechanism of migraine prophylaxis and prevention of postischemic neuronal damage. J Cereb Blood Flow Metab 2005;25(9):1225–35.
- Kaube H, Goadsby PJ. Anti-migraine compounds fail to modulate the propagation of cortical spreading depression in the cat. Eur Neurol 1994;34(1):30–5.
- Shields KG, Goadsby PJ. Propranolol modulates trigeminovascular responses in thalamic ventroposteromedial nucleus: a role in migraine? Brain J Neurol 2005;128(Pt 1):86–97.
- Silberstein S, Saper J, Berenson F, Somogyi M, McCague K, D'Souza J. Oxcarbazepine in migraine headache: a double-blind, randomized, placebo-controlled study. Neurology 2008;70(7): 548–55.
- Hoffmann U, Dilekoz E, Kudo C, Ayata C. Oxcarbazepine does not suppress cortical spreading depression. Cephalalgia 2011;31(5):537–42.
- Cutrer FM. Antiepileptic drugs: how they work in headache. Headache 2001;41 Suppl 1:S3–10.
- Shank RP, Doose DR, Streeter AJ, Bialer M. Plasma and whole blood pharmacokinetics of topiramate: the role of carbonic anhydrase. Epilepsy Res 2005;63(2–3):103–12.



- Durham PL, Niemann C, Cady R. Repression of stimulated calcitonin gene-related peptide secretion by topiramate. Headache 2006;46(8):1291–5.
- Garcia-Estevez DA, Pardo-Parrado M, Silvarrey-Rodriguez S. [Frequent episodic migraine and calcitonin gene-related peptide. Influence of treatment with topiramate and zonisamide on levels of the peptide]. Rev Neurol 2017;65(4):153–6.
- Chen G, Manji HK, Hawver DB, Wright CB, Potter WZ. Chronic sodium valproate selectively decreases protein kinase C alpha and epsilon in vitro. J Neurochem 1994;63(6):2361–4.
- Li Y, Zhang Q, Qi D, Zhang L, Yi L, Li Q, et al. Valproate ameliorates nitroglycerin-induced migraine in trigeminal nucleus caudalis in rats through inhibition of NF-small ka, CyrillicB. J Headache Pain 2016:17:49.
- Solomon GD. Comparative efficacy of calcium antagonist drugs in the prophylaxis of migraine. Headache 1985;25(7):368–71.
- 51. Ye Q, Yan LY, Xue LJ, Wang Q, Zhou ZK, Xiao H, et al. Flunarizine blocks voltage-gated Na(+) and Ca(2+) currents in cultured rat cortical neurons: a possible locus of action in the prevention of migraine. Neurosci Lett 2011;487(3):394–9.
- Ye Q, Wang Q, Yan LY, Wu WH, Liu S, Xiao H, et al. Flunarizine inhibits sensory neuron excitability by blocking voltage-gated Na+ and Ca2+ currents in trigeminal ganglion neurons. Chin Med J 2011;124(17):2649–55.
- Bisol LW, Brunstein MG, Ottoni GL, Ramos FL, Borba DL, Daltio CS, et al. Is flunarizine a long-acting oral atypical antipsychotic? A randomized clinical trial versus haloperidol for the treatment of schizophrenia. J Clin Psychiatry 2008;69(10):1572–9.
- 54. Berilgen MS, Bulut S, Gonen M, Tekatas A, Dag E, Mungen B. Comparison of the effects of amitriptyline and flunarizine on weight gain and serum leptin, C peptide and insulin levels when used as migraine preventive treatment. Cephalalgia 2005;25(11): 1048–53.
- Jackson JL, Cogbill E, Santana-Davila R, Eldredge C, Collier W, Gradall A, et al. A comparative effectiveness meta-analysis of drugs for the prophylaxis of migraine headache. PLoS One 2015;10(7):e0130733.
- Ozyalcin SN, Talu GK, Kiziltan E, Yucel B, Ertas M, Disci R. The efficacy and safety of venlafaxine in the prophylaxis of migraine. Headache 2005;45(2):144–52.
- Gray AM, Pache DM, Sewell RD. Do alpha2-adrenoceptors play an integral role in the antinociceptive mechanism of action of antidepressant compounds? Eur J Pharmacol 1999;378(2):161–8.
- Ni YG, Miledi R. Blockage of 5HT2C serotonin receptors by fluoxetine (Prozac). Proc Natl Acad Sci U S A 1997;94(5): 2036–40.
- Yucel Y, Coskun S, Cengiz B, Ozdemir HH, Uzar E, Cim A, et al. Association of polymorphisms within the serotonin receptor genes 5-HTR1A, 5-HTR1B, 5-HTR2A and 5-HTR2C and migraine susceptibility in a Turkish population. Clin Psychopharmacol Neurosci 2016;14(3):250–5.
- Aurora SK, Gawel M, Brandes JL, Pokta S, Vandenburgh AM, Group BNAEMS. Botulinum toxin type a prophylactic treatment of episodic migraine: a randomized, double-blind, placebocontrolled exploratory study. Headache 2007;47(4):486–99.
- Freitag FG, Diamond S, Diamond M, Urban G. Botulinum toxin type A in the treatment of chronic migraine without medication overuse. Headache 2008;48(2):201–9.
- Mathew NT, Frishberg BM, Gawel M, Dimitrova R, Gibson J, Turkel C, et al. Botulinum toxin type A (BOTOX) for the prophylactic treatment of chronic daily headache: a randomized, double-blind, placebo-controlled trial. Headache 2005;45(4):293–307.
- Silberstein S, Mathew N, Saper J, Jenkins S. Botulinum toxin type A as a migraine preventive treatment. For the BOTOX Migraine Clinical Research Group. Headache 2000;40(6):445–50.

- Silberstein SD, Stark SR, Lucas SM, Christie SN, Degryse RE, Turkel CC, et al. Botulinum toxin type A for the prophylactic treatment of chronic daily headache: a randomized, doubleblind, placebo-controlled trial. Mayo Clin Proc. 2005;80(9): 1126–37
- Aurora SK, Dodick DW, Turkel CC, DeGryse RE, Silberstein SD, Lipton RB, et al. OnabotulinumtoxinA for treatment of chronic migraine: results from the double-blind, randomized, placebocontrolled phase of the PREEMPT 1 trial. Cephalalgia 2010;30(7):793–803.
- Diener HC, Dodick DW, Aurora SK, Turkel CC, DeGryse RE, Lipton RB, et al. OnabotulinumtoxinA for treatment of chronic migraine: results from the double-blind, randomized, placebocontrolled phase of the PREEMPT 2 trial. Cephalalgia 2010;30(7):804–14.
- Dodick DW, Turkel CC, DeGryse RE, Aurora SK, Silberstein SD, Lipton RB, et al. OnabotulinumtoxinA for treatment of chronic migraine: pooled results from the double-blind, randomized, placebo-controlled phases of the PREEMPT clinical program. Headache 2010;50(6):921–36.
- Erbguth FJ. From poison to remedy: the chequered history of botulinum toxin. J Neural Transm 2008;115(4):559

 –65.
- Scott AB. Botulinum toxin injection into extraocular muscles as an alternative to strabismus surgery. Ophthalmology. 1980;87(10): 1044–9.
- Binder WJ, Blitzer A, Brin MF. Treatment of hyperfunctional lines of the face with botulinum toxin A. Dermatol Surg 1998;24(11): 1198–205.
- Silberstein SD, Gobel H, Jensen R, Elkind AH, Degryse R, Walcott JM, et al. Botulinum toxin type A in the prophylactic treatment of chronic tension-type headache: a multicentre, double-blind, randomized, placebo-controlled, parallel-group study. Cephalalgia 2006;26(7):790–800.
- Ondo WG, Vuong KD, Derman HS. Botulinum toxin A for chronic daily headache: a randomized, placebo-controlled, parallel design study. Cephalalgia 2004;24(1):60–5.
- Jackson JL, Kuriyama A, Hayashino Y. Botulinum toxin A for prophylactic treatment of migraine and tension headaches in adults: a meta-analysis. JAMA 2012;307(16):1736–45.
- Gaul C, Holle-Lee D, Straube A. [Botulinum toxin type A in headache treatment: established and experimental indications]. Nervenarzt 2016;87(8):853–9.
- Escher CM, Paracka L, Dressler D, Kollewe K. Botulinum toxin in the management of chronic migraine: clinical evidence and experience. Ther Adv Neurol Disord. 2017;10(2): 127–35
- Aoki KR. Review of a proposed mechanism for the antinociceptive action of botulinum toxin type A. Neurotoxicology. 2005;26(5): 785–93.
- Burstein R, Zhang X, Levy D, Aoki KR, Brin MF. Selective inhibition of meningeal nociceptors by botulinum neurotoxin type A: therapeutic implications for migraine and other pains. Cephalalgia. 2014;34(11):853–69.
- Aoki KR. Evidence for antinociceptive activity of botulinum toxin type A in pain management. Headache. 2003;43 Suppl 1:S9–15.
- Whitcup SM, Turkel CC, DeGryse RE, Brin MF. Development of onabotulinumtoxinA for chronic migraine. Ann N Y Acad Sci. 2014;1329:67–80.
- 80. Simpson LL. The origin, structure, and pharmacological activity of botulinum toxin. Pharmacol Rev. 1981;33(3):155–88.
- Blasi J, Chapman ER, Link E, Binz T, Yamasaki S, De Camilli P, et al. Botulinum neurotoxin A selectively cleaves the synaptic protein SNAP-25. Nature 1993;365(6442):160–3.
- Mustafa G, Anderson EM, Bokrand-Donatelli Y, Neubert JK, Caudle RM. Anti-nociceptive effect of a conjugate of substance



P and light chain of botulinum neurotoxin type A. Pain 2013;154(11):2547-53.

- 83. Verderio C, Grumelli C, Raiteri L, Coco S, Paluzzi S, Caccin P, et al. Traffic of botulinum toxins A and E in excitatory and inhibitory neurons. Traffic 2007;8(2):142–53.
- Drinovac V, Bach-Rojecky L, Lackovic Z. Association of antinociceptive action of botulinum toxin type A with GABA-A receptor. J Neural Transm 2014;121(6):665–9.
- Habermann E. Inhibition by tetanus and botulinum A toxin of the release of [3H]noradrenaline and [3H]GABA from rat brain homogenate. Experientia 1988;44(3):224–6.
- Aoki KR, Francis J. Updates on the antinociceptive mechanism hypothesis of botulinum toxin A. Parkinsonism & related disorders. 2011;17 Suppl 1:S28–33.
- Gazerani P, Pedersen NS, Staahl C, Drewes AM, Arendt-Nielsen L. Subcutaneous Botulinum toxin type A reduces capsaicininduced trigeminal pain and vasomotor reactions in human skin. Pain 2009;141(1–2):60–9.
- Gazerani P, Staahl C, Drewes AM, Arendt-Nielsen L. The effects of Botulinum toxin type A on capsaicin-evoked pain, flare, and secondary hyperalgesia in an experimental human model of trigeminal sensitization. Pain 2006;122(3):315–25.
- Kosaras B, Jakubowski M, Kainz V, Burstein R. Sensory innervation of the calvarial bones of the mouse. J Comp Neurol 2009;515(3):331–48.
- Schueler M, Messlinger K, Dux M, Neuhuber WL, De Col R. Extracranial projections of meningeal afferents and their impact on meningeal nociception and headache. Pain 2013;154(9):1622-31.
- Schueler M, Neuhuber WL, De Col R, Messlinger K. Innervation of rat and human dura mater and pericranial tissues in the parietotemporal region by meningeal afferents. Headache 2014;54(6): 996–1009.
- Zhang X, Strassman AM, Novack V, Brin MF, Burstein R. Extracranial injections of botulinum neurotoxin type A inhibit intracranial meningeal nociceptors' responses to stimulation of TRPV1 and TRPA1 channels: are we getting closer to solving this puzzle? Cephalalgia 2016;36(9):875–86.
- Phillips MI, de Oliveira EM. Brain renin angiotensin in disease. J Mol Med 2008;86(6):715–22.
- Tronvik E, Stovner LJ, Schrader H, Bovim G. Involvement of the renin-angiotensin system in migraine. Journal of hypertension supplement: official journal of the International Society of Hypertension. 2006;24(1):S139

 –43.
- Schrader H, Stovner LJ, Helde G, Sand T, Bovim G. Prophylactic treatment of migraine with angiotensin converting enzyme inhibitor (lisinopril): randomised, placebo controlled, crossover study. BMJ 2001;322(7277):19–22.
- Ito H, Takemori K, Suzuki T. Role of angiotensin II type 1 receptor in the leucocytes and endothelial cells of brain microvessels in the pathogenesis of hypertensive cerebral injury. J Hypertens 2001;19(3 Pt 2):591–7.
- Montastruc P, Dang-Tran L, Carvajal A, Rostin M, Montastruc JL. Naloxone reverses the effects of enalapril and enalaprilic acid on the pressor responses to afferent vagal stimulation. Neuropeptides 1985;6(6):537–42.
- Allen AM, Moeller I, Jenkins TA, Zhuo J, Aldred GP, Chai SY, et al. Angiotensin receptors in the nervous system. Brain Res Bull 1998:47(1):17–28
- Saavedra JM. Brain angiotensin II: new developments, unanswered questions and therapeutic opportunities. Cell Mol Neurobiol 2005;25(3-4):485-512.
- Nishimura Y, Ito T, Saavedra JM. Angiotensin II AT(1) blockade normalizes cerebrovascular autoregulation and reduces cerebral ischemia in spontaneously hypertensive rats. Stroke 2000;31(10): 2478–86.

- Reuter U, Chiarugi A, Bolay H, Moskowitz MA. Nuclear factorkappaB as a molecular target for migraine therapy. Ann Neurol 2002;51(4):507–16.
- Lorenzo O, Ruiz-Ortega M, Suzuki Y, Ruperez M, Esteban V, Sugaya T, et al. Angiotensin III activates nuclear transcription factor-kappaB in cultured mesangial cells mainly via AT(2) receptors: studies with AT(1) receptor-knockout mice. J Am Soc Nephrol 2002;13(5):1162–71.
- 103. Pfaffenrath V, Diener HC, Fischer M, Friede M, Henneicke-von Zepelin HH, Investigators. The efficacy and safety of Tanacetum parthenium (feverfew) in migraine prophylaxis—a double-blind, multicentre, randomized placebo-controlled dose-response study. Cephalalgia 2002;22(7):523–32.
- Pittler MH, Ernst E. Feverfew for preventing migraine. Cochrane Database Syst Rev 2004(1):CD002286.
- Diener HC, Pfaffenrath V, Schnitker J, Friede M, Henneicke-von Zepelin HH. Efficacy and safety of 6.25 mg t.i.d. feverfew CO2extract (MIG-99) in migraine prevention—a randomized, doubleblind, multicentre, placebo-controlled study. Cephalalgia 2005;25(11):1031–41.
- Johnson ES, Kadam NP, Hylands DM, Hylands PJ. Efficacy of feverfew as prophylactic treatment of migraine. Br Med J 1985;291(6495):569–73.
- Tassorelli C, Greco R, Morazzoni P, Riva A, Sandrini G, Nappi G. Parthenolide is the component of tanacetum parthenium that inhibits nitroglycerin-induced Fos activation: studies in an animal model of migraine. Cephalalgia 2005;25(8):612–21.
- 108. Materazzi S, Benemei S, Fusi C, Gualdani R, De Siena G, Vastani N, et al. Parthenolide inhibits nociception and neurogenic vasodilatation in the trigeminovascular system by targeting the TRPA1 channel. Pain 2013;154(12):2750–8.
- Benemei S, Fusi C, Trevisan G, Geppetti P. The TRPA1 channel in migraine mechanism and treatment. Br J Pharmacol 2014;171(10):2552-67.
- Jain NK, Kulkarni SK. Antinociceptive and anti-inflammatory effects of Tanacetum parthenium L. extract in mice and rats. J Ethnopharmacol 1999;68(1–3):251–9.
- Groenewegen WA, Heptinstall S. A comparison of the effects of an extract of feverfew and parthenolide, a component of feverfew, on human platelet activity in-vitro. J Pharm Pharmacol 1990;42(8):553

 –7.
- Mittra S, Datta A, Singh SK, Singh A. 5-Hydroxytryptamineinhibiting property of Feverfew: role of parthenolide content. Acta Pharmacol Sin 2000;21(12):1106–14.
- Sommer C. Is serotonin hyperalgesic or analgesic? Curr Pain Headache Rep 2006;10(2):101–6.
- Srikiatkhachorn A, Puangniyom S, Govitrapong P. Plasticity of 5-HT serotonin receptor in patients with analgesic-induced transformed migraine. Headache 1998;38(7):534–9.
- Grossman W, Schmidramsl H. An extract of Petasites hybridus is effective in the prophylaxis of migraine. Alternative medicine review: a journal of clinical therapeutic 2001;6(3):303–10.
- Diener HC, Rahlfs VW, Danesch U. The first placebocontrolled trial of a special butterbur root extract for the prevention of migraine: reanalysis of efficacy criteria. Eur Neurol 2004;51(2):89-97.
- Lipton RB, Gobel H, Einhaupl KM, Wilks K, Mauskop A. Petasites hybridus root (butterbur) is an effective preventive treatment for migraine. Neurology 2004;63(12):2240–4.
- Agosti R, Duke RK, Chrubasik JE, Chrubasik S. Effectiveness of Petasites hybridus preparations in the prophylaxis of migraine: a systematic review. Phytomedicine 2006;13(9–10):743–6.
- 119. Fiebich BL, Grozdeva M, Hess S, Hull M, Danesch U, Bodensieck A, et al. Petasites hybridus extracts in vitro inhibit COX-2 and PGE2 release by direct interaction with the enzyme



- and by preventing p42/44 MAP kinase activation in rat primary microglial cells. Planta Med 2005;71(1):12–9.
- Thomet OA, Wiesmann UN, Schapowal A, Bizer C, Simon HU. Role of petasin in the potential anti-inflammatory activity of a plant extract of petasites hybridus. Biochem Pharmacol 2001;61(8):1041–7.
- Wang GJ, Wu XC, Lin YL, Ren J, Shum AY, Wu YY, et al. Ca2+ channel blocking effect of iso-S-petasin in rat aortic smooth muscle cells. Eur J Pharmacol 2002;445(3):239–45.
- 122. Wu SN, Chen H, Lin YL. The mechanism of inhibitory actions of S-petasin, a sesquiterpene of Petasites formosanus, on L-type calcium current in NG108-15 neuronal cells. Planta Med 2003;69(2): 118–24.
- 123. Wang GJ, Shum AY, Lin YL, Liao JF, Wu XC, Ren J, et al. Calcium channel blockade in vascular smooth muscle cells: major hypotensive mechanism of S-petasin, a hypotensive sesquiterpene from Petasites formosanus. J Pharmacol Exp Ther 2001;297(1): 240–6
- Schoenen J, Jacquy J, Lenaerts M. Effectiveness of high-dose riboflavin in migraine prophylaxis. A randomized controlled trial. Neurology 1998;50(2):466–70.
- Boehnke C, Reuter U, Flach U, Schuh-Hofer S, Einhaupl KM, Arnold G. High-dose riboflavin treatment is efficacious in migraine prophylaxis: an open study in a tertiary care centre. Eur J Neurol 2004;11(7):475–7.
- Nambiar N, Aiyappa C, Srinivasa R. Oral riboflavin versus oral propranolol in migraine prophylaxis: an open label randomized controlled trial.. Neurol Asia 2011(16):223–9.
- 127. Riboflavin. Monograph. Altern Med Rev. 2008;13(4):334-40.
- Meissner K, Fassler M, Rucker G, Kleijnen J, Hrobjartsson A, Schneider A, et al. Differential effectiveness of placebo treatments: a systematic review of migraine prophylaxis. JAMA Intern Med 2013;173(21):1941–51.
- 129. Gasparini CF, Sutherland HG, Griffiths LR. Studies on the pathophysiology and genetic basis of migraine. Curr Genomics 2013;14(5):300–15.
- Goadsby PJ, Holland PR, Martins-Oliveira M, Hoffmann J, Schankin C, Akerman S. Pathophysiology of migraine: a disorder of sensory processing. Physiol Rev 2017;97(2):553–622.
- Welch KM, Ramadan NM. Mitochondria, magnesium and migraine. J Neurol Sci 1995;134(1–2):9–14.
- Lodi R, Iotti S, Cortelli P, Pierangeli G, Cevoli S, Clementi V, et al.
 Deficient energy metabolism is associated with low free

- magnesium in the brains of patients with migraine and cluster headache. Brain Res Bull 2001;54(4):437–41.
- Boska MD, Welch KM, Barker PB, Nelson JA, Schultz L. Contrasts in cortical magnesium, phospholipid and energy metabolism between migraine syndromes. Neurology 2002;58(8): 1227–33
- Mauskop A, Altura BT, Cracco RQ, Altura BM. Deficiency in serum ionized magnesium but not total magnesium in patients with migraines. Possible role of ICa2+/IMg2+ ratio. Headache 1993;33(3):135–8.
- Sarchielli P, Coata G, Firenze C, Morucci P, Abbritti G, Gallai V.
 Serum and salivary magnesium levels in migraine and tensiontype headache. Results in a group of adult patients. Cephalalgia 1992;12(1):21–7.
- Gupta VK. Magnesium therapy for migraine: do we need more trials or more reflection? Headache 2004;44(5):445–6.
- 137. Wang F, Van Den Eeden SK, Ackerson LM, Salk SE, Reince RH, Elin RJ. Oral magnesium oxide prophylaxis of frequent migrainous headache in children: a randomized, double-blind, placebocontrolled trial. Headache 2003;43(6):601–10.
- Koseoglu E, Talaslioglu A, Gonul AS, Kula M. The effects of magnesium prophylaxis in migraine without aura. Magnes Res 2008;21(2):101–8.
- 139. Tarighat Esfanjani A, Mahdavi R, Ebrahimi Mameghani M, Talebi M, Nikniaz Z, Safaiyan A. The effects of magnesium, L-carnitine, and concurrent magnesium-L-carnitine supplementation in migraine prophylaxis. Biol Trace Elem Res 2012;150(1–3):42–8.
- Peikert A, Wilimzig C, Kohne-Volland R. Prophylaxis of migraine with oral magnesium: results from a prospective, multi-center, placebo-controlled and double-blind randomized study. Cephalalgia 1996;16(4):257–63.
- Pfaffenrath V, Wessely P, Meyer C, Isler HR, Evers S, Grotemeyer KH, et al. Magnesium in the prophylaxis of migraine—a doubleblind placebo-controlled study. Cephalalgia 1996;16(6):436–40.
- Silberstein SD, Goldberg J. Menstrually related migraine: breaking the cycle in your clinical practice. J Reprod Med 2007;52(10): 888–95.
- Taylor FR. Nutraceuticals and headache: the biological basis. Headache 2011;51(3):484–501.
- Mauskop A, Altura BM. Role of magnesium in the pathogenesis and treatment of migraines. Clin Neurosci 1998;5(1):24–7.

