

Wild bees along an urban gradient: winners and losers

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Abstract The variability of bee communities along an urban gradient has a potentially strong impact on pollinator-plant interactions. We investigated changes in bee species richness and community composition in the city centre and suburbs of Poznań, Poland. During 2006–2008 we recorded 2,495 individuals from 104 species. The most abundant species was *Andrena haemmorhoa*. Other abundant species were *Bombus pascuorum*, *Andrena vaga*, *Andrena nitida*, *Bombus terrestris*. Several species appeared to be more abundant in the city centre whereas others showed an opposite tendency. We have found that the urbanized landscape can act as a filter for the bee community since some specific ecological traits facilitate colonization of the city centre. Small-bodied species that start their activity later in the season and are not solitary preferred the city centre. However, bee species diversity and richness remained stable across the urban gradient indicating some advantages of being a city dwelling bee. We suggest that a city can be a very important habitat for a diverse bee fauna and that conservation of a wide range of habitats of different urbanization levels seems to be the most suitable strategy for conservation of bee diversity in cities.

Keywords Hymenoptera · Apoidea · Bees · Urbanization · City · Poznań

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Introduction

In recent years, interest in the urban bee fauna has increased (McIntyre and Hostetler 2001; Frankie et al. 2005; McFrederick and Le Buhn 2006; Fetridge et al. 2008; Matteson et al. 2008; Wojcik et al. 2008; Pawelek et al. 2009). Urbanization causes a drastic and persistent change in the landscape and environment which, in turn, transforms interspecific interactions within communities (McKinney 2006). Replacement of natural habitats by human-dominated areas results in habitat loss and fragmentation. The isolation of a habitat patch may also affect the abundance and diversity of insect species. Habitat fragments isolated in human-dominated regions are less likely to be colonized by the native biota (Kozlov 1996; Van Dyck and Matthysen 1999). Fragmentation and destruction of semi-natural habitats and replacement with exotic plants may also be the cause of the changes of insect populations. For example an exotic rich winter foraging resource in urban parks and gardens give a possibility of changes in pollinator phenology (Stelzer et al. 2010). Moreover plant species richness often increases in urban areas compared to more natural areas (Grimm et al. 2008).

Many species will have to survive in human-modified areas if they are to survive at all and anthropogenic habitats offer more opportunities for conservation than was previously thought (Gascon et al. 1999). Moreover, focussing public attention on city-dwelling species provides greater opportunities for researchers to demonstrate the importance of conservation to society. It is clear that the perception of wildlife by society is crucial for effective conservation of biodiversity (e.g. Clucas et al. 2008; Home et al. 2009; Mulder et al. 2009). Therefore, in light of these facts, knowledge on wildlife in urban areas seems to be of high importance for biodiversity conservation in heavily modified areas.

Bees play a very important role in plant pollination, both in the natural environment and in human-dominated habitats. They also act as hosts and prey for other species. Therefore, the response of bee communities to urbanization seems to be crucial for the maintenance of biodiversity within cities. For bees, urbanization means a change in the availability of nesting sites as well as the quality and accessibility of food plants. It can also affect bees through habitat fragmentation (McIntyre and Hostetler 2001; Cane 2005a; Frankie et al. 2005; Ahrné et al. 2009). Urban environments are mosaics of habitats that are regularly disturbed and communities need to be dynamic to adapt to disturbances imposed by human activities (Sattler et al. 2010). However, little is still known about the effects of urbanization on the richness and composition of bee communities (Cane 2005a; Hernandez et al. 2009). Moreover, further research needs to be made on trait variation in relation to urbanization. There may be some species-specific traits which facilitate existence in a urban environments and urbanization may affect different species differently (Bańkowska 1980, 1981; Niemelä et al. 2000; McIntyre et al. 2001). Urbanization tends to alter species composition from that of the surrounding landscape (Grimm et al. 2008; Matteson et al. 2008; Kadlec et al. 2009).

The aims of our study were (1) to identify the bee species present within the city centre of Poznań, Poland; (2) describe the response of different bee species to increasing urbanization and (3) to find species-specific traits which facilitate existence in an urban environment. We put forward a hypothesis stating that bee species diversity decreases as urbanization increases and that city centre bees differ in terms of their ecological traits from suburban-dwelling bees. Moreover, we expect that some of species-specific traits (e.g. foraging or nesting behavior) can be helpful in selecting winners and losers among bee species that inhabit urbanized landscapes.

Materials and methods

Study area

The study was carried out in Poznań (52°25'N, 16°58'E), a city with 560,000 residents, over 3 years (2006–2008). Bees were sampled on eight sites (50 × 50 m) located within the urban matrix representing a wide spectrum of urban environments (Fig. 1). For this purpose we selected large green areas, potentially available for many bee species with the aim of identifying the full set of species present in the city. We were not able to include all the green areas in the city but included the largest ones which are probably the most important for the studied group. The

plots (Fig. 1) were along an urbanization gradient and comprised: Plot 1—The garden behind the church surrounded by concentrated settlement of tenement houses (CS1); Plot 2—The garden in front of The Museum of Utility Art in the Royal Castle surrounded by concentrated settlement of tenement houses (CS2); Plot 3—Green area between concentrated settlement of tenement houses (CS3); Plot 4—River meadow close to concentrated settlements (RV); Plot 5—Botanical Garden (BG); Plot 6—Dendrological Garden (DG); Plot 7—Green area in a sparse housing estate (HE); Plot 8—Home garden located in a district of detached houses (HG).

On the basis of the percentage of built up areas within a 500 m radius from the sites, percentage of tree cover within 500 m radius, percentage of green areas cover (including tree cover) within 500 m radius and distance from the city centre we divided the eight plots into city centre ($n = 4$) and suburb ($n = 4$) classes. As the variables were correlated we used principal component analysis (PCA) to combine the four variables into one component explaining 89% of variance of original variables. Positive values of the component (hereafter referred to as “urbanization”) represent urbanized habitat whereas negative values represent suburban habitats. For some of the analyses all the plots were divided on the basis of the “urbanization” component into urban (Plots no. 1–4 in Table 1) and suburban (5–8) plots.

Bee sampling

Yellow Möericke traps were used to catch the specimens as they had previously been used successfully for the collection of bees in urban fragments (Banaszak 2004). Within each site four yellow pan traps were used. Insects were collected from traps every 7–10 days from April to September for 3 years (2006–2008). The collected specimens were temporarily stored in 70% ethanol until pinned for identification. Direct searching along transects, which covered the most attractive resource patches, was also conducted at each site. On transects, bees were netted by hand twice each month from April to September. Within the plot the surveyor walked at a slow speed among any potentially attractive resource patches and collected bees during an observational period of 30 min. Species that could not be identified in the field were kept for identification later. In total, 187 samples were collected over 3 years (from 14 to 44 samples per plot).

All individual bees were pinned in the laboratory, sorted and identified to species. Ecological traits for each species were compiled from primary literature, catalogs, and reference works (Banaszak 1993; Scheuchl 1995; Schmid-Egger and Scheuchl 1997; Banaszak et al. 2001; Pesenko et al. 2002; Pawlikowski and Celary 2003). For each

Fig. 1 Location of study plots in Poznań (City centre: *CS1*, *CS2*, *CS3* concentrated settlements of tenement houses, *RV* river meadow close to concentrated settlements; Suburbs: *HG* Home garden located in a district of detached houses, *HE* sparse housing estate, *DG* Dendrological Garden, *BG* Botanical Garden)

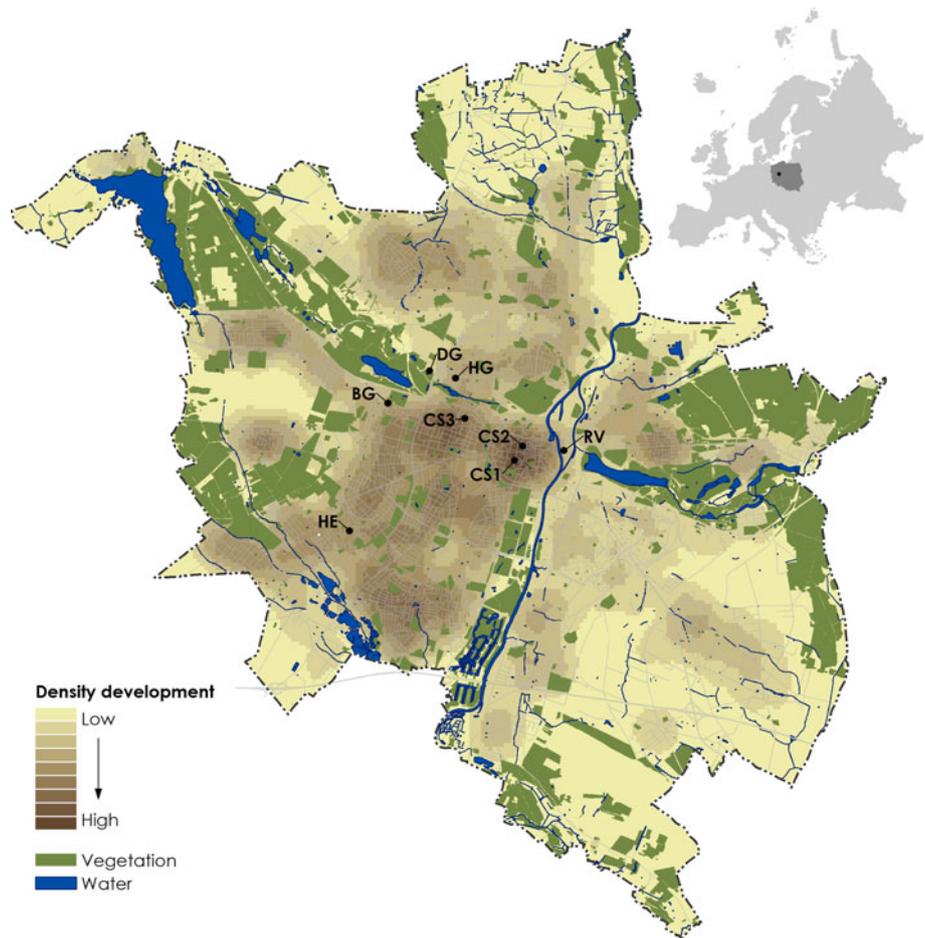


Table 1 Characteristics of the eight plots and values of the component “urbanization” extracted by PCA

No.	Plot	% Built up areas <i>r</i> = 500 m	% trees, <i>r</i> = 500	% green, <i>r</i> = 500	Distance to centre (m)	Component “urbanization”
1	CS1	80	5	10	250	1.190
2	CS2	90	5	10	700	1.217
3	CS3	70	10	20	1,800	0.640
4	RV	50	5	35	1,050	0.487
5	BG	30	25	50	2,730	−0.533
6	DG	40	20	50	4,900	−0.672
7	HE	20	40	65	3,420	−1.206
8	HG	15	30	70	3,480	−1.124

species mean body length, nesting place (cavity or soil), social behavior (solitary, eusocial or parasitic), floral specificity [e.g. oligolectic (single plant species or genus), polylectic (many plant species)], phenology—month of first activity (e.g. April, June, July) was determined.

Statistical analysis

We used rarefaction curves implemented in EstimateS 800 software (Colwell 2005) to analyze species diversity and

species richness of bees (Gotelli and Colwell 2001). We used the MaoTau estimator as a function of the number of individuals included in the analysis (individual-based rarefaction curve) from the two localities (centre vs. suburbs). The estimator generates the expected number of species for a given number of sampled individuals in the two classes (Colwell 2005). We used individual-based rarefaction since it is independent of possible differences in mean number of individuals per sample between the classes (Gotelli and Colwell 2001). Next, we made an

attempt to estimate total species richness of the bee community in the two classes. For this purpose we estimated the number of unseen species (i.e. missed during sampling) with the help of abundance-based coverage estimator (ACE). The ACE approach uses abundances of rare species to estimate number of all species in a given class (Chao et al. 2006).

We used an unconstrained ordination method: detrended correspondence analysis (DCA) implemented in CANOCO 4.5 for gradient length diagnostics, and canonical correspondence analysis (CCA) for comparison of bee communities with ecological traits (Lepš and Šmilauer 2003). For the set of 187 samples we used distance to the city centre and % built-up areas in the vicinity (radius = 200 m) of sites as two explanatory variables in CCA. The day of the season (1 = 1st March) of each sample was used as a third explanatory variable in order to control temporal variability within the community. Moreover, we used year as covariables to exclude possible between-year variations in the community. Species data were log-transformed prior to analysis. A Monte-Carlo test with 5,000 permutations was used to test the significance of the canonical axes. We restricted the number of bee species presented on the CCA biplot to 62 species for which the three explanatory variables (mentioned above) explained at least 1% of their variability in the sites. Finally, general additive models (GAM) were used as a visualization method to show the variability of species richness, abundance and Shannon diversity index among sites along the two first axes of the CCA (Lepš and Šmilauer 2003).

We made an attempt to investigate species-specific traits explaining their preference for or avoidance of urbanization. For this purpose we divided all the bees into two categories: centre-dwelling (more abundant in the centre compared to the suburbs) and suburb-dwelling (reverse pattern). Available ecological traits for all the species ($n = 104$) were used to explain species-specific preferences or avoidance of urbanization with the help of a generalized linear model (GLM) with binomial error distribution and logit link function (Quinn and Keough 2002) implemented in SPSS 16.0. As explanatory variables we used mean body length (covariate), social characteristic (factor) and month of first activity during the year (covariate). Akaike information criterion (AIC) was used to select the most parsimonious model from the set of competing models (Johnson and Omland 2004).

Finally, we compared the bee fauna recorded in Poznań with the non-urban bee community reported from Wielkopolski National Park, located in the vicinity of Poznań. For this purpose we used detrended correspondence analysis (DCA) in CANOCO (Lepš and Šmilauer 2003).

Results

Species composition

We collected a total of 2,495 individuals from 104 species (Banaszak-Cibicka 2009); the Chao1 species richness estimator, corrected for unseen species in the samples, suggested 124 species. All of the collected species were native species. The most abundant species (accounting for 11% of all individuals collected) was *Andrena haemorrhoa* (Fabricius, 1781). Other abundant species were *Bombus pascuorum* (Scopoli, 1763), *Andrena vaga* (Panzer, 1799), *Andrena nitida* (Müller, 1776), *Apis mellifera* (Linnaeus, 1758), *Bombus terrestris* (Linnaeus, 1758); these five species accounted for 29% of individuals collected. These species are eusocial bees (*Bombus pascuorum*, *Apis mellifera*, *Bombus terrestris*) or early spring solitary bees (*Andrena haemorrhoa*, *Andrena vaga*, *Andrena nitida*). Twenty-two species were represented by only a single individual. The most represented families were Andrenidae (26% of species, 38% of individuals), Halictidae (22% of species, 18% of individuals) and Apidae (12% of species, 28% of individuals).

Most bees were soil nesting (57% of species, 62% of individuals), polylectic (74% of species, 88% of individuals) and solitary (65% of species, 56% of individuals). Parasitic species made up a small proportion of the urban bee community (12% of species, but only 0.9% of individuals). Oligolectic species, which collect pollen from within a single family or genus of plants, were also rare (15% of species, 10% of individuals). The ecological traits of the recorded species are given in the Table 4 (Appendix).

Diversity and species richness

The expected cumulative number of bee species as a function of the number of sampled individuals was similar between city centre and suburbs (Fig. 2). For 600 sampled individuals the expected cumulative number of species was ca 66 for both urban classes. Estimated species richness, corrected for species unseen in the samples, was also similar between the two urban classes (Fig. 2).

Winners and losers on the urban gradient

Bee species showed very different responses to urbanization (Fig. 3). Some were recorded predominantly in the city centre [e.g. *Hylaeus hyalinatus* (Smith, 1842); *Hylaeus angustatus* (Schenck, 1859); *Evylaeus nitidulus* (Fabricius, 1804); *Megachile pilidens* (Alfken, 1923); *Megachile rotundata* (Fabricius, 1784)] while others showed the opposite pattern [*Evylaeus lucidulus* (Schenck, 1861);

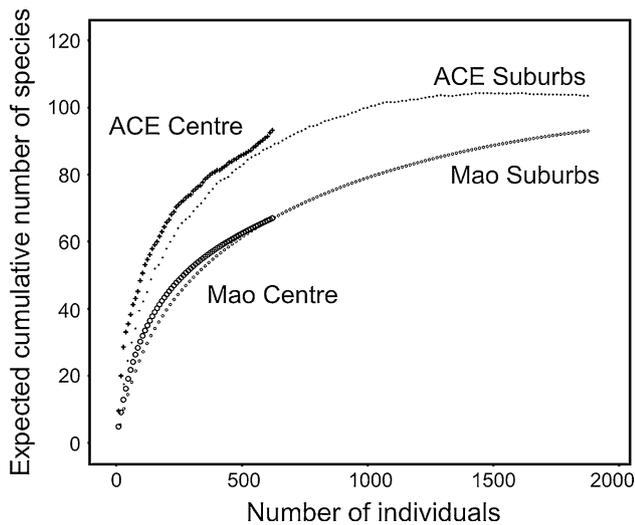


Fig. 2 Expected cumulative number of bee species (expressed with Mao Tau estimator) as a function of number of sampled individuals in the centre and suburb zones of Poznań, Central Poland. The abundance-based coverage estimator (ACE), corrected for species unseen in the samples is given for each habitat

Evyllaes calceatus (Scopoli, 1763); *Panurgus calcaratus* (Scopoli, 1763)]. The distribution and abundance of some species seemed to be unaffected by the urbanization gradient [e.g. *Seladonia subaurata* (Rossi, 1792)]. The distribution of species included in the analysis along the two canonical axes was not random (Monte Carlo test of significance of all canonical axes, trace = 1.025, F ratio = 2.80, $P = 0.0002$, 5,000 permutations). The abundance of bees at sites decreased towards the city centre and a similar tendency, although less distinct, was recorded for the number of species per site (Fig. 3). However, species diversity found in particular samples, expressed by the Shannon index, did not show clear variability along the urbanization gradient (Fig. 3).

Ecological traits of winners and losers on the urban gradient

Species-specific ecological characteristics were usually associated. Social behavior was associated with nesting place (Chi-square test, $\chi^2 = 126.6$; $df = 6$; $P < 0.0001$) and with floral specificity ($\chi^2 = 110.9$; $df = 4$; $P < 0.0001$). Therefore, only three of them (mean body length, social behavior and month of first activity) were used in modeling. Ecological traits included in the modelling significantly explained species-specific response to urbanization. The full model, containing social pattern, body length and time of first activity was the most parsimonious among the set of seven competing models (Table 2). The model was highly statistically significant.

The probability that a given bee species was more abundant in the city centre compared to the suburbs depended positively on its phenology: species that start their activity late (e.g. June–July) were significantly more associated with the city centre than species that start activity earlier (Table 3; Fig. 4). Solitary-living bees were significantly associated with the suburbs compared to species belonging to the two remaining social categories (eusocial bees and parasites). Mean body length, despite being non-significant (Table 3) was included in the most parsimonious model and, according to AIC, contributed to explaining species-specific responses to urbanization in 104 bee species.

Urban versus non-urban bee fauna

Bee communities recorded in the botanical and dendrological gardens in Poznań city were the most similar to the community reported from Wielkopolski National Park (WNP, Cierznik 2003) whereas communities from the plots CS1 and CS2 were the less similar from that reported in the natural habitat. In general, distance along the first axis of DCA between the bee fauna from the Centre and WNP was smaller than the distance between the bee fauna from Suburbs and WNP (Fig. 5).

Discussion

General description of species composition in relation to previous research

The 104 bee species observed in Poznań during the study comprised 40% of the 259 species recorded from the region (the Wielkopolska-Kujavian Lowland, Banaszak 1982). The total number of bee species recorded to date in Poznań (Banaszak 1973, 1976, 1982; Wójtowski and Szymaś 1973a, b; Wójtowski and Feliszek 1977) is 184, i.e. 71% of the species recorded from the region. These data indicate that a city can be an important habitat for a diverse bee fauna.

The dominant species, *Andrena haemorrhoa*, *Andrena vaga*, *Andrena nitida*, *Andrena fulva*, *Andrena ventralis*, *Osmia rufa* and *Anthophora plumipes*, are early spring species which appear at the end of April and in May. The dominants from the *Evyllaes* and *Bombus* genus are eusocial species and *Apis mellifera*, accounted for 5.4% of the collected individuals.

We also found some southern species which are rare in Poland: *Megachile pilidens*, *Andrena florea*, *Andrena mitis* and *Osmia mustelina*. These have not been recorded from the Wielkopolski National Park (situated 15 km south of Poznań) or from the Wielkopolska-Kujavian Lowland

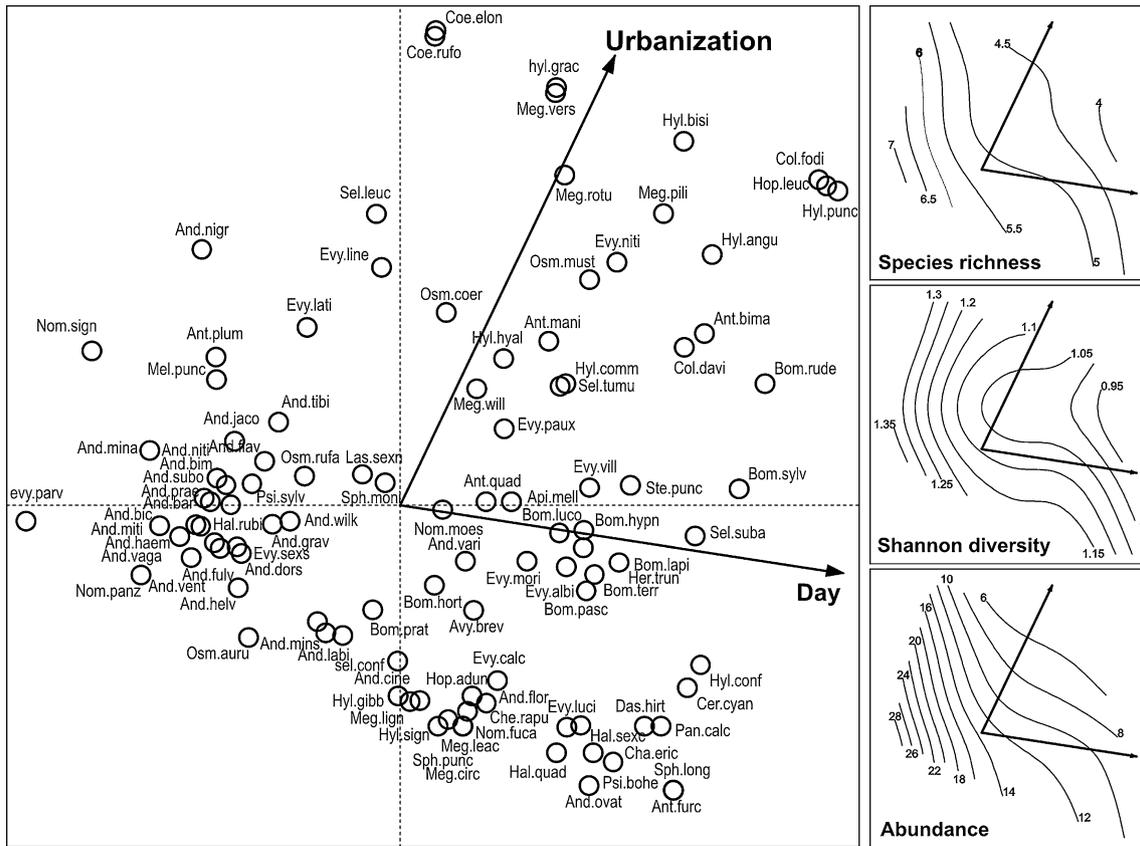


Fig. 3 Canonical correspondence analysis (CCA) biplot showing distribution of 62 bee species in relation to the urbanization gradient (expressed with component extracted from PCA—see “Methods” for details) and time used as explanatory variables along the two first canonical axis (large subplot). Abbreviations of species names use the

first three letters of genus and the first four letters of species scientific names. Variability of the three diversity indices characterizing the bee community (species richness, abundance and Shannon diversity index) in relation to the habitat characteristics is shown with isolines fitted with a General Additive Model (smaller subplots)

Table 2 Akaike information criterion (AIC) scores of the eight competing generalized linear models with binomial error distribution and logit link function explaining preference or avoidance of 104 bee species to urbanization

No.	Model	<i>k</i>	AIC	ΔAIC	χ^2	<i>P</i>
1	Soc. + length + phenol.	3	99.064	0	25.65	<0.0001
2	Soc. + phenol.	2	99.366	0.302	23.35	<0.0001
3	Phenol.	1	102.057	2.993	16.66	<0.0001
4	Soc.	1	119.630	20.566	1.09	0.581
5	Length	1	114.311	15.247	4.41	0.036
6	Soc. + length	2	117.126	18.062	5.59	0.133
7	Length + phenol.	2	101.503	2.439	19.21	<0.0001
8	Intercept only	0	116.503	17.439		

For each model number of predictors (*k*), value of Akaike Information Criterion (AIC), value of AIC in relation to best model (ΔAIC), test statistic (χ^2), and its significance (*P*) are given. *Soc.* social behavior, *length* mean body length, *phenol.* phenology, month of first activity

except for *Megachile pilidens* which was found on xeric grasslands in the valley of the River Vistula. It is possible that urban areas which are warmer than surrounding countryside may provide better living conditions for these bees.

Ecological characteristics of species found in Poznań

The majority of bee individuals (62%) collected in Poznań nest in the soil and the diversity and abundance of cavity-nesting species was relatively low (9% of individuals). In

Table 3 Parameter estimates of a generalized linear model with binomial error distribution and logit link function, assessed as most parsimonious according to AIC (see Table 2 for details) explaining preference or avoidance by 104 bee species of urbanization

Effect	<i>B</i>	Wald	<i>P</i>	OR	95% CI for OR	
Intercept	-6.851	10.538	0.001	0.001	0.00002	0.066
Social behavior: parasites	-0.191	0.039	0.844	0.826	0.122	5.581
Social behavior: eusocial	1.762	5.568	0.018	5.822	1.348	25.146
Social behavior: solitary	0			1		
Mean body length	-0.124	2.197	0.138	0.883	0.749	1.041
Month of first activity	1.298	14.205	0.000	3.661	1.864	7.189

The table presents *B* coefficient, Wald statistic and its significance, odds ratio (OR) and its 95% confidence intervals. The reference category for the parameter estimates is 0 (i.e. avoidance of city centre)

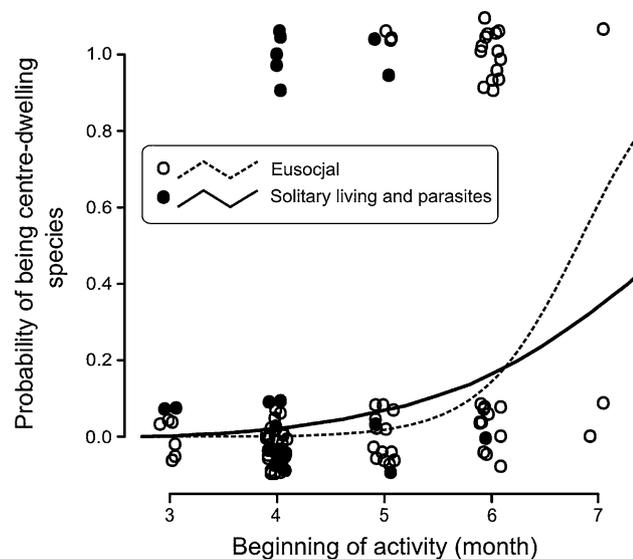


Fig. 4 The effect of species-specific social characteristics and month of first activity on the preferences of species to the city centre as assessed by logistic regression. Jittering was added to aid visualization. Note that fits are not adjusted for the body length effect that was included in the most parsimonious model on the basis of AIC

contrast, soil-nesting species and individuals in New York city gardens were relatively scarce (Matteson et al. 2008). A higher abundance of cavity-nesting bee species in urban areas has been shown in other papers (Cane 2005a; Zanette et al. 2005; Cane et al. 2006; Hinners 2008; Matteson et al. 2008). In our study, bee species from the family Andrenidae were the most abundant. Species of this genera are

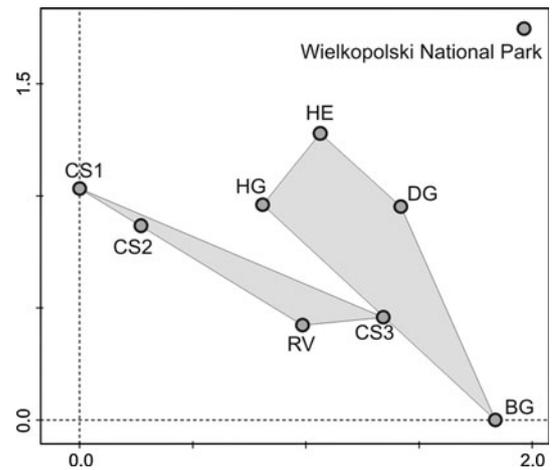


Fig. 5 Position of eight samples from Poznań city and sample from Wielkopolski National Park along the first two axes of detrended correspondence analysis (DCA)

largely ground-nesters and early spring fliers. In contrast to our study in many works that reported diversity of bee families, species from Andrenidae were rare in urban habitat patches (Owen 1991; Antonini and Martins 2003; Matteson et al. 2008; Wojcik et al. 2008). Scarcity of Andrenidae likely reflects a lack of appropriate flowering plants, especially trees, in early spring (Matteson et al. 2008). We suppose that the lack (or low number) of available food resources is the most important driver of low number of Andrenidae recorded in New York by Matteson et al. (2008). Our sites included the Botanical and Dendrological gardens where there were appropriate floral resources for these bees. However, the abundance of bee species from the family Andrenidae decreased with increasing urbanization (decreased percentage of tree cover in city centre). The relatively high abundance of Andrenidae also caused the proportion of soil-nesting species to be high in Poznań, as compared to other cities.

Cleptoparasites made up 12.5% of the collected species but only 0.9% of individuals. In residential urban areas in Berkeley, California, Frankie et al. (2005) found only 7.9% of species to be cleptoparasites and in New York City urban gardens, parasites made up 5.6% of collected species (Matteson et al. 2008). Cane (2005a, b) noted that cleptoparasitic species are largely absent in urban bee faunas and suggested that such species may require large host populations to persist. Despite the abundance of *Andrena* hosts collected in Poznań there were few cleptoparasites of the genus *Nomada* (four species, seven individuals).

Species traits promoting colonization of urbanized habitat

In agreement with our hypothesis, species found in the city centre and suburbs differed to some extent in their

ecological traits. The probability that a given bee species was more abundant in the city centre compared to the suburbs depended positively on its activity pattern during the year: species that start their activity late (e.g. June–July) more often preferred the city centre than species that start activity earlier. Low numbers of solitary-living early spring bees in the city centre are likely due to a lack of appropriate floral resources during the flight times of these bees. These species prefer trees as their main floral resource and in Poznań, trees are more abundant in parks and gardens and bigger housing estates outside the city centre.

Solitary-living bees preferred the city centre less than to social species. Previous authors have shown that social species are more abundant in urban areas (e.g. Banaszak et al. 1978), probably because sociality permits behavioural and ecological flexibility. Additionally, social insects are considered to be species rich and ecologically dominant (Chapman and Bourke 2001).

Mean body length, while non-significant (Table 2) was included in the most parsimonious model and, according to AIC, contributed to explaining species specific responses to urbanization in 104 bee species. Small species were more abundant in the city centre. This may be related to habitat and food resource availability along the urbanization gradient. In general, green areas are smaller in Poznań city centre compared to the suburbs which may promote the smaller bees in such habitats. However, the importance of this trait seems to be rather weakly supported by the data and needs to be verified in future.

The floral diversity of urban habitats, such as small parks and gardens is often dominated by exotic and horticultural varieties (Thompson et al. 2003; Frankie et al. 2005; Gaston et al. 2005). Therefore, there is an expectation that generalist (polylectic) bee species may increasingly dominate urban locations. Generalist bee species with broad tolerances are favoured in urban areas (Cane 2005b; Eremeeva and Sushchev 2005) and many urban bee surveys indicate that floral specialists are scarce in urban habitats (Cane 2005a; Frankie et al. 2005; Cane et al. 2006; McFrederick and Le Buhn 2006). The most abundant oligolectic species was *Andrena vaga* which collects pollen from willow (*Salix* spp.) that grow in parks and gardens. However, the association between floral specificity and response to urbanization needs further study.

Advantages and disadvantages of being a city dwelling bee

The abundance of bees appeared to decrease towards the city centre. However, species diversity did not show a clear trend along the urban gradient. Therefore, we did not confirm the hypothesis of bee species diversity decreasing

as urbanization increases. It has been shown that the increase of buildings associated with urbanization and the loss of vegetation cover have a negative effect on the abundance of bees (Zanette et al. 2005). On the other hand, urban areas also include flower rich green areas that can provide forage for a high diversity of wild bees (Frankie et al. 2005; Gaston et al. 2005; Loram et al. 2007). Cities may also contain materials and sites for nesting, such as bare soil, dead stems and cavities in man-made structures (McFrederick and Le Buhn 2006; Ahrné et al. 2009). Furthermore, urban areas have extremely high spatial habitat heterogeneity produced by many different land uses and plant cultivation at small spatial scales (Thompson et al. 2003) especially compared to the simplified and homogenous structure of the modern agriculture landscape. The mosaic of land use patterns may increase species diversity by increasing habitat diversity (Zerbe et al. 2003). This spatial heterogeneity can support even greater species richness than surrounding natural areas (Cierzniaik 2003).

Most studies have compared urban, suburban or remnant habitat patches to natural areas and most of them documented lower species richness and abundance in urban areas than natural ones (McIntyre and Hostetler 2001; Fetridge et al. 2008; Matteson et al. 2008). However, in some groups of bees, such as bumble bees, species richness increased in urban areas relative to the surrounding natural habitat (Goulson et al. 2002; McFrederick and Le Buhn 2006; Winfree et al. 2007; Matteson and Langellotto 2009). There are few studies that have surveyed different site types within a city matrix along an urbanization gradient. Ahrné (2008) reported lower species richness of bumble bees and cavity nesting bees in areas containing a higher percentage of impervious surfaces. Her study concluded that urbanization is negatively correlated with bee species richness. In contrast, a study in Boulder County, Colorado showed that neither the number of species nor their abundance differed significantly among different levels of urbanization (Kearns and Oliveras 2009). The specific impact of urbanization on species richness will vary, depending on many geographical, historical and economic factors that are unique to each city (McKinney 2008).

Management implication and conclusions

Green areas within the city have become increasingly important places for conserving biodiversity. They can also be designed with the goal of attracting specific pollinators, such as bees (Owen 1991; Tommasi et al. 2004; McFrederick and Le Buhn 2006; Tait 2006; Fetridge et al. 2008; Matteson et al. 2008; Wojcik et al. 2008; Kearns and Oliveras 2009; Pawelek et al. 2009; Matteson and Langellotto 2010), and thus can contribute to conservation of local fauna, especially when utilizing native plant species

(McIntyre and Hostetler 2001; McKinney 2002; Cane et al. 2006). We recommend that the results of our study should be taken into account in landscape planning within urban areas.

Our results also show that, in contrast to some expectations and published literature, the diversity of the bee community can remain relatively stable across an urban gradient. This stability leads to a suggestion that the distinct man-made landscape transformation in the city centre of Poznań also has important advantages for the studied insects and gives them the opportunity to increase in number and ensure existence of a species-rich community. However, we have found that the urbanized landscape can act as a filter for the bee community since some specific ecological traits facilitate colonization of the city centre. As a consequence, city zones across the urbanization gradient differed in term of species composition. Therefore, efforts targeted at bee conservation (e.g. distribution of protected areas) should not be concentrated in one city

zone; instead protection across a wide range of habitats of different levels of man-made transformation seems to be the most appropriate strategy for bee diversity conservation in cities.

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Appendix

See Table 4.

Table 4 Bee species occurring in Poznań—2006–2008 (from Banaszak-Cibicka 2009)

Species	Abundance	% of total	Sociality	Pollen specificity	Nest substrate
<i>Colletes daviesanus</i> (Smith, 1846)	19	0.7	S	O	S
<i>Colletes fodiens</i> (Fourcroy, 1785)	10	0.4	S	O	S
<i>Hylaeus angustatus</i> (Schenck, 1859)	15	0.6	S	P	C
<i>Hylaeus bisinuatus</i> (Förster, 1871)	5	0.2	S	P	C
<i>Hylaeus communis</i> (Nylander, 1852)	18	0.7	S	P	C
<i>Hylaeus confusus</i> (Nylander, 1852)	1	0.1	S	P	C
<i>Hylaeus gibbus</i> (Saunders, 1850)	4	0.1	S	P	C
<i>Hylaeus gracilicornis</i> (Morawitz, 1867)	1	0.1	S	P	C
<i>Hylaeus hyalinatus</i> (Smith, 1842)	59	2.3	S	P	C
<i>Hylaeus punctatus</i> (Brulle, 1832)	1	0.1	S	P	C
<i>Hylaeus signatus</i> (Panzer, 1798)	2	0.1	S	P	C
<i>Andrena barbilabris</i> (Kirby 1802)	6	0.2	S	P	S
<i>Andrena bicolor</i> (Fabricius, 1775)	11	0.4	S	P	S
<i>Andrena bimaculata</i> (Kirby 1802)	4	0.1	S	P	S
<i>Andrena cineraria</i> (Linnaeus, 1758)	1	0.1	S	P	S
<i>Andrena dorsata</i> (Kirby 1802)	18	0.7	S	P	S
<i>Andrena flavipes</i> (Panzer, 1799)	49	1.9	S	P	S
<i>Andrena florea</i> (Fabricius, 1793)	2	0.1	S	O	S
<i>Andrena fulva</i> (Müller, 1766)	73	2.9	S	P	S
<i>Andrena gravida</i> (Imhoff, 1899)	14	0.5	S	P	S
<i>Andrena haemorrhoea</i> (Fabricius, 1781)	277	11.1	S	P	S
<i>Andrena helvola</i> (Linnaeus, 1758)	30	1.2	S	P	S
<i>Andrena jacobii</i> (Perkins, 1921)	4	0.1	S	P	S
<i>Andrena labiata</i> (Fabricius, 1781)	4	0.1	S	P	S
<i>Andrena minutula</i> (Kirby, 1802)	4	0.1	S	P	S
<i>Andrena minutuloides</i> (Perkins, 1914)	12	0.4	S	P	S
<i>Andrena mitis</i> (Schmiedeknecht, 1838)	1	0.1	S	P	S
<i>Andrena nigroaenea</i> (Kirby, 1802)	5	0.2	S	P	S

Table 4 continued

Species	Abundance	% of total	Sociality	Pollen specificity	Nest substrate
<i>Andrena nitida</i> (Müller, 1776)	137	5.5	S	P	S
<i>Andrena ovatula</i> (Kirby, 1802)	1	0.1	S	O	S
<i>Andrena praecox</i> (Scopoli, 1763)	11	0.4	S	P	S
<i>Andrena subopaca</i> (Nylander, 1848)	15	0.6	S	P	S
<i>Andrena tibialis</i> (Kirby, 1802)	10	0.4	S	P	S
<i>Andrena vaga</i> (Panzer, 1799)	164	6.5	S	O	S
<i>Andrena varians</i> (Rossi, 1792)	1	0.1	S	P	S
<i>Andrena ventralis</i> (Imhoff, 1832)	87	3.4	S	P	S
<i>Andrena wilkella</i> (Kirby, 1802)	2	0.1	S	O	S
<i>Panurgus calcaratus</i> (Scopoli, 1763)	7	0.2	S	O	S
<i>Halictus quadricinctus</i> (Fabricius, 1776)	2	0.1	S	P	S
<i>Halictus rubicundus</i> (Christ, 1791)	5	0.2	E	P	S
<i>Halictus sexcinctus</i> (Fabricius, 1775)	9	0.3	S	P	S
<i>Seladonia confusa</i> (Smith, 1853)	2	0.1	E	P	S
<i>Seladonia leucahenea</i> (Ebmer, 1972)	1	0.1	E	P	S
<i>Seladonia subaurata</i> (Rossi, 1792)	11	0.4	E	P	S
<i>Seladonia tumulorum</i> (Linnaeus, 1758)	37	1.4	E	P	S
<i>Lasioglossum sexnotatum</i> (Kirby, 1802)	22	0.8	S	P	S
<i>Evylaeus albipes</i> (Fabricius, 1781)	27	1.0	E	P	S
<i>Evylaeus brevicornis</i> (Schenck, 1863)	9	0.3	E	P	S
<i>Evylaeus calceatus</i> (Scopoli, 1763)	62	2.4	E	P	S
<i>Evylaeus laticeps</i> (Schenck, 1868)	66	2.6	E	P	S
<i>Evylaeus linearis</i> (Schenck, 1869)	20	0.8	E	P	S
<i>Evylaeus lucidulus</i> (Schenck, 1861)	3	0.1	S	P	S
<i>Evylaeus morio</i> (Fabricius, 1793)	38	1.5	E	P	S
<i>Evylaeus nitidulus</i> (Fabricius, 1804)	51	2.0	E	P	S
<i>Evylaeus parvulus</i> (Schenck, 1853)	1	0.1	S	P	S
<i>Evylaeus pauxillus</i> (Schenck, 1853)	63	2.5	E	P	S
<i>Evylaeus sexstrigatus</i> (Schenck, 1868)	3	0.1	S	P	S
<i>Evylaeus villosulus</i> (Kirby, 1802)	7	0.2	S	P	S
<i>Sphecodes longulus</i> (Hagens, 1882)	1	0.1	CP	[P]	[S]
<i>Sphecodes monilicornis</i> (Kirby, 1802)	2	0.1	CP	[P]	[S]
<i>Sphecodes puncticeps</i> (Thomson, 1870)	2	0.1	CP	[P]	[S]
<i>Dasygaster hirtipes</i> (Harris, 1780)	37	1.4	S	O	S
<i>Anthidium manicatum</i> (Fabricius, 1775)	11	0.4	S	P	–
<i>Stelis punctulatisima</i> (Kirby, 1802)	3	0.1	CP	[P]	[C]
<i>Heriades truncorum</i> (Linnaeus, 1758)	9	0.3	S	O	C
<i>Chelostoma rapunculi</i> (Lepeletier, 1841)	2	0.1	S	O	C
<i>Hoplitis adunca</i> (Panzer, 1798)	1	0.1	S	O	C
<i>Hoplitis leucomelena</i> (Kirby, 1802)	1	0.1	S	P	C
<i>Osmia aurulenta</i> (Panzer, 1799)	4	0.1	S	P	C
<i>Osmia coerulescens</i> (Linnaeus, 1758)	8	0.3	S	P	C
<i>Osmia mustelina</i> (Gerstaecker, 1869)	1	0.1	S	O	C
<i>Osmia rufa</i> (Linnaeus, 1758)	72	2.8	S	P	C
<i>Chalicodoma ericetorum</i> (Lepeletier, 1841)	1	0.1	S	O	C
<i>Megachile circumcincta</i> (Kirby, 1802)	9	0.3	S	O	S
<i>Megachile leachella</i> (Curtis, 1828)	1	0.1	S	P	–
<i>Megachile ligniseca</i> (Kirby, 1802)	5	0.2	S	P	S

Table 4 continued

Species	Abundance	% of total	Sociality	Pollen specificity	Nest substrate
<i>Megachile pilidens</i> (Alfken, 1923)	5	0.2	S	P	S
<i>Megachile rotundata</i> (Fabricius, 1784)	5	0.2	S	P	C
<i>Megachile versicolor</i> (Smith, 1844)	1	0.1	S	P	C
<i>Megachile willughbiella</i> (Kirby, 1802)	15	0.6	S	P	C
<i>Coelioxys elongata</i> (Lepeletier, 1841)	1	0.1	CP	[P]	S
<i>Coelioxys rufocaudata</i> (Smith, 1854)	1	0.1	CP	[P]	S
<i>Anthophora bimaculata</i> (Panzer, 1798)	1	0.1	S	P	S
<i>Anthophora furcata</i> (Panzer, 1798)	1	0.1	S	O	S
<i>Anthophora plumipes</i> (Pallas, 1772)	64	2.5	S	P	S
<i>Anthophora quadrimaculata</i> (Panzer, 1806)	8	0.3	S	P	S
<i>Melecta punctata</i> (Fabricius, 1775)	2	0.1	CP	[P]	[S]
<i>Nomada fucata</i> (Panzer, 1798)	2	0.1	CP	[P]	[S]
<i>Nomada moeschleri</i> (Alfken, 1913)	1	0.1	CP	[P]	–
<i>Nomada panzeri</i> (Lepeletier, 1841)	1	0.1	CP	[P]	[S]
<i>Nomada signata</i> (Jurine, 1807)	3	0.1	CP	[P]	[S]
<i>Ceratina cyanea</i> (Kirby, 1802)	4	0.1	S	P	C
<i>Bombus bohemicus</i> (Seidl, 1838)	2	0.1	CP	[P]	[H]
<i>Bombus hortorum</i> (Linnaeus, 1761)	21	0.8	E	P	H
<i>Bombus hypnorum</i> (Linnaeus, 1758)	59	2.3	E	P	H
<i>Bombus lapidarius</i> (Linnaeus, 1758)	73	2.9	E	P	H
<i>Bombus lucorum</i> (Linnaeus, 1761)	60	2.4	E	P	H
<i>Bombus pascuorum</i> (Scopoli, 1763)	165	6.6	E	P	H
<i>Bombus pratorum</i> (Linnaeus, 1761)	19	0.7	E	P	H
<i>Bombus ruderarius</i> (Muller, 1776)	10	0.4	E	P	H
<i>Bombus sylvarum</i> (Linnaeus, 1761)	9	0.3	E	P	H
<i>Bombus sylvestris</i> (Lepeletier, 1832)	2	0.1	CP	[P]	[H]
<i>Bombus terrestris</i> (Linnaeus, 1758)	130	5.2	E	P	H
<i>Apis mellifera</i> (Linnaeus, 1758)	136	5.4	E	P	H

Abundance total number of specimens collected across all sites and all years. *% of total* percentage of total collection ($n = 2,495$) identified as this species. *Sociality* classification as either E—eusocial, CP—cleptoparasitic, S—solitary. *Pollen specificity* classification as either polylectic (collecting pollen from multiple plant families) or oligolectic (collecting pollen from a single plant family or genus); [P] indicates parasitic species that do not collect pollen. *Nest substrate* classification of the nest substrate of each nonparasitic species as C—cavity, H—hive, S—soil; letters in brackets ([]) indicate the nest substrate of the host of a parasitic species; dashes (–) indicate attribute is unknown. The list of species was arranged in systematic order

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