Assessment of Microzooplankton Grazing on *Heterosigma akashiwo* Using a Species- Specific Approach Combining Quantitative Real-Time PCR (QPCR) and Dilution Methods

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In the original version of this article, Table 2 contains incorrect probe sequence for Hs and DNA target for pGEM plasmid. The correct version of Table 2 is provided here.

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Table 2 List of primer and probe sequences used for QPCR analysis of the raphidophytes *Chattonella subsalsa*, *C. cf. verruculosa*, *Fibrocapsa japonica*, and *Heterosigma akashiwo*, and for amplification of the pGEM plasmid

DNA Target	Primer/Probe	Sequence (5'-3')	References
<i>Chattonella subsalsa</i> 18S rDNA	Cs 1350F*	CTAAATAGTGTGGGTAATGCTTAC	Coyne <i>et al.</i> 2005
	Cs 1705R*	GGCAAGTCACAATAAAGTTCCAA	
	Raph Probe	CAACGAGTACTTTCCTTGGCCGGA	
<i>Chattonella cf. verruculosa</i> 18S rDNA	Cv 1561F	ATGCATACAGCGAGTCTAGA	Handy <i>et al.</i> 2006
	Cv 1780R	TCACTCCGAAAAGTGTC AAC	
	Cv Probe	CAAGAGTACCCAGGCCTCTCGACC	
<i>Fibrocapsa japonica</i> 18S rDNA	Fc 1350F	TGCTTTAGTCATTGTGTGCAG	This study
	Fc 1705R	ACCACAACTAATGAGGAGGC	
	Fc Probe	CCCAGGCCTACCGCCAAGGTTGTA	
<i>Heterosigma akashiwo</i> 18S rDNA	Hs 1350F	CTAAATAGTGTCGGTAATGCTTCT	Coyne <i>et al.</i> 2005
	Hs 1705R	GGCAAGTCACAATAAAGTTCCAT	
	Hs Probe	CAACGAGTAACGACCTTTGCCGGA	
pGEM plasmid DNA	M13F	CCCAGTCACGACGTTGTAAAACG	Coyne <i>et al.</i> 2005
	pGEM R	TGTGTGGAATTGTGAGCGGA	
	pGEM Probe	CACTATAGAATACTCAAGCTTGCATGCCTGCA	

*F: Forward primer, R: reverse primer